

Longitudinal Associations between Prenatal Exposure to Phthalates and Steroid Hormones in Maternal Hair Samples from the SEPAGES Cohort

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1 2 3	Longitudinal associations between prenatal exposure to phthalates and steroid hormones in maternal hair samples from the SEPAGES cohort
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43 Abstract

44 We aimed to assess phthalate-hormone associations in the new-generation SEPAGES 45 cohort (2014-2017, France) using improved exposure and outcome assessments. 46 Metabolites from 7 phthalate compounds and the replacement di(isononyl)cyclohexane-47 1,2-dicarboxylate (DINCH) were measured in within-subject pools of repeated urine 48 samples collected at the second and third pregnancy trimesters (≈ 21 samples/trimester). 49 Metabolites from 5 steroid hormones were measured in maternal hair samples collected 50 at delivery reflecting on cumulative levels over the previous weeks-months. Adjusted 51 linear regression and bayesian weighted quantile sum (BWQS) mixture models were 52 performed. Each doubling in third trimester urinary MBzP concentrations was 53 associated with average increases of 13.3% (95%CI: 2.65, 24.9) for ∑cortisol, 10.0% (95%CI: 0.26, 20.7) for ∑cortisone, 17.3% (95%CI: 1.67, 35.4) for 11-54 55 dehydrocorticosterone and 16.2% (95% CI: 2.20, 32.1) for testosterone, together with a 56 suggestive 10.5% (95%CI: -1.57, 24.1) increase in progesterone levels. Each doubling 57 in second trimester urinary di-isononyl-phthalate (Σ DiNP) concentrations was inversely 58 associated with testosterone levels (-11.6%; 95%CI: -21.6, -0.31). For most hormones, a 59 non-significant trend towards a positive phthalate mixture effect was observed at the 60 third but not the second trimester. Our study showed that exposure to some phthalate 61 metabolites, especially MBzP, may affect adrenal and reproductive hormone levels 62 during pregnancy. 63

64 Keywords: endocrine disruption, phthalate, DINCH, DiNP, MBzP, cortisol,

65 testosterone, pregnancy

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Synopsis (30 words max.): This study implementing improved exposure and outcome
assessment methods showed that some phthalate metabolites can influence adrenal and
reproductive hormone levels during pregnancy.

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1. Introduction

72 Maternal steroid hormone regulation during pregnancy is critical to maintain a healthy pregnancy and achieve an optimal fetal development.^{1,2} For example, the 73 74 equilibrium between glucocorticoids (e.g., cortisol) and its precursor progesterone is 75 critical to maintain pregnancy and switch the maternal immune response towards fetal tolerance.^{3,4} Contrastingly, a disequilibrium in progesterone and/or glucocorticoids 76 77 predisposes to inflammation, which may lead to placental insufficiency and poor fetal growth.³ Regarding the hypothalamus-pituitary-adrenal (HPA) axis, corticotropin-78 79 releasing hormone (CRH) and cortisol play a major role in the timing of labor, and have been associated with prematurity and low birth weight in epidemiologic studies.^{2,5} In 80 81 relation to the hypothalamus-pituitary-gonadal (HPG) axis, women presenting higher 82 testosterone levels, such as cases of polycystic ovarian syndrome (PCOS) also show a higher risk of preterm birth, fetal growth restriction and pregnancy complications.⁶⁻⁹ 83 84 Overall, disruptions in steroid hormone levels during pregnancy may result in adverse pregnancy and birth outcomes.³ 85

86 Phthalates are a family of high-volume chemicals used in a wide range of 87 commercial products. However, some of them are considered endocrine disruptors and can interfere with hormonal regulation.¹⁰ High molecular weight phthalates (> 250 Da; 88 89 ester side-chain lengths of five or more carbons) are employed as plasticizers in the 90 production of polyvinyl chloride (PVC) plastics, and are found in a variety of products such as food contact materials,¹¹ building and construction materials (e.g., vinyl 91 92 flooring, floor tiles, wall coverings and furniture upholstery), medical devices (tubing, catheters, blood/dialysis bags), and toys.^{12,13} Low molecular weight phthalates (< 250 93 94 Da; ester side-chain lengths of one to four carbons) are used as solubilizing agents in the 95 formulation of cosmetics and personal care products (e.g., fragrances), and as coatings

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of some pharmaceuticals.¹³ Despite some regulations in Europe, recent birth cohorts and 96 97 biomonitoring studies show that phthalate exposure is still widespread with more than 90% of the European population showing detectable concentrations in urine.^{14,15} In 98 99 addition, newer phthalates such as bis(2-propylheptyl) phthalate (DPHP) and the non-100 phthalate substitute 1,2-Cyclohexane dicarboxylic acid diisononyl ester (DINCH) have recently entered the market, showing increasing concentrations in humans over time.^{14,16} 101 102 Since these new chemicals have a similar structure than those they replace and could potentially exert deleterious effects,^{17,18} there is a need for continuous biomonitoring 103 and surveillance.^{19,20} 104

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106 Phthalates and their metabolites have been shown to interfere with steroid 107 hormone regulation in laboratory animals through a variety of mechanisms that depend on age and sex.^{10,21} Through the construction of an adverse outcome pathway (AOP) 108 109 network, Baken et al. has shown that exposure to specific phthalates (diethyl-hexyl 110 phthalate (DEHP), monoethyl-hexyl phthalate (MEHP) and dibutyl-phthalate (DBP) 111 among others) could be related to hormonal alterations and reproductive problems in experimental animals.²¹ These effects depended on sex and exposure timing and can be 112 113 mediated through peroxisome-proliferator activated receptors and other pathways implicating the estrogenic (ER) and glucocorticoid (GR) receptors.²¹ In addition to the 114 115 ability of some phthalates to interact with the (HPG) axis, the adrenal gland has recently emerged as another target of phthalate toxicity, with DEHP and DBP being able to 116 influence adrenal gland histology and steroidogenesis in both male and female rats.^{22,23} 117 118 However, mechanistic data on the impact of phthalates on the HPA axis is still limited.²⁴ 119

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121 The human evidence is very limited, with some studies in pregnant women 122 finding associations between specific phthalate metabolites and either increased or decreased testosterone levels in maternal serum^{25,26} or urine,²⁷ and reduced serum 123 progesterone levels.^{25,28} One study showed increased cord blood cortisol levels related 124 125 to several phthalate metabolites among women carrying female fetuses, while reduced 126 cortisol levels were reported among those who carried male fetuses in association with DEHP metabolites.²⁹ Another study found that prenatal exposure to DEHP metabolites 127 128 was associated with reduced cord blood cortisol and cortisone levels and increased adrenal androgens.³⁰ These previous studies showed associations that varied by 129 130 exposure at different pregnancy trimesters and in some cases were sex-specific.

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132 Limitations of previous observational studies include the assessment of phthalate 133 metabolites in spot urine samples, which leads to exposure misclassification and 134 attenuation bias due to the short-half life and large within-day and within-week variability of phthalate metabolites.^{31,32} Another shortcoming is that most studies used a 135 unique or few $blood^{25,26,28-30}$ or urine²⁷ sample(s) to evaluate hormone levels, since 136 many of them are influenced by circadian rhythms and time-specific confounders.^{33,34} 137 138 Distinct collection protocols or biospecimens are needed to provide complementary 139 information about these hormone levels over an extended period of time (i.e., weeks or 140 months) while being less influenced by circadian rhythms and transient stressors (e.g., 141 allergies, colds, transient moods).

Our objective was to study the longitudinal associations between prenatal exposure to phthalates and maternal steroid hormones in a new-generation cohort. We relied on the prospective collection of multiple urine samples during pregnancy to improve exposure characterization.^{31,32} In addition, we collected hair samples at delivery that allowed to reliably measure cumulative levels of steroid hormones over the
previous weeks to months.^{35–37}

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149 **2. Methods**

150 2.1 Study population

We used data from the French mother-child cohort SEPAGES that recruited 484 pregnant women at eight obstetrical ultrasonography practices of the Grenoble metropolitan area from July 2014 to July 2017. Inclusion criteria were: singleton pregnancies up to gestational week 19, being older than 18 years, living in the metropolitan area of Grenoble and planning to deliver in one of the four Grenoble maternity clinics. The research protocol was presented to 3360 women, with an eligibility and participation rate of 69% and 21%, respectively.³⁸

The current study included mothers with adrenal and reproductive hormones assessed in hair samples collected at delivery and urinary phthalate metabolites measured at the second trimester (T2) (N = 382) and third (T3) trimester (N = 378) of pregnancy (Supporting Information Figure S1). The SEPAGES study was approved by the Ethics Committee (CPP) Sud-Est V and the National Commission on Informatics and Liberty (CNIL). After study staff explained all details to participants, pregnant women signed the written informed consent.

165 2.2 Repeated urine collection and exposure assessment

Pregnant women were asked to collect 3 urine samples per day (morning, midday and evening), for 7 consecutive days, twice during pregnancy: at the second trimester [median (P25, P75) gestational age: 17 weeks (16-18)], and during the third trimester [33 weeks (31-34)]. Women showed excellent compliance to the urine collection protocol: the median (P25, P75) number of biospecimens collected per 171 pregnant woman was 21 (20, 21) samples/trimester at both T2 and T3 collection weeks. 172 Women stored urine samples in their freezer until the SEPAGES field workers picked 173 them up at the end of each collection week for transport to a certified biobank (ISO 174 9001 standard, Grenoble University Hospital, bb-0033-00069). For each of the two 175 prenatal urine collection weeks, within-subject pools were conducted by combining 176 equal volumes of all spot urine samples obtained over a collection week (Supporting 177 Information Figure S2). This pooling strategy, that does not consider urinary dilution of 178 each spot sample, was previously validated as a good proxy of the urine concentrations 179 that would have been obtained in the pool of the whole volume of all individual spot samples collected.³⁹ We have indeed showed that, when an equal volume of each 180 181 individual urine sample is pooled, as was the case of the SEPAGES cohort, 182 standardization by specific gravity or creatinine is not needed, and can be even counterproductive for some compounds such as bisphenol A.³⁹ 183

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185 Aliquots of weekly pools were stored at -80 °C before being sent on dry ice 186 with a temperature sensor to the Norwegian Institute of Public Health (Oslo, Norway), a 187 certified European laboratory that has participated in recent inter-laboratory comparisons.⁴⁰ The concentration of 13 phthalate and 2 non-phthalate plasticizer 188 189 metabolites were measured (Supporting Information Table S1) using high performance liquid chromatography coupled to mass spectrometry.⁴¹ The limits of detection (LOD) 190 191 and quantification (LOQ) ranged from 0.07 to 0.4 ng/ml and 0.20-0.50 ng/ml, 192 respectively. Urine specific gravity was assessed in each weekly pool using a handheld 193 Atago PAL 10-S refractometer.

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197 Around delivery (1-3 days post-partum), at the maternity ward, mothers 198 collected their own maternal hair samples using material and protocol provided by 199 SEPAGES fieldworkers. Once collected, hair samples were stored at ambient 200 temperature by the maternity staff until it was transported to the biobank where they 201 were stored at -80°C. Hair samples from 408 SEPAGES mothers were sent to the 202 Applied Metabolomics Research Laboratory of the IMIM-Hospital del Mar Medical 203 Research Institute (Barcelona, Spain), where the first 3 cm (scalp side, corresponding to the hair growth of the third trimester^{42,43}) were used to assess ten steroid hormones (5 204 hormones and 5 metabolites, Table 2) using a previously validated liquid 205 chromatography-tandem mass spectrometry (LC-MS/MS) methodology.⁴⁴ 206 The hormones and metabolites assessed were: cortisol and its metabolites 20a-207 dihydrocortisol and 20\beta-dihydrocortisol; cortisone and its metabolites 20a-208 209 dihydrocortisone, 20β-dihydrocortisone and β-cortolone; 11-dehydrocorticosterone; 210 testosterone; and progesterone. The selection of these hormones was based on their reliable measurement in hair samples,⁴⁴ their relevance to pregnancy, and their 211 212 representativeness of both HPG and HPA hormonal axes. During the analytical 213 procedure, 2 samples broke during centrifugation and 10 had a sample weight below the 214 minimum required. Additionally, 12 samples did not pass analytical quality controls for 215 most hormones and were excluded from our analysis. Thus, data on steroid hormones 216 was available for 384 women. Figure S3 graphically depicts the relationships among 217 these hormones (Supporting Information).

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222 **2.4 Statistical Analysis**

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2.4.1 Phthalate biomarkers

224 Concentrations below the LOD and between the LOD-LOQ were imputed by 225 values randomly selected between 0 and LOD and between LOD and LOQ, 226 respectively, considering the underlying distribution of concentrations.^{45,46}

If needed (i.e., sample processing and analytical conditions affecting the 227 228 measured concentrations), phthalate metabolite concentrations were standardized using a two-step approach to correct for between-sample variations in the processing and 229 chemical analysis of urine samples.^{47,48} The following sampling conditions were 230 231 considered: time of urine sample transportation between the home of participants to the 232 biobank, the time that samples were maintained at 4 °C in the course of the pooling 233 process, and analytical batches. First, the associations between each natural log-234 transformed phthalate biomarker and the three above-mentioned conditions were 235 estimated running separate linear regressions adjusted for maternal age, education, pre-236 pregnancy body mass index (BMI), parity, date, season, pregnancy trimester of sample 237 collection and urinary specific gravity. Second, we used the measured phthalate 238 concentrations and the estimated effects of processing/assay conditions associated with 239 phthalate concentrations (p-value < 0.20) to predict standardized concentrations, that is, concentrations that would have been obtained if all samples had been processed under 240 241 the same conditions and assayed in the same batch. Details have been described in Guilbert et al., (2021) and Mortamais et al., (2012).^{47,48} Since high correlations 242 243 (Spearman rho >0.80) between these standardized and non-standardized phthalate concentrations have been previously reported,⁴⁹ the current work provides descriptive 244 245 statistics only for standardized concentrations, which were used in all phthalatehormone analyses. In the case of phthalates metabolites belonging to a same parentcompound, molar sums (μ mol/L) were computed (i.e., Σ DEHP, sum of diisononyl phthalate metabolites (Σ DiNP) and Σ DINCH). Urinary phthalate metabolite concentrations were natural log-transformed to reduce the skewness of distributions and the influence of extreme values.

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2.4.2 Hormone biomarkers

Some steroids presented a moderate percentage of non-detected values (range between 0 and 26%). As for exposure biomarkers, non-detects were imputed by values randomly selected between 0 and the LOD, considering the estimated distribution of the quantified hormone concentrations.^{45,46} Such method results in a balanced distribution of imputed values, providing unbiased or minimally biased parameter estimates when 30% or less of data are below detection limits.⁴⁶

258 To reduce the number of statistical comparisons, we computed the molar sum of 259 cortisol and cortisone and their metabolites by summing their concentrations divided by 260 their molecular weight. Five outcomes were thus included in the main analysis: 261 Σ cortisol, Σ cortisone, 11-dehydrocorticosterone, testosterone and progesterone. In 262 addition, two hormones ratios were computed: the Σ cortisol to Σ cortisone ratio as an indicator of corticoid activity that may be predictive of hypertensive disorders;⁵⁰ and the 263 264 testosterone to Σ cortisol ratio as an indicator of the general (im)balance between the mutually inhibiting HPG and HPA axes that has been related to antisocial and risk-265 taking behaviors.⁵¹ The two ratios were considered as secondary outcomes to be 266 267 interpreted alongside with the primary outcomes. All hormones and ratios were naturallog transformed to reduce the skewness of distributions.⁵¹ 268

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2.4.3 Covariates and confounding factors

271 Data on potential covariates were self-reported using questionnaires 272 administered by a fieldworker during study visits and clinical examinations. 273 Confounding factors and outcome predictors were selected a priori based on causal knowledge using a Directed Acyclic Graph (DAG)⁵² (Supporting Information Figure 274 275 S4). All models were adjusted for: maternal age at conception (continuous), pre-276 pregnancy body mass index (BMI, underweight/normal weight vs. overweight/obese 277 mothers), education level (below vs. Master's degree or above), active smoking during 278 pregnancy (smoking at any trimester vs. non-smoker), passive tobacco exposure 279 (exposed to indoor tobacco smoke ≥ 1 hour/week at any trimester vs. non-exposed), parity (nulliparous vs. uniparous/multiparous), maternal anxiety/depression score at 280 281 third trimester using the French version of the Hospital Anxiety and Depression scale (continuous),⁵³ infant sex and season of hair collection (winter vs. remaining seasons 282 283 since hair samples collected in winter showed significantly lower corticosteroid levels 284 compared to other seasons).

We additionally controlled for technical variables such as hair analysis batch (dichotomous) and time elapsed (continuous) between hair sampling and hormone measurement. The slight number of missing values for some covariates (see Table 1 footnote) was singly imputed with the variable median or mode value.

289 2.4.4 Associations between phthalates and hormones

The linearity of the relationship between phthalate metabolites and hormones was tested using generalized additive models (GAMs). Most associations showed effective degrees of freedom below $2,^{54,55}$ supporting linear relationships and so, the modelling of phthalates as continuous variables (Supporting Information Figure S5). Additionally, the few possible non-linear associations identified were not statistically significant (GAM p-value > 0.05). Adjusted linear regression models were then performed for each phthalatehormone pair at both the second and third trimester of pregnancy, since previous studies showed associations may differ by trimester.^{27–29} Regression estimates were expressed as percent change (PC) for each 2-fold increase in standardized urinary phthalate biomarkers using the following formula: $([e^{(\ln 2 \times \beta)} - 1] \times 100)$.

A phthalate-sex interaction term was added to the model to identify potential sex-specific associations, since previous studies also showed effect modification by this variable.^{27,29} A p-value below 0.10 for the interaction term was considered as evidence of effect modification,⁵⁶ in which case a sex-stratification of the specific association was conducted.

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2.4.5 Mixture model

Given the possibility of cumulative phthalate effects,⁵⁷ the overall mixture effect 307 308 of prenatal exposure to phthalates and DINCH on maternal hair hormone levels was 309 estimated using adjusted Bayesian Weighted Quantile Sum (BWQS) regression. BWQS 310 is a quantile-based approach that combines multiple independent variables additively 311 into a weighted index, with weights estimating the contribution of each component to the mixture.⁵⁸ This model represents a novel Bayesian extension of the classic WQS 312 313 regression, counteracting its main limitations including higher stability of estimates, 314 higher statistical power (no data splitting into training and validation sets), and being 315 able to estimate mixture associations considering multiple chemicals with individual positive, negative, and/or null effects simultaneously.⁵⁹ Phthalate and DINCH 316 317 metabolite concentrations were modeled in quartiles. Similar to linear regressions, 318 BWQS effect estimates (Beta1 and 95% Credible Intervals) were expressed as the PC in 319 hormone levels for each quartile increase in the mixture: [(exp(Beta1) - 1) * 100].

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321 2.4.6 Sensitivity Analyses

The robustness of the main models was tested after: 1) additional adjustment for urine specific gravity (continuous), to rule out any potential influence of urine dilution;⁶⁰ 2) exclusion of extreme exposure and hormone values corresponding to percentiles 1 and 99 of their respective distributions.

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2.4.7 Interpretation and software

328 Statistical significance was set as p-value <0.05. A p-value between 0.05 and 329 0.10 was considered as possibly suggestive of an association. However, results were 330 interpreted not solely depending on statistical significance, but considering patterns of associations and the previous evidence available to contextualize the findings.⁶¹ Thus, 331 332 associations with p-value <0.10 will only be considered meaningful if there is enough 333 previous toxicological or epidemiological support or if it follows a pattern of 334 associations (i.e., a non-isolated association). Regression analyses were carried out with 335 Stata version 14.2 (Stata Corp), while GAM ("mgcv" package) and BWQS models using RStudio version 4.0.3 (RStudio Team, 2020). 336

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338 3. Results

339 3.1 Characteristics of the study population

Mothers showed a high education level (58% reached Master's degree or higher), 16% were overweight/obese ($\geq 25 \text{ Kg/m}^2$), and conceived at a mean (SD) age of 32.6 (3.9) years (Table 1). About half of the mothers (44%) were nulliparous at the time of conception, and there was a slightly higher number of boys (54%) born in the 344 cohort. The prevalence of maternal active smoking was low (6%) compared to data in
345 France,³⁸ while 17% of the women were exposed to environmental tobacco smoke.

346 Phthalates and DINCH metabolites were quantified in more than 95% of the 347 urine samples at both pregnancy trimesters. The highest median urinary concentrations 348 were observed for the sum of DEHP metabolites and the low molecular weight 349 metabolite MEP (Supporting Information Table S2). Phthalate metabolites showed low-350 to-moderate estimates of Spearman correlation (rho range: 0.27-0.59) between the two 351 pregnancy trimesters (Supporting Information Figure S6). In general, phthalate exposure levels were lower or similar to other contemporary European cohorts.^{14,62} 352 353 Extended description of phthalate exposure levels in the SEPAGES cohort has been 354 previously reported.¹⁴

355 Regarding hormones measured in hair samples, progesterone showed the highest 356 levels (median: 76.5 ng/g), followed by Σ cortisone (median: 32.9 ng/g) and Σ cortisol (median: 6.1 ng/g) (Table 2). Σ Cortisol and Σ cortisone were highly correlated with 357 358 their respective metabolites (rho > 0.84), with the exception of \sum cortisone and β -359 cortolone that were moderately correlated (rho: 0.43). Σ Cortisol and Σ cortisone levels 360 were also highly correlated between them (rho: 0.86). While \sum cortisol and \sum cortisone 361 moderately correlated with 11-Dehydrocorticosterone (rho: 0.56 and 0.67, respectively), 362 their correlations with testosterone and progesterone levels tended to be lower 363 (Supporting Information Figure S7).

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Table 1. Characteristics of the study population (N=382).

	Included (N=382)	Excluded (N=102)	D l a	
Characteristics	N (%) or Mean (SD)	N (%) or Mean (SD)	P-value ^a	
Mother's age at conception of index pregnancy (years)	32.6 (3.9)	32.2 (3.8)	0.41	
Mother's anxiety/depression symptoms at 3 rd	10.4 (4.8)	11.0 (4.6)	0.29	

trimester ^b			
Mother's pre-pregnancy BMI (kg/m ²)	22.3 (3.6)	22.8 (4.4)	0.09
< 25	320 (83.8%)	78 (76.5%)	
≥25	62 (16.2%)	24 (23.5%)	
Mother's education			0.40
Less than Master's degree	162 (42.4%)	48 (47.1%)	
Master's degree or higher	220 (57.6%)	54 (52.9%)	
Mother's tobacco consumption during pregnancy (=consumers ≥1 cigarette/day)	23 (6.0%)	6 (5.9%)	0.96
Mother's passive tobacco exposure (= exposed)	64 (16.8%)	24 (23.5%)	0.12
Parity			0.07
Nulliparous	167 (43.7%)	55 (53.9%)	
Uniparous/Multiparous	215 (56.3%)	47 (46.1%)	
Season of hair collection			0.09
Winter	75 (19.6%)	28 (27.5%)	
Remaining seasons	307 (80.4%)	74 (72.5%)	
Child sex (= boys)	206 (53.9%)	52 (53.1%)	0.88
Time elapsed from sample collection to assessment (years)	4.3 (0.78)	-	-
Analytical batch of hair analysis		-	-
Batch 1	288 (75.4%)	-	-
Batch 2	94 (24.6%)	-	-

366 Abbreviations: BMI (Body Mass Index); N (Number); SD (Standard deviation).

367 Note: Percentage of missing covariates for included participants: maternal anxiety/depression score
 368 (5.0%), education (0.5%), pre-pregnancy BMI (0.5%), active (8.1%) and passive (5.5%) smoking. The
 369 remaining covariates had no missing values.

^a Categorical variables compared using the two-sample proportion test and continuous variables
 compared using the two-sample t test.

^b Hospital Anxiety and Depression score rated on a scale of 21. Anxiety and depression symptoms were
 summed, so the scores should be interpreted on a scale of 42.

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Table 2. Hormone levels in the study population (N=382).

Hormon	LOD	% >LOD	P25	Median	P75	
\sum Cortisol (nn	\sum Cortisol (nmoles/g) ^a		-	0.009	0.017	0.035
	Cortisol (ng/g)	0.06	98	0.73	1.71	4.06
	20α-dihydrocortisol (ng/g)	0.07	85	0.23	0.63	1.48
	20β-dihydrocortisol (ng/g)	0.01	98	1.85	3.69	6.71
\sum Cortisone (r	\sum Cortisone (nmoles/g) ^a		-	0.050	0.091	0.156
	Cortisone (ng/g)	0.10	95	4.77	12.0	22.5
	20α-dihydrocortisone (ng/g)	0.02	99	3.80	8.27	15.7
	20β-dihydrocortisone (ng/g)	0.02	97	2.60	5.74	9.82
	β -cortolone (ng/g)	0.50	74	<lod< td=""><td>4.54</td><td>7.66</td></lod<>	4.54	7.66
11-Dehydrocorticosterone (ng/g)		0.10	88	0.91	1.82	2.98
Testosterone	Testosterone (ng/g)		89	0.23	0.42	0.65
Progesterone (ng/g)		0.10	100	30.8	76.5	166.6

^a Molar sum of cortisol and cortisone metabolites. Median ∑Cortisol and ∑Cortisone levels expressed as
 ng/g were approximately 6.1 and 32.9 ng/g, respectively.

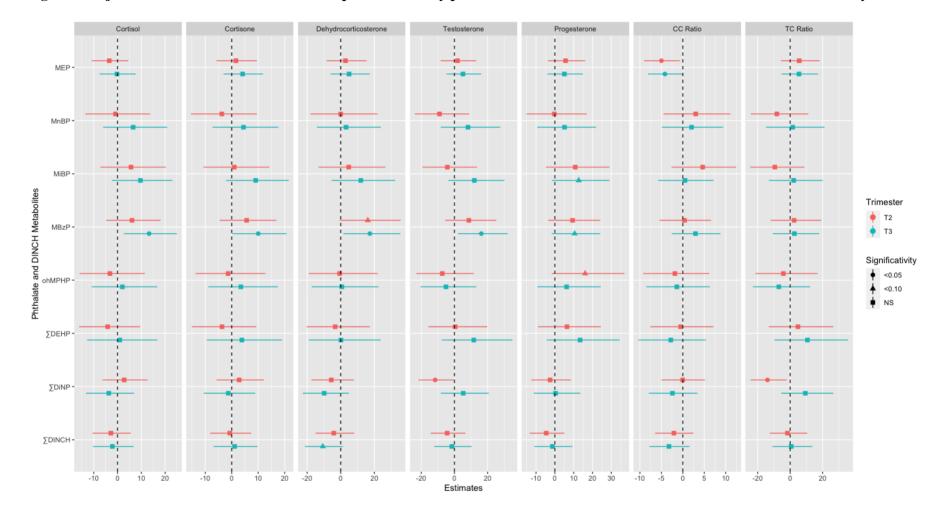
- 380
- 381 3.2 Associations in the whole population

382 MBzP showed the most consistent pattern of associations with hair hormones 383 (Figure 1 and Supporting Information Table S3). Second trimester exposure to MBzP 384 tended to be associated with higher levels of all hormones, but only reaching borderline-385 statistical significance for 11-dehydrocorticosterone levels (PC= 16.1%; 95%CI: -1.60, 386 35.6). Third trimester MBzP exposure was robustly and positively associated with most 387 of the hormones investigated. Each doubling in third trimester urinary MBzP 388 concentrations was associated with average increases of 13.3% (95%CI: 2.65, 24.9) for 389 ∑cortisol, 10.0% (95%CI: 0.26, 20.7) for ∑cortisone, 17.3% (95%CI: 1.67, 35.4) for 390 11-dehydrocorticosterone and 16.2% (95%CI: 2.20, 32.1) for testosterone, together with 391 a suggestive 10.5% (95%CI: -1.57, 24.1) increase in progesterone levels (Figure 1 and 392 Table S3). No associations with hormone ratios were found for MBzP at either the 393 second or third trimester.

Although without reaching statistical significance, third trimester urinary MiBP concentrations tended to be positively associated with most of the hormones assessed in hair samples (Figure 1 and Table S3), with a suggestive association (i.e., p-value < 0.10) being observed for progesterone (PC= 12.7%; 95%CI: -1.51, 28.9).

Second trimester urinary \sum DiNP concentrations were inversely associated with testosterone levels (PC= -11.6%; 95%CI: -21.6, -0.31) and the TC ratio (PC= -14.0%; 95%CI: -24.4, -2.15). Third trimester \sum DiNP exposure did not follow the same pattern nor was associated with any hormone outcome.

402 Although maternal MEP exposure was associated with a lower CC ratio at both 403 second (PC= -4.98%; 95%CI: -9.05, -0.72) and third (PC= -4.18%; 95%CI: -8.12, - 404 0.07) trimesters, this association appeared isolated, since MEP was not associated with
405 any of the primary hormone outcomes examined. Additionally, no meaningful
406 associations were observed for MnBP and ∑DEHP at any pregnancy trimester (Figure 1
407 and Supporting Information Table S3).



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Note: Geometric figures (circles, triangles and squares) and error bars represent linear regression effect estimates with their corresponding 95% confidence intervals expressed as percent change (PC) in hormone levels for each doubling in exposure biomarkers. Models adjusted for: maternal age at conception (continuous), pre-pregnancy body mass index (BMI, normal weight vs. overweight/obese), education level (below vs. Master's degree or above), active smoking during pregnancy (smoker vs. nonsmoker), passive tobacco exposure (exposed vs. non-exposed), parity (nulliparous vs. uniparous/multiparous), maternal anxiety/depression score at third trimester (continuous), infant sex and season of hair collection (winter vs. remaining seasons), hair analysis batch (dichotomous) and time elapsed (continuous) between hair sampling and hormone measurement. Abbreviations: 11-DHC (11-dehydrocorticosterone); CC (Cortisol to Cortisone ratio); TC (Testosterone to Cortisol ratio).

418 3.3 Sex-specific associations

419 Few sex-specific associations were observed (Supporting Information Table S4). 420 Sex-stratified models showed that the previously observed second trimester inverse 421 \sum DiNP-testosterone association was driven by boys (PC= -21.9%; 95%CI: -34.3, -422 7.15), while a null association was observed in girls (PC= 0.28%; 95%CI: -15.7, 19.3).

423 A sex-specific pattern was identified for MnBP, a compound not associated with 424 any hormone when boys and girls were studied together. In women carrying male 425 fetuses, second trimester MnBP exposure was positively associated with 11-426 dehydrocorticosterone (PC= 40.4%; 95%CI: 3.53, 91.9), together with non-significant 427 increases in Σ cortisol (PC= 14.9%; 95%CI: -7.30, 41.4) and Σ cortisone (PC= 8.67%; 428 95%CI: -9.88, 32.0) levels, while women carrying female fetuses showed opposite 429 trends including significantly lower 11-dehydrocorticosterone levels (PC= -22.6%; 430 95%CI: -40.5, 0.00) (Table S4).

431

432 3.4 Mixture associations

433 BWQS models showed that the phthalate mixtures measured at the second and 434 third trimesters of pregnancy were not significantly associated with any of the hormones 435 investigated (Table 3). Notwithstanding, several suggestive positive associations were 436 observed for the third trimester mixture and testosterone (PC= 17.0%; 95%CI: -3.63, 437 41.6), Σ cortisone (PC= 11.0%; 95%CI: -4.28, 27.3) and progesterone (PC= 10.3%; 438 95%CI: -7.36, 30.2) (Table 3). Although in general phthalates showed similar mixture 439 weights, MBzP and MiBP tended to show slightly higher weights, in line with single-440 pollutant models (Supporting Information Figure S8). The phthalate mixture at the third 441 trimester also tended to be associated with a lower CC ratio, which was mostly driven

- 442 by MEP in accordance with single-pollutant models (Supporting Information Figure
- 443 S8).
- 444

445 Table 3. Mixture associations using Bayesian Weighted Quantile Sum (BWQS)

446 regression models.

Hormones	Second trimester	Third trimester
normones	PC (95% CrI)	PC (95% CrI)
∑Cortisol	-2.33 (-18.3, 16.2)	5.74 (-9.89, 23.5)
∑Cortisone	0.06 (-14.0, 16.7)	11.0 (-4.28, 27.3)
11-Dehydrocorticosterone	-0.39 (-22.2, 27.5)	5.55 (-17.2, 32.9)
Testosterone	-4.50 (-22.3, 17.5)	17.0 (-3.63, 41.6)
Progesterone	4.64 (-12.9, 26.9)	10.3 (-7.36, 30.2)
CC Ratio	-3.22 (-13.4, 7.68)	-5.50 (-13.6, 3.44)
TC Ratio	-3.15 (-23.5, 23.3)	10.3 (-10.3, 35.9)

447

448 Note: Percent change (PC) in hormone levels per quartile increase of the phthalate mixture, with its 449 related 95% Credible Interval (95% CrI). The weight contributed by each chemical compound to the 450 most relevant mixture associations can be consulted in Figure S8. Mixture models were adjusted for the 451 same set of covariates included in single-pollutant models.

452

453 3.5 Sensitivity Analyses

The additional inclusion of urine specific gravity as a covariate in the models did not materially change associations (Supporting Information Table S5).

455 not materially change associations (Supporting information Table S5).

456 The exclusion of extreme values attenuated the isolated associations observed 457 between MEP and the CC ratio that were no longer significant (Supporting Information

458 Figure S9). Associations observed at the third trimester for MBzP did not materially

459 change, while associations for third trimester MiBP were notably strengthened, showing

- 460 significant associations with \sum cortisol (PC= 13.6%; 95%CI: 0.92, 27.95), \sum cortisone
- 461 (PC= 15%; 95%CI: 2.71, 28.65), 11-dehydrocorticosterone (PC= 26.1%; 95%CI: 6.34,
- 462 49.42), testosterone (PC= 16.9%; 95%CI: 0.45, 36.12) and progesterone (PC= 15.9%;
- 463 95%CI: 0.81, 33.24) levels (Supporting Information Table S6 and Figure S9).

464

465

466 **4. Discussion**

467 This cohort is the first to rely on within-subject pools of multiple urine samples 468 to assess phthalate exposure during gestation and to examine its association with 469 maternal hair samples collected at delivery to assess cumulative hormone levels. At the 470 second trimester, Σ DiNP concentrations were associated with lower testosterone levels 471 in all the sample, and especially among women carrying male fetuses. At the third 472 trimester, urinary MBzP concentrations were robustly and consistently associated with 473 higher levels of all the adrenal and reproductive hormones investigated. Third trimester 474 MiBP exposure followed a similar pattern of positive associations with all hormones, 475 that reached statistical significance after the exclusion of extreme values.

476

In line with single-pollutant results, a non-significant trend towards a positive association with most hormones was observed for the phthalate mixture at the third - but not second – trimester. Nevertheless, the lack of a significant mixture association seems to imply that, in this study population, specific phthalate metabolites (Σ DiNP, MBzP and to some extent MiBP) rather than a "cocktail effect" are the drivers of changes in specific hormones.

483

484

4.1 Biological plausibility and literature comparison

485 MBzP and other phthalate monoesters exert a dose-dependent increase in the 486 gene expression of peroxisome proliferator-activated receptor gamma (PPAR γ) in 487 mouse Granulosa cells at environmentally relevant concentrations,⁶³ which may reduce 488 aromatase expression, leading to increased testosterone levels by reducing its

conversion to estradiol.²¹ Although this hypothesis could explain the increased 489 490 testosterone levels reported in our study, it does not explain the MBzP-related 491 systematic increase observed in all adrenal and reproductive hormones. Our MBzP 492 findings could result from one of the following modes of action: 1) A central 493 simultaneous increase in both hypothalamic gonadotropin and corticotropin releasing 494 hormones or their pituitary downstream hormones (for which we did not find supporting 495 evidence); 2) A peripheral increase in the rate-limiting step of steroidogenesis, that is, 496 an increased cholesterol delivery to the inner mitochondrial membrane by Steroidogenic 497 Acute Regulatory Proteins (StAR), or upregulation of CYP450scc (cholesterol side-498 chain cleavage enzyme) that converts cholesterol into pregnenolone, the common precursor of all steroid hormones (Supporting Information Figure S3).⁶⁴ We found in 499 vitro and epidemiological evidence supporting that MBzP can upregulate P450scc in 500 placental cells and tissue at human-relevant concentrations,^{65,66} while no relevant data 501 502 was found in relation to StAR. Although to our best knowledge this MBzP-upregulation 503 of P450scc has not been studied in other steroid organs with high P450scc expression such as the adrenal glands and ovary,⁶⁷ a molecular docking study supported the ability 504 of MBzP to interact with P450scc.⁶⁸ Therefore, a MBzP-driven increase in 505 506 steroidogenic hormones through a multi-organ (placenta, adrenal glands and/or ovary) 507 upregulation of P450scc appears as a biologically plausible explanation that could be 508 further investigated in future experimental studies.

Although the current work is difficult to compare with previous studies due to differences in matrices for hormone assessment (use of blood^{25,26,29} or urine²⁷ instead of hair), our MBzP results are coherent with Pacyga et al., who reported a trend towards positive associations between maternal prenatal urinary MBzP concentrations and the sum of urinary testosterone metabolites measured three times during pregnancy among

434 U.S. women.²⁷ Other studies reported non-significant associations with serum 514 515 testosterone and progesterone, although the trend in estimates was again toward higher 516 hormone levels.^{25,26} Regarding corticosteroids, a positive association was reported 517 between first trimester urinary MBzP concentrations and cord blood cortisol to cortisone ratio measured at birth among 553 Chinese mother-newborn pairs.²⁹ Despite 518 519 the small number of studies, the literature appears supportive of possible associations between prenatal MBzP exposure and increased adrenal and reproductive hormone 520 521 levels during pregnancy.

MiBP showed a pattern of increased steroidogenesis similar to MBzP, that was notably strengthened after exclusion of extreme exposure and hormone values. Compared to MBzP, both the epidemiological^{25–27,29} and toxicological evidence for hormonal effects during pregnancy is more limited and equivocal.⁶⁹ Although MiBP has also been shown to interact with the P450scc enzyme, the specific study tested very high doses in adult male mice,⁷⁰ being not comparable to our work. Additional data will help to confirm or rule out our MiBP results.

529 DiNP is a phthalate replacement for DEHP. Although it is a less potent antiandrogenic chemical than DEHP,⁷¹⁻⁷⁴ DiNP still shows endocrine disrupting abilities. In 530 531 adult female mice, DiNP exposure altered steroid hormone levels and ovarian 532 folliculogenesis at moderate doses, with DiNP decreasing serum testosterone levels at a low to moderate dose of 100 µg/kg/day.^{75,76} In the current work we also observed 533 534 decreased testosterone concentrations in response to higher DiNP exposure, especially 535 among women carrying male fetuses. Since the human male fetus is able to synthesize its own testicular testosterone between the first and second trimester,⁷⁷ we hypothesize 536 537 this association could reflect an anti-androgenic effect that may not only affect the 538 mother but also the developing male fetus. Although biologically plausible, this

association needs to be confirmed in future studies given the limited toxicologic andepidemiologic data available for this phthalate replacement.

541 MnBP is known to exert anti-androgenic actions in male^{21,78,79} and female 542 reproductive organs.^{78,80–82} However, interspecies differences in glucocorticoids⁸³ 543 precluded us from comparing our MnBP results to the rodent studies available.^{23,84} 544 Since comparison with other epidemiological studies is neither possible, the MnBP-545 related increase in 11-dehydrocorticosterone levels among women carrying male fetuses 546 is difficult to interpret and needs to be confirmed in future studies.

Although MEP was negatively associated with the CC ratio at both second and third trimesters, which could indicate a higher conversion of cortisol to its less active metabolite cortisone, no associations were observed between MEP and cortisol or cortisone individually. The lack of toxicological support, together with the fact that when outliers were excluded the MEP-CC ratio associations disappeared, diminished our confidence in this potential association.

553

554

4.2 Hair vs. other matrices for hormone assessment

555 Compared to saliva, urine and blood, cortisol in hair is thought to reflect mid to 556 long-term cumulative levels which are less affected by circadian rhythms and timevarying stressors.⁴³ Although there are still some doubts about whether hair cortisol 557 represents long or mid-term cumulative concentrations,^{85,86} hair seems to provide 558 559 information at least on the previous weeks, constituting an advantage over saliva, urine and blood spot samples which only represents the past few hours.⁸⁷ In a recent study 560 561 with a longitudinal design comparing hair and multiple saliva samples, Singh et al., 562 2023 confirmed that hair cortisol is a reliable retrospective biomarker of at least the previous six weeks.⁸⁷ Recent evidence suggests that not only corticosteroids but other 563

steroids such as reproductive hormones can be assessed in hair samples, possibly
 leading to an improved hormone assessment in clinical and population studies.⁴²

566

567 In the current study, hair corticosteroids and reproductive hormones were 568 assessed in the last 3 centimeters of maternal hair obtained around delivery. Assuming 569 an average hair growth of 1 cm per month, the samples would reflect cumulative 570 hormone levels over the previous 3 months (the third trimester), or at least the previous six weeks.^{43,87} Indeed, we observed more associations when phthalate metabolites were 571 572 assessed at the third compared to the second trimester. Since phthalates have short halflives, a possible explanation is that phthalate exposure at the third trimester may be 573 574 more biologically relevant to the cumulative period reflected by hair hormone analysis.

575

576 4.3 Clinical relevance

Regarding the extrapolation of population-based results to the clinic, it should be taken into account that even moderate disturbances in the hormonal milieu can have relevant effects particularly during fetal development.^{2,3,5,88} Additionally, exposure to phthalates and their replacements is pervasive with virtually all human populations showing quantifiable levels.^{14–16} In this setting, small shifts in the mean of a distribution can produce large effects in their tails.^{89–92}

583

The ovarian synthesis of maternal testosterone increases about 70% throughout pregnancy.⁹³ Notwithstanding, testosterone levels are especially elevated in cases of pre-eclampsia and PCOS (Kallak et al., 2017). Since these hyperandrogenic pregnancy states have been related to preterm birth, fetal growth restriction and pregnancy complications including diabetes,^{6–9} as well as a higher risk of metabolic and behavioral problems in the offspring,^{94,95} the positive associations observed with MBzP (and suggested for MiBP) could be of clinical relevance, especially at high-end exposure levels.

592 At the end of gestation, glucocorticoid levels increase up to 20-fold compared to 593 those of mid-pregnancy, and pregnancy is thus conceptualized as a period of transient hypercortisolism.³ Indeed, the placenta acts as a stress-sensitive organ that increasingly 594 595 secretes CRH levels that in turn increase adrenocorticotrophic hormone (ACTH) 596 secretion at the maternal pituitary, with consequently increased cortisol levels at the 597 adrenal glands, which become gradually hypertrophic throughout pregnancy. The 598 activation of the HPA axis, especially the release of CRH by the placenta, has been 599 proposed to function as a biological clock that can determine a preterm, term, or postterm delivery.² Thus, our findings of increased hair cortisol and cortisone in response to 600 601 MBzP exposure could have an impact on the timing of labor, possibly leading to an 602 increased risk of preterm birth as shown by a recent meta-analysis [OR (95%CI) = 1.18 (1.01, 1.37)].⁹⁶ 603

604

605

4.4 Strengths and limitations

606 The main strengths of this work are the improved methodologies that limited bias 607 arising due to daily variations occurring in both the phthalate metabolites and hormones 608 examined. This new-generation cohort is the first to perform a within-subject pooling of designed to reduce phthalate 609 multiple repeated urine samples. exposure misclassification and attenuation bias.^{31,32} At the same time, our study is the first in the 610 611 field of endocrine disruption to use maternal hair samples obtained at delivery to 612 measure cumulative adrenal and reproductive hormone levels over the third trimester of pregnancy.^{42,43} Among limitations are the medium-size of the cohort, especially when 613

614 examining sex-stratified associations, although we anticipate a substantially increased statistical power compared to similar-sized cohorts collecting few urine samples.³¹ 615 616 Another limitation was the lack of data for related steroid hormones such as estradiol or 617 the adrenal androgen dehydroepiandrosterone (DHEA). Finally, although we controlled 618 for the most relevant covariates reported in the literature, we cannot fully exclude the 619 possibility of residual or unmeasured confounding (e.g., diet quality or medications 620 containing phthalates). Additionally, the low number of women reporting hair dyes 621 during pregnancy (n = 7) prevented us from adjusting models with this variable. 622 However, we relied on a 7-day recall questionnaire completed during the second and 623 third pregnancy trimesters to estimate hair dye treatments, which may have probably 624 underestimated its use over the whole pregnancy.

625

Overall, this prospective study with improved exposure and outcome assessment showed that maternal third trimester urinary MBzP concentrations were robustly and consistently associated with increased levels of all the adrenal and reproductive hormones evaluated in maternal hair samples at delivery, deserving further confirmation and investigation of its possible clinical implications.

631

632 Supporting Information

633 Phthalate biomarkers assessed in SEPAGES; Urinary phthalate metabolite 634 concentrations; Adjusted main associations for single-pollutant models; Sex-stratified 635 phthalate-hormone associations; Sensitivity analysis of main model with additional 636 adjustment for specific gravity; Sensitivity analysis of main model with exclusion of 637 outliers; Study population flow-chart; Cohort design figure; Directed acyclic graph 638 (DAG) for covariate selection; Generalized additive models (GAMs); Spearman 639 correlation heatplot of phthalate metabolites; Spearman correlation heatplot of hair
640 hormones; Relative weights of BWQS mixture models; Figure of sensitivity analysis
641 excluding outliers.

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661

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675

676 **Conflict of interest**

677 The authors declare no actual or potential competing financial conflicts of interest.

678

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