

Monogenic diabetes

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Monogenic diabetes

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Abstract

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Monogenic diabetes includes several clinical conditions generally characterized by early-onset diabetes, like neonatal diabetes, maturity-onset diabetes of the young (MODY) and diabetes-associated syndromes. However, patients with apparent type 2 diabetes may actually have monogenic diabetes. Indeed, the same monogenic diabetes gene depending of the functional impact of its variants can contribute to different forms of diabetes with early or late onset. Same spectrum of diabetes phenotypes can also be observed for a single specific pathogenic mutation shared by different carriers, even of the same family. Monogenic diabetes is mostly due to impaired function or development of pancreatic islets with defective insulin secretion in absence of obesity. The most prevalent monogenic diabetes form is MODY that may account for 0.5-5% of patients diagnosed with non-autoimmune diabetes, but is probably underdiagnosed due to insufficient genetic testing. Most cases with neonatal diabetes or MODY have an autosomal dominant form of diabetes. More than 40 monogenic forms of diabetes have been identified so far, the most prevalent being deficiencies for GCK (glucokinase) and HNF1A (hepatocyte nuclear factor 1 alpha). Some of these genes (including GCK and HNF1A) are actionable, namely carriers of a pathogenic mutation should benefit from precision medicine. This precision medicine includes specific treatments of hyperglycemia, the monitoring of associated extrapancreatic phenotypes, and/or the follow up of clinical trajectories especially during pregnancy, which increases patients' quality of life. Next-generation sequencing has made affordable and cost efficient the genetic diagnosis of monogenic diabetes that enables effective genomic diabetic medicine.

Introduction

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Monogenic diabetes is a heterogeneous group of disorders that include various clinical conditions associated with hyperglycemia, which are mostly due to impaired function or altered development of islets of Langerhans (including pancreatic beta cells that secrete insulin) and are linked to a single, pathogenic, genetic event. Monogenic diabetes comprises a large spectrum of clinical conditions, such as neonatal diabetes (a very rare form of monogenic diabetes at 1 in 90,000 live births 57 that occurs within the first 6 months of life), maturity-onset diabetes of the young (MODY), an early-onset diabetes occurring during childhood, adolescence or young 59 adulthood (age of onset under 25 years according to the original definition) with 60 strong family history, and diabetes-associated syndromes (e.g. Wolfram syndrome, Wolcott-Rallison syndrome and Mitchell-Riley Syndrome), the common denominator 62 of which is early-onset diabetes. Diabetes-associated syndromes are multisystem diseases that cause neonatal diabetes or diabetes with a later onset¹. Diabetes may 64 be not recognized as a first manifestation of these syndromes¹. 65 The first gene identified to cause monogenic diabetes (GCK, encoding glucokinase, a 66 alucose sensor for the pancreas) was discovered in 1992^{2,3}. Since then, several 67 dozens of genes harboring rare, coding or even non-coding mutations causing 68 monogenic diabetes have been identified, following various modes of inheritance 69 (Table 1). Some of these genes are medically actionable, which means that pathogenic mutations in these genes are related to health conditions with known 71 72 medical recommendations to act upon with the carriers (e.g. follow-up of clinical trajectories, treatment of hyperglycemia). This advocates for an enhanced screening for monogenic diabetes worldwide. 74 Monogenic diabetes phenotypes overlap with both type 1 and 2 diabetes mellitus, 75 and same genes can confer risk to a common form of diabetes or can cause 76 monogenic diabetes in another inherited context. Type 1 diabetes is due to 77 autoimmune beta-cell destruction, usually leading to absolute insulin deficiency, and 78 results from polygenic predisposition⁴. Mutations in several genes linked to 79 autoimmune diabetes have been found to cause monogenic diabetes (i.e. apparent 80 type 1 diabetes), mostly with a recessive pattern of inheritance (Figure 1). On the 81 other hand, type 2 diabetes is due to a progressive loss of adequate beta-cell insulin secretion (i.e. mostly seen in adults) frequently on the background of insulin 83

resistance, and results from polygenic predisposition like type 1 diabetes⁴. 84 Nonetheless, the dominant deficiency of many genes causing monogenic diabetes 85 has been found in patients with apparent type 2 diabetes, namely with age of onset 86 above 40 years⁵. In fact, the diagnosis of type 2 diabetes can be referred as a 87 diagnosis of exclusion and therefore type 2 diabetes is very heterogeneous. 88 Research on monogenic diabetes has truly revolutionized our understanding of the 89 pathophysiology of diabetes and the genetics of common diseases⁶. Furthermore. 90 monogenic diabetes has provided one of the best examples of the vision of precision 91 92 medicine so far. This Primer focuses on monogenic forms of diabetes primarily due to impaired function or development of pancreatic islets. It does not address potential 93 94 diabetes due to monogenic forms of insulin resistance such as lipodystrophy.

Epidemiology

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MODY may account for about 2-5% of patients diagnosed with non-autoimmune diabetes (as estimated in European patient cohorts, mainly from childhood diabetes registries), with a global prevalence of ~100 per million in white European populations⁷. However, the true prevalence of monogenic diabetes is still unknown. It may be largely underestimated in many countries due to poor access to genetic testing, no access to this diagnostic procedure or lack of awareness. In fact, genetic testing is not widely available even in most affluent countries: for instance, it was estimated that >80% of MODY cases in the United Kingdom (UK) may go undiagnosed⁸. Hopefully, recent access in next-generation sequencing facilities has improved MODY discovery rates at least in young patients with typical MODY clinical features⁹. In the UK, the apparent prevalence of MODY is four times higher in South West of England, where genetic testing is more widely implemented than in other UK regions such as Northern Ireland where it is not the case⁸. It is unlikely that ethnic differences explain these differences, but rather lower doctors' awareness of MODY in some UK regions. In the general population from the United States, 3 in 200 individuals from the general population have been shown to carry a variant that has been reported previously to cause MODY although the majority of them were not known to have diabetes yet¹⁰. A recent study in different countries around the Mediterranean Sea has screened for

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pathogenic mutations in genes linked with monogenic diabetes in patients with an

early age of diagnosis: in the entire study population, a genetic diagnosis of monogenic diabetes was established in 18% of the participants. In most countries, the pathogenic mutations were mostly harboured by GCK (7%) and HNF1A (5%). However, diagnosis rates differed widely by country of inclusion, with the highest rates observed in Greece (40%), Algeria (30%) and France (28%), and the lowest in Tunisia (8%) and Turkey (12%) while no variant of interest were found in the Moroccan patients¹¹. Differences in ethnicity, but also in patients' ascertainment, may explain this variability, knowing that the inclusion criterion was an age below 40 years at diabetes diagnosis (without insulin treatment and obesity at diagnosis, and with no autoantibodies to at least three islet cell antibody tests), or self-reported family history of diabetes consistent with autosomal dominant inheritance. Recently, a comprehensive genetic investigation in South India revealed a monogenic etiology in 15% of the cases: however, in contrast to European populations, the distribution of MODY genes was very different with a lack of pathogenic mutations in GCK^{12} . Although the true prevalence of monogenic diabetes is still unclear in adult patients with history of early-onset diabetes and absence of obesity, its putative occurrence among patients with typical type 2 diabetes (i.e. with age of onset > 40 years and overweight or obesity) has been largely ignored until recently. In a recent study, 6% of patients of European origin with apparent type 2 diabetes followed by a French diabetes clinic were found to carry pathogenic mutations in genes linked to monogenic diabetes⁵. These patients with MODY mutations were impossible to identify using clinical criteria as they did not differ from other patients with T2D⁵. Furthermore, among them, 2% carried pathogenic mutations in actionable MODY genes (i.e. their discovery should lead to an attempt to change diabetes care)⁵. In addition, in an Italian study, ~3% of adult patients with early-onset type 2 diabetes carried pathogenic mutations in MODY genes¹³. This was in line with a recent study done in US children with type 2 diabetes¹⁴. The main conclusion is that hidden

Mechanisms/pathophysiology

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MODY exists among patients with apparent type 2 diabetes and only genetic

diagnosis can recognize them, and when possible can enable precision medicine.

Pancretic islets, or islets of Langerhans, are islands of endocrine cells scattered throughout the pancreas¹⁵. Human pancreatic islets include 60% beta cells that produce insulin in response to a glucose challenge, 30% alpha cells that produce glucagon, with the remainder 10% made up of delta cells (somatostatin-producing), PP cells (pancreatic polypeptide-producing), and epsilon cells (ghrelin-producing)¹⁵. Endocrine cells are randomly distributed in human islets¹⁵. In pancreatic beta cells, insulin is packaged into secretory granules and released in response to high glucose (and other nutrient) levels, neurotransmitters (that are released in response to food) and incretin hormones¹⁵. Many genes causing monogenic diabetes have been found via multiple genetic strategies and technologies that have evolved according to the state-of-the-art in genetics (Table 1; Box 1). MODY inheritance mostly follows an autosomal dominant mode whereas neonatal diabetes and diabetes-associated syndromes can be dominant or recessive disorders. Linkage analyses (mostly using microsatellites), followed by Sanger sequencing of candidate genes in diabetes-associated linkage genomic peaks were very successful in the discovery of various types of MODY (e.g. dominant GCK-, HNF1A-, HNF4A- or CEL-diabetes)2,3,16-18. In suspected recessive forms of monogenic diabetes, linkage analyses were mostly based on the discovery of runs of homozygosity (ROH), which are genomic regions where identical haplotypes are inherited from each (consanguineous) parent. Surprisingly perhaps, single candidate gene strategy (using Sanger sequencing), which requires a prior functional hypothesis that particular genes of interest contain pathogenic mutations that may cause monogenic diabetes, was also very successful. More recently, the advent of next-generation sequencing technology (including whole-exome and whole-genome sequencing) has enabled the discovery of new monogenic diabetes genes. It is striking that the vast majority of monogenic diabetes genes are well expressed in pancreatic islets and beta cells, and play an important role in the development and/or function of pancreatic beta cells, notably insulin secretion (Table 2; Figure 2). In fact, all compartments of the pancreatic beta cells can be impacted in monogenic diabetes. We are showing below the main genes causing monogenic

Nucleus

diabetes compartment by compartment.

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A plethora of genes whose mutations cause monogenic diabetes encode transcription factors that are highly expressed in pancreatic islets and beta cells. Among these, the most frequently mutated is HNF1A, whose deficiency leads to MODY or even to adult-onset non-autoimmune diabetes. HNF1A is mainly expressed in pancreatic beta cells and in hepatocytes. Pathogenic mutations in this gene lead to impaired transactivation function, protein expression, DNA binding, and/or subcellular (nuclear) localization 19. HNF1A is an important transcription factor for the maintenance of beta-cell function, as well as for the development and maturation of pancreatic islets²⁰. HNF1A directly binds to the promoter region of the insulin gene INS, positively regulating the transactivation of the latter²⁰. Importantly, the bi-allelic deficiency of PDX1 or PTF1A, and the dominant deficiency of GATA6 cause neonatal diabetes linked with pancreatic agenesis, along with a spectrum of extrapancreatic features. However, penetrance can be incomplete²¹. PDX1, PTF1A and GATA6 are transcription factors that play a key role in pancreatic development and pancreatic cell fate specification²²⁻²⁴. Notably, PTF1A does not seems to be expressed in adult and fetal pancreatic beta cells; however, it is expressed in the very early stages of pancreatic development, being required for pancreatic specification and exocrine versus endocrine fate decision²³.

Endoplasmic reticulum

Several monogenic diabetes genes (*i.e. WFS1*, *CISD2*, *MIA3* [previously called *TANGO1*], *MANF*, *YIPF5*, *DNAJC3*, *EIF2AK3*, *EIF2B1*, *PPP1R15B* and *IER3IP1*; Table 1) encode proteins that play a key role in endoplasmic reticulum, in particular in the maintenance of endoplasmic calcium homeostasis (Figure 2). Strikingly, all these genes are ubiquitously expressed and their bi-allelic deficiency systematically causes severe early-onset diabetes associated with extrapancreatic features including deafness, hearing loss, renal tract abnormalities, short stature, hepatic dysfunction, microcephaly, epilepsy and other neurodevelopmental disorders. It has been shown that the deficiency of these genes results in endoplasmic reticulum stress indicative of increased unfolded protein response (UPR) signaling, leading to insulin secretion failure or even apoptosis of pancreatic beta cells^{25,26}. Recent studies (focused on the deficiency of *YIPF5* and *MIA3*) have also suggested that endoplasmic reticulum stress may be caused by impaired transport from the endoplasmic reticulum through the Golgi apparatus²⁶.

Plasma membrane

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Several monogenic diabetes genes encode channel subunits (ABCC8, KCNJ11, and KCNK16) or transporters (SLC2A2, SLC19A2) that are embedded in the plasma membrane (Figure 2). ABCC8 and KCNJ11 encode the subunits of an ATP-sensitive potassium channel that are highly expressed in the pancreatic islets and beta cells, although their expression is also high in other organs and tissues such as the cerebellum and pituitary (Table 2). The ATP-sensitive potassium channel encoded by KCNJ11 and ABCC8 plays a key role in insulin granule exocytosis from pancreatic beta cells. Indeed, an increase in glucose metabolism leads to an increase in intracellular ATP/ADP ratio due to enhanced glycolysis and Krebs cycle activity, which in turn results in closure of the ATP-sensitive potassium channel and membrane depolarization⁷. This activates voltage-dependent calcium channels leading to a rise in intracellular calcium, which stimulates insulin release. Activating (or gain-of-function) homozygous or heterozygous mutations in ABCC8 or KCNJ11 lead to neonatal diabetes, MODY or even late-onset diabetes, since the mutated ATP-sensitive potassium channel remains open despite high glucose levels, thereby leading to inhibition of insulin secretion in mutation carriers⁷. An activating mutation in KCNK16 encoding another potassium channel has recently been suggested to cause MODY²⁷. Contrary to ABCC8 and KCNJ11, the expression of this gene is specific to pancreatic beta cells. A pathogenic mutation in this gene was found to cause a profound increase in potassium current, altering beta-cell calcium flux, and decreasing glucose-stimulated insulin secretion²⁷. Very rare forms of monogenic diabetes were attributed to pathogenic mutations in SLC2A2 and SLC19A2 encoding glucose and thiamine transporters^{28,29}. Recessive SLC2A2- and SLC19A2-diabetes (known as Fanconi-Bickel syndrome and Rogers syndrome, respectively) lead to severe extrapancreatic features including failure to thrive, hepatomegaly secondary to glycogen accumulation, galactose intolerance, proximal tubular nephropathy, phosphaturia, aminoaciduria and hypophosphatemia for SLC2A2 deficiency, and megaloblastic anaemia, and sensorineural deafness for SLC19A2 deficiency^{28,29}. Isolated early-onset diabetes was also reported in dominant-SLC19A2 deficiency³⁰.

Mitochondria

The mitochondrial m.3243A>G mutation is one of the most frequent point mutations of the mitochondrial DNA³¹. This mutation causes a syndrome known as MELAS for

Mitochondrial Encephalopathy with Lactic Acidosis and Stroke-like episodes, or MIDD for Maternally-Inherited Diabetes and Deafness. This mutation can actually lead to a broad spectrum of phenotypes more or less severe, including hearing loss, diabetes, seizure, stroke-like episodes, muscle weakness and heart disease³¹. The molecular mechanisms underlying m.3243A>G-associated phenotypes may be due to a complex interplay between m.3243A>G mutant load, mitochondrial DNA copy number, nuclear genetic factors, epigenetics, and environmental exposures³².

[H2] Cytosol

In the cytosol, the most commonly mutated gene in dominant monogenic diabetes is *GCK* encoding glucokinase, which is the predominant glucose-phosphorylating enzyme in pancreatic beta cells and in hepatocytes³³. GCK has a relatively low affinity for glucose and displays positive cooperativity for this substrate, despite the fact that it is a monomeric enzyme³³. Functional analyses of the kinetic properties of the GCK mutants found in MODY have shown that the relative enzymatic activity was impaired, leading to a decrease in the glycolytic flux in pancreatic beta cells³³. This translates *in vivo* as a glucose-sensing defect and a right shift in the dose response curve of glucose-induced insulin secretion³³. *GCK* mutations are pathogenic through different mechanisms including impaired enzymatic activity, altered protein stability and/or increased interaction with the glucokinase regulatory protein^{34,35}. In the liver, a decrease in glycogen synthesis and storage, and an increase in gluconeogenesis after standard meals were reported in patients deficient for GCK; contributing to postprandial hyperglycemia³³. Pathogenic homozygous mutations in *GCK* also cause neonatal diabetes³⁶.

Insulin granules

Dominant or bi-allelic pathogenic mutations in *INS* were found to cause "neonatal" diabetes, (usually in infancy rather than in the first months of life), MODY or even late-onset diabetes. Most of the pathogenic, coding *INS* mutations described so far have been shown to cause severe proinsulin misfolding, endoplasmic reticulum retention, increased UPR signaling and pancreatic beta-cell apoptosis³⁷. Homozygous, pathogenic, non-coding *INS* mutations were reported in patients with neonatal diabetes^{38,39}. These mutations decreased insulin biosynthesis through distinct mechanisms, including *INS* gene deletion, severe dysfunction of transcriptional activity due to mutations located in the *INS* promoter, lack of the

translation initiation signal, and altered mRNA stability because of the disruption of a polyadenylation signal^{38,39}.

Exocrine pancreas

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A dominantly inherited monogenic form of diabetes is caused by mutations in CEL encoding the carboxyl ester lipase, which is expressed by pancreatic acinar cells^{18,40}. Patients with a pathogenic *CEL* mutation develop childhood-onset exocrine pancreas dysfunction followed by diabetes during adulthood. Clinical investigations have reported that pre-diabetic mutation carriers exhibit significantly reduced first-phase insulin response with early β-cell dysfunction prior to development of diabetes¹⁸. However, it is unclear how CEL mutations cause diabetes. Studies with in vitro and in vivo human pluripotent stem cell-based disease models and pancreatic cell lines have demonstrated co-localization of insulin and CEL protein in mutant beta-like cells, which supports a crosstalk between exocrine and endocrine compartments⁴¹. The presence of the mutant CEL in insulin-expressing cells from patients with this form of diabetes provides further evidence that an abnormal relationship between pancreatic exocrine and endocrine cells likely underlies the beta-cell dysfunction in this form of diabetes⁴¹. Specifically, the mutant CEL protein is taken up and remained insoluble in beta cells with consequent development of endoplasmic reticulum stress that contributes to defects in secretory function and proliferation 40,41. Whether CEL is taken up by beta cells through a paracrine or endocrine pathway within the pancreas and if circulating CEL in human plasma is preferentially taken up by beta cells compared to other cell types requires further investigation.

Diagnosis, screening and prevention

Clinical presentation

According to its original definition, MODY is inherited following an autosomal dominant pattern (*i.e.* 50% of offspring are carriers of heterozygous, pathogenic DNA mutations and are in theory affected) and occurs in lean individuals before the age of 25 years who do not always need insulin therapy⁴². Hyperglycemia and related symptoms can be mild especially at disease onset and can deteriorate with increasing disease duration, depending on the gene and the nature of the mutations causing hyperglycemia^{7,43}.

In practice, MODY should be strongly suspected in children and young adults with mild hyperglycemia at onset in the absence of typical features of both type 1 and type 2 diabetes. The former includes the coexistence of autoimmunity and the need of insulin treatment at diagnosis (possibly presenting with ketoacidosis), while obesity and other features related to insulin resistance including acanthosis nigricans, and sometimes early-onset atherogenic dyslipidemia and essential hypertension are among the latter^{6,7,33,44}. Finally, although the possibility of a *de novo* mutation should always be considered, a strong family history of diabetes reinforces the suspicion of MODY. While these criteria are valid, they are far from exhaustive. Indeed, several reports have now made it clear that the picture is much more heterogeneous, with MODY also occurring in patients (both children and adults) with excess adiposity and other features of insulin resistance and/or in middle-aged individuals^{7,45}. As a consequence, the "old definition" 42 misses at least a guarter of patients with MODY and therefore should be abandoned^{8,46,47}. The marked heterogeneity in the clinical presentation of MODY is largely the consequence of the multiple genetic etiologies of this form of diabetes. However, pathogenic, heterozygous mutations in several genes, including ABCC8, KCNJ11, INS, HNF1B, GATA4 and GATA6, are involved in MODY but also in severe cases of

consequence of the multiple genetic etiologies of this form of diabetes. However, pathogenic, heterozygous mutations in several genes, including *ABCC8*, *KCNJ11*, *INS*, *HNF1B*, *GATA4* and *GATA6*, are involved in MODY but also in severe cases of neonatal diabetes (Table 1); at the same time, rare and frequent variants in some of these genes also contribute to shaping the risk for common multifactorial type 2 diabetes. Thus, the different forms non-autoimmune diabetes belongs to a continuous spectrum of disease rather than being distinct entities^{6,48}. In fact, it has been shown that, even in the same family, the same heterozygous mutation can lead to neonatal diabetes, MODY, late-onset non-autoimmune diabetes or even normal glucose tolerance at an advanced age^{21,49–51}, an observation that clearly highlights the incomplete penetrance of pathogenic mutations in monogenic diabetes. Notably, a high polygenic risk score, summarizing the effect of several hundred genetic variants associated with type 2 diabetes, was found to be a significant determinant of an earlier age of diabetes diagnosis in patients with a pathogenic mutation of *HNF1A*⁵¹. Therefore, the polygenic background (as captured by a polygenic risk score for common type 2 diabetes) could be one of the causes of this variable penetrance of rare, pathogenic mutations in monogenic diabetes.

Furthermore, very recently, it was reported that approximately 20% of the pathogenic mutations found in patients indicated as having MODY were actually harbored by

genes or loci typically associated with syndromic presentations; these particularly include the (i) maternally inherited mitochondrial m.3243A>G variant, (ii) heterozygous, pathogenic mutations in *HNF1B* and (iii) bi-allelic, pathogenic mutations in *WFS1*^{44,52,53}. These observations challenge the concept of MODY and diabetes-associated syndromes being distinct clinical entities. It is now clear that MODY (especially due to *WFS1*, *SLC19A2* and *GCK* deficits) without any atypical presentation can be recessively inherited (*i.e.* with homozygous mutations or compound heterozygous pathogenic variants), especially but not exclusively in population with high rates of consanguinity⁵⁴. As mentioned above, these new findings are likely the consequences of the variable penetrance and expressivity of mutations in genes causing monogenic diabetes and further support the existence of a continuous spectrum of disease, with very blurred boundaries between each of the different diabetes subtypes thus far described as distinct entities^{45,55}.

All this indicates the need to reconsider the current classification of monogenic diabetes which appears to be too rigid and inconsistent with both clinical and molecular evidence. A practical proposal is to group the various forms of diabetes due to genetic mutations under the deliberately generic umbrella of "monogenic diabetes", to which the following notations are added consistently with each specific case: (i) the disease-causing gene; (ii) the type of hereditary transmission (e.g. dominant, recessive) and (iii) the peculiar features of clinical presentation (e.g. neonatal onset of hyperglycemia, presence of syndromic features). For example, MODY due to pathogenic, heterozygous mutations in GCK would be "dominant GCK-diabetes" while neonatal hyperglycemia due to pathogenic, homozygous mutations in GCK would be "recessive, neonatal, GCK-diabetes". As an additional example, diabetes due to pathogenic, heterozygous mutations in HNF1B would be defined as "dominant HNF1B-diabetes" or "dominant, syndromic HNF1B-diabetes" depending on the absence or presence of renal, urinary, genital or gonadal abnormalities.

Identification of patients for genetic screening

The traditional approach to identifying candidates for MODY genetic screening utilized the original clinical criteria for diagnosis⁴². However, these criteria are neither sensitive nor specific enough for identifying the majority of individuals with monogenic forms of diabetes. Large cases series have shown that in some populations pathogenic mutations are found in only 10% of all individuals suspected

to have MODY by clinical criteria (e.g. early-onset non-autoimmune diabetes with normal weight)^{12,56}. Subsequently, attempts were made to further refine the diagnosis of MODY using a risk calculator, mainly developed in populations of European origin. This MODY risk calculator, utilizing variables such as current age, age at diagnosis. gender, ongoing treatment, duration of insulin treatment, family history of diabetes, body mass index, and glycated hemoglobin A1c (HbA1c) levels⁵⁷, tend to perform well in distinguishing individuals with MODY from those with type 1 diabetes. However, they are less useful in differentiating MODY from type 2 diabetes, particularly in populations (e.g. South Asian) with a high prevalence of youth-onset type 2 diabetes⁵⁸. Considering the limitations of available diagnostic criteria and risk scores, and the fact that universal genetic screening of all youth-onset diabetes is unfortunately impractical in large parts of the world, the identification of candidates for screening is best achieved using a combination of phenotypic and biochemical variables. There is some evidence, albeit based on models with inherent assumptions, that a phenotype-

Patient age

needs confirmation with larger studies.

Diabetes diagnosed within the first 6 months of life is most likely due to monogenic defects, as autoimmune type 1 diabetes is rare at this age⁶⁰. More than 50% of children with neonatal diabetes have been found to carry pathogenic mutations in *ABCC8, KCNJ11* or *INS*^{60,61}. Diabetes due to these mutations is mostly diagnosed within the first 6 months, although it can occasionally present beyond 6 months⁶⁰. Diabetes due to pathogenic mutations in *HNF1A* and *HNF4A* tend to present with hyperglycemia before the age of 25 years, although the penetrance of disease by this age is only around 60%⁶². Individuals with dominant *GCK*-diabetes may be diagnosed at any age, and depending on this, may be treated as type 1 diabetes, type 2 diabetes or even gestational diabetes. The phenotype of nonprogressive hyperglycemia in these cases only becomes apparent with the passage of time. They are not at risk of developing chronic complications of diabetes⁴³.

based screening strategy for monogenic diabetes is cost-effective⁵⁹; however, this

Family history of diabetes

According to the originally described clinical criteria, MODY is characterised by severe young-onset diabetes and a positive family history of young-onset diabetes in

several consecutive generations. However, the majority of patients with type 2 diabetes will also report a positive family history of diabetes in either or both parents¹³. Even so, a family history of non-insulin treated young onset diabetes in an individual diagnosed with type 1 diabetes should raise the suspicion of monogenic diabetes.

Obesity and markers of insulin resistance

Presence of overweight or obesity, and markers of insulin resistance (e.g. acanthosis nigricans and skin tags) would suggest a diagnosis of type 2 diabetes rather than monogenic diabetes in such cases. It should, however, be noted that type 2 diabetes can occur even in the absence of obesity or clinically evident insulin resistance, particularly in ethnic groups such as South Asians.

Presence of extrapancreatic features

Presence of multi-organ involvement (*e.g.* renal cysts in dominant *HNF1B*-diabetes; hepatic, renal and skeletal abnormalities in Wolcott-Rallison syndrome; deafness, optic atrophy, diabetes insipidus and genitourinary abnormalities in Wolfram syndrome; deafness and megaloblastic anemia in Rogers' syndrome) should prompt genetic testing for elucidation of the molecular defect. Most of these syndromic forms of diabetes are due to bi-allelic, pathogenic mutations (in *EIF2AK3* [Wolcott-Rallison syndrome], *WFS1/CISD2* [Wolfram syndrome] and *SLC19A2* [Rogers' syndrome] genes, respectively) but also heterozygous, pathogenic mutations in *HNF1B* (Table 1). As diabetes may be the first manifestation of many of these syndromes and may remain the only manifestation for a considerable period of time⁵³, it is important to look for mutations in syndromic diabetes genes in all patients with suspected monogenic diabetes⁴⁴. On the contrary, extra pancreatic features such as kidney failure and pancreatic exocrine dysfunction may precede diabetes by decades in dominant *HNF1B*-diabetes and dominant *CEL*-diabetes, respectively^{18,63}.

Biochemical variables

Absence of islet autoantibodies, presence of endogenous insulin production (as evidenced by detectable C-peptide levels distant from the time of diagnosis) and modestly elevated blood glucose levels have been suggested as criteria for selection of individuals for genetic testing^{47,64–66}. However, another study concluded that no single clinical criterion was able to identify all patients with monogenic diabetes⁶⁷. It

should also be noted that while these criteria have been shown to perform well in distinguishing monogenic diabetes from type 1 diabetes, they are unlikely to reliably distinguish monogenic diabetes from type 2 diabetes. Some studies have even suggested that offering genetic screening for MODY could be considered for all children with diabetes irrespective of other clinical characteristics⁶⁸. This approach, however, is not practical in resource-constrained settings.

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Some clinical pointers to differentiate between monogenic diabetes and the more common type 1 and type 2 diabetes are given in Figure 3.

Genetic screening for monogenic diabetes

In the candidates described above, genetic screening for monogenic diabetes is carried out by sequencing the coding parts of genes known to be involved in this disease. In the 2000s and even before, this screening was performed via Sanger sequencing which was very labor-intensive and expensive. Since the advent of nextgeneration sequencing, genetic screening for monogenic diabetes has become increasingly cost-effective and less laborious. In this context, two sequencing technologies are mostly used today in genetic diagnostic laboratories: (i) targeted enrichment of a set of genes (or gene panels) known to be involved in monogenic diabetes; and (ii) whole-exome sequencing (i.e. sequencing of ~2 % coding part of the genome). Thanks to their targeted nature, gene panels inherently lead to a smaller amount of data to handle and lower cost, and decrease the risk of secondary findings when compared to whole-exome sequencing^{69,70}. However, as new genes are discovered, the gene panel must be re-designed and re-validated. This is not necessary with whole-exome sequencing. Also, while whole-exome sequencing leads to bioinformatic challenges resulting from the large amount of data, it enables the identification of pathogenic variants in genes that were not considered at prescription (i.e. years after the initial assessment) and it fuels research of new monogenic diabetes genes. Gene panels and whole-exome sequencing are both limited in their capacity to detect copy-number variations (e.g. the deletion of one or several entire exons which are common in dominant HNF1B-diabetes) or low-level heteroplasmic mitochondrial variants (e.g. m.3243A>G variant causing monogenic diabetes)^{69,71}. Therefore, multiplex ligation-dependent probe amplification (MLPA) and Sanger sequencing of mitochondrial DNA are often used to supplement these

techniques^{69,71}. It is likely that whole-genome sequencing will substitute these technologies in the next five years, although the issues of cost and large amount of data need to be solved. Since 2015, the standards and guidelines of the American College of Medical Genetics and Genomics (ACMG) have enabled a rigorous interpretation of sequence variants, which have been followed by all genetic diagnostic laboratories worldwide⁷². These criteria use various types of variant evidence including population data (e.g. extremely low frequency of variants in the Genome Aggregation Database), computational data (e.g. use of in silico predictive programs of variants), functional data (by *in vitro* or *in vivo* assays), and segregation data⁷². More specific guidelines have been gathered by the ClinGen Monogenic Diabetes Expert Panel VCEP for interpreting muat. In dominant monogenic diabetes, it has recently been recommended to exclude BLK, KLF11 and PAX4 from monogenic diabetes testing as the evidence in support of these gene-disease associations is still controversial; furthermore, it was recommended that all variants with a frequency higher than 0.001 in any ancestry included in the Genome Aggregation Database should be excluded from the analysis⁷³.

Prevention of monogenic diabetes

Prevention in the classical sense of the term is not possible for monogenic diabetes at present. The identification of a pathogenic mutation early in life give the opportunity of diets low on fast acting carbohydrates. That may delay the onset of diabetes and hence lower risk for diabetes-associated late complications. It is indeed likely that environmental influences might play a role in the development of diabetes in many of these cases. For instance, an individual harbouring a mutation in *HNF1A* (involved in pancreas development) might develop diabetes at an earlier age (and therefore be at higher risk of complications) if the beta cell reserve is placed under strain by the development of obesity and insulin resistance. Correction of these environmental factors might favourably modify the presentation of monogenic diabetes at least in some cases. No studies looking at this issue have been reported to our knowledge.

Management

The optimal treatment of young patients with emerging MODY has been discussed since the 1970s⁷⁴. In 1991, one year before the discovery of *GCK* deficiency in MODY, it was noted that MODY was very heterogeneous with some patients responding well to sulfonylureas (*i.e.* the only diabetes treatment along with insulin therapy in the USA at that time) and others not⁷⁵. It remains, that at that time nearly all children discovered worldwide with hyperglycemia were immediately treated with insulin by pediatricians. That included the numerous children that carried pathogenic mutations in *GCK* who were mistakenly treated with daily injections of insulin with poor results³.

Progressively, the recognition of MODY as an entity different from type 1 diabetes and the identification of subtypes of MODY and other monogenic forms of diabetes has led to a better understanding of monogenic diabetes pathophysiology and elucidated clinical characteristics and trajectory of patients with monogenic diabetes. From this point, specific treatments based on the nature of the genetic defect have emerged and have been progressively introduced as the genetic diagnosis of

monogenic diabetes became more popular among the diabetes community (Figure 4). Given the outstanding efficiency of this personalized treatment, monogenic diabetes can be seen as a one-of-a-kind achievement of precision medicine that is only limited by the lack of provider knowledge of genetics and consequent lack of

availability of genetic counseling and testing in many territories.

Dominant GCK-diabetes

Dominant *GCK*-diabetes is characterized by sustained and non-manageable modest fasting and post prandial hyperglycemia, which occurs in the very first years of life and is actually only modulated by age. Indeed, it is not rare that elderly patients deficient for *GCK* show HbA1c (*i.e.* a metric that reflects glucose control in the last couple of months) higher than 7.5%, which is above the HbA1c targets of an optimal diabetes treatment⁴³. This has led to an avalanche of non-useful medications given to these patients in absence of genetic diagnosis, which are expensive and with potential adverse effects.

Diabetes treatment in patients deficient for *GCK* is unnecessary for two reasons. First, in these patients, no treatment can modify blood glucose that is firmly controlled

by GCK enzymatic activity. Second, it has been unambiguously demonstrated that

these patients do not develop any typical complications of diabetes (e.g. retinopathy,

nephropathy)⁴³; thus, there is no reason to manage a form of diabetes that is not harmful for health. Remarkably, discontinuation of diabetes treatment does not change glucose homeostasis or other medical outcomes of these patients, whatever the duration of the condition before genetic recognition⁷⁶. Asymptomatic, mild fasting hyperglycaemia (usually 5.6–8.0 mmol/l) related to dominant *GCK*-diabetes should be only monitored without any attempt to fix it. However, dominant *GCK*-diabetes should be taken seriously in the management of pregnancies.

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Indeed, when a mother with diabetes carries a pathogenic GCK mutation while the fetus does not carry the mutation, maternal hyperglycemia causes fetal hyperinsulinemia and increased risk of macrosomia⁷⁷. On the contrary, when the mother and her fetus share the same pathogenic GCK mutation, the glucose threshold to trigger insulin secretion is similar in the mother and her fetus, leading to normal fetal insulin levels and average birth weight⁷⁷. Furthermore, if the mother presents with normal blood glucose levels and the father carries a pathogenic GCK mutation that was transmitted to the fetus, maternal blood glucose levels are insufficient to stimulate adequate fetal insulin secretion to sustain optimal growth, leading to increased risk of low birth weight⁷⁸. Finally, if the mother presents with normal blood glucose levels and the father carries a pathogenic GCK mutation that was not transmitted to the fetus, fetal insulin secretion and birth weight are normal⁷⁷. Therefore, insulin therapy should be given to a pregnant woman deficient for GCK only when frank hyperglycemia occurs and/or when diagnostic ultrasonography shows accelerated fetal growth (as elevated fetal insulin secretion leads to increased risk of fetal macrosomia and associated obstetric complications)⁷. In fact, it would be important to diagnose dominant GCK-diabetes in all women with gestational diabetes (which is rarely done unfortunately), as the optimal diabetes care greatly depends on whether the fetus has inherited the pathogenic GCK mutation or not, rather than the degree of maternal blood glucose. It has been stated that around 5% of women with gestational diabetes carry a pathogenic mutation causing MODY (and not only in GCK)⁷⁹, but actually no large genetic study of women with GDM has systematically addressed this issue vet.

Dominant *GCK*-diabetes should be considered in pregnant women with mild fasting hyperglycemia (5.6–8.0 mmol/L), often during the first trimester, and with normal body mass index. If fasting hyperglycemia is found in the second trimester, MODY might be suspected in case abnormal 120-min oral glucose tolerance test above 7.5

mmol/l with HbA1c being either normal or moderately increased (5.6–7.6%). These patients usually present with a history of gestational diabetes in a previous pregnancy, a record of mild hyperglycemia before conception or after the delivery and a strong family history of type 2 diabetes at early ages. It is also possible to suspect dominant *GCK*-diabetes with history of fetal macrosomia in previous pregnancies, or of insulin therapy only during the previous pregnancy that has been removed after delivery⁷⁹.

Dominant *HNF1A*- or *HNF4A*-diabetes

HNF4A and HNF1A- or HNF4A-diabetes often causes severe, early-onset diabetes, which is frequently revealed at puberty (*i.e.* when insulin resistance can appear)³³. In absence of genetic diagnosis, patients deficient for HNF4A or HNF1A are unfortunately generally treated by insulin injections⁷. If glucose homeostasis is badly controlled, dominant HNF1A- or HNF4A-diabetes shares the same long-term risks as type 1 diabetes, namely diabetic kidney disease and retinal complications. However, as both HNF4A and HNF1A modulate the liver metabolism of sulfonylureas, the intake of oral sulfonylureas has been shown to be particularly efficient at low dosage in the patients deficient for HNF4A or HNF1A, which diminishes the risk of hypoglycemia⁸⁰. These two forms of monogenic diabetes are thus optimally treated with low-dose sulfonylureas because of their increased insulin secretion and their increased insulin sensitivity.

Unfortunately, as the genetic diagnosis is often much delayed in age, the current

Unfortunately, as the genetic diagnosis is often much delayed in age, the current practice is still to introduce oral sulfonylureas after decades of insulin therapy, especially when diabetes occurs in adolescence or young adulthood and is confused with type 1 diabetes. This is tragic as it is important to perform the switch from insulin to sulfonylureas as early as possible. Indeed, at any age, the successful switch of insulin therapy into sulfonylureas in patients deficient for *HNF1A* or *HNF4A* is highly dependent of the residual insulin secretion that can be measured by the C-peptide which is the residue of the processing of proinsulin to insulin, reflecting insulin endogenous production⁸¹. Furthermore, body mass index, disease duration and HbA1c levels help predict the efficacy of sulfonylureas in patients deficient for *HNF1A* or *HNF4A*⁷⁶. As insulin secretion progressively declines in these patients, it is of paramount importance to recognize these conditions as early as possible. When the

treatment with sulfonylureas is not sufficiently efficient, it has been proposed to treat these patients with glucagon-like peptide 1 (GLP1) analogs that may work well according to recent studies⁸².

With regard to pregnancy, fetal growth is affected by maternal glycemic control in dominant *HNF1A*-diabetes, and by both fetal mutation status and maternal hyperglycemia in dominant *HNF4A*-diabetes⁷⁹. Importantly, sulfonylureas cross the placental barrier and can lead to macrosomia and hyperinsulinemic hypoglycemia, particularly in the third trimester⁷⁹. Therefore, it is recommended to switch to insulin at pre-pregnancy (when possible) or to switch to insulin at the end of the first trimester if the glucose control is adequate during the first months of pregnancy. The overall principles of insulin treatment in these pregnant women with dominant *HNF1A*- or *HNF4A*-diabetes are actually similar to the treatment of other pregnant women with diabetes. However, due to the risk of higher birth weight and fetal macrosomia, the pregnancy of these women should be regularly monitored by sonography to detect the fetal growth since 28th week of gestation every two weeks⁷⁹. If macrosomia is identified, induction of delivery or caesarian between 35 and 38th week of gestation should be considered⁷⁹.

Importantly, dominant *HNF1A*-diabetes can be associated with liver adenomatosis (in 6.5% of the cases according to a study), a rare disease that can lead to severe abdominal hemorrhage⁸³. Therefore, it is recommended to screen carriers in the families for liver tumours, as preventive surgical or non-surgical procedures can be discussed to prevent the risk of liver bleeding, especially when adenomas are numerous or large⁸³.

Dominant KCNJ11- or ABCC8-diabetes

As mentioned above, pathogenic, activating, heterozygous mutations in the two ATP-dependent potassium channel genes *KCNJ11* and *ABCC8*, which cause neonatal diabetes, can also lead to MODY or even typical type 2 diabetes. Their treatment of choice is relatively high dosage of sulfonylureas as SUR1 (encoded by *ABCC8*) is the ligand of this class of drugs which generally enables long-term excellent glucose control^{84,85}. However, if the transfer from insulin is successful for most patients with dominant *KCNJ11*-diabetes, the good response to sulfonylurea is best predicted by the *in vitro* response of the specific mutation and the duration of diabetes⁸⁶. This result strongly supports the early genetic testing and early treatment change⁸⁶.

Neurological features, especially epilepsy, impaired cognition, motor function and muscular tone, visuo-motor integration, and attention deficits are frequent in children and adults carrying pathogenic mutations in *KCNJ11* or *ABCC8*⁸⁷. A recent metareview of 34 publications showed that the sulfonylurea glibenclamide significantly improved neurological abnormalities in these patients, but earlier treatment initiation was associated with greater benefits. Hypotonia was the symptom that responded best⁸⁷.

Other forms of monogenic diabetes

There is no specific treatment for the other types of monogenic diabetes. These patients are often treated with insulin injections⁸⁸. This is the case for dominant *HNF1B*-diabetes. Unlike patients with dominant *HNF1A*- or *HNF4A*-diabetes, those deficient for *HNF1B* do not respond adequately to treatment with sulfonylureas, possibly due to comorbid pancreatic hypoplasia and some degree of hepatic insulin resistance⁸⁹. However, the genetic diagnosis of monogenic diabetes should be carefully followed in many cases by the monitoring of extra-pancreatic development abnormalities: more commonly in heart in patients deficient for *GATA4* or *GATA6*, and in the renal and genitourinary organs in patients deficient for *HNF1B*^{21,49,63}. It is noteworthy that in many cases the impaired development of the heart found in *GATAs* mutations has led to surgery at birth, a history that should be elicited while evaluating early-onset diabetes in order to guide the genetic screening.

Specific lifestyle and dietary modifications are not different from common forms of type 1 diabetes and type 2 diabetes with reasonable physical activity (avoiding hypoglycemia) and a balanced diet avoiding high glycemic index food⁹⁰. Of note, most patients with monogenic diabetes are not obese (Figure 3), and thus weight loss is generally not an issue to be fixed by lifestyle modifications.

Quality of life

Monogenic diabetes is genetically and phenotypically heterogeneous and the clinical care of its different subtypes is diversely demanding. However, the genetic diagnosis of monogenic diabetes usually increases the quality of life of patients and their family and decreases their anxiety about their future children^{91,92}. This is obvious for dominant *GCK*-diabetes, as the prognosis of glucokinase deficiency is benign and, thus, diagnosing this mild form of hyperglycemia is "good news". However, the

condition should not be forgotten especially in women considering future pregnancies. For dominant *HNF1A*- and *HNF4A*-diabetes, low dosage sulfonylureas with or without GLP1 analogues has greatly improved the quality of life of these patients⁷, and it is likely that it will be beneficial for the long-term health of these patients. The situation is the same for patients dominant *KCNJ11*- or *ABCC8*-diabetes. Indeed, in contrast to patients with common type 2 diabetes, there is no long-term beta-cell desensitization to sulfonylureas and excellent glucose control remains for decades⁸⁵. Unfortunately, for patients with one of the other forms of monogenic diabetes, insulin therapy is generally required which is as challenging as in patients with type 1 diabetes. With regard to long-term disability, the most worrying situation concerns dominant *HNF1B*-diabetes as infertility and even end-stage renal failure are possible⁶³. Close follow up of these patients is necessary.

Studies in 2011 and 2014 have evaluated the cost-effectiveness of neonatal diabetes and MODY genetic testing, respectively 91,92. Main outcome measures were costs and quality-adjusted life years (QALYs) based on lifetime risk of complications and treatments, expressed as the incremental cost-effectiveness ratio (ICER). Genetic diagnosis targeting KCNJ11 and ABCC8 was particularly cost-efficient with a good 0.32 QALYs at 10 years and 0.70 at 30 years, along with an ICER of \$200,000 per QALY⁹². Testing GCK, HNF1A and HNF4A in patients between the ages of 25–40 years with presumed type 2 diabetes yielded an average gain of 0.012 QALY and resulted in an ICER of \$205,000 per QALY, considering a MODY prevalence of 2%⁹². An increase in MODY prevalence from 2 to 6% would lead to an ICER of about \$50,000 per QALY, which is frequently used as the threshold for assessing costeffectiveness of a health intervention⁹¹. It is noteworthy that this study was based on the expensive and labor-intensive Sanger sequencing (i.e. \$2,580 per individual tested). The decrease in the cost of genetic testing (e.g. using next-generation sequencing methodologies that are much more economical and quick to run) from \$2,580 to \$700 led to an ICER of \$50,000 per QALY⁹¹. A more recent article has confirmed that the widespread implementation of genetic diagnosis along with screening of biomarkers (i.e. negativity for islet cell autoantibodies and positivity for C-peptide) could improve the lives of patients with MODY while saving the health system money (increased average quality of life [0.0081 QALY] and decreased costs [-\$735] per patient)⁹³. Adding cascade genetic testing (i.e. testing first-degree

relatives of probands) further improved the lives of patients and substantially decreased costs⁹³.

Outlook

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Monogenic diabetes research has truly revolutionized our understanding of the pathophysiology of diabetes and the genetics of common diseases⁶: it has been one of the first proof of concepts that a very prevalent and complex disorder could be caused by a single gene defect. Later on, further models of complex traits including obesity but also heart diseases or Alzheimer disease have emerged, showing a continuum between rare and common forms of diseases. As mentioned above, depending of the nature of the DNA variant, the same monogenic diabetes genes can contribute to different forms of diabetes with early or late onset. This spectrum of diabetes phenotypes can also be observed for a single specific pathogenic mutation shared by different carriers. Therefore, a boundary between monogenic diabetes and type 2 diabetes is less obvious than the community thought 20 years ago. These new findings have dramatic consequences on diabetes management: a non-negligible part of patients with "typical" common forms of type 2 diabetes actually carry a pathogenic mutation in a monogenic diabetes gene, and thus they may potentially be treated differently than those who have common, truly polygenic type 2 diabetes⁵. Monogenic diabetes research (along with genome-wide association studies related to type 2 diabetes) has also radically modified our understanding of the pathophysiology of non-autoimmune diabetes. Indeed, before 1992, when dominant GCK-diabetes was discovered, insulin resistance was considered as the primary abnormality eventually leading to abnormal glucose control with secondary exhaustion of pancreatic insulin-secreting cells. Monogenic diabetes research has demonstrated for the first time that inherited defects in insulin secretion can cause diabetes, even at a young age, without the need for severe insulin resistance or concurring obesity. Far later, genome-wide association studies (in combination with other genomic analyses or in vitro assays) have also shown that (i) type 2 diabetes-associated single nucleotide polymorphisms (SNPs) are enriched in regulatory regions that are active in pancreatic islets and not in tissues related to insulin resistance^{94,95}; (ii) susceptibility genes for common type 2 diabetes (i.e. close to type 2 diabetesassociated SNPs) are involved in insulin secretion far more frequently than in insulin

- resistance⁹⁶. In this respect, monogenic diabetes genes and susceptibility genes for common type 2 diabetes are mostly involved in the development and function of pancreatic beta cells.
- If monogenic diabetes research in the past 30 years has been very successful, several gaps remain to be filled. Several hypotheses could explain why the etiology of hyperglycemia in some patients with atypical diabetes remain unclear even after state-of-the-art attempt of genetic diagnosis:
- (i) These patients may actually have a polygenic form of type 2 diabetes with very early age of onset (*e.g.* due to the presence of type 2 diabetes in both paternal and maternal lineages). Polygenic risk scores for type 2 diabetes derived from genome-wide association studies should be systematically analyzed as these MODY-like patients maybe in the upper range of the polygenic risk scores, which borders the risk scores for diabetes of pathogenic mutations in monogenic diabetes genes⁹⁷.
- 749 (ii) Alternatively, these patients may carry a deleterious mutation in a non-coding 750 regulatory region of a monogenic diabetes gene. An elegant study showed that 751 deleterious homozygous non-coding mutations in a distal *PTF1A* enhancer led to 752 neonatal diabetes due to pancreatic agenesis⁹⁸. Therefore, whole-genome 753 sequencing (along with genomic analyses) would be the only way to elucidate these 754 cases.
- 755 (iii) Of course, it is possible that other, as yet unknown but very rare monogenic 756 diabetes genes remain to be identified.

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- That advocates for a graded approach in the diagnosis of monogenic diabetes including, first, whole-exome sequencing with a focus on known monogenic diabetes genes. If negative, the mutation analysis should be extended to candidate genes for diabetes or should be included in a gene-centric analysis, including other unelucidated cases and elderly normal glucose controls, in order to find new monogenic diabetes genes. In parallel, whole-genome sequencing and DNA microarray genotyping may be performed. The situation is quite different in non-European ancestries (e.g. Asians, Africans) where only a minority of cases have been elucidated, so far¹¹. A specific effort of screening these populations by all possible means (i.e. whole-exome sequencing, whole-genome sequencing, DNA microarrays) is needed to make further progress in these ethnicities.
- Despite strong initial hopes with GCK activators⁹⁹, no drug directly derived from monogenic diabetes research has been developed yet and introduced into the

market. GCK activators failed since their effect on insulin secretion was modest (with diminished long-term efficacy) and they had adverse effects on the liver, with the development of steatosis and hypertriglyceridemia directly linked to increased liver glucokinase activity¹⁰⁰. To our knowledge, no other promising drug target directly coming from monogenic diabetes research has emerged so far. Ironically, the first ever anti-diabetic drug family, the sulfonylureas, which directly targets SUR1 encoded by *ABCC8*, has a much better long term efficacy in monogenic diabetes than in patients with common type 2 diabetes⁴⁵. Of note, families where three generations of patients with apparent type 2 diabetes are well controlled by sulfonylureas may actually carry a mutation in *ABCC8*, *KCNJ11*, *HNF1A* or *HNF4A*⁵⁵. Further efforts may provide new clues. This is very important since the majority of patients with type 2 diabetes fail to achieve treatment objectives (in term of glucose control, weight management etc.), and innovative molecules restoring normal insulin secretion are lacking. More efforts directed to the pathways involving monogenic diabetes genes in pancreatic beta cells may reveal novel drug targets.

The priorities for basic and clinical research and precision medicine centered on monogenic diabetes may be summarized as follows: (i) developing economical, quick and reliable gene screening methodologies and high throughput pipelines that can be disseminated in every country to diagnose most of the patients with monogenic diabetes whatever their age is and their phenotype looks like; (ii) making more progress in developing specific treatments for monogenic diabetes gene mutation carriers as their long-term health (especially in dominant *HNF1A*- or *HNF4A*-diabetes) is still uncertain – an effort that implies having an efficient follow-up of every patient diagnosed with monogenic diabetes; (iii) further advancing our knowledge on pancreatic beta-cell biology by leveraging the outcomes of monogenic diabetes research.

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- A.D., M.V.); Diagnosis/Screening/Prevention (A.B., R.U., V.M., V.T.); Management
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811 Conflict of Interest

The authors declare no competing interests.

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Vionnet, N. *et al.* Nonsense mutation in the glucokinase gene causes early-onset non-insulin-dependent diabetes mellitus. *Nature* **356**, 721–722 (1992). **First original** article that highlighted pathogenic mutations in *GCK* leading to one of the most prevalent forms of monogenic diabetes

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Steele, A. M. *et al.* Prevalence of vascular complications among patients with glucokinase mutations and prolonged, mild hyperglycemia. *JAMA* **311**, 279–286 (2014) **Original article that showed a very low prevalence of vascular complications in patients with long-term dominant** *GCK***-diabetes**

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Saint-Martin, C., Bouvet, D., Bastide, M. & Bellanné-Chantelot, C. Gene Panel Sequencing of Patients With Monogenic Diabetes Brings to Light Genes Typically Associated With Syndromic Presentations. *Diabetes* **71**, 578–584 (2022). **Recent original article that highlighted the incomplete penetrance of pathogenic mutations in syndromic monogenic diabetes genes. Indeed, these mutations were found in patients with a suspicion of non-syndromic monogenic diabetes.**

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1205 Tables

Table 1 | Identification and inheritance mode of monogenic diabetes genes

Cons	Inharitanaa mada	Leastion	Dof
Gene	Inheritance mode	Location	Ref.
MODY (without other atypi		7012	[2,3
GCK	AD/AR	7p13	16
HNF1A	AD	12q24.31	17
HNF4A	AD	20q13.12	101
PDX1	AD	13q12.2	102
HNF1B	AD	17q12	103
NEUROD1	AD	2q31.3	18
CEL	AD	9q34.13	
INS	AD	11p15.5	37
ABCC8	AD	11p15.1	104
KCNJ11	AD	11p15.1	50
GATA6	AD	18q11.2	21
WFS1	AD/AR	4p16.1	105
TRMT10A	AR	4q23	106
PCBD1	AR	10q22.1	107
GATA4	AD	8p23.1	49
APPL1	AD	3p14.3	108
RFX6	AD	6q22.1	109
MAFA	AD	8q24.3	110
SLC19A2	AD/AR	1g24.2	30
ONECUT1	AD	15q21.3	111
m.3243A>G	maternally	Mitochondria	52
KCNK16	AD	6p21.2	27
Neonatal Diabetes (withou		[0μ21.2	
HNF1B	AD	17q12	[101
GCK	AR		36
		7p13	113
KCNJ11	AD/AR	11p15.1	114
ABCC8	AD/AR	11p15.1	115
INS	AD/AR	11p15.5	116
GATA6	AD	18q11.2	49
GATA4	AD	8p23.1	1.0
Syndrome including Neon		Т	1117
m.3243A>G	maternally	Mitochondria	102
HNF1B	AD	17q12	118,119
AIRE*	AR/AD	21q22.3	120
PDX1	AR	13q12.2	
SLC2A2	AR	3q26.2	28
WFS1	AR/AD	4p16.1	121,122
SLC19A2	AR	1q24.2	29
EIF2AK3	AR	2p11.2	123
FOXP3*	XR	Xp11.23	124
PTF1A	AR	10p12.2	125
GLIS3	AR	9p24.2	126
CEL	AD	9q34.13	18
PAX6	AR	11p13	127
STAT5B*	AR	17q21.2	128
CISD2	AR	4q24	129
RFX6	AR	6q22.1	130
NEUROD1	AR	2q31.3	131
NEUROG3	AR	10q22.1	132
IER3IP1	AR	18q21.1	133
ZFP57	AR	6p22.1	134
MNX1	AR	7q36.3	135
GATA6	AD		21
		18q11.2	136
DNAJC3	AR	13q32.1	137
STAT3*	AD	17q21.2	106
TRMT10A	AR	4q23	1.00

GATA4	AD	8p23.1	49
NKX2-2	AR	20p11.22	138
CDKN1C	AD	11p15.4	139
PPP1R15B	AR	1q32.1	140
LRBA*	AR	4q31.3	141
IL2RA*	AR	10p15.1	142
FOXA2	AD	20p11.21	143
CNOT1	AD	16q21	144
YIPF5	AR	5q31.3	145
EIF2B1	AD	12q24.31	146
MIA3	AR	1q41	147
STAT1*	AD	2q32.2	148
MANF	AR	3p21.2	149
ONECUT1	AR	15q21.3	111

1208 **AD**, autosomal dominant inheritance; **AR**, autosomal recessive inheritance; **XR**, X-1209 linked recessive inheritance

*Genes causing auto-immune diabetes

Additional MODY genes have been described in *BLK*, *KLF11* and *PAX4* but the evidence in support of these gene-disease associations has been refuted⁷³.

Table 2. | Expression of monogenic diabetes genes and nature/location of encoded proteins

	•		TPM	TPM	TPM	
Gene	Protein type	Location	adult	fetal	adult	Other hits
			beta cell	beta cell	islet	
ABCC8	Channel subunit	Membrane	1010	232	226	Brain (cerebellum), pituitary
KCNJ11	Channel subunit	Membrane	34	28	11	Skeletal muscle, pancreas, brain (cerebellum), heart, thryroid, stomach, pituitary
KCNK16	Channel subunit	Membrane	182	97	68	None
SLC2A2	Transporter	Membrane	10	1	10	Liver, Small intestine
SLC19A2	Transporter	Membrane	9	9	11	Ubiquitous
GCK	Enzyme	Cytosol	30	68	25	Liver
APPL1	Protein binding	Cytosol	42	17	14	Ubiquitous
CEL	Enzyme	Cytosol	-	-	86	Pancreas
CNOT1	Protein/RNA binding		-	-	71	Ubiquitous
LRBA*	Protein binding	Cytosol	62	43	19	Ubiquitous
INS	Hormone	Granule	205700	83300	37000	None
PAX6		Nucleus	261	333	122	Brain (cerebellum)
PCBD1	Enzyme	Nucleus	53	113	48	Ubiquitous
RFX6		Nucleus	79	107	18	None
PDX1		Nucleus	118	105	18	Pancreas
	Protein binding	Nucleus	107	27	54	Ubiquitous
NKX2-2		Nucleus	104	42	16	Brain, pituitary
NEUROD1	Transcription factor	Nucleus	189	167	31	Brain (cerebellum)
GLIS3		Nucleus	136	111	35	Thyroid
STAT3*	Transcription factor	Nucleus	110	32	81	Ubiquitous
FOXA2	Transcription factor	Nucleus	42	43	23	Liver, lung, stomach, pancreas, colon, thyroid
NEUROG3	Transcription factor	Nucleus	0	35	0	Brain (hippocampus), small intestine, colon
MAFA	Transcription factor	Nucleus	97	13	26	Skeletal muscle, testis
STAT5B*	Transcription factor	Nucleus	26	18	17	Ubiquitous
STAT1*	Transcription factor	Nucleus	43	10	68	Ubiquitous
MNX1	Transcription factor	Nucleus	35	21	9	Pancreas, colon, pituitary, stomach, small intestine
HNF1B	Transcription factor	Nucleus	5	23	16	Kidney, liver, lung, pancreas, bladder, colon, small intestine, stomach, testis
HNF1A	Transcription factor	Nucleus	10	22	5	Liver, kidney, pancreas, colon, small intestine, stomach
HNF4A	Transcription factor	Nucleus	5	12	15	Liver, kidney, pancreas, colon, small intestine, stomach
GATA6	Transcription factor	Nucleus	-	-	14	Ubiquitous
GATA4	Transcription factor	Nucleus	1	0	3	Pancreas, ovary, heart, testis, stomach, artery
	Enzyme	Nucleus	2	2	4	Ubiquitous
		Nucleus	-	-	4	Liver, pancreas, testis
FOXP3*	•	Nucleus	0	2	0	Blood, testis, spleen, skin, small intestine, lung
PTF1A	Transcription factor	Nucleus	0	0	0	Pancreas
ZFP57	Transcription factor	Nucleus	0	0	0	Brain (spinal cord, substantita nigra)
AIRE*	Transcription factor	Nucleus	_	-	0	Brain
IL2RA*	Protein binding	Nucleus	0	0	1	Spleen, small intestine, lung
WFS1	Protein binding	ER	112	19	28	Ubiquitous
MANF		ER	93	46	86	Ubiquitous
	Protein binding	ER	56	71	35	Ubiquitous
	Protein binding	ER	49	8	23	Ubiquitous
EIF2B1	Protein binding	ER	42	29	19	Ubiquitous
MIA3	Protein binding	ER	43	21	37	Ubiquitous
	Protein binding	ER	59	21	47	Ubiquitous
YIPF5	Protein transport	ER	-	-	41	Ubiquitous
CISD2	Protein binding	ER	15	8	28	Ubiquitous
EIF2AK3	Protein binding	ER	15	12	24	Ubiquitous
LII 2/11/0	i. Totom binding	<u>-1\</u>	10	14	47	Obiquitodo

^{*}Genes causing auto-immune diabetes

1216	TPM in adult and fetal pancreatic beta cells are shown according to RNA-seq data published
1217	in ref ¹⁵⁰ . TPM in adult pancreatic islets are shown according to RNA-seq data published in
1218	ref ¹⁵¹ .
1219	ER , endoplasmic reticulum; TPM , transcript per million
1220	

Figure 1 | Overlapping between monogenic diabetes, type 1 diabetes and type 2 diabetes

Genes harbouring rare pathogenic mutations leading to monogenic diabetes, type 1 diabetes and/or type 2 diabetes. #autosomal recessive inheritance; *autosomal dominant inheritance; *X-linked recessive inheritance



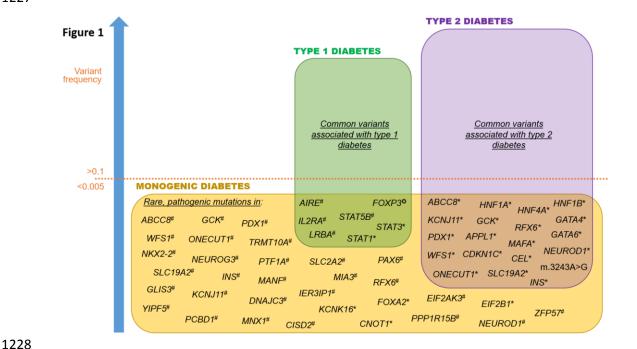


Figure 2. | Monogenic diabetes genes in pancreatic beta cells

Monogenic diabetes genes impact all compartments of pancreatic beta cells including nucleus, endoplasmic reticulum, plasma membrane, mitochondria etc.

*Genes linked with auto-immune diabetes. Only genes that are expressed in human islets or beta cells are shown in this figure

Insulin granules

Figure 3 | Differentiation between monogenic diabetes, type 1 diabetes and type 2 diabetes, based on clinical presentation and ancestry

Figure 3

Clinical presentation features that differentiate monogenic diabetes, type 1 diabetes and type 2 diabetes at the time of diagnostic. Ideas of magnitude have been shown in this flow chart based on multiple articles in the literature^{5,9,11,152,153} and on the definition of type 1 diabetes, type 2 diabetes and monogenic diabetes.

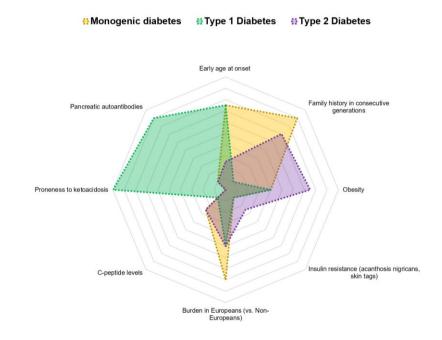
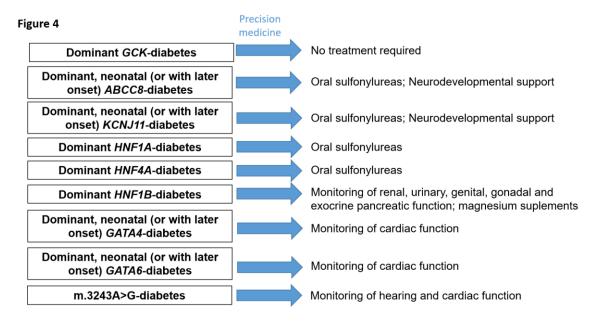


Figure 4 | Actionable genes linked with monogenic diabetes

Precision medicine can be applied in several monogenic forms of diabetes.



1248 1249	Boxes
1250 1251 1252	Box 1 Monogenic diabetes-associated genes and technology used for their discovery
1253	Linkage analyses followed by Sanger sequencing of candidate genes in
1254	diabetes-associated linkage genomic peaks
1255	GCK, HNF1A, HNF4A, CEL, INS, WFS1, SLC19A2, CISD2, IER3IP1
1256	Candidate gene strategy
1257	PDX1, HNF1B, NEUROD1, KCNJ11, ABCC8, GATA4, NEUROG3, AIRE,
1258	SLC2A2, PAX6, STAT5B, ZFP57
1259	 Homozygosity mapping followed by Sanger sequencing of candidate
1260	genes in runs of homozygosity (ROH)
1261	EIF3AK3, FOXP3, PTF1A, GLIS3
1262	Candidate variant strategy
1263	m.3243A>G
1264	Next-generation sequencing
1265	GATA6, MNX1, DNAJC3, STAT3, APPL1, MAFA, KCNK16, PPP1R15B,
1266	LRBA, IL2RA, FOXA2, CNOT1, YIPF5, EIF2B1, MIA3, STAT1, MANF
1267	 Linkage analyses and next-generation sequencing
1268	TRMT10A, PCBD1, CDKN1C
1269	 Homozygosity mapping and next-generation sequencing
1270	RFX6, NKX2-2, ONECUT1