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1 **Multi-modal SARS-CoV-2 proteins interactomics unveil spatial patterns of**
2 **invasion featuring 151 contacts with human proteins essential for infection**

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30 **Summary**

31 From a comprehensive proximity-dependent biotinylation (BioID)-based proximal SARS-CoV-2 interactome, we
32 generated a 3D map positioning both SARS-CoV-2 and host proteins within cell volumes. Then, a subset of the
33 host proteome proximal to the viral polypeptides was probed for direct contacts by a semi-quantitative split-
34 nanoluciferase assay. We thus provide an intensity map of 414 direct virus-host protein contacts, of which 151
35 involved host proteins essential for SARS-CoV-2 infection. Among the 70 virus-host complexes modeled by
36 AlphaFold2, 10 scored positive, providing as many candidate interfaces relevant for structure-driven screening
37 of PPIs-disrupting compounds. Based on our data, we selected and showed that USP13 inhibition potentially
38 inhibits SARS-CoV-2 replication. This innovative and multi-pronged pipeline provides a resource of
39 unprecedented depth. Its capacity in identifying virus-host PPIs highly relevant for the screening of antiviral
40 compounds and its versatility for any emerging human pathogens could make this pipeline a prime tool for viral
41 preparedness.

42 **Keywords:** Protein-Protein Interactions, SARS-CoV-2, Systems Biology, BioID, split-nanoluciferase, Alphafold2,
43 3D interactive topology.

45 **Introduction**

46 The COVID-19 pandemic revealed the need to improve our reactivity in developing effective countermeasures
47 when facing emerging pathogens. Controlling the spread of a pathogen in an immunologically naive population
48 requires prevention, containment, as well as the timely implementation of therapeutics to limit transmission and
49 severe health effects. In addition, after over three years, the COVID-19 pandemic persists as a serious public
50 health concern, especially in countries with low vaccination rates. Considering the series of COVID-19 waves
51 due to the emergence of SARS-CoV-2 variants, the control of COVID-19 disease remains challenging. Further
52 characterization of SARS-CoV-2 pathogenesis at a molecular level is still needed to combat viral mechanisms
53 through antiviral therapeutics. Identifying host proteins to target is necessary for long-term COVID-19
54 management due to their lower mutation rate compared to the viral ones. Therefore, determining virus-host
55 protein interfaces and developing targeted therapies to disrupt their contact appears promising for host-targeted
56 therapeutics.

57 Here, we describe an interactomics pipeline that we applied to SARS-CoV-2 and that: (i) can be deployed as
58 soon as the genome of any pathogen is sequenced; (ii) identifies host-pathogen promiscuity within cellular

59 territories; (iii) identifies direct pathogen/host contacts possibly essential for infection; and (iv), provide protein-
60 protein interaction (PPI) interfaces of strong therapeutic interest.

61 SARS-CoV-2 is a positive-sense RNA virus replicating in the cytoplasm and strongly depending on intracellular
62 membranous organelles for its productive life cycle. Nine of the twenty-nine viral proteins are localized in
63 intracellular organelles and exhibit transmembrane domains^{1,2}. Thus, membranes- and organelles-adapted
64 biochemical protocols are best suited to study SARS-CoV-2 proteins interactions within the host cell.

65 Proteomic studies relying on classical affinity purification methods followed by mass spectrometry (AP-MS)-
66 based identification of protein complexes have been performed to define the interplay of SARS-CoV-2 proteins
67 with host cell components³⁻⁶. One major limitation of AP-MS is the requirement of maintaining protein-protein
68 interactions (PPIs) throughout the purification process, which necessitates the use of a gentle lysis buffer to
69 solubilize physically associated proteins without disrupting PPIs, hence precluding the detection of PPIs within
70 poorly soluble compartments. The proximity-dependent biotinylation technique (BioID)⁷ allows the identification
71 of proximal PPIs of a given BirA_{R118G}(BirA*)-tagged protein in living cells through their covalent biotin labeling,
72 eliminating the requirement for the preservation of PPIs upon verification process. Therefore, proximal partners
73 of the BirA*-tagged bait proteins are captured regardless of their solubility and PPI affinity. Since SARS-CoV-2
74 heavily relies on membranous organelle-associated processes, proximal interactomics pipelines, such as BioID,
75 are best suited to explore the protein environment in which SARS-CoV-2 polypeptides operate to hijack host
76 cells². We used BioID-based proximal interactomic approaches to delineate the interplay of SARS-CoV-2/human
77 proteins (**Fig.S1**) and better understand the invasion features of SARS-CoV-2 within cells. This dataset allowed
78 us to construct a 3D map of the approximate relative localizations of virus and host proteins within the cell volume
79 (**Fig.S1**). Featuring a user-friendly interface, this 3D map is an invaluable tool for unraveling the SARS-CoV-2-
80 mediated hijacking of specific regions of the host proteome.

81 Orthogonal approaches detecting binary PPIs provide insight on the physical contacts between viral and host
82 proteins, which are underlying virus-driven hijacking of the host proteome. Therefore, we assessed direct virus-
83 host protein contacts by implementing a split-NanoLuciferase protein complementation assay performed in
84 mammalian cells (mN2H)⁸. PPI scoring based on interaction-mediated luciferase signals gave rise to interaction
85 maps highlighting the prevailing virus-host contacts among proximal host proteins (**Fig.S1**). The systematic
86 exploration of direct virus-host contacts among virus-proximal host proteins revealed 414 direct interactions
87 between 26 viral and 118 host proteins. Intriguingly, 41 of these host proteins (35%) have been reported as
88 critical for SARS-CoV-2 infection in 3+ CRISPR-Cas9 screens, corroborating the central role of direct virus
89 interactors for SARS-CoV-2 infection and replication^{9,10}. Subjecting several virus-host complexes to *in silico*

90 structure prediction using AlphaFold2 Multimer¹¹ or structure-based homology (**Fig.S1**), we obtained multiple
91 high confidence structures. Moreover, using an in-house assay for sensitive and quantitative detection of SARS-
92 CoV-2 infection in cell lines, we confirmed the potential of USP13 chemical inhibition to combat SARS-CoV-2
93 replication (**Fig.S1**), thus providing a strong rationale for a structure-directed search of USP13-NSP13 inhibitors.

94 Overall, our dataset templates the first interactive 3D modeling of the viral-host proteomes interface in the cell
95 volume, providing an unparalleled picture of the SARS-CoV-2 invasion pattern. Our pipeline culminates in
96 identifying high-confidence virus-host protein interfaces, paving the way for identifying PPI-disrupting
97 compounds with high therapeutic potential. Significantly, being applicable to any virus or intracellular pathogen
98 as soon as the genome is sequenced, this pipeline could represent a major asset for responding rapidly to
99 emerging pathogens and improving preparedness efficiency.

101 **Results**

102 **The landscape of SARS-CoV-2 proteins' cellular environment and physical interactions cores**

103 We used BioID to capture the neighbors of 27 SARS-CoV-2 polypeptides: (i) 15 of the 16 non-structural proteins
104 that are produced in infected cells from the proteolytic cleavage of the Orf1a/b polyproteins. NSP11 was excluded
105 due to its small size (12 AAs); (ii) the four structural proteins: Spike (S) glycoprotein, Envelope (E) protein,
106 Membrane (M) glycoprotein, and Nucleocapsid (N) protein; (iii) eight accessory proteins: Orf3a, Orf3d, Orf6,
107 Orf7a, Orf7b, Orf8, Orf9b, Orf9c (**Fig.S1** and **supp table 1**). Following biochemical assays showing that the
108 BirA*-viral bait proteins phenocopy their untagged counterparts in impairing the innate immune responses
109 (**Fig.S2-S4**), we proceeded to the identification of proximal host factors of the 27 SARS-CoV-2 proteins. Our
110 experiments were performed either in basal conditions (n=3 with N- and C-ter BirA* tag) or following poly(I:C)
111 induction (n=3) to mimic a cellular antiviral response and subsequent host cell proteome remodeling. A total of
112 243 SARS-CoV-2 exploratory BioID samples were generated, and the BioID background was determined with
113 30 negative control runs performed using cells expressing BirA* alone, covering all the experimental settings.
114 High confidence interactors (p -values <0.01) were defined based on fold change (FC) over background and
115 detection in three experimental replicates. Aggregating all identified neighbors across the different replicates and
116 conditions, we obtained 10,125 high confidence proximal interactions between the 27 SARS-CoV-2 proteins and
117 2,593 host partners.

118 To benchmark our proximal interactomics dataset, we compared our results to all previously reported SARS-

119 CoV-2 interactions data in the BioGRID database (<https://thebiogrid.org/>). We observed a high overlap with the
120 proximal interactomic datasets even though the experimental conditions vary substantially with regard to the
121 used systems (BioID, TurboID and BioID2), cell lines (HEK293 and A549), purification protocols and analytical
122 tools. In total, 4,738/10,125 (47%) of our PPIs have been identified in at least one other proximal interactomic
123 study. Conversely, the percentages of the reported proximal interactions that are present in our dataset are
124 81.4% (2903/3565; BioID in HEK293 address only part of the SARS CoVo2 proteome¹²), 38.5% (3003/7810;
125 TurboID in A549¹³) and 24.7% (172/695 BioID2 in A549¹⁴). In addition, 1089 of our 10,125 proximal PPIs were
126 recovered in the body of SARS-CoV-2 viral proteins interactions studies obtained by different methods, *i.e.* AP-
127 MS, AP-WB, Y2H, crystallography (**supp table 2**). Combining our SARS-CoV-2 interactome datasets with the
128 existing ones, two validated core datasets can be distinguished (**supp table 3**): (i) a proximal core, supported
129 by other proximity-dependent biotin labeling studies, encompassing 4696 proximal interactions between 27 viral
130 bait proteins and 1,361 host proteins; and (ii), a physical core, *i.e.* the interactions confirmed by 1+ other method
131 detecting proteins complexes, including direct and indirect PPIs. The physical core encompasses 1088
132 interactions between 659 host proteins and 27 viral bait proteins (**Fig.1A**). Of these physical core interactions,
133 330 were identified only in our proximal interactome, highlighting the benefit of this SARS-CoV-2-host map.

134 **Functional novelties contributed by our SARS-CoV-2-host proximal interactomics**

135 Gene Ontology (GO) and pathways enrichment analyses of the proximal interactome of each viral bait unravel
136 their cell space and point to targeted processes or pathways (**supp table 2, supp table 4, Fig.S5**). It confirms,
137 *e.g.* the prominent targeting of the vesicular compartments reported in previous interactomics studies and
138 expected given the reliance of SARS-CoV-2 replication and transcription on ER-derived double-membrane
139 vesicle formation. We focused on the novelties provided by our dataset from the GO enrichment analysis. We
140 thus compared the GO terms associated with Biological Processes (BP), Cellular Components (CC) and
141 Pathways (KEGG) emerging specifically in our dataset against the combination of GO terms from all the other
142 interactomic datasets (**supp table 5**). It resulted in the identification of 33 novel pathways overrepresented in
143 the vicinity of viral polypeptides, a number of which relate to reported SARS-CoV-2 infection traits, such as cilium
144 assembly¹⁵, RNA-related processes and various lipids biosynthesis pathways¹⁶ (**supp table 5**). These data thus
145 provide a shortlist of novel PPIs and host factors assigned to pathways known to be altered upon SARS-CoV-2
146 infection.

147 To further characterize the links between functionally relevant host proteins within the proximal landscape of viral
148 polypeptides, we intersected our BioID dataset with SARS-CoV-2-related CRISPR-Cas9 studies available at
149 BioGRID ORCS <https://orcs.thebiogrid.org/>. It appears that 187 host factors required for SARS-CoV-2-induced

150 cytotoxicity in ≥ 3 CRISPR-Cas9 screens are present in our BioID dataset, connected to 27 viral bait proteins
151 through 694 proximal interactions (**supp table 2**). The distribution of interactions involving essential host factors
152 is as follows: (i) 105 PPIs are laying both in the physical and proximal cores connecting 52 essential host factors
153 to 18 baits (**Fig. 1B**). Of note, this set of PPIs represent super-high confidence PPIs, *i.e.* detected 3+ times by
154 at least two orthogonal methods; (ii) 41 PPIs are in the physical core only and connect 16 essential host factors
155 to 16 viral proteins (**Fig.1C**); (iii) 297 PPIs are in the proximal core only, connecting 91 essential host factors
156 with to 20 viral proteins (**Fig.1D**); and (iv), 261 PPIs were uniquely detected in our interactome, connecting 129
157 essential host factors to 27 viral proteins (**Fig.1E**). Note that among the 187 essential host factors, several are
158 involved in both the physical, the proximal cores through different PPIs they engage with viral proteins.

159 **Insights in the SARS-CoV-2 proteome hijacking upon anti-viral cellular response**

160 Antiviral cellular response induced by the poly(I:C) treatment expectedly remodeled the proximal landscape of
161 viral proteins with $\sim 10\%$ to $\sim 95\%$ of the interactors being impacted, depending on the viral protein (**supp table**
162 **6**). Three distinct remodeling patterns could be distinguished in response to infection-mimicking treatment,
163 excluding the viral proteins not reacting (less than 20% of differential hits) or those with less than 30 hits total:
164 (i) net enrichment for S, N, and NSP2; (ii) net depletion for NSP10, Orf3a, Orf6, NSP6, NSP7, Orf9b, Orf3d, E
165 and M; and (iii), exchange for NSP4, NSP12 and Orf9c (**Fig. 2A and supp table 6**). S and N proteins enter the
166 host cells with the incoming viral particles, and both gained proximal proteins in an antiviral cellular state. S
167 mostly gained ER proteins (93/102), which is a primary target of the virus. The N neighborhood gained the RISC-
168 loading complex components (DICER1, TARBP2, PRKRA), as well as proteins involved in the interferon
169 signaling pathway (IFIT1, IFIT2, IFIT3, EIF2AK2 and TRIM26). Additional novel links between viral factors and
170 major antiviral response pathway components emerged. We detected the recruitment of ISG20 and TOMM70
171 into close proximity of Orf9b, which is in accordance with its role in the inhibition of IFN-I response to infection¹⁷.
172 Additionally, we detected ZAP (ZC3HAV1) and TARBP2 near N; HERC5 and STAT1 near NSP2; and PARP9
173 near NSP16 (**supp table 6**).

174 **A comprehensive three-dimensional SARS-CoV-2 interactome**

175 Since BioID bait proteins label proximal partners within a spherical volume with a $\sim 10\text{-}20$ nm radius¹⁸, proximal
176 interactomics data capture information about the vicinity of the host factor in the physical space of living cells.
177 Based on these properties, we developed a methodology to approximate the relative positions of viral and host
178 proteins inside a theoretical cellular space (see Methods). By modeling the pairwise BioID proximal interactions
179 into a weighted graph structure and then applying a force-directed graph layout algorithm in three dimensions,

180 we are able to arrive at a set of 3D coordinates for the graph's vertices, representing the positions of viral and
181 host proteins. We generally consider these positions as indicative of the centers of extended cellular regions
182 where molecules of each protein reside. The set of coordinates allows us to infer relative distances between
183 nodes within our abstract 3D space, compare proximities and observe general regions of viral interaction. While
184 difficult to establish a precise, geometric correspondence between our generated 3D volume and the cell's
185 physical volume due to experimental as well as graph modeling limitations (such as in the case of multiple protein
186 localizations which cannot be accurately captured by a single node location), we consider our approximation of
187 the spatial organization of viral and host proteins as forming a "sketch" of their interplay inside the actual physical
188 cell volume.

189
190 The global shape of the 3D virus-host network reveals a cluster of 13 viral proteins (Orf3a, Orf3d, M, S, E, NSP3,
191 NSP4, NSP6, Orf8, Orf7a, Orf7b, Orf6 and Orf9c) gathered with human proteins, surrounded by 14 viral proteins
192 dragged out by different sets of partners (**Fig.2B**). From the density of nodes in the 3D map, we thus inferred
193 that removing the host proteins connected to eight or more viral proteins would avoid potential compartment-
194 dependent proximal background. The overall density of the host proteins in the 3D space shows a sharp
195 decrease with radial distance from the cluster's centroid (**Fig.S6**). When restricting to proteins with a maximum
196 of seven viral interactions, we observe a density peak and a smooth decrease with radial distance from the
197 centroid (**Fig.S6**). We use the radius of the density inflection point to define a threshold for the cluster size or
198 dense core. This enables us to identify a list of 790 host factors residing within the dense core boundary, which
199 are in their vast majority assigned to membrane categories (325 to the ER membrane, 165 to the vesicle
200 membrane, 109 to the Golgi membrane and 109 to the plasma membrane region) (**supp table 7**). These
201 membrane-rich organelles are the known primary location of nine viral proteins (Orf3a, M, S, E, NSP3, NSP4,
202 NSP6, Orf7a and Orf7b)^{1,2}, all of which are included in this core. Expectedly, when removing host proteins with
203 degree 8 or higher, the dense core was relaxed while keeping membrane-bound host proximal partners of a
204 specific set of viral proteins (**Fig.2B**). Hence, this operation facilitates the extraction of discrete proximity-based
205 hypotheses for each viral protein. The resulting 3D virus-host interaction map is available in an interactive web
206 interface (temporarily accessible at <http://62.217.122.54/>, to be deployed at www.sars-cov-2-interactome.org
207 upon publication) to visualize and explore the SARS-CoV-2-host proteins spatial organization. The regions of
208 pathways-filtered subnetworks can also be captured, revealing their positioning relative to the SARS-CoV-2
209 proteins, thus highlighting coordination between viral proteins to exploit host cells (**Fig. 2C**). This is well
210 exemplified for protein translation, spatially concentrated near NSP1, NPS7, NSP10 and Orf8, with the
211 translation initiation complex restricted to the vicinity of NSP1, and ribosomal subunits close to Orf8, thereby

distinguishing discrete virus-host interplays regulating translation. In addition, SNARE proteins involved in membrane fusion are specifically near the cluster of viral membrane factors. Other processes are distributed throughout the virus-host proximal network of interactions, such as the cell junction or viral processes.

Assembling data from previous studies in the literature and combining it with our BioID dataset enabled us to get a full picture of the SARS-CoV-2-host interplay identified to date by utilizing the same modeling and 3D visualization approach. The overall network shape determined by our dataset is preserved, indicating that the graph layout approach provides consistent viral-host neighborhoods from all SARS-CoV-2 interactomics studies. Each released interactome is separately accessible in this global SARS-CoV-2-host 3D map. The expected underrepresentation of membrane-associated PPIs obtained from the AP-MS dataset compared to proximal interactomes emerges in the visualization (“Global comparative view” tab at <http://62.217.122.54>). At a first glance, the membrane-rich central region of the network is equally enriched in CRISPR-Cas9 hits than regions enriched in soluble factors, stressing the importance of those host proteins for SARS-CoV-2 infection. GO-term based filtering of SARS-CoV-2 targeted host factors provides a rather comprehensive picture of the spatial organization of the SARS-CoV-2 proteins in relation to specific parts of the cellular machinery, thus strongly increasing the interpretability of the identified SARS-CoV-2 host interactions. The 3D representation of SARS-CoV-2-host proximal interactions is a pioneering step forward toward a more realistic picture of SARS-CoV-2 invasion patterns in cells, and PPIs network representation in general.

Exploring physical contacts of the viral proteins in the proximity interactome dataset

Combining orthogonal PPI detection methods is a well-established option to increase the accuracy of large-scale PPIs maps. We complemented our BioID proximal interactomics dataset with a split-nanoluciferase assay, which senses binary PPIs by protein-fragment complementation of the nanoluciferase enzyme in human cells (mN2H), and offers excellent detection performance⁸. PPIs screens by mN2H are supervised, systematic (matrix-based) and semi-quantitative (luciferase-based intensity map interactions), providing a strong orthogonality to the BioID (see methods). We sampled two sets of factors from our BioID dataset for mN2H-based PPIs profiling with the SARS-CoV-2 proteins (**Fig. 3A**): (i) 144 host factors highly enriched with less than 4 viral bait proteins (based on log₂ fold change over background and q-value significance); and (ii) 92 host factors scoring positive in 3+ CRISPR-Cas9 screens. The selected host factors were tested against all 27 viral proteins by mN2H, resulting in comparative virus-host interaction profiles among the selected proximal factors. The full mN2H matrix interrogated about 6372 viral-host protein pairs for direct interaction (**supp tables 8 and 9**), of which 587 correspond to proximal protein pairs as identified by BioID (**supp table 9**).

242 Among the tested viral-host protein pairs, regardless of their status in BioID, 6% (414/6372) showed direct
243 interactions, involving 118 host proteins. In the protein pairs detected by BioID 21% (123/587) were direct,
244 indicating a 3.5-fold enrichment (**supp table 9**). The mN2H SARS-CoV-2-host contactome determined in this
245 study is provided in **Fig.3B**. It significantly extends our recently released SARS-CoV-2-human contactomes^{9,10},
246 offering a wealth of novel SARS-CoV-2-host contacts. Significantly, the mN2H-based systematic PPIs profiling
247 uniquely identified 151 direct interactions involving 41 host factors essential for SARS-CoV-2 lethality in 3+
248 CRISPR-Cas9 screens, of high therapeutic interest.

249 Signal intensities detected in split-nanoluciferase assays correlate with the interaction strength¹⁹, allowing
250 qualification of interactions according to their distance from the positive PPIs threshold. From the comparative
251 intensity map, distinct sets of interactions easily emerged, such as NSP1 and NSP7 proteins targeting three
252 subunits of the EIF3 translation complex (EIF3L, EIF3G, EIF3H), or the binding of mitochondrial factors by Orf9b
253 (**Fig.4A**). Other host proteins critical for infection (based on CRISPR hits) are primarily contacted by a single
254 viral protein (**supp table 9, Fig.4A**). The intensity map of SARS-CoV-2-host interactions affords an advanced
255 level of precision of the SARS-CoV-2-host interplay with a subset of the human proteome, including proteins
256 essential for virus-induced cytotoxicity.

257 Hierarchical clustering of mN2H PPIs profiles of SARS-CoV-2 proteins essentially reflects the 3D map (**Fig.4B**),
258 suggesting that direct contacts shape the virus-host spatial organization. It also reveals a specialized
259 manipulation of the host machinery by neighboring viral proteins in the 3D map. PPI profile-based clustering
260 separates viral proteins into two main groups: the first one consists of most of the non-structural proteins that
261 each exhibit few contacts with host proteins, while the second cluster consists of membrane-associated viral
262 proteins that engage in the majority of direct interactions with the host. The most striking host cluster corresponds
263 to proteins involved in ER organization, which is co-opted by all membrane-associated viral factors (**Fig.4B**).
264 Other host clusters appear specific to a given viral protein, such as proteins involved in translation initiation (4
265 members of the EIF family) or in phosphatidylinositol processes (FLII and WIP1) that are especially associated
266 with NSP1. NSP3 clusters with proteins involved in negative regulation of proteins' metabolic process (SENP1,
267 RTN4, FXR1 and FXR2). Orf3d targeted proteins involved in intramembrane lipid transporter activity (ATP9A,
268 PLSCR1), while Orf9b binds to many mitochondrial proteins (BCL2L13, MFF, MRPL34, NDUFB8 and
269 TMEM242).

270 **Intraviral interactions profiling**

271 In the course of the virus life cycle, viral proteins cooperate to orchestrate the reprogramming of the cell

272 machinery. To provide a picture of viral proteins cooperation and expand on previous yeast two-hybrid based
273 datasets^{9,10}, we explored the intrinsic capacities of the SARS-CoV-2 proteins to interact with each other using
274 mN2H-based PPI profiling (**supp table 10, Fig.4C**). A number of viral heterodimers were detected, including the
275 well-characterized NSP16-NSP10^{20,21}, N-NSP3²². Of note, no binary interactions between the core replicase
276 complex proteins (NSP7-NSP8-NSP12) were detected, possibly due to inappropriate the conformation of the
277 proteins when tested in a pairwise manner, as these proteins are part of a more complex structure needing all
278 three proteins to achieve the hexameric architecture of this complex²³. A hub of intra-viral PPIs emerges within
279 the set of organelles-associated viral factors lying close together in the 3D network and the hierarchical clusters
280 (NSP3, NSP4 and NSP6, E, S, M, Orf7a and Orf7b) (**Fig.4D**). Thus, direct intra-virus interactions are likely
281 building cooperative anchorages within these cellular regions upon infection. The intraviral PPI data thus helps
282 to refine the SARS-CoV-2-host interactome. Our SARS-CoV-2-host interactions resource as such provides multi-
283 level insights into the SARS-CoV-2 cellular invasion pattern, both through the spatial positioning of viral proteins
284 relative to host proteome and the direct virus-host and virus-virus contacts underlying such organization during
285 the course of the virus life cycle.

286 **Predictive value of the BioID-based 3D interaction map**

287 When plugged into the BioID data-driven 3D map, the mN2H positive host proteins were closely confined around
288 their viral partners (despite the fact that they were not used for the 3D layout modeling) (**Fig.5A**). We examined
289 the distances between the viral proteins and host factors based on four categories: (i) non-interacting, non-
290 proximal proteins (PPIs not detected by either method); (ii) proximal proteins (PPIs detected by BioID only) (iii)
291 direct-contact proteins within the BioID dataset (PPIs detected by both BioID and mN2H) (iv) *de-novo* identified
292 direct-contact proteins (PPIs detected by mN2H only). We report that directly interacting proteins detected by
293 mN2H and identified within the 6372 viral-host protein pairs tested in mN2H were placed at shorter distances on
294 average than non-interacting proteins (based on mN2H negative signal). This remained true when restricting to
295 the subset of proteins detected by BioID, as well as not detected by BioID: mN2H-positive interactions showed
296 an average distance lower than mN2H-negative ones in both cases (**Fig.5B, Fig.S6E-F**). While the mN2H-tested
297 matrix showed an average viral-host protein distance of 0.58 in 3D model coordinates, pairs identified by mN2H
298 only, BioID only and mN2H & BioID were at average distances of 0.45, 0.29 and 0.25, respectively. These results
299 support the predictive power of the 3D network layout in foretelling volumes enriched in direct PPIs.

300 Identifying the interaction interface of host-virus complexes provides an important asset to developing
301 rationalized PPIs-disrupting compounds. We assessed the power of our mN2H matrix of direct interactions to
302 provide virus-host interaction interfaces by *in silico* prediction of the structure of virus-host complexes using

AlphaFold-Multimer¹¹. On a selection of 70 virus-host PPIs scoring positive by mN2H and using pDockQ scoring^{24,25}, 10/70 mN2H-positive pairs scored positive (pDockQ >0.23) (**supp table 11**), providing as many high-confidence interfaces (**Fig. S7**) which can serve as a basis for the development of PPI targeting compounds.

Significance of the SARS-CoV-2-host contacts to lead to the identification of therapeutic targets

We also applied a search for structural homology between virus-host binary interacting pairs (i.e. pairs that scored positive in mN2H) and human complexes with known structures. The previously reported and found in this study interaction between NSP13 and the human deubiquitinase USP13²⁶, could be derived by homology-based structural modeling with the human UBP14/PRS7 complex (**Fig.5C**). To estimate the potential of targeting NSP13/USP13 with PPIs-disruptive compounds, the involvement of USP13 in SARS-CoV-2 infection has to be demonstrated, given it has not been detected in CRISPR screens.

To address this and meet the need for high-throughput assays that allow screening for compounds with antiviral activity, we developed a time- and cost-effective quantitative readout of SARS-CoV-2-infection in cell lines, which relies on the complementation of the split-nanoluciferase enzyme upon fusion between infected cells, a hallmark of SARS-CoV-2-induced cytopathogenic effect²⁷⁻²⁹. The assay relies on the auto-assembly of two peptides of the nanoluciferase enzyme (a large N-terminal fragment NanoLg and a small C terminal peptide NanoSm, **see method**). Briefly, the assay is based on infection by SARS-CoV-2 of a co-culture of two SARS-CoV-2-sensitive cell lines, each stably expressing one the complementary fragment (VeroE6-NanoLg or VeroE6-NanoSm, respectively). Infection by SARS-CoV-2 induces the formation of syncytia, where complementary nanoluciferase fragments assemble to form an active enzyme (**Fig.5D**). A robust and dose-dependent nanoLuciferase signal was detected upon infection with serial dilutions of the strain Wuhan of SARS-CoV-2 (hCoV-19/France/GES-1973/2020, EPI_ISL_414631) of the co-culture, but not in separated monoculture of each cell line, attesting the performance of this assay (**Fig.S8**). Using this fusion-induced nanoluciferase complementation assay (NanoFuse) in VeroE6 cells, we tested the effect of the spautin-1, a previously reported USP13 inhibitor²⁶. We observed a dose-dependent decrease of SARS-CoV-2 infection upon spautin-1 treatment (**Fig. 5E**). The widely used protease inhibitor GC-376, assessed in the same assay had a 50% inhibitory concentration (IC₅₀) of 2.3 μM, in accordance with previous reports³⁰ which ascertains that the nanoluciferase complementation assay properly reflects an active SARS-CoV-2 infection (**Fig.5E**). Spautin-1 inhibited SARS-CoV-2 infection more efficiently than GC376, with a calculated IC₅₀ of ~0.8 μM, while no toxicity of the compound was observed (**Fig. S8**). The dose-response profile suggests that USP13 is important but non-essential for SARS-CoV-2 replication since the inhibition plateaued at 80%, which aligns with its lack of detection in the CRISPR- screens. Nevertheless, our data confirmed the involvement of USP13 in infection by SARS-CoV-2, thus deserving further

334 screening of PPI disruptive compounds. We have also developed a nano-luciferase complementation assay to
335 detect SARS-CoV-2, whose sensitivity and simplicity enable high-throughput screening of SARS-CoV-2
336 interfering compounds.

337 **Discussion**

338 The wealth of SARS-CoV-2 interactomics data sketches the complexity of any virus-host interplay. Our data
339 improves the current knowledge of cell hijacking by SARS-CoV-2 first by providing a high coverage of the virus-
340 host landscape, merging data from 27 SARS-CoV-2 polypeptides in basal and infection-mimicking conditions.
341 Combined with the other SARS-CoV-2 interactomes, our dataset significantly increases the number of SARS-
342 CoV-2-host links validated by ≥ 2 orthogonal methods (physical core of virus-host PPI), while overlap with only
343 proximity-dependent interactomes defines a validated proximal core. Our study also identifies 5,114 novel high-
344 confidence proximal interactions. From our BioID dataset, we generated an innovative representation, allowing
345 visualization of the relative spatial distribution of virus and host proteins. BioID identifies proteins within a 10-20
346 nm spherical radius around a bait protein, which can be leveraged to position baits and interactors relative to
347 each other. The occupation volumes near viral baits can thus be depicted, providing a geometric map of SARS-
348 CoV-2 proteins in the cell. It represents a conceptual advance towards a more realistic description of the virus
349 invasion pattern within the cell's volume and possible strategies to counteract it. Combining the results of all
350 SARS-CoV-2-host interactomes generated to date paves the way for the perturbation of regions emerging as
351 central in cell hijacking, thus offering an avenue for advanced therapeutic paths. In the longer term, the
352 customizable online dashboard will achieve a basal 3D cellular repository obtained by incrementing all human
353 proximal interactomic studies to this initial framework, expanding the scope and reach of this approach. While
354 representing the proteins as fixed points in space presents a limitation since it does not adequately capture
355 protein movement inside the cell or multiple localizations, our representation can still serve to produce indicative
356 centroids of volumes where proteins could be localized and a first attempt to depict this has been implemented
357 as an option in the online dashboard. A more complete modeling would probably entail abandoning the graph
358 structure and assigning 3D probability densities instead of 3D points to each protein, something akin to a
359 probability density "cloud". These densities could then have multiple peaks representing distinct localizations.
360 Using such an extended location formalism is a promising direction for exploiting the wealth of BioID data for a
361 more accurate and realistic cell volume depiction, which we have left for future work.

362 Beyond the big picture, our work is unique in that it systematically captures the direct virus-host contacts within
363 a subset of the proximal interactome, identifying 414 direct virus-host PPIs. These direct PPIs are characterized
364 by shorter than average edges in the geometric SARS-CoV-2-host map, supporting the predictive power of the

365 map on direct contact probability as forebode³¹.

366 Among the 235 cellular proteins sampled from BioID dataset, 118 displayed direct contacts, of which 41 are
367 essential high confidence factors for SARS-CoV-2-induced toxicity previously identified in 3+ CRISPR-Cas9
368 screens. The two only recently released contactome maps^{9,10}, based on yeast two-hybrid, remarkably showed
369 that the direct targets of viral proteins are part of network communities enriched in proteins genetically associated
370 with the risk of developing severe COVID-19¹⁴, and that such direct SARS-CoV-2 targets are enriched in gene
371 products differentially expressed in samples from COVID-19 patients¹⁵, respectively. Importantly, our pipeline
372 was thus the only one leading to the capture of direct contacts between SARS-CoV-2 proteins and human
373 proteins essential for infection, those PPIs being attractive targets for anti-viral therapeutics exploration. We
374 believe it is likely due to the ability of our pipeline to identify viral proximal factors by BioID independently of their
375 subcellular localization and solubility. Indeed, of the 151 direct PPIs engaging the 41 essential host proteins, 91
376 (60%) involve SARS-CoV-2 proteins associated with membrane-associated sub-cellular compartments (NSP3,
377 NSP4, NSP6, S, E, M, Orf3a, Orf7a and Orf7b), and 31 of 41 (76%) targeted essential host proteins are
378 associated with intracellular organelles.

379 Profiling by mN2H revealed a clustering of the viral proteins with the relative strength of the interactions
380 highlighting the strongest ones, thereby helping to uncover the main drivers of SARS-CoV-2 mediated cell
381 hijacking. This is exemplified by the set of transmembrane proteins (TMED2, TMEM41B, TMEM106B,
382 TMEM242, TMEM33 and TMEM159) exhibiting distinct interaction patterns with viral proteins. The TMEM41B,
383 a poorly characterized ER-localized protein, prominently binds to Orf9b and Orf3a and has been shown to be
384 essential for infection by SARS-CoV-2, and by other human coronaviruses (HCoV-OC43, HCoV-NL63, and
385 HCoV-229E³²) as well as by some flaviviruses³³. It represents a cellular regulation hub co-opted by multiple
386 viruses that is worth countering. Weak interactions represent low affinity or labile PPIs, which may also be
387 functionally relevant. Thus, prioritizing PPIs to be further investigated should consider both the interaction profiles
388 (interaction strength and redundancy) and host protein essentiality for infection.

389 Knowledge of the structure of SARS-CoV-2-host complexes would be a clear asset for screening PPIs-disruptive
390 compounds. The mN2H dataset provides a resource for PPIs modeling, which we trained using a selection of
391 N2H-identified virus-host complexes. This analysis identified 10 virus-host complexes of strong existence
392 probability (DockQ>0.23) and provided structure models. The AlphaFold2-predicted structure of those proteins
393 that are structurally known are close to their experimentally defined structures. Examples include the Rab6A and
394 Rab7A proteins structured in complexes with NSP4 of SARS-CoV-2, which are perfectly superimposable with
395 their experimentally determined crystal structures.

396 The structure of virus-host protein complexes can also be predicted by homology-based modeling, when
397 homology is found with structurally characterized co-complexes. This was the case for NSP13/USP13 interaction
398 by structure-based homology with the human UBP14/PRS7 complex. To assess the role of USP13 in SARS-
399 CoV-2 infection, we developed a quick, cost-effective and sensitive assay for measuring SARS-CoV-2 infection,
400 based on the complementation of the nanoluciferase enzyme mediated by cell fusion following infection. We
401 then characterized USP13 as important for SARS-CoV-2 replication since its inhibition by Spautin-1 drastically
402 impaired SARS-CoV-2 replication with an IC₅₀ of ~0.8 μM. These data corroborate a previous study²⁶ and point
403 to Spautin-1 as a potential adjuvant therapy. Together, these results validate the NSP13/USP13 interaction as
404 relevant for structure-driven screening of PPIs disruptive drugs. Importantly, they provide proof of principle that
405 our proteomic BioID/mN2H pipeline represents a valuable resource for selecting relevant PPIs for antiviral
406 compound screening. This pipeline is easily adaptable to all emerging human pathogens to prompt the
407 identification of therapeutic targets and, therefore, should account for improved preparedness.

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542 **Methods**

543 **Plasmids**

544 SARS-CoV-2 viral proteins coding sequences were cloned by Gateway recombination system (Life
545 Technologies) into pDEST pcDNA5 FRT/TO N-ter and C-ter BirA*Flag vectors for the BioID (Coyaud et al.,
546 2018), and into pciNeo-N2 or pciNeo-C2⁸ using the collection of Gateway-compatible entry clones reported in
547 ³⁴ (see Supp Table 1 for sequences). For N2H, cDNA encoding the human proteins to be tested were available
548 pDONR vectors from the human ORFeome collection v8.1 from the Center for Cancer Systems Biology (CCSB)
549 (ORFeome Collaboration, 2016)) or inhouse made clones. Each hORF was introduced into pciNeo-N1 pDEST
550 plasmids by Gateway reaction. All clones were sequence verified.

551 **Cell lines**

552 HEK293 cells stably were stably transfected by the different BirA*tag- encoding plasmids as in (Coyaud et al.
553 2018). Briefly, Flp-In™ T-REx™ HEK293 cells were grown in Dulbecco's Modified Eagle's Medium (DMEM,
554 Gibco) supplemented with 10% fetal bovine serum (FBS, Sigma-Aldrich), GlutaMAX™ and Penicillin-
555 Streptomycin (1x). Using the Flp-In system (Invitrogen), Flp-In™ T-REx™ HEK293 stably expressing BirA*Flag
556 or FlagBirA* alone (for control samples), or N- and C-terminally tagged viral bait proteins were generated by co-
557 transfecting pOG44 with each pcDNA5 FRT/TO BirA*Flag-viral protein encoding plasmid and selected with 200
558 µg/ml hygromycin B

559 **Confocal microscopy**

560 Each N-ter tagged viral protein expression was induced by 1µg/mL tetracycline and 50µM biotin for 24 hrs (in
561 basal or poly(I:C) conditions) in stably transfected Flp-In™ T-REx™ HEK293 cells. Briefly, cells were grown on
562 poly-lysine coated coverslips, rinsed with PBS before fixation with 4% paraformaldehyde for 15 min at RT. After
563 fixation, they were washed 3 times with PBS 0.1% Triton X-100. Cells were then blocked in PBS 0.01% Triton,
564 and 5% bovine serum albumin (BSA) for 30 min at 4°C. Transfected cells were then incubated overnight at 4°C
565 with monoclonal anti-FLAG M2 mouse antibody (dilution 1:2,000, Sigma-Aldrich). After 3 washes for 10 min with
566 PBS 0.01% Triton X-100, cells were incubated for 1 h at 37 °C with the Alexa Fluor 647-conjugated secondary
567 antibody donkey anti-mouse IgG (H+L) (1:2,000, Thermo Fisher Scientific) and streptavidin-Alexa Fluor 488
568 (1:20,000; Invitrogen). Cells were rinsed with PBS, and nuclei were stained with 1 µg/ml 4',6-diamidino-2-
569 phenylindole DAPI (Thermo Fisher Scientific) in PBS for 5 min. Finally, cells were washed, desalted and mounted
570 using Dako Fluorescent Mounting Medium (Agilent Dako, Santa Clara, CA, USA). Acquisition was performed on

571 a Zeiss LSM700 confocal microscope connected to a Zeiss Axiovert 200 M equipped with an EC Plan-Neofluar
572 40×/1.30 numerical aperture and an oil immersion objective (Carl Zeiss AG, Oberkochen, Germany). Processing
573 of the images was performed using Zeiss Zen 2 software and assembled using Adobe Illustrator.

574 **IFN-dependent luciferase activity assays**

575 ISRE reporter HEK293 cells referred to as STING37 (Lucas-Hourani et al., 2013) were plated in a 24-well plate
576 (2.5×10^5 per well). After 24h, cells were transfected using Lipofectamine 2000 (11668-019, Invitrogen by
577 Thermo Fisher Scientific) with 100 ng of pCIneo-MAVS³⁵ or the empty vector (pCIneo) with or without 200 ng of
578 pFlag-BirA plasmids expressing viral proteins. 24 h post-transfection cells were lysed by Passive Lysis buffer
579 (E1941, Promega), and luciferase activity was measured with Bright-Glo Luciferase assay system (E2650,
580 Promega).

581 **Bio-ID sample generation**

582 BioID samples were prepared from Flp-In™ T-REx™ HEK293 expressing each BirA*-flagged SARS-CoV-2
583 protein or BirA* alone as in³⁶. Briefly, three independent replicates of two 150 cm² plates of sub-confluent (60%)
584 cells were incubated for 24 hrs in complete media supplemented with 1 µg/ml tetracycline (Sigma), 50 µM biotin
585 (Thermo Fisher Scientific). For poly(I:C) treatment conditions, poly(I:C) (Sigma Sigma-Aldrich; P1530) was
586 transfected at 2µg/mL for 5hrs using PolyJet reagent prior to tetracycline and biotin 5 hours before tetracycline
587 and biotin addition. Cells were collected and pelleted (300 x g, 3 min), washed twice with PBS, and dried pellets
588 were snap frozen. Each cell pellet was resuspended in 5 ml of lysis buffer (50 mM Tris-HCl pH 7.5, 150 mM
589 NaCl, 1 mM EDTA, 1 mM EGTA, 1% Triton X-100, 0.1% SDS, 1:500 protease inhibitor cocktail (Sigma-Aldrich),
590 1:1,000 Turbonuclease (BPS Bioscience) and incubated on an end-over-end rotator at 4°C for 1 hour, briefly
591 sonicated to disrupt any visible aggregates, then centrifuged at 45,000 x g for 30 min at 4°C. Supernatant was
592 transferred to a fresh 15 mL conical tube. 25 µl of packed, pre-equilibrated Streptavidin Ultralink Resin (Pierce)
593 were added, and the mixture incubated for 3 hours at 4°C with rotation. Beads were pelleted by centrifugation at
594 300 x g for 2 min and transferred with 1 mL of lysis buffer to a fresh eppendorf tube. Beads were washed once
595 with 1 mL of lysis buffer and twice with 1 mL of 50 mM ammonium bicarbonate (pH=8.3), then transferred in
596 ammonium bicarbonate to a fresh centrifuge tube and washed two more times with 1 ml of ammonium
597 bicarbonate buffer. Tryptic digestion was performed by incubating the beads with 1 µg MS-grade TPCK trypsin
598 (Promega, Madison, WI) dissolved in 200 µl of 50 mM ammonium bicarbonate (pH 8.3) overnight at 37°C. The
599 following morning, 0.5 µg MS-grade TPCK trypsin was added to the beads and incubated 2 additional hours at
600 37°C. Following centrifugation at 2,000 x g for 2 min, the supernatant was collected and transferred to a fresh

601 eppendorf tube. Two additional washes were performed with 150 μ L of 50 mM ammonium bicarbonate and
602 pooled with the first eluate. The sample was lyophilized and resuspended in buffer A (2% ACN 0.1% formic acid).
603 1/3rd of each sample was analyzed per mass spectrometer run.

604 **BioID data acquisition**

605 MS samples were prepared from three biological replicates of each bait protein fused either with an N-terminal
606 or a C-terminal BirA*Flag epitope tag in basal condition or following poly(I:C) (Sigma-Aldrich; P1530) transfection
607 at 2 μ g/mL for 5hrs using PolyJet reagent (Signagen) prior to tetracycline and biotin induction, and analyzed on
608 a Thermo Q-Exactive mass spectrometer. Samples were separated by online reversed-phase chromatography
609 using a Thermo Scientific Easy-nLC1000 system equipped with a Proxeon trap column (75 μ m ID x 2 cm, 3 μ m,
610 Thermo Scientific) and a C18 packed-tip column (Acclaim PepMap, 75 μ m ID x 50 cm, 2 μ m, Thermo Scientific).
611 The digested peptides were separated using an increasing amount of acetonitrile in 0.1% formic acid from 2 to
612 30% for 2 hours at a 300 nL/min flow rate. A voltage of 2.4 kV was applied by the liquid junction to electrospray
613 the eluent using the nanospray source. A high-resolution mass spectrometer Q-Exactive™ Thermo Scientific™
614 was coupled to the chromatography system to acquire the 10 most intense ions of MS1 analysis (Top 10) in
615 data-dependent mode. The MS analyses were performed in positive mode at a resolving power of 70,000 FWHM,
616 using an automatic gain control target of 3e6, the default charge state was set at 2 and a maximum injection
617 time at 120 ms. For full scan MS, the scan range was set between m/z 300 to 1600. For ddMS2, the scan range
618 was between m/z 200 to 2000, 1 microscan was acquired at 17,500 FWHM, an AGC was set at 5e4 ions, and
619 an isolation window of m/z 4,0 was used. The mass spectrometry proteomics data have been deposited to the
620 ProteomeXchange Consortium via the PRIDE³⁷ partner repository with the dataset identifier PXD033452
621 (reviewer username: pxd033452@ebi.ac.uk; pw: Z1tHrDTc)

622 **BioID data analysis**

623 The proteins were identified by comparing all MS/MS data with the Homo sapiens proteome database (Uniprot,
624 release March 2020, Canonical+Isoforms, comprising 42,360 entries + viral bait protein sequences added
625 manually), using the MaxQuant software version 1.5.8.3. The digestion parameters were defined using trypsin
626 with 2 maximum missed cleavages. The oxidation of methionine and N-terminal protein acetylation were defined
627 as variable modifications. The Label-free quantification (LFQ) was done keeping the software's default
628 parameters. As for initial mass tolerance, 6 ppm was selected for MS mode, and 20 ppm was set for
629 fragmentation data to match MS/MS tolerance. The identification parameters of the proteins and peptides were
630 performed with a false discovery rate (FDR) at 1%, and a minimum of 2 unique peptides per protein. The LFQ

631 values from the 30 control runs (regrouping FlagBirA* and BirA*Flag alone samples, from stable and transiently
632 transfected cell lines, and w/wo poly(I:C) pretreatment) were collapsed to the three highest values for each given
633 ID. These three values were defined as the control group for comparison with viral bait proteins triplicates. The
634 statistical analysis was done by Perseus software (version 1.6.2.3). Briefly, the LFQ intensity of each sample
635 was downloaded in Perseus, and the data matrix was filtered by removing the potential contaminants, reverse
636 and only identified by site. The data were then transformed using the $\log_2(x)$ function. Before statistical analysis,
637 81 groups (27 bait proteins, N-ter, C-ter and poly(I:C) for each) were defined with 3 replicates per group. Only
638 preys with detected values in all three replicates of a given viral bait protein were kept for further analysis. Missing
639 values were then replaced from the normal distribution separately for each column. Two-sample Student's T-
640 test was then performed comparing all three biological replicates of each bait and condition against the three
641 control runs. High-confidence proximal interactors were defined by permutation-based FDR with a cut-off of 0.01.
642 Perseus output with all experimental values is reported in **Supp Table 2** (tab B).

643 The matrix (**Supp Table 2**) shows the average \log_2 fold change against control and the corresponding p- and q-
644 values for each bait and condition. The poly(I:C) data were also compared against the control (green font when
645 significant) and the corresponding condition (N-terminal tag for all viral bait proteins, except for NSP1). The
646 poly(I:C) column indicates the status of each interaction when compared to the basal condition. 'Decreased' and
647 'Increased' were defined when the \log_2 fc against the basal condition was $<(-1)$ or $>(+1)$, respectively, and the
648 p-value comparing the 3 replicates of the two conditions below 0.05. 'Unchanged' corresponds to \log_2 fc values
649 between -1 and $+1$, and empty cells depict interactors undetected in the basal and poly(I:C) condition. The
650 InDegrees column depicts the number of bait proteins detecting a given interactor, regardless of the condition
651 (N-ter, C-ter or poly(I:C)). This criterion was chosen to filter out the most connected interactors (8+), likely to be
652 organelle-specific background not filtered using the BirA* alone control samples. 2D networks presented in the
653 different figures were generated using the Cytoscape software (v.3.9.1; <https://cytoscape.org/>).

654 **Modeling the virus-host interplay in 3D from the BiOLD Data set**

655 In order to approximate the regions where proteins mainly reside in the cell volume, we used computational
656 techniques for the layout of graphs in three dimensions. A custom force-directed and multi-stage algorithm was
657 employed due to its general applicability and capability for the generation of high-quality layouts^{38,39-41}. Another
658 advantage offered by the class of force-directed algorithms, making it suitable for our specific case, is the
659 inherent physical attributes and the flexibility of tuning provided by the underlying physics-inspired model
660 producing the layout coordinates. In this approach, the observed BiOLD proximal interactions (when present)
661 modulate the strength of an attractive force between proximal protein pairs, while repulsive forces universal

among all node pairs ensure the separation of regions and the expansion into the available volume. In this way, host proteins previously known to have proximal interactions are pulled close to each other and viral proteins with overlapping sets of host interactors are also brought into relative proximity. Interactors for which there is no pairwise proximal relation are pushed apart, their placement determined by their own set of individual proximal interactions. By simultaneously aggregating all the proximity interactors and their interactions subject to these forces, a global approximate picture of the relative spatial organization of viral proteins with respect to the host proteome is emerging.

Interactome network setup and visualization

The starting point for constructing the interactome network is the BioID high confidence PPI data table, each row of the table containing data of a single bait-target interaction. Aggregating all the PPI data, we obtained a total of 10,125 interactions involving 27 baits and 2,593 targets (total of 2,620 nodes) to which we also appended 290 mN2H positive interactions not detected by BioID (these latter PPIs are not participating in the layout and therefore do not influence the node coordinates, but are used for displaying purposes). Based on this dataset, we construct an undirected graph where interactors and interactions are represented as the graph's vertices (nodes) and edges (links) respectively. In addition to links due to the observed BioID bait-target interactions, we augment the graph with 3,492 edges representing previously known target-target interactions. We thus obtain a final graph with a total of 2,620 vertices and 13,617 edges.

To visualize the graph, our objective is to determine an optimum placement of the nodes in three-dimensional space that would minimize edge crossings and reveal the interactome's structure to the best possible extent. We employ a custom, multi-stage, force-directed graph layout algorithm in 3D, based largely on the spring-electric model outlined by Hu³⁸. We modulate the strength of the attractive (spring) forces in the corresponding physical model with a factor w given by the combination of the BioID enrichment ratio (or fold change, fc) according to the formula: $w = \left[\sum_i \log_2(fc)_i \right]^p$, where $p=1/2$ and $i = \{\text{N-ter, C-ter, poly(I:C)}\}$, when present for the interaction.

For reported target-target interactions where this information is not available we set this factor equal to 1. We also ignore the aforementioned purely mN2H positive (non-BioID) interactions in order to preserve the orthogonality of the two approaches. The node coordinates in 3D obtained by minimizing the energy of the spring-electric physical model are ingested by a plotting package that generates a 3D visualization of the network, assigning a point to each node and a line connecting a pair of nodes when there is an interaction between the corresponding interactors. Our code was written in Python 3.8⁴² and makes use of several modules,

691 primarily: NetworkX for graph operations ⁴³, NumPy for numerical computations ⁴⁴, pandas for data manipulation
692 ⁴⁵ and Plotly for visualization ⁴⁶.

693 **Interactive web application**

694 In order to make our results available to the community in a usable and informative manner, we have
695 implemented a web application, accessible at <http://www.sars-cov-2-interactome.org/>. It contains the network
696 visualization, as well as multiple options for selecting and filtering the dataset to view sub-networks, providing
697 the ability to focus on particular areas of interest and explore various levels of detail. The interactive 3D
698 visualization of the interactome is a main ingredient of the dashboard, as it enables navigation within a browser
699 window, zooming and rotating in space, giving an intuitive illustration of the relative localizations of proteins inside
700 the cell volume. In addition, we integrate information about the previously reported SARS-CoV-2 status of
701 interactors and interactions, as well as previously reported interactions between the targets (cf. **Supp Table 3**
702 for details). The network customization options contain various filters, separated broadly into two levels: on the
703 first level, there are options for: (i) focusing on targets of higher specificity by filtering out targets with a total
704 number of bait interactions (or degree) above a configurable threshold (set to 7 by default); (ii) restricting to
705 targets based on known relevance for SARS-CoV-2 (based on total number of CRISPR screens the given
706 interactor has been identified as functionally important for SARS-CoV-2 infection); (iii) filtering interactions based
707 on their poly(I:C) treatment result; or (iv), selecting particular baits of interest and their interactions only. When
708 multiple filters are activated at this level, the resulting dataset is formed by the intersection of the selected criteria.
709 In addition, at this level, we provide options for displaying the previously reported target-target interactions, as
710 well as highlighting previously reported and N2H-validated bait-target interactions. At the second level, there is
711 the functionality to select any number of annotations of interest from three main categories (pathways, biological
712 processes, cellular components) in order to generate an annotated subnetwork, while remaining subject to the
713 filtering selections from the preceding level. In this second level, multiple annotation selections are combined so
714 that the resulting network is the union of the individually annotated subnetworks. Specific targets of interest can
715 also be selected individually, or added to an already generated subnetwork. Notably, there is an option to select
716 any number of annotations for which to display their approximate localization overlaid on the network, either as
717 3D density volumes, or as labels positioned at the volume's centroids ('label cloud'). At any given time, the
718 interactive visualization reflects the user's filtering selections, offering an intuitive view of the particular subset of
719 data under consideration, while the filtered dataset is also visible in table format below the network and
720 downloadable. Finally, there are options for mostly cosmetic attributes of the network visualization, such as the
721 ability to toggle text labels and edges, present a faded view of the full network in the background (thus giving a
722 better global view of the location of nodes and edges in the interactome), and display 3D spherical shapes for

the interactors (as opposed to flat 2D circular markers), better representing their physical size and actual location in space. A summary panel of the network with a breakdown of the totals for nodes and edges is presented alongside the network, as well as a panel displayed upon clicking on a node or edge with corresponding metadata, including AlphaFold protein structure predictions. A detailed usage tutorial is also provided as part of the website. The dashboard web application has been implemented using the Dash module in Python 3.8.

Mammalian cell-based N2H assay

HEK293T cells were seeded at 6×10^4 cells per well in 96-well, flat-bottom, cell culture microplates (Greiner Bio-One, #655083), and cultured in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal calf serum at 37 °C and 5% CO₂. Twenty-four hours later, cells were transfected with 100 ng of each N1- and N2/C2-expressing plasmids using linear polyethylenimine (PEI MAX 40000; Polysciences Inc; Cat# 24765) to co-express the protein pairs fused with complementary NanoLuc fragments, F1 and F2⁸. Twenty-four hours after transfection, the culture medium was removed, and 30 μL of 100× diluted NanoLuc substrate⁴⁷ was added to each well of a 96-well microplate containing the transfected cells. Luciferase enzymatic activity was measured using a CentroXS luminometer (Berthold; 2 s integration time). The luciferin substrate used (Q-108) in all the bioluminescence experiments was obtained in a concentrated solution, from the corresponding O-acetylated derivative hikarazine-108, following acidic hydrolysis as previously described^{48,49}. The binary virus-host PPIs were assessed and were independently performed two times, each made in duplicates to quadruplicates. PPIs were monitored using the cellular proteins (CP) fused the F1 nanoluciferase fragment at their N-terminus (N1-CP), while the viral proteins were fused to the nanoluciferase F2 fused either at N-, their C-terminus or both. Configuration of the virus-host PPIs matrix was either N1N2 or N1C2, and was selected from the viral bait Bir-A* fusion configuration leading to host partners identification in BioID.

To score positive PPIs from the tested matrixes, we applied the following pipeline:

1-Log₂-transformed averaged RLU signals were normalized to the mean columns signal and mean row signals (Log₂ RLU PPI - mean Log₂ column - mean Log₂ rows). Column and row averages reflect the overall level of viral and cellular protein interaction signals. A Z-score (relative to the mean and standard deviation of the full Normalized PPIs matrix) was applied on normalized data to obtain normal distribution and to calculate a P-value. A significant P-value (< 0.05) or equivalently a Z-score upper to 1.64 were scored as positive PPIs. In this way, positive PPIs are distinguished over non-interacting pairs with high stringency, leading to the identification of high-confidence PPIs.

Hierarchical clustering of virus-host PPIs was performed using the R software by using euclidean distance

753 parameter of the dist function. Clustering was then performed by ward.D method with R software by using
754 pheatmap function.

755 **A novel sensitive assay to measure SARS-CoV-2 infection**

756 cDNA encoding for the nanoluciferase fragment 1-158 (NanoLg) and the 13 aa long c-terminal Nanoluciferase
757 peptide (aa 159-171, Nano-Sm) were cloned in the pLVX-puro vector (Clontech) in fusion with GFP11 (at the C-
758 ter of LN) or GFP1-10 (at the N-ter of SN) respectively. Lentiviral vectors were produced by co-transfection of
759 the pLVX-puro NanoLg or pLVX-puro NanoSm plasmids with gag-pol packaging plasmid psPAX2 (Addgene,
760 #12260) and VSV-G envelope expressing plasmid in HEK293T cells using calcium phosphate precipitation.
761 Lentiviral titers were measured by p24 ELISA according to the manufacturer's instructions (Clontech).

762 10^5 cells Vero E6 were transduced with $5 \cdot 10^6$ TU NanoLg or NanoSm expressing lentiviruses for 4 hours, then
763 cells were grown in DMEM 5% SVF + 1% Penicillin-Streptomycin + 8 μ g/ml puromycin. Transduced cells were
764 then amplified under puromycin treatment.

765 For the SARS-CoV-2 infection assay, Vero E6-NanoLg and Vero E6-NanoSm were plated separately at 25000
766 cells/well or co-cultured (12500 of each cell lines mixed/well) in white 96 well plates (Greiner Bio-One, #655083).
767 One day after, cells were infected by serial dilutions of SARS-CoV-2 strain 1973 (hCoV-19/France/GES-
768 1973/2020, EPI_ISL_414631) in 50 μ l of DMEM, then incubated for 24 hours. Infection rates were measured by
769 lysing cells after removal of the medium in 30 μ l of 100-fold diluted NanoLuc substrate (Promega, #N1110).
770 Luciferase enzymatic activity was measured using a CentroXS luminometer (Berthold; 2 s integration time).

772 **Small Molecule Inhibition**

773
774 GC376 (Sigma Aldric) and Spautin-1 (Calbiochem) were dissolved in DMSO to make a 100 mM stock solution.
775 A 40 mM solution was prepared from the stock in DMSO, and then serial 2-fold dilutions were prepared in DMSO.
776 The final range of inhibitors was from 40 mM to 0.63 mM (*i.e.* 8 2-fold dilutions), which is 200 times higher than
777 the tested concentrations. Each compound dilution was diluted 1000-fold in DMEM (giving 2X concentrated
778 chemicals with equal DMSO concentration for each)

779 Vero E6 NanoLg + Vero E6 NanoSm co-cultures (12500 each) plated the day before in 96 white plates were
780 infected with 500 PFU of the previously described SARS-CoV-2 strain in 50 μ l DMEM for 1 hour. Then 50 μ l of
781 the 2X concentrated compounds were added. Cells were incubated for 24 hours. Infection rate was measured
782 by nanoluciferase activity reading as described above.

783 Each compound was measured in biological triplicates, each containing 3 to 8 technical replicates.

784 The results are presented as RLU \pm standard deviations. Dose-response curves were plotted using Prism v9
785 (GraphPad) in which IC₅₀ values were calculated with the variable slope model.

786 Cell viability was measured using the Cell-Titer-Glo[®] Luminescent Cell Viability Assay kit (Promega, G7750).

788 **AlphaFold-Multimer *in silico* structure prediction**

789
790 We produced predicted complex structures for the selected pairs of viral-host proteins by running the AlphaFold
791 source code v2.3.2 (git commit hash 2e6a78f) from <https://github.com/deepmind/alphafold> in multimer mode with
792 default parameters and with full databases via the provided Docker script. For each pair, we ran using input host
793 protein sequence FASTA files from <https://www.uniprot.org/> and generated 5 relaxed structures (from different
794 random seeds) for each of the 5 multimer models. We kept the top-ranked relaxed structure in terms of model
795 confidence in order to compute a predicted DockQ (pDockQ) score²⁵. Following the CAPRI definitions for the
796 correspondence between DockQ values and accuracy of predicted structures, we deem structures with pDockQ
797 ≥ 0.23 as positive hits for the purposes of our *in silico* validation.

799 **Figure legends**

800 **Fig 1. The physical core of the SARS-CoV-2-host interactome. (A)** Physical core representing the SARS-
801 CoV-2 interactions of our BioID dataset that have been identified by orthogonal, physical PPI detection methods.
802 Bait proteins are represented as black nodes, the proximal host proteins by nodes color coded following the
803 number of CRISPR hits as indicated in the color scale for the whole panel; **(B-E)** subnetworks of virus-host PPIs
804 identified in our BioID dataset and involving host factors scoring in 3+ independent CRISPR screens, subdivided
805 following their profile in the literature **(B)**: In proximal and physical cores; **(C)**: in Physical core; **(D)**: in Proximal core;
806 **(E)**: in Our BioID only).

807 **Fig 2. SARS-CoV-2/human proximal network representation. (A)** poly(I:C) remodeling of SARS-CoV-2
808 proteins proximal partners; **(left)** graphical pipeline; **(middle)** distribution of proximal interactors changes
809 for each viral bait protein. Horizontal axis depicts the percentage of the whole interactome of each bait
810 significantly changed upon poly(I:C) treatment. Red and blue bars for increased and decreased proximal host
811 partners, respectively. Number of unique host factors are shown on the graph. Bait names are colored according
812 to their status, as depicted in the graphical pipeline; **(right)** Volcano plots representing one bait of each category
813 (details are available in Supplemental table S6). **(B)** Global data-driven SARS-CoV-2/human proximal network
814 projection on 2D and its relaxation upon removal of the host proteins targeted by eight or more viral baits. Pxl

815 stands for proximal interactions **(C)** Spatial localization of functional groups of proteins in the proximal network
816 relative to the viral proteins (black nodes), as indicated. **(D)** Localization of human proteins essential for SARS-
817 CoV-2 induced toxicity (3+ CRISPR hits) in the 3D SARS-CoV-2 /Host proximal interaction maps.

818 **Fig 3. mN2H orthogonal validation of a subset of high confidence interactors.** **(A)** Selection of the
819 interactors and experimental pipeline for the mN2H validation. **(B)** Full SARS-CoV-2/Human contactome network
820 identified in this study by mN2H.

821 **Fig 4. mN2H interaction profiling.** **(A)** The relative strength of PPIs, given by the distance of PPI to the positive
822 threshold, is shown for host factors scoring positive with one or more viral factors. Their corresponding number
823 of CRISPR Hits is indicated **(B)** Hierarchical clustering of the SARS-CoV-2 proteins and human host proteins
824 basis of their mN2H interaction profiles. The heatmap represents the signal intensity derived from the relative
825 strength of positive interactions (*i.e.* distance to the threshold). **(C)** Intra-virus interactions heat map, intraviral
826 interactions were tested in triplicate using three different complementary NanoLuc configurations, the score
827 corresponding to the numbers of positive interactions found. **(D)** Network representation of intraviral interactions.
828 Thickness of the lines is relative to the distance from the threshold, the green proteins appear associated with
829 organelles that are in close proximity to each other in the 3D network.

830 **Fig 5. Organization and validation of the BioID and mN2H networks.** **(A)** Snapshot of the 3D network
831 depicting spatially resolved and colored clouds enriched in a subset of GO categories. Merged data from this
832 study and the literature are graphically coded as described in the legend. **(B)** Violin plot of the viral-host distances
833 in the 3D map according to the PPIs status, as described (Student t-test; *p*-values as indicated). **(C)** Modeling of
834 the USP13/NSP13 complex and validation of USP13 as a host target. Molecular docking of USP13/NSP13
835 interaction. **(D)** Schematic of infection-induced cell fusion assay using nanoluciferase complementation. **(E)**
836 Measured IC50 for Spautin-1 using the nanoluciferase complementation assay. Briefly, a co-culture of Vero E6-
837 NanoLg and Vero E6-NanoSm, plated at equal density the day before infection, was infected at a MOI of 0.01
838 with the Wuhan strain of SARS-CoV-2. Increasing concentrations of Spautin-1 (red) and GC376 (blue) were
839 added at the time of infection. Luciferase was measured 24 hours post-infection as a read-out of infection-
840 induced syncytia formation.

841 **Supplemental Figures legends**

842 **Fig.S1. Overview of the pipeline for identification of SARS-CoV-2/human interactome** **(A)** Pipeline of the
843 SARS-CoV-2-host interactome identification and potential mining.

844 **Fig.S2. Expression, localization and biotinylation capacities of BirA*Flag-tagged SARS-CoV-2 proteins.**
845 Confocal imaging BirA*Flag proteins expressed in 293 Flp-In TReX in basal culture. Staining of DAPI, biotin
846 (streptavidin- AlexaFluor488; proximal interactors) and anti-FLAG (anti-mouse AlexaFluor647; bait proteins).

847 **Fig.S3. Expression of individual BirA* Flag-tagged SARS-CoV-2 proteins.** N-ter and C-ter BirA* Flag-tagged
848 SARS-CoV-2 expressing cells were lysed with Lämmli buffer, and western Blot were realized with an equivalent
849 volume of whole cell extract. BirA* Flag-tagged proteins were revealed with anti-FLAG antibody (anti-mouse
850 HRP secondary antibody), and anti-GAPDH (anti-mouse HRP secondary antibody) was used for protein loading
851 control.

852 **Fig.S4. Effect of individual BirA* Flag-tagged SARS-CoV-2 proteins on IFN activation.** HEK293-ISRELuc
853 reporter cells were co-transfected with BirA*Flag alone or the indicated BirA*Flag SARS-CoV-2 proteins and **(A,**
854 **B)** MAVS expression plasmid **(C, D)** pcDNA3. IFN was measured as luciferase (RLU) Fold change over ISRELuc
855 activity without MAVS co-expression.

856 **Fig.S5. Metascape analysis of the enriched targeted categories per viral protein.** Metascape Express
857 Analysis report was used to automatically select the functional categories enriched for each viral. The detailed
858 output of the functional enrichment analysis, including the gene list assigned to each category for each bait
859 protein, is provided in Supp Table 4.

860 **Fig.S6. Metrics and features of the BioID-based network and 3D map.** Analysis of the effect of filtering out
861 host proteins with BioID degree greater than seven on the network topology: **(A)** Histogram of host protein degree
862 and dependence of the network's total number of nodes and edges. The majority of host proteins have one or
863 two viral interactions. After applying the degree cutoff, around 82% and 53% of nodes and edges respectively
864 are retained. **(B)** Histogram of viral protein degree before and after filtering for highly specific interactors. We
865 observe that the specificity ratio is generally higher for viral proteins with smaller total number of interactions;
866 Distance-based analysis of the network's 3D representation: **(C)** Host target density and total population
867 dependence with radial distance from dense core center for the full and filtered (host protein degree ≤ 7)
868 networks. Identification of dense core size via the radial location of the density interpolated line inflection point
869 for the case of the filtered network. Host proteins below the cutoff are considered to belong inside the dense
870 core. **(D)** Distance histograms, densities and averages (vertical dashed lines) for all viral-host protein pairs tested
871 in mN2H, including breakdown by mN2H +/- status, and subset of BioID+ interactions. mN2H+ positive pairs are
872 found at lower average distances, their distribution deviating significantly from the overall mN2H matrix
873 distribution and showing similarity to the BioID+ distribution. **(E)** Distance histograms, densities and averages

(vertical dashed lines) restricted to mN2H-tested BioID+ protein pairs including breakdown by mN2H+/- status. Despite having no influence on the construction of the 3D layout, mN2H+ pairs exhibit a lower average distance, reinforcing the predictive power of the map; **(F)** Distance histograms, densities and averages (dashed lines) restricted to mN2H-tested BioID- protein pairs and breakdown by mN2H+/- status. Despite having no influence on the construction of the 3D layout, N2H+ pairs exhibit a lower average distance, reinforcing the predictive power of the map.

Fig.S7. AlphaFold-Multimer predicted structures of SARS-CoV-2-host complexes. Cartoon representation of the SARS-CoV-2 /Human protein complexes with a pDockQ score >0.23. The viral proteins are shown in light violet, the host proteins in light pink. Protein surfaces are shown. When no structure prediction was available, the resulting unstructured representations obtained by AlphaFold were not shown for the corresponding proteins, owing to the lack of information they bring. The *.pdb files corresponding to each predicted complex are provided as hypertext links in **Supp table 11**.

Fig.S8. Benchmarking of the Nanoluciferase auto-complementation system for SARS-CoV-2 infection measurement. Vero E6-NanoLg and Vero E6-NanoSm, either cultured separately or co-cultured, equal density were infected with serial dilutions of the Wuhan strain of SARS-CoV-2. Luciferase was measured 24 hours post-infection as a read-out of infection-induced syncytia formation.

Supplemental tables

Supp. Table 1. List and sequences of SARS-CoV-2_{wuhan} ORFs used in this study. Each gene synthesizes cDNA encoding viral proteins from the early SARS-CoV-2 strain WuhanHu-1 (GenBank MN908947⁵⁰) where obtained from³⁴ as gateway pENTRY clones and transferred in the BioID or mN2H-dedicated plasmids

Supp. Table 2. SARS-CoV-2 BioID High data. Tab A. Full high-confidence proximal interactors. The number of baits detecting a given prey protein is indicated in the 'Indegree' column. The 'Abundance vs background' column represents the ratio between the detected prey in given bait assays against the control group (see Tab for RawQuant Perseus data). The 'poly(I:C) vs. basal' shows the log₂ fold LFQ intensity changes between the preys detected after poly(I:C) treatment enrichment and the corresponding basal condition. Over 1 is considered as either enrichment or gain (if absent from the basal condition), between 1 and -1 corresponds to unchanged, and below -1 shows decreased or lost hits. When no value is indicated, it corresponds to interactions not detected in the poly(I:C) and the corresponding basal condition. The reported interactions columns display the bait-prey pairs previously identified in the literature in other proximal interactomics or orthogonal physical PPIs detection studies (see Tab C-1 and C-2 respectively for details and references). For each cellular protein, the

904 number of CRISPR screens where they scored as necessary for SARS-CoV-2 infection is indicated. Tab B. Raw
905 output of the Perseus analysis of MaxQuant LFQ results. Tab D ToppCluster analysis for enriched/ gained
906 interactors. Tab E. ToppCluster analysis for enriched/ gained interactors after poly(I:C) treatment. Tab F.
907 Reported interactions between the viral proximal factors

908 **Supp. Table 3. Overlap of our SARS-CoV-2 BioID Datasets with reported SARS-CoV2 interactomes**

909 **Supp. Table 4. Functional annotation of high confidence proximal interactors for each bait viral protein**
910 . Functional annotations are generated using a compiled list of proximal interactors from N-ter, C-ter and poly(I:C)
911 conditions.

912 **Supp. Table 5. GO-term analysis of the BioID dataset compared with dataset combining all other**
913 **interactomes**

914 **Supp. Table 6. Poly:IC remodeling of SARS-CoV-2 proximal interactomes.** Fold change of proximal
915 interactions is given for each host factor per viral protein, and represented by volcano plots.

916 **Supp. Table 7. Factors of the dense core** (less than 0.3 of the centroid in the 3D map coordinates)

917 **Supp. Table 8. mN2H raw data and normalization.** Mean RLU of quadruplicates (Fold Change or FC matrices)
918 or duplicates to quadruplicates (CRISPR matrices) of mN2H PPIs are given along with the normalization pipeline
919 for mN2H positive signals scoring

920 **Supp. Table 9. Summary of the mN2H data.** The distance to the positive PPIs threshold is given for each virus-
921 host PPIs tested. PPIs detection in previous proximal interactomics and physical interactomics studies is given,
922 as well as the number of CRISPR Hits of the host factor.

923 **Supp. Table 10. Intra-virus mN2H raw data and normalization.**

924 **Supp. Table 11. Predictions of SARS-CoV-2-host complexes.** pDockQ scores (Tab A) and confidence scores
925 (Tab B) of mN2H positive virus-host PPIs obtained by AlphaFold-Multimer. pdb files of the corresponding
926 predicted virus-host complex structures are accessible through hypertext links.

927 **Supp. Table 12. Raw data of the IFN activation signaling pathway upon expression of SARS-CoV-2**
928 **proteins.**

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956 **Author contributions**

957 E.MN.L., P.S-T., A-C.G., E.C., Y.J. and C.D. obtained and cloned the SARS-CoV-2 polypeptides coding
958 sequences. E.MN.L. and P.S.T. produced BioID vectors and E.MN.L. conducted all BioID sample purification.
959 E.C. realized the fluorescence microscopy experiments and cell cultures for the BioID part. J-P.G. ensured all
960 mass spectrometry acquisition and contributed to the data analyses conducted by E.C. K.B., Y.R. and A.K.
961 realized the quality controls and luciferase activity assays. G.D., M.A., M.D.S. realized the mN2H experiments.
962 F.D., F.A., S.M., C.D. set up the novel luciferase-based infection assay, F.D. and C.D. performed the follow-up
963 experiments. Y.S., N.S. and P.A. performed the structural modeling analyses. N.P. and S.V. ensured the
964 bioinformatic statistical analyses. P.F-B., P.A., A-C.G., S.VD.W., Y.J., Y.S., C.D. and E.C. contributed to the
965 project development through the VirHostOme consortium assembled *ad hoc* by P.F.-B to aggregate international
966 interactomics research efforts. E.C., Y.J., C.D., Y.S. and E.MN.L conceived the project. C.D. and E.C. secured
967 funding. Y.S. conceived and coded the online 3D network interactive visualization interface, with inputs from
968 E.C., C.D. and Y.J. and support of G.A.P. E.C., C.D., Y.S., M.A., A.K. and G.D assembled the figures and wrote
969 the paper, with scientific and editorial inputs from Y.J., P.F-B. and E.MN.L.

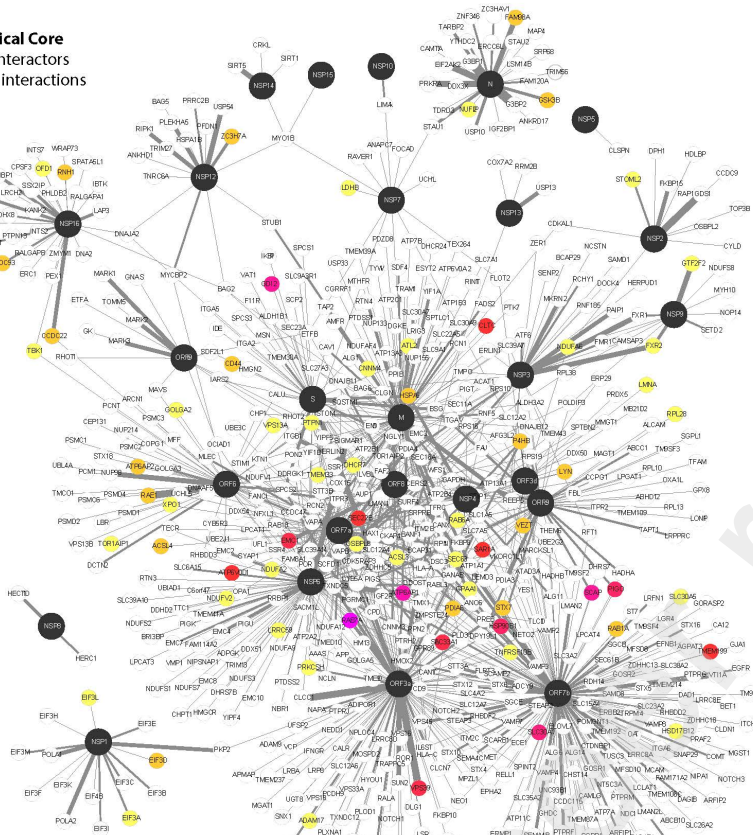
970 **Declaration of interests**

971 The authors declare no competing interests.

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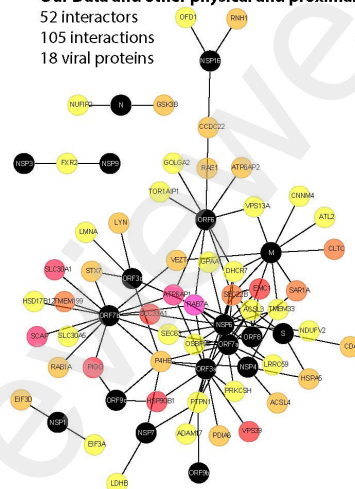
Fig.1

A Physical Core
659 interactors
1088 interactions

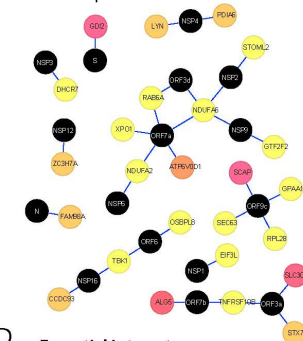


#CRISPR screens where essential
0-2 3 4 5 13

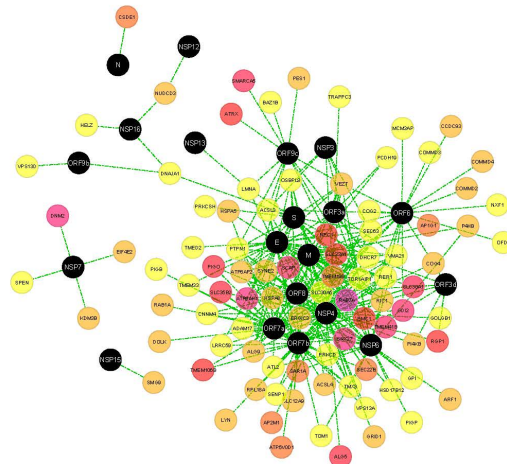
B Essential interactors
Our Data and other physical and proximal
52 interactors
105 interactions
18 viral proteins



C Essential interactors
Our Data and other physical
16 interactors
41 interactions
16 viral proteins



D Essential interactors
Our Data and other proximal
91 interactors
297 interactions
20 viral proteins



E Essential interactors
Our Data only
129 interactors
261 interactions
27 viral proteins

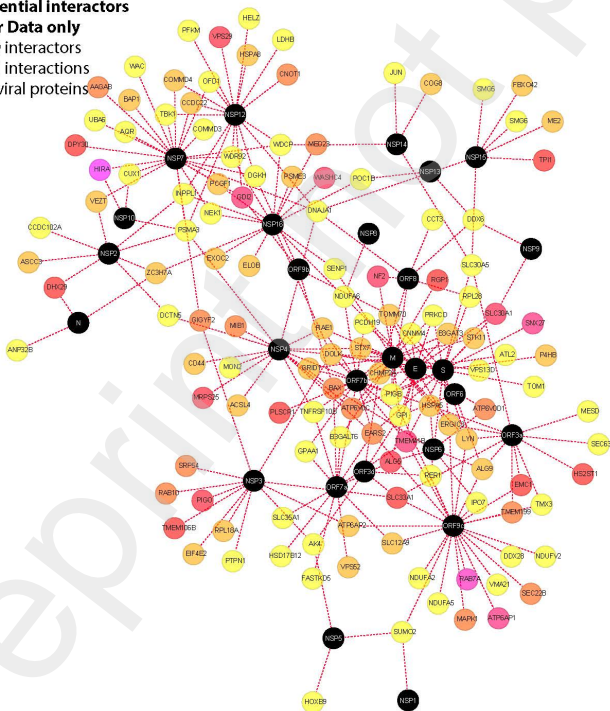


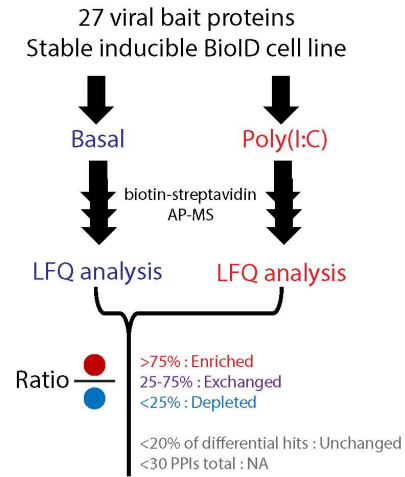
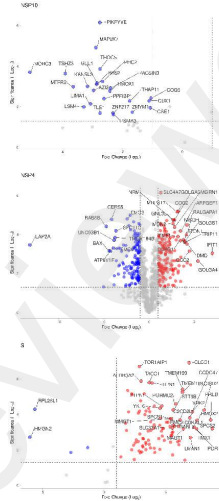
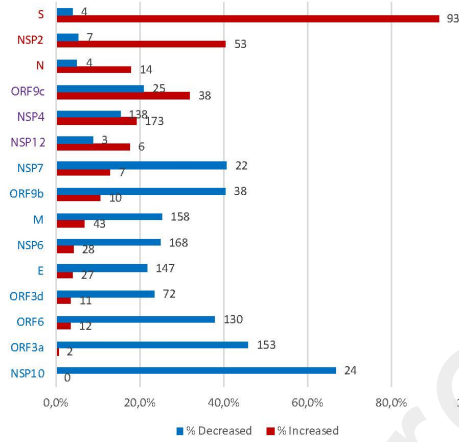
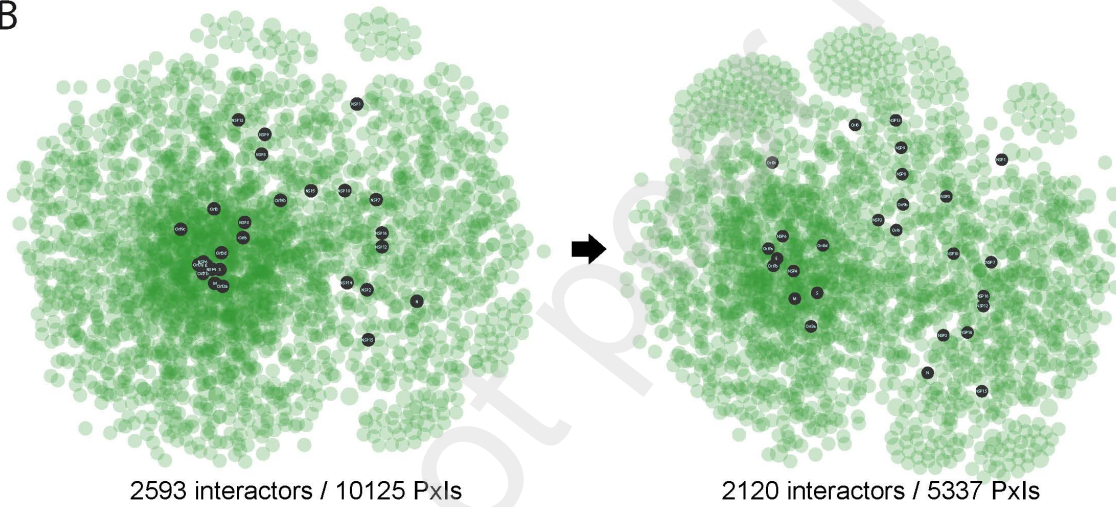
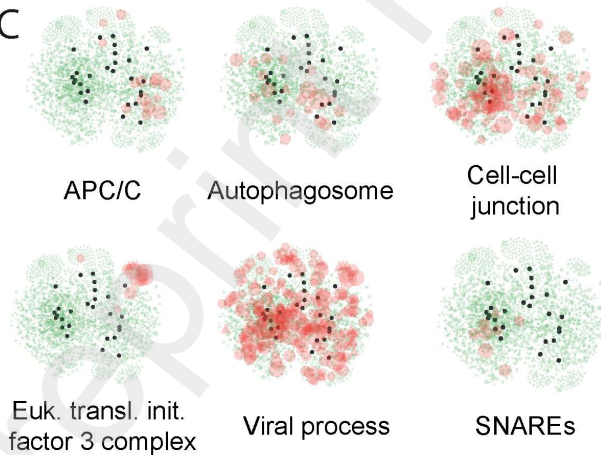
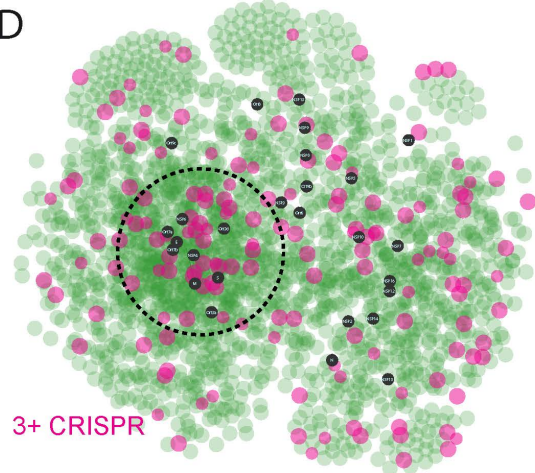
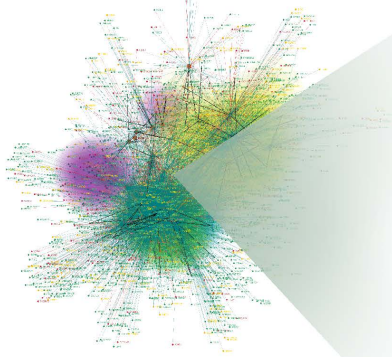
Fig.2**A****Poly(I:C) impact on proximal interactions****B****C****D**

Fig.3

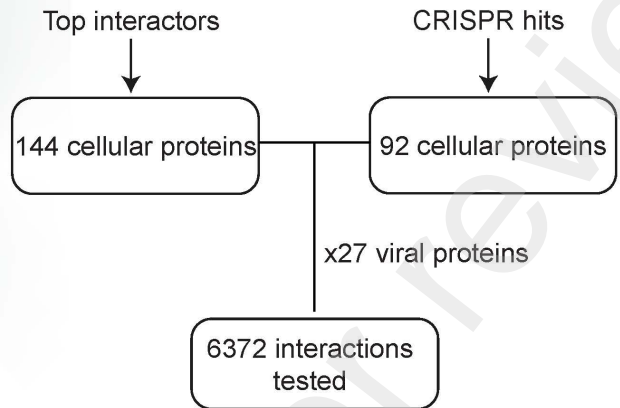
A

BioID high confidence dataset

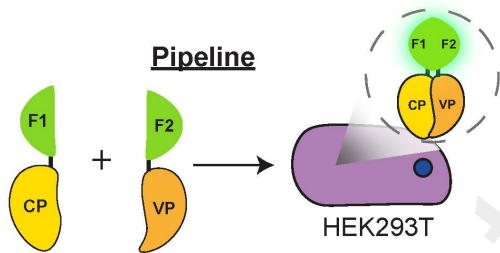


mN2H-based physical contact detection

Selection



Pipeline



Scoring

Total positive

6%

BioID positive

21%

B

mN2H-positive interactions
414 PPIs
25 viral proteins
118 host factors

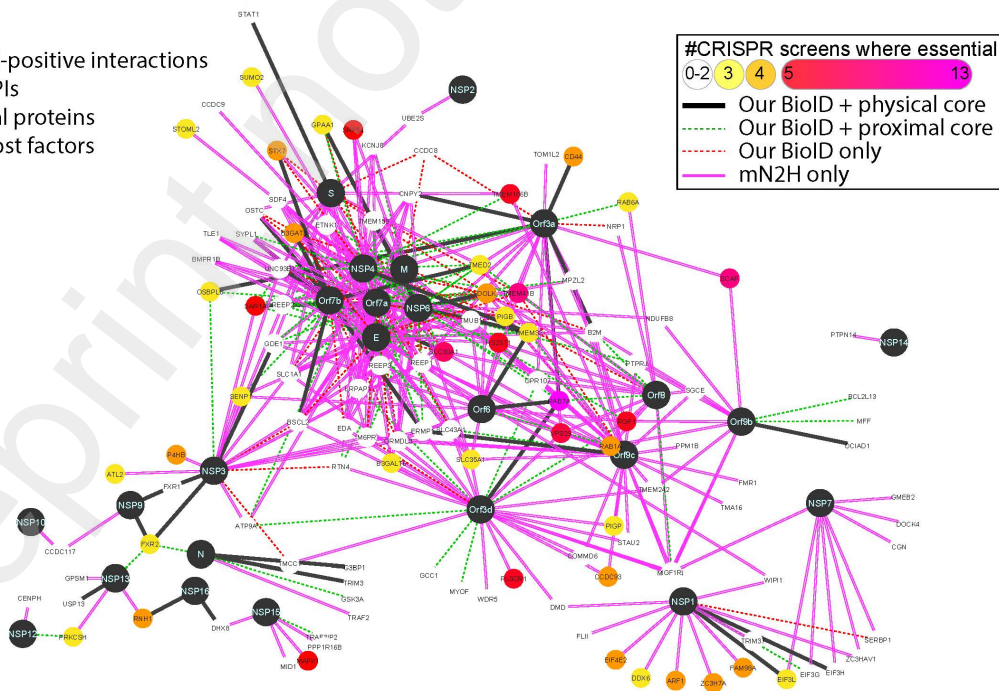


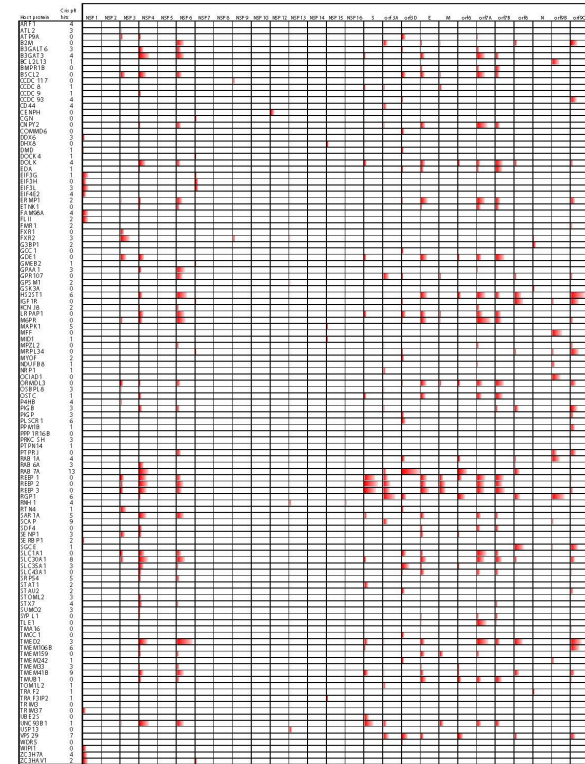
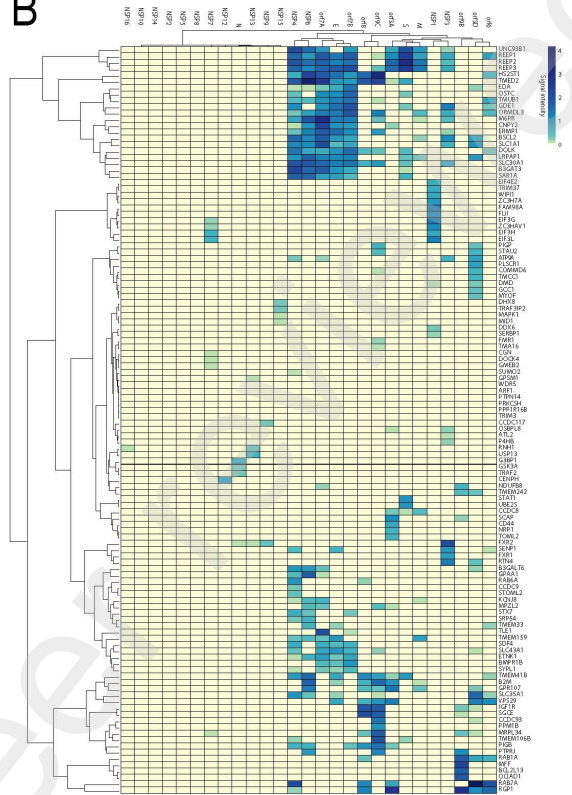
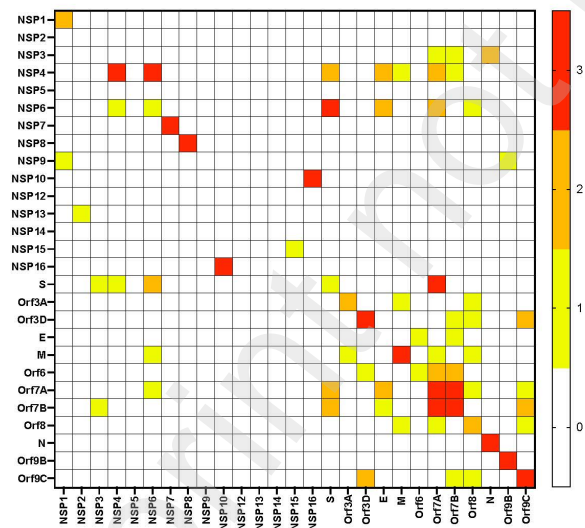
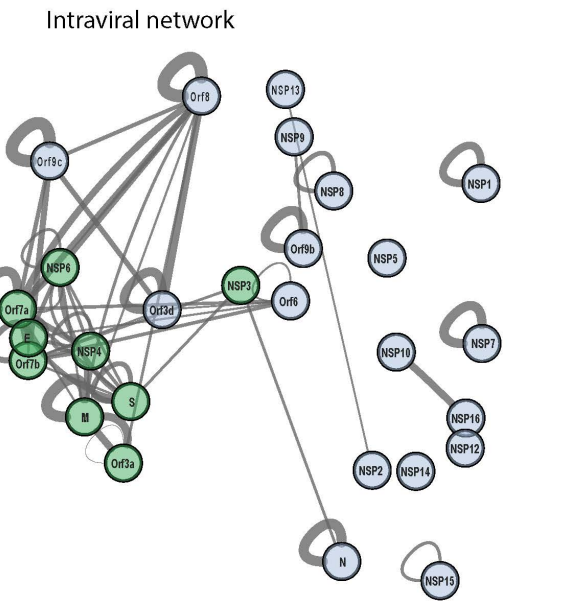
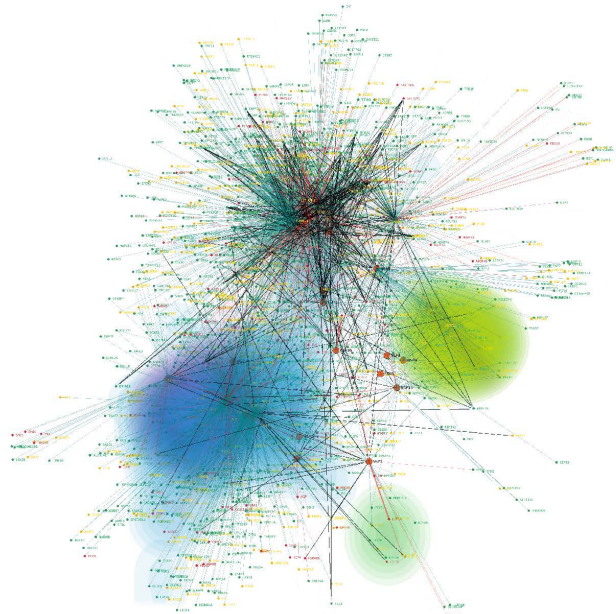
Fig.4**A****B****C****D**

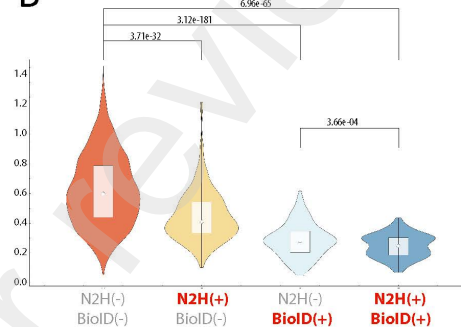
Fig.5

A

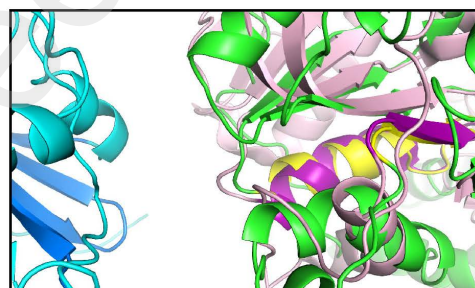
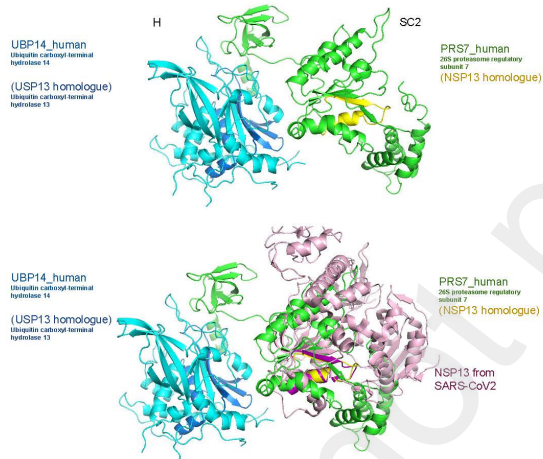


Symbol	Description	[Total %]
●	Baits	[27]
●	Targets, Novel	[1030 63%]
●	Targets, Weakly related to SARS-CoV-2	[481 29%]
●	Targets, Strongly related to SARS-CoV-2	[127 8%]
■	Region: G0:0970652 HAAIS complex	
■	Region: G0:0936064 ciliary basal body	
■	Region: 1268679 Eukaryotic Translation Initiation	
■	Region: G0:0006614 SR-dependent cotranslational protein targeting ...	
—	Poly(I:C) Increased	[406]
—	Poly(I:C) Unchanged / decreased / N/A	[2664]
—	N2H validated	[293]

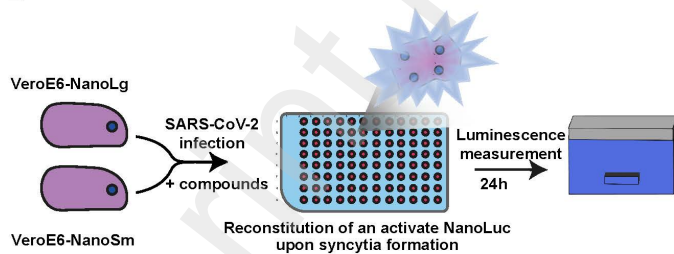
B



C



D



E

