

Astrocytes and Microglia as Major Players of Myelin Production in Normal and Pathological Conditions

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6 Abstract

7 Myelination is an essential process that consists in ensheathment of axons by myelin. In the 8 central nervous system (CNS), myelin is synthesized by oligodendrocytes. The proliferation, migration and differentiation of oligodendrocyte precursor cells constitute a prerequisite before 9 mature oligodendrocytes extend their processes around the axons and progressively generate a 10 multilamellar lipidic sheath. Although myelination is predominately 11 driven bv 12 oligodendrocytes, the other glial cells including astrocytes and microglia, also contribute to this process. The present review is an update of the most recent emerging mechanisms involving 13 astrocyte and microglia in myelin production. The contribution of these cells will be first 14 15 described during developmental myelination that occurs at the early postnatal period and is critical for the proper development of cognition and behavior. Then, we will report the novel 16 findings regarding the beneficial or deleterious effects of astroglia and microglia, which 17 18 respectively promote or impair the endogenous capacity of oligodendrocyte progenitor cells to induce spontaneous remyelination after myelin loss. Acute delineation of astrocyte and 19 microglia activities and cross-talk should uncover the way towards novel therapeutic 20 21 perspectives aimed at recovering proper myelination during development or at breaking down the barriers impeding the regeneration of the damaged myelin that occurs in CNS demyelinating 22 diseases. 23

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During the last decades, the production of myelin in the central nervous system (CNS) 28 has been the focus of a multitude of studies in the context of development or regeneration. 29 Oligodendrocytes, the cells synthesizing myelin in the CNS, were initially found in hind-jawed 30 fishes (Zalc and Colman, 2000;Hartline and Colman, 2007;Zalc, 2016). These cells derive from 31 oligodendrocyte progenitor cells (OPCs) occuring in several waves. In mice, the first wave 32 arises at embryonic day (E) 12.5 in the ventral domain of the ventricular wall in the developing 33 34 brain and spinal cord. The second one emerges more dorsally at E15.5 in both regions and the last one arises specifically in the perinatal cortex around birth (Pringle and Richardson, 35 1993;Timsit et al., 1995;Trousse et al., 1995;Spassky et al., 1998;Olivier et al., 2001). 36 Unexpectedly, the destruction of one of these waves in the telencephalon induces the remaining 37 cells to compensate for the lost oligodendrocytes thus suggesting functional similarity. In 38 agreement with this observation, ventrally derived oligodendroglial cells strongly decline early 39 after birth in the postnatal dorsal forebrain and are replaced by dorsally derived cells, which 40 thus constitute more than 80% of oligodendroglial cells present in the adult corpus callosum 41 and cerebral cortex (Kessaris et al., 2006;Tripathi et al., 2011). Myelination itself progresses 42 according to typical sequences, which are spatially and temporally determined. In human, after 43 16 weeks of gestation, myelin can be detected in the fasciculus cuneatus, and shortly after 44 occurs in the cerebellar and pyramidal tracts. Then, myelination proceeds rapidly during the 45 first year from the occipital to the fronto-temporal lobes. Regions devoted to basic homeostasis 46 are proposed to be myelinated before areas involved in more intricate tasks like the frontal 47 cortex. Late myelinated areas are typically myelinated at a lower level than early myelinating 48 49 regions (Brody et al., 1987;Kinney et al., 1988). More recent data derived from serial reconstructions of the adult mouse cortex showed that the degree of myelination can even vary 50 51 along a single axon. Thus, neurons located in superficial cortical layers exhibit both myelinated-52 and large unmyelinated segments, contrasting with the pattern observed for instance in the 53 spinal cord or the optic nerve where myelinated axons display regular internodes (Tomassy et 54 al., 2014).

55 The finding that the CNS is able to regenerate myelin lost upon various types of insults was observed much earlier than the characterization of oligodendrocyte and myelin generation. 56 57 Indeed, almost 60 years ago, occasional myelin sheaths was visualized a short time after 58 demyelination was induced into the subpial cord by repeated withdrawal and reinjection of 59 cerebrospinal fluid (Bunge et al., 1961). However, the demonstration that central remyelination 60 was able to restore secure conduction in the demyelinated spinal cord and to lead to functional recovery was provided only two decades later (Smith et al., 1979). Since then, thousands of 61 62 publications were dedicated to this process notably in the aim to investigate how new OPCs are recruited to the demyelinating area and differentiate into myelinating oligodendrocytes (Gallo 63 and Armstrong, 2008;Clemente et al., 2013;Domingues et al., 2016;Lloyd et al., 2017). 64

Although in both development and repair, the oligodendrocyte is the pivotal cell type for myelin production, its function is widely influenced by the other glial cells, namely astrocytes and microglia. This review is aimed at revisiting the present knowledge regarding communication between oligodendroglial, astroglial and microglial cells in the context of myelin development and regeneration with a specific focus on most recent advances.

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71 Astrocyte-oligodendrocyte communication in developmental myelin production

In the mouse spinal cord and cerebral cortex, the first mature astrocytes expressing the glial fibrillary acidic protein (GFAP) can be detected at the embryonic day (E) 12.5 and 16,5 respectively, thus before start of OPC maturation and axon myelination taking place during the first three postnatal weeks (Miller et al., 1985;Qian et al., 2000). Like neurons and oligodendrocytes, astrocytes are derived from the neuroepithelium from which they migrate throughout the CNS along radial glia processes (Molofsky and Deneen, 2015).

78

Astrocyte-derived trophic factors and cytokines control oligodendrogenesis

79 The crucial role of astrocytes was initially related to the finding that these cells are the 80 main producers of platelet-derived growth factor (PDGF). Acting through its receptor PDGFRa, PDGF promotes OPC proliferation and mobility and, also inhibits OPC differentiation towards 81 mature oligodendrocytes, by generating a graded response requiring activation of the 82 phosphoinositide 3-kinase and/or phospholipase C γ pathways. Astrocyte-derived PDGF was 83 thus proposed to drive the clock that times oligodendrocyte development (Raff et al., 84 1988; Richardson et al., 1988; McKinnon et al., 2005). Besides PDGF, astrocytes secrete other 85 growth factors. Brain-derived neurotrophic factor (BDNF) is namely required to potentiate 86 myelination during the early postnatal development as indicated by BDNF knockout 87 heterozygous mice, which exhibit delayed CNS myelination (Cellerino et al., 1997). In the same 88 line, under stress conditions such as those occurring during prolonged cerebral hypoperfusion 89 at birth, astrocyte-derived BDNF also supports OPC maturation in a TrkB-dependent manner 90 as shown in vitro in sublethal CoCl2 exposition of OPC / astrocyte primary co-cultures and, in 91 vivo, in a transgenic mouse line in which BDNF expression was specifically downregulated in 92 astrocytes (Miyamoto et al., 2015). The observation that ciliary neurotrophic factor (CNTF) 93 synthesis by astrocytes starts at the onset of myelination in the rodent optic nerve (Stockli et 94 95 al., 1991; Dobrea et al., 1992) led to suggest its physiological role on myelination, as well. The strong pro-myelinating effects of CNTF and other members of the family such as leukemia 96 inhibitory factor (LIF) in myelinating co-cultures further supported this hypothesis. CNTF acts 97 on oligodendrocytes by favoring their final maturation via a mechanism involving the 130 kDa 98 glycoprotein receptor common to the CNTF family and transduced through the Janus kinase 99 (JAK) pathway (Stankoff et al., 2002). In contrast, other astrocyte-derived secreted proteins 100 negatively regulate oligodendrocyte biology. For instance, the chemokine CXCL1 transiently 101 expressed at a high level during spinal cord development signals through the chemokine 102 receptor CXCR2 expressed by immature OPCs. Migrating OPCs were proposed to enter the 103 presumptive white matter where they may encounter an environment in which astrocytes 104 105 transiently and locally express high CXCL1 levels. Through CXCR2, CXCL1 may inhibit PDGF-stimulated OPC migration before subsequently increasing OPC proliferation in concert 106 107 with PDGF (Tsai et al., 2002).

108

Astrocytes control the availability of cues regulating oligodendrocyte production

Besides the secretion of trophic factors, astrocytes are also able to control the bioavailability of oligodendrocyte-regulating cues (Fig. 1A). The best documented exemple was provided by the capacity of astrocytes to control the concentration of the morphogen Sonic Hedgehog (Shh) during OPC production in the optic nerve. Shh participates in the proliferation and migration of OPCs during the colonization of this nerve (Merchan et al., 2007). The multiligand receptor megalin, a member of the low density lipoprotein receptor family able to bind Shh, was found to be exclusively expressed by astrocytes according to a dynamic pattern paralleling optic nerve colonization by OPCs arising from the optic chiasm and migrating to the retina. Indeed, when OPCs start their migration throughout the nerve at E14,5 in the mouse, megalin is more widely distributed in the region close to the optic chiasm whereas the distribution is reversed at E16.5 when the first OPCs reach the retina. Since, thereafter, megalin was found to be weakly and uniformly expressed all along the nerve, this receptor was proposed to control Shh internalization and its subsequent release at the suitable concentration during the various steps leading to oligodendrogenesis (Ortega et al., 2012).

123 Astrocyte integrity is required for proper myelination

Consistent with the above findings, the requirement of astrocyte integrity for the normal 124 development of oligodendrocytes was shown by the analysis of transgenic mouse strains devoid 125 of GFAP expression. Although viable, GFAP knockout mice exhibit abnormal myelination 126 including the presence of actively myelinating oligodendrocytes in adults, nonmyelinated axons 127 in optic nerve, reduced myelin thickness in spinal cord and ultrastructural defects such as 128 loosening of myelin sheaths (Liedtke et al., 1996). More recently, the invalidation of the 129 transmembrane protein CD38, which possess ADP-ribosyl cyclase activity and is highly 130 expressed in astrocytes, was similarly reported to result in altered astrocyte maturation and 131 delayed oligodendrocyte differentiation during the postnatal development (Hattori et al., 2017). 132 Furthermore, physical interactions between oligodendrocytes and astrocytes appear to be also 133 required for myelination as shown by analysis of the role of connexins (Cx) 30 and Cx43 present 134 on astrocytes and forming gap junctions with Cx32 and Cx47 present on oligodendrocytes. 135 First, Cx47 mutants that cause the Pelizaeus-Merzbacher-like disease do not efficiently form 136 functional channels with Cx43 (Orthmann-Murphy et al., 2007). Second, the deletion of both 137 Cx47 and Cx30 in mice induces severe myelination defects including vacuole formation and 138 thin myelin sheaths as well as a decreased number of oligodendrocytes (Tress et al., 2012). A 139 mechanism possibly involved is proposed to be disruption of the metabolic support provided 140 by astrocytes to oligodendrocytes in agreement with the gap junction-mediated unidirectional 141 flux that transports the molecules preferentially from astrocytes to oligodendrocytes (Robinson 142 et al., 1993). 143

144 Astrocytes are indispensable for axon myelination

Accurate delineation of the molecular mechanisms supporting the role of astrocytes in 145 myelination itself has never ceased for the last decades. Among the most relevant findings, the 146 requirement of astrocytes for alignment and adherence of oligodendrocyte processes to axons 147 was shown by using cocultures of retinal ganglion cells and optic nerve oligodendrocytes that 148 led to propose a mechanism implicating neuroaminidase-mediated removal of polysialic acid 149 (PSA) from both cell types (Meyer-Franke et al., 1999). In agreement with this result, abolition 150 of the postnatal downregulation of PSA in oligodendrocytes from transgenic mice expressing 151 the polysialyltransferase ST8SiaIV under the control of the proteolipid protein promoter, 152 153 indicated that PSA downregulation is required for efficient differentiation of oligodendrocytes, as well as for myelin formation and maintenance. However, in vivo implication of astrocytes is 154 still lacking (Fewou et al., 2007). Another link between astrocytes and myelination was 155 established by the demonstration that electrical activity in premyelinated axons increases 156 myelination after OPCs mature to a promyelinating stage via the activity-dependent release of 157 ATP from axons. ATP acts in a paracrine manner on astrocytes via the purinergic P2 receptor 158 159 and induces the release of the LIF cytokine that ultimately stimulates myelination (Ishibashi et al., 2006). A specific myelinating CNS coculture system provided evidence that astrocytes are
implicated rather in promotion of rapid myelin growth than in initiation of myelination
(Watkins et al., 2008).

163

Astrocytes provide lipids for myelin sheath production

164 Still consistent with their implication in myelination, astrocytes were also proposed to supply lactate to oligodendrocytes. Lactate constitutes a source of energy and a precursor of 165 lipid synthesis including cholesterol altogether necessary for myelin production. Although 166 astrocytes and oligodendrocytes are both in direct contact with blood vessels and may thus take 167 up lactate from blood, the hypothesis is that lactate released from astrocytes through the 168 monocarboxylate transporter (MCT) 4 may be supplied to oligodendrocytes possibly importing 169 the molecule through MCT1 (Sanchez-Abarca et al., 2001;Rinholm et al., 2011). The 170 requirement for local synthesis of cholesterol is consistent with the fact that myelin comprises 171 about 80% of the brain cholesterol content and that peripheral cholesterol entry into the brain 172 is largely precluded by the blood-brain barrier. During postnatal myelination, de novo synthesis 173 of cholesterol is carried out by both astrocytes and oligodendrocytes. Astrocyte-derived 174 cholesterol is then distributed to oligodendrocytes via lipoproteins contained in apolipoproteins, 175 mostly the apolipoprotein E (ApoE) in the CNS, and secreted via the ATP-binding cassette 176 transporter ABCA1 (Saher and Stumpf, 2015). Astrocytes provide a substantial fraction of the 177 lipids incorporated into CNS myelin and in the absence of astrocyte lipid synthesis, 178 oligodendrocytes are unable to finalize CNS myelination, leading to hypomyelinated and 179 slower-conducting fibers in adulthood. Indeed, the specific inactivation of an essential co-180 activator of the transcription factor SREBP, the SREB cleavage activating protein (SCAP) in 181 oligodendrocytes, resulted in lipid biosynthesis and myelination delay nevertheless recovering 182 in adulthood. In contrast, when the enzyme was deleted in astrocytes or in both astrocytes and 183 oligodendrocytes, a persistent hypomyelination was observed. Thus, extracellular lipids seem 184 to be supplied by astrocytes under conditions of compromised oligodendrocyte lipid synthesis. 185 Moreover, full myelin synthesis requires an astrocyte lipid supply in addition to endogenous 186 oligodendrocyte lipid synthesis (Camargo et al., 2017). 187

Besides providing lipids for synthesis of myelin sheaths during development, astrocytes, 188 namely those contacting the nodes of Ranvier, were also reported to reversibly modify myelin 189 thickness and nodal gap length thus appropriately regulating conduction velocity in individual 190 axons. In support of this observation, the reduction of exocytosis induced in transgenic mice 191 expressing a dominant-negative fragment of the vesicle-associated membrane protein 2 192 (VAMP2) in astrocytes, exhibited detachment of adjacent paranodal loops of myelin from the 193 axon, increased nodal gap length, thinning of myelin sheath in the optic nerve and finally 194 decrease in visual acuity. These data led to propose that thrombin-dependent proteolysis of the 195 196 cell adhesion molecule neurofascin 155 that attaches myelin to the axon, is inhibited by 197 vesicular release of thrombin protease inhibitors from perinodal astrocytes, which likely involves these cells in myelin remodeling necessary for optimal electrical conduction (Dutta et 198 199 al., 2018).

A last evidence of the remarkable and specific relationship between astrocytes and myelination is finally provided in a model of neuromyelitis optica, a chronic inflammatory demyelinating disease characterized by the destruction of astrocytes and their foot processes in early lesions. Indeed, in a model of this disease, the death of oligodendrocytes was found to

- 204 occur before granulocyte or macrophage / microglia infiltration, but only a few hours after the
- death of astrocytes induced in a complement-dependent manner (Wrzos et al., 2014).
- 206

207 Microglia-oligodendrocyte communication in developmental myelin production

Considered as resident myeloid cells of the CNS, microglia have been only recently characterized as the progeny of yolk sac-derived macrophages that start to enter the CNS at E9 through the blood vasculature before closure of the blood-brain barrier that occurs by E13 in mice (Ginhoux et al., 2010;Schulz et al., 2012;Kierdorf et al., 2013). Thereafter, at least under non-pathological conditions, microglia cells are autonomously maintained through proliferation (Askew et al., 2017). Microglia is crucial during neurodevelopment namely via its interaction with neuronal cells for wiring and neural circuit regulation (Thion et al., 2018).

Distinct patterns of secreted molecules for microglia and astrocyte-mediated control of OPC generation

217 The first arguments supporting a role for microglia in developmental myelination were 218 mainly derived from microglia-oligodendrocyte cocultures. They showed that microglia was 219 able to stimulate the synthesis of sulfatide, a myelin-specific galactolipid, and to increase myelin-specific proteins, MBP and PLP in oligodendrocytes (Hamilton and Rome, 1994). 220 221 Microglia activity was mimicked by conditioned medium derived from microglia cultures 222 suggesting that the cells acted via the secretion of appropriate molecules. Together with promoting cell maturation, microglia was found to prevent OPC apoptosis by upregulating 223 nuclear factor-kappa B (NF-kB) p65 subunit via the recruitment of PI-3 kinase to the PDGFRa 224 thus leading to a synergism with PDGF (Nicholas et al., 2001). 225

226 Although both microglia- and astrocyte-conditioned media were able to prevent OPC 227 degeneration induced by growth factor deprivation in the culture medium during a short time, 228 microglia-derived medium was unable to support OPC survival in the long-term compared to 229 astrocytes. These differential effects were supported by distinct patterns of cytokine/growth factors correlated with differentially activated intracellular signaling pathways in OPCs (Pang 230 et al., 2013). More specifically, mechanisms underlying enhanced microglia-induced 231 oligodendrocyte differentiation were proposed to be related to IGF-1. IGF-1 levels were more 232 than six-fold higher in microglia- than in astrocyte-derived medium and its expression was 233 consistently detected in amoeboid microglial cells located in the corpus callosum until the 234 seventh postnatal day, which corresponds to the time of morphological change of microglia 235 towards the ramified phenotype present in adulthood unless microglia become activated. In a 236 consistent manner, exposure of microglia cultures to lipopolysaccharide leading to microglia 237 activation, increased IGF-1 secretion (Kaur et al., 2006) and IGF-1 knockout mice display 238 reduced oligodendrocyte survival, differentiation and maturation (Ye et al., 2002). Moreover, 239 IGF-2 also derived from microglia was found to prevent TNFa-induced death of mature 240 oligodendrocytes in vitro (Nicholas et al., 2002). 241

242

Unique phenotype of neonatal microglia implicated in developmental myelination

As soon as the end of 1970's, the existence of a specific subpopulation of microglial cells had been proposed in the early postnatal corpus callosum (Imamoto and Leblond, 1978). Unexpectedly, the pharmacological suppression of microglia activation by using the antiinflammatory drug minocycline, was reported to significantly inhibit oligodendrogenesis both *in vitro* in neurosphere culture and *in vivo* in the subventricular zone (SVZ) of the dorsal forebrain. This inhibition was mediated by the blockade of several proinflammatory cytokines including IL-1β, IL-6, TNF- α , and IFN- γ . In addition, the observation that activated microglia significantly increased O4⁺ cells but decreased PDGFRa⁺ OPCs (Fig. 1B) again confirmed microglia activity on oligodendrocyte maturation (Shigemoto-Mogami et al., 2014).

More recent data provided by two independent groups led to a major advance in the 252 understanding of the identity and role of this microglial subopulation that appeared to highly 253 proliferate and display a rhomboid shape in contrast to microglia detected at the same time in 254 the neighboring cortex and exhibiting small processes comparable to adult microglia 255 256 (Hagemeyer et al., 2017; Wlodarczyk et al., 2017). Presently, neonatal microglia implicated in developmental oligodendrogenesis appear as a cell population with a dramatically different 257 gene expression profile compared to the one from adult healthy mice. Based on the observation 258 that during neuroinflammation, microglia expressing the integrin complement receptor CD11c 259 are a major source of IGF-1, Wlodarczyk and collaborators detected a substantial increase of 260 CD11c⁺ microglia during the first days after birth before a sharp decrease mainly in highly 261 myelinating areas such as the developing corpus callosum and cerebellum. IGF-1 depletion in 262 this subset led to a reduction in brain weight, a decrease in the expression of myelin proteins 263 including PLP, MBP, MAG and MOG, and was finally associated with higher frequency of less 264 myelinated fibers in the corpus callosum. Thus, as the major source of IGF-1, the CD11c⁺ 265 microglia subset was proposed to play a critical role in primary myelination and neuronal 266 support in the neonatal CNS (Wlodarczyk et al., 2017). Interestingly, the comparison of 267 transcriptomes from sorted CD11c⁺ and CD11c⁻ microglia revealed that neonatal microglia, 268 naive adult microglia and microglia derived from the experimental autoimmune 269 encephalomyelitis (EAE) model of demyelination formed distinct global expression clusters. 270 Unexpectedly, CD11c⁺ and CD11c⁻ subpopulations were relatively close together indicating 271 that the major difference was related to developmental age rather than subpopulation 272 273 phenotype. The identification of gene expression profiles associated with microglia from the different conditions showed that under demyelinating conditions, microglia cells were enriched 274 for immune system genes consistent with their activated state. On the contrary, in the neonatal 275 brain, microglia gene profile included genes involved in nervous system development 276 277 displaying a neurogenic phenotype. Remarkably, upon a quite total ablation of microglia in the 278 adult brain by using an inducible mouse strain, clusters of highly proliferating microglia were detectable throughout the CNS. Although expressing CD11c and nestin, the proliferating cells 279 did not display a neurogenic gene expression profile confirming that the myelinogenic CD11c⁺ 280 subset of microglia is unique to neonatal CNS (Wlodarczyk et al., 2017). 281

282 Independently, Hagemeyer and collaborators similarly identified this amoeboid microglia subset that they called 'the fountain of microglia' specifically located in myelinating 283 regions from postnatal day (P)1 to P8, displaying a high expression of the activation marker 284 Mac3 and dramatically collapsing at P9. Microglia found in the postnatal subcortical (with an 285 amoeboid shape) and cortical regions (with multiple cellular processes) were found to share 286 their early origin from the same CX3CR1⁺ CNS endogenous precursors without any 287 contribution from circulating blood monocytes. The pharmacological blockade of the receptor 288 for the secreted cytokine, colony stimulating factor 1 (CSF1), known to efficiently deplete 289 microglia (Elmore et al., 2014), showed that early postnatal microglia is required for the proper 290 induction of oligodendrocyte progenitors and subsequent myelination. Transcriptomic analysis 291

of the studied subpopulation revealed genes also detected in disease models such as CLEC7a, 292 SPP1, IGF-1, ANXA5, ITGAX, and GPNMB. An additional RNA sequencing analysis led to 293 identify change in gene expression profile between P7 and P10, the former being related to 294 phagocytosis, migration, priming of microglia more particularly described during aging and 295 disease while the latter was associated with apoptosis and necrosis according to a profile further 296 increased in adulthood. Thus, a specificity of neonatal microglia in P7 corpus callosum was the 297 expression of SPP1, CLEC7A and CD206 since none of them could be detected in cortical 298 microglia (Staszewski and Hagemeyer, 2019). Importantly, functional microglia is also 299 required for OPC homeostasis in the adult brain in agreement with the severe white matter 300 301 abnormalities observed in patients suffering from microgliopathies (Hagemeyer et al., 2017).

302

The extracellular matrix at the crossroads of microglia and OPC activity

The investigation of how OPCs integrate signals from the matrix and other glial cells 303 led to a growing interest for the adhesion G protein-coupled receptor (aGPCR) family, the 304 second largest class of GPCRs playing crucial roles in developmental processes (Mehta and 305 Piao, 2017). The finding that the genetic loss of one of its members, GPR56, was phenocopyied 306 by the specific deletion of the enzyme microglia-derived transglutaminase 2 (TG2) in microglia 307 both resulting in the decrease of OPC cell division and number of mature oligodendrocytes, as 308 well as to hypomyelination during postnatal CNS development, led to identify GPR56 as the 309 receptor of TG2. In agreement with the ability of TG2 to bind ECM proteins to exert its 310 crosslinking activity, the activation of GPR56 by TG2 was found to require the ECM protein 311 laminin to promote OPC proliferation according to a mechanism leading to the dissociation of 312 GPR56 N- and C-terminal fragments allowing the agonist to initiate G-protein signaling and 313 subsequently RhoA activation. Thus, microglia TG2, extracellular laminin and OPC GPR56 314 provide an additional evidence of the critical role of microglia / oligodendrocyte 315 communication for developmental oligodendrogenesis (Giera et al., 2015). Remarkably, 316 laminin is also involved in OPC proliferation via promoting the metalloproteinase-mediated 317 cleavage of dystroglycan, one of its OPC surface receptors (Leiton et al., 2015). Moreover, 318 other ECM glycoproteins play a crucial role in myelination as, for instance, anosmin-1 319 characterized as an important modulator of oligodendrocyte progenitor lineage progression 320 likely via FGFR1/ERK1/2 signaling activation, which results in the control of MBP expression, 321 myelin formation and conduction velocity (Murcia-Belmonte et al., 2016). The enhanced 322 differentiation of induced pluripotent stem cells into myelin-expressing oligodendrocytes 323 promoted by the functionalization of culture substrates using brain ECM prepared from 324 325 decellularized human brain tissue further supports the critical activity of ECM in myelination (Cho et al., 2019). 326

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328 Influence of astrocytes during remyelination

329

Reactive astrocytes provide pro-regenerative trophic factors

As in other diseases, astrocytes become reactive after CNS demyelination. Astrocyte reactivity includes hypertrophy of the cells and upregulation of intermediate filament proteins such as GFAP and vimentin. The requirement of astrocytes for oligodendrocyte-mediated remyelination has been established more than 15 years ago (Blakemore et al., 2003;Talbott et al., 2005) and more recently reported to depend on the presence of androgen hormones in male

mice (Bielecki et al., 2016). The delay in astrocyte activation in mice lacking IL-1 β and the 335 finding that activated microglia release II-1 β , were the first arguments supporting the idea that 336 microglia activation is a pre-requisite for astrocyte activation. In a consistent manner, 337 microglia-derived IL-1ß is able to upregulate CNTF. Added to astrocyte cultures, CNTF 338 induces astrocyte activation (Herx et al., 2000;Albrecht et al., 2003), which promotes fibre 339 myelination in vitro (Nash et al., 2011). In vivo, astrocytes derived from mice infected with the 340 mouse hepatitis virus, a model for multiple sclerosis (MS), secrete high levels of CNTF during 341 the remyelination phase while CNTF injection in the spinal cord upregulates the transcripts 342 encoding FGF2, a potent OPC mitogen (Albrecht et al., 2003). The loss of astrocytic gp130 343 receptor, the ubiquitous signal transducer for CNTF, consistently exacerbates both 344 demyelination and the proinflammatory T cell infiltration in the EAE model via apoptosis of 345 astrocytes, decrease of CNS regulatory Foxp3-expressing T cells and increase of IL-17-, IFNy-346 and TNF-producing T cells. The SHP2/Ras/ERK pathway was then identified as the involved 347 intracellular mechanism (Haroon et al., 2011). 348

IL-1 β also stimulates the astroglial production of LIF promoting survival of 349 oligodendrocytes and thus decreasing disease severity in EAE mice (Butzkueven et al., 2002). 350 In a pathological context where tumor necrosis factor (TNF) is a key component of the 351 inflammatory response, LIF can alternatively be released upon TNFR2-mediated activation of 352 the PI3K-PKB/Akt pathway in primary astrocytes. The selective stimulation of TNFR2 on 353 astrocytes cocultured with OPCs promoted OPC differentiation into mature oligodendrocytes 354 while the process was blocked in the presence of LIF neutralizing antibodies (Fischer et al., 355 2014). Reactive astrocytes also produce BDNF supporting oligodendrogenesis and regeneration 356 after white matter damage. In vitro, conditioned medium from astrocytes restored the process 357 of OPC maturation even under an hypoxic stress known to block OPC differentiation unless the 358 medium was specifically treated in order to remove BDNF. Similarly, in vivo, the conditional 359 astroglial deletion of BDNF led to a highly reduced number of newly generated 360 oligodendrocytes and thus to larger white matter damage in animals subjected to prolonged 361 cerebral hypoperfusion (Miyamoto et al., 2015). 362

The multitude of factors secreted by astrocytes are proposed to induce a rapid change in 363 the lesion environment sensed by OPCs, which thereby become activated. Upon demyelination, 364 OPC activation is associated with a reversion of gene expression profile of adult OPCs towards 365 a profile more closely resembling that of neonatal OPCs. A notable increase in the expression 366 of genes associated with innate immune system functions (including IL1ß and CCL2) was 367 identified, as well, together with an increased ability of activated OPCs to migrate and 368 369 differentiate compared with non-activated ones. Activated OPCs finally express transcription 370 factors such as TCF4 and Sox2, which serve to maintain OPC in the cell cycle and to prime 371 these cells for further differentiation (Moyon et al., 2015;Franklin and ffrench-Constant, 2017).

372

Pro-inflammatory secreted molecules associated with reactive astrocytes

All the above activities derived from reactive astrocytes are likely involved in the spontaneous regenerative process of remyelination that can occur with a remarkable efficiency, not only in experimental models of demyelination but also in humans suffering from MS (Prineas et al., 1993;Patrikios et al., 2006;Patani et al., 2007;Franklin and ffrench-Constant, 2017). However, in chronic lesions, remyelination mostly fails or occurs only in the periphery of the plaques (Prineas and Connell, 1979;Barkhof et al., 2003;Bramow et al., 2010), where

OPC migration and / or differentiation are likely impaired (de Castro et al., 2013;Franklin and 379 ffrench-Constant, 2017). Such failure has been associated with deleterious activities of 380 astrocytes classically related to a more severe level of reactivity and the secretion of damaging 381 molecules (Fig. 2). One of these molecules, $TNF-\alpha$, was mainly detected in fibrous astrocytes 382 at the periphery of chronic active MS lesions and mostly transcribed in highly demyelinated 383 plaques (Hofman et al., 1989;Selmaj et al., 1991;Bitsch et al., 2000). Although it is not yet clear 384 whether TNF- α is internalized or produced by astrocytes (Selmaj et al., 1991; Aranguez et al., 385 1995), the direct physical contact between astrocytes and oligodendrocytes was proposed to be 386 necessary for oligodendrocyte apoptosis by TNF- α in glial cell cultures. One possibility for 387 contact-dependent cell killing is through gap junctions known to couple astrocytes and OLs 388 (Nagy and Rash, 2000;Orthmann-Murphy et al., 2007) and previously suggested for 389 propagation of cell injury (Lin et al., 1998; Farahani et al., 2005; Froger et al., 2010; Kim et al., 390 2011). 391

In the same manner, the knockout of IFN- γ receptor in astrocytes was found to decrease 392 chemokine expression and inflammatory cell infiltration thus preventing demyelination and 393 consequently lowering clinical signs, namely by alleviating both Th1- and Th17-mediated 394 adoptive EAE (Ding et al., 2015). Together with suggesting an important role for IFN- γ 395 signaling in astrocytes during autoimmune CNS inflammation, this phenotype was consistent 396 with the delay in disease recovery observed in transgenic EAE mice ectopically expressing IFN-397 γ in astrocytes during the recovery stage and also with the dramatic reduction of 398 oligodendroglial repopulation in the demyelinated lesions of the cuprizone model upon CNS 399 delivery of IFN- γ (Lin et al., 2006). The detection of IFN- γ in astrocytes of active chronic MS 400 plaques and its ability to induce class II major histocompatibility antigen HLA-DR-Ia in 401 astrocytes *in vivo* suggested that IFN- γ may play an important role in the development of MS 402 lesions (Traugott and Lebon, 1988; Hashioka et al., 2009). 403

Another cytokine secreted by astrocytes is CXCL10 that is namely detectable around active MS lesions (Ransohoff et al., 1993;Omari et al., 2005;Carter et al., 2007). Its production increases before the onset of clinical symptoms, maintains at high levels during the peak of the disease and decreases during remission in the EAE model (Glabinski et al., 1997;Fife et al., 2001). *In vitro*, CXCL10 is upregulated in an astrocyte phenotype endowed with an inhibitory role in glial/axonal ensheathment without impairment of OPC proliferation / differentiation or process extension (Nash et al., 2011).

411 Among the other molecules highly expressed by reactive astrocytes, endothelin-1 also stands as a strong inhibitor of remyelination. This neuropeptide acts by promoting Notch 412 activation in OPCs through induction of Jagged1 expression in reactive astrocytes (Hammond 413 et al., 2014). In the same line, lesion formation was reported to be associated with increased 414 expression of fibroblast growth factor 9 (FGF9) by astrocytes acting not via a direct effect on 415 the oligodendroglial lineage, but via an off target effect mediated by soluble astrocyte-derived 416 factors inhibiting terminal differentiation of myelinating oligodendrocytes. In vitro, this activity 417 418 is associated with the appearance of multi-branched 'pre-myelinating' oligodendrocytes that 419 extend processes able to interact with axons without forming any myelin sheaths, reminiscent of the oligodendrocyte phenotype observed in chronically demyelinated MS lesions. Astrocyte-420 derived factors included metalloproteases associated with ECM remodelling and the pro-421 422 inflammatory chemokines CCL2 and CCL7 that contribute to the development of inflammatory 423 responses in the CNS (Lindner et al., 2015).

424 **Reactive astrocytes secrete extracellular matrix deleterious for remyelination**

425 Apart from cytokines, growth factors and neuropeptides, ECM molecules are also secreted by reactive astrocytes and highly modify the lesion environment in MS, consequently 426 influencing the behavior of OPCs (Maier et al., 2005;van Horssen et al., 2005;van Horssen et 427 al., 2006;Clemente et al., 2011;Pu et al., 2018). Both the receptors present at the surface of 428 OPCs and the ECM molecules secreted by reactive astrocytes are susceptible to change the 429 environment from permissive to inhibitory for remyelination (de Castro et al., 2013). Indeed, 430 except the interaction reported between astroglial laminin and oligodendroglial $\alpha 6 \beta 1$ integrin 431 shown to attenuate oligodendrocyte death in vitro (Corley et al., 2001), ECM accumulation 432 appears as an important factor in tissue regeneration failure. For instance, the expression of N-433 cadherin on the surface of both oligodendrocytes and astrocytes led to the first in vitro evidence 434 that this molecule was in part responsible for the poor migration-promoting properties induced 435 by astrocytes on oligodendrocytes suggesting that the inhibition of cell-cell adhesion could 436 improve repopulation of MS lesions by OPCs. Indeed, the blocking of cadherin function by 437 specific peptides reduced adhesion of oligodendroglia to astrocyte monolayers, diminished the 438 duration of the contact between oligodendrocyte processes and individual astrocytes, and 439 increased the migration of oligodendrocytes on astrocyte monolayers (Schnadelbach et al., 440 2000). In the same line, astrocytes in culture produce fibronectin as fibril-like structures in 441 inflammatory conditions, known to inhibit myelin formation in different experimental 442 paradigms (Siskova et al., 2009). In agreement with this hypothesis, fibronectin rapidly 443 accumulates as an acute response to demyelination in MS and disappears during remyelination, 444 whereas astrocyte-released aggregated fibronectin persists in chronic lesions (Stoffels et al., 445 2013). Similarly, the ECM-associated glycoprotein anosmin-1 can also be detected in the core 446 of chronic active and inactive MS plaques where it is suggested to prevent OPC colonization 447 rather than inhibit their differentiation via a mechanism possibly involving the FGF-2 receptor 448 FGFR1 consistently detected in the periplaque of chronic lesions (Bribian et al., 2006;Bribian 449 et al., 2008;Clemente et al., 2011). 450

Major CNS ECM proteoglycans have also been characterized in MS lesions. In active 451 plaque edges, the chondroitin sulfate proteoglycans (CSPG) versican, aggrecan, neurocan and 452 the dermatan sulfate proteoglycan increase in correlation with astrocytosis. In contrast, these 453 molecules accumulate in foamy macrophages in active plaque centers, suggesting their 454 engulfement together with myelin. In inactive lesions and normal-appearing white matter, 455 proteoglycans are decreased and display undetectable abnormal heterogeneous aggregation, 456 457 respectively (Sobel and Ahmed, 2001). The roles of CSPG during development in particular in myelination and the different localizations of CSPG according to the type of MS lesions both 458 support the idea that these molecules comprising part of the astrogliotic scar, are critical in 459 inhibition of OPC processes outgrowth and OPC differentiation (Siebert and Osterhout, 2011). 460 The hypothesis is consistent with the recent design of a novel CSPG synthesis inhibitor reducing 461 CSPG deposition into the lesion microenvironment and found to rescue OPC process outgrowth 462 in vitro and to accelerate remyelination following focal demyelination in mice (Keough et al., 463 2016). Moreover, the glycosaminoglycan hyaluronan characterized by its ability to bind CSPG 464 465 is deposited in early MS lesions in a low molecular weight form by lymphocytes and microglia / macrophages whereas it is deposited by astrocytes in a higher molecular weight form in chronic 466 lesions. There, hyaluronan degraded by hyaluronidases expressed by OPCs inhibits OPC 467 maturation and thus remyelination (Back et al., 2005) through a mechanism requiring the Toll-468 like receptor 2 expressed by oligodendrocytes and upregulated in MS lesions (Sloane et al., 469

2010). Evaluated in the Theiler's murine encephalomyelitis model, the spatio-temporal course
of ECM alterations confirmed the correlation of matrix accumulation with astrogliosis still
supporting a mainly astrocytic origin of ECM deposits. The data led to propose disturbed
aggregation or post-translational modifications of matrix molecules leading to impairment of
their regular degradation rather than changes in their transcription level (Haist et al., 2012).

475 The dual activity of astrocytes in the demyelinated tissue

The dual activity of astrocytes described above not only reflects more or less severe 476 levels of astrocyte reactivity, but also clearly reminds the duality that characterizes the glial 477 478 scar itself, which is considered as a rearrangement of tissue structure, astrocyte proliferation, and pronounced overlap of astrocyte processes resulting in the disruption of individual astrocyte 479 domains (Zamanian et al., 2012;Anderson et al., 2016;Adams and Gallo, 2018). While 480 traditionally viewed as a barrier to axon regeneration, the glial scar has been nevertheless 481 reported to be beneficial for restricting leukocyte migration outside the damaged tissue 482 (Faulkner et al., 2004; Okada et al., 2006; Herrmann et al., 2008; Voskuhl et al., 2009). Moreover, 483 in contrast to the initial dogma, its deleterious effect on axon regeneration was recently 484 questioned. Indeed, genetically targeted loss-of-function approaches leading to prevention of 485 astrocyte scar formation, attenuation of scar forming astrocytes or deletion of chronic astrocyte 486 scars, failed to result in spontaneous regrowth of transected CNS axons following spinal cord 487 injury lesions. These data were corroborated by results of RNA sequencing revealing the 488 expression of multiple axon-growth supporting molecules namely in astrocytes (Anderson et 489 al., 2016). 490

Astrocyte ability to exhibit different phenotypes obviously participates in this 491 heterogeneity (Zamanian et al., 2012;Liddelow and Barres, 2017). Indeed, despite a core set of 492 genes up-regulated in different injury models such as ischemic stroke and neuroinflammation, 493 50% of altered gene expression is specific to a given injury. The first transcriptomic analyses 494 provided evidence for the existence of two subtypes of reactive astrocytes (Zamanian et al., 495 2012) afterwards named 'A1' and 'A2'. A1 astrocytes induced by classically activated 496 neuroinflammatory microglia following exposure to lipopolysaccharide have been described as 497 harmful by inhibiting OPC proliferation and differentiation. A2 astrocytes identified during the 498 regeneration stage in an ischemia model, were found to up-regulate genes thought to be 499 protective. In line with this observation, analysis of MS lesions reported the detection of A1 500 mostly in active lesions whereas A2 were found during remyelination (Haindl et al., 2019). 501 However, it may be more likely that the occurrence of different astrocytic phenotypes and the 502 existence of suitable levels of various factors expressed at particular time points may achieve 503 successful repair. In agreement with this hypothesis, investigation of the astrocytic reaction at 504 various steps of lesion progression in the rat EAE model led to detect simultaneously in shadow 505 plaques all features of a glial scar and densities of OPCs and mature oligodendrocytes, which 506 507 were quite comparable to the densities observed in unaffected white matter (Haindl et al., 2019).

The first line response of astrocytes to myelin injury consisting in myelin phagocytosis was similarly considered to be either beneficial or detrimental on the lesion pathology, depending on the inflammatory context. Indeed, although myelin debris phagocytosis was mainly reported to be performed by microglia/macrophages, myelin-positive hypertrophic astrocytes can be observed at sites of acute myelin breakdown. The uptake was proposed to rely on receptor-mediated endocytosis and to result in astroglial NF-kB activation and secretion of 514 specific chemokines. The latter leads to recruitment of immune cells shown *in vitro* in rodent 515 and validated in human disease. Although microglia and monocyte recruitment may be 516 beneficial for myelin clearance in a non-inflammatory environment, it increases tissue damage 517 in demyelinating conditions driven by inflammation such as MS since additional recruitment of 518 lymphocytes and microglia/macrophages exacerbates the inflammatory process (Ponath et al., 519 2017).

Finally, the finding that transplanted immature astrocytes are unable to become reactive 520 after CNS injury (Jiang et al., 2013) and that only immature (but not mature) astrocytes are 521 neuroprotective and suppress endogenous astrocyte activation and glial scar formation 522 presently stands as an interesting novel concept (Chen et al., 2015). Thus, transplanted human 523 524 pluripotent stem cell-derived astrocytes at a defined immature stage were shown to regulate the 525 differentiation of endogenous OPCs, to promote myelinogenesis and to improve behavioral outcome in the model of periventricular leukomalacia in neonate mice via a mechanism 526 depending on the tissue inhibitor of metalloproteinases TIMP-1 (Jiang et al., 2016). 527

528 Key studies supporting detrimental roles of astrocytes on oligodendrocytes and 529 myelin

Several publications using astrocyte ablation have improved our knowledge about the 530 main detrimental roles of reactive astrocytes in the context of CNS demyelination. For instance, 531 OPC transplantation in ethidium bromide-demyelinated animals in the presence or absence of 532 astrocytes indicated that astrocyte-free regions favour Schwann cell differentiation whereas the 533 presence of astrocytes delayed the interaction of OPCs with the demyelinated axons (Blakemore 534 et al., 2003). Similarly, the depletion of astrocytes via intracallosal injection of La-aminoadipate 535 in cuprizone-treated animals revealed a notable increase in the percentage of myelinated areas, 536 decrease in Iba-1⁺ microglia staining and collapse in the expression of genes related to either 537 recruitment of microglia classicaly triggered by astrocytes or suppression of OPC 538 differentiation (Madadi et al., 2019). The considerable amount of fibronectin produced by 539 astrocytes and the ability of astrocytic fibronectin to become aggregated after treatment with 540 lipopolysaccharide led to conclude to the deleterious effects of such aggregates on 541 542 oligodendrocyte differentiation and myelin regeneration in vivo, in agreement with the detection of a low level of fibronectin aggregates in remyelinated MS lesions (Stoffels et al., 2013). The 543 signaling molecule endothelin-1 (ET-1) expressed by reactive astrocytes in MS and murine 544 demyelinated lesions is a negative regulator of OPC differentiation and remyelination acting by 545 promoting Notch activation in OPCs during remyelination (Hammond et al., 2014). 546

547 Moreover, the CCL2 chemokine produced by astrocytes plays an important role in the continued recruitment of immune cells and activation of glial cells in the CNS during chronic 548 EAE (Kim et al., 2014). Similarly, the adhesion molecule VCAM-1 expressed by astrocytes is 549 essential for T cell entry into the CNS parenchyma from EAE animals (Gimenez et al., 2004). 550 551 As previously mentionned, myelin uptake is an early response of astrocytes in diseases with prominent myelin injury that results in recruitment of immune cells possibly increasing tissue 552 damage in demyelinating conditions driven by inflammation (Ponath et al., 2017). In the same 553 554 line, astrocytes constitute one of the cell types that have the capacity to express molecules 555 required for antigen presentation under inflammatory conditions required for T cell reactivation targeting the myelin sheath-forming oligodendrocytes (Waisman and Johann, 2018). Finally, 556 557 although in the healthy brain, perivascular astrocyte end-feet closely ensheath the 558 microvasculature, activation of perivascular astrocytes may alter their involvement in the blood-

brain barrier formation as suggested by the damage of perivascular end-feet reported in MS

560 patients. The revisit of this observation in the EAE model showed that reactive astrocytes detach

from the blood vessels and lose their contacts with both blood vessels and neuronal synapses, which was proposed to contribute to the neurological impairment and the cognitive decline occurring in EAE/MS as well as to the neurodegenerative disease progression (Eilam et al.,

- 564 2018).
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566 Microglia in remyelination

In the context of myelin repair, microglia and their hematopoietic cell-derived counterparts, macrophages, both contribute to the regenerative process through key effects including the phagocytosis of myelin debris, the secretion of cytokines, chemokines, growth factors or soluble mediators and the recruitment/differentiation of progenitors at the lesion site. Although microglia and macrophages can now be distinguished by using a few specific markers (Bennett et al., 2018), they have been indistinguishable for a long time. Therefore, 'microglia/macrophages' will be used for most presently available data.

574 Microglia/macrophage phenotypes upon CNS demyelination

575 As resident immune cells of the CNS, microglia/macrophages become activated as soon 576 as they detect any insult. This activation implicates changes in morphology and transcriptional 577 profiles of the cells. The failure of microglia to convert from a 'resting state' with a branched-578 like morphology to an 'activated state' with an amoeboid-like morphology was namely 579 observed upon knocking down either the β -galactoside-binding lectin Galectin-3 (Reichert and 580 Rotshenker, 2019) or one of the receptors of the secreted proteins Sonic hedgehog, the protein 581 Boc (Zakaria et al., 2019).

582 Like astrocytes, microglia/macrophages are endowed with a dual activity supported by the existence of different phenotypes observed under various in vitro conditions and that have 583 been named, respectively the classical activated state (M1) associated with pro-inflammatory 584 activities and the alternative polarized state (M2) involved in OPC recruitment and 585 differentiation associated with anti-inflammatory activities (Fig. 3A). The characterization of 586 these phenotypes performed in the model of focal demyelination induced by stereotaxic 587 injection of lysolecithin into the corpus callosum led to the identification of a few specific 588 markers namely including iNos, TNF, CD16/CD32 for the classical activated state and Arg1, 589 IGF1 and CD206 for the alternative state. The switch from an M1- to an M2-dominant response 590 was detected in microglia/macrophages at initiation of remyelination (Miron et al., 2013). It 591 was more recently characterized as a process requiring necroptosis of pro-inflammatory 592 microglia and subsequent repopulation to a pro-regenerative state (Lloyd et al., 2019). In vitro 593 experiments revealed the ability of M2 cell conditioned media to promote OPC differentiation 594 whereas the depletion of M2 cells in focal demyelinating lesions prevented OPC differentiation. 595 In addition, parabiosis experiments detected a higher number of M2 cells in lesions of aged 596 mice in which remyelination was increased by coupling to a younger mouse. Finally, the 597 blockade of activin-A secreted by M2 cells fully prevented OPC differentiation in demyelinated 598 cerebellar slice cultures (Miron et al., 2013). 599

Microglia/macrophage activation cannot be nevertheless regarded as a simple 600 dichotomy of M1 or M2, but undoubtedly as a much broader reactive phenotype spectrum. In 601 agreement with this observation, in the Theiler's induced demyelinating disease model, the 602 endogenous cannabinoid 2-AG induces pro-phagocytic response of microglia and OPC 603 differentiation, without any detectable regulation of the main markers used to identify classic 604 and alternatively activated microglia, respectively (Mecha et al., 2019). In the same line, myelin 605 debris were consistently proposed to promote the emergence of a novel phenotype that differs 606 from conventional M1 and M2. Indeed, myelin-induced macrophages appear foamy, displaying 607 a striking deficiency of the expression of the ATP-binding cassette transporter ABCA1 required 608 609 for lipid efflux in response to myelin debris loading, thus suggesting that this homeostatic mechanism characterizing non myelin-induced macrophages is overwhelmed. The impaired 610 capacity of foamy macrophages for apoptotic and necrotic cell clearance is consistent with this 611 hypothesis and likely contributes to the development of secondary injury (Wang et al., 2015b). 612

In agreement with this observation, a growing number of recent publications support the 613 existence of a wide spectrum of microglia activation states evidenced by single cell and bulk 614 RNA sequencing approaches characterizing this process not as a simple phenotypical change 615 but as a dynamic response involving spatially and transcriptionally distinct subpopulations. 616 High-dimensional single-cell mapping in EAE mouse CNS led to a unique activation profile 617 comprising universal markers of microglia/macrophage activation including CD44, CD86, 618 programmed death-ligand 1 (PDL1) as well as a decreased CD14 expression and increased 619 major histocompatibility complex II (MHC II) and stem cell antigen-1 (Sca-1) expressions 620 notably not detected in old mice or Alzheimer's disease models (Mrdjen et al., 2018). Single-621 cell RNA sequencing of microglia from LPC-mediated lesions revealed multiple gene 622 expression profile clusters of activated microglia/macrophages sharing common genes such as 623 apoE and unique genes such as Ccl4 or Cxcl10 according to the clusters (Hammond et al., 624 2019). Multiple activation states specifically reflecting microglia (and not macrophages) was 625 also identified upon LPC-mediated demyelination suggesting that microglia can have multiple 626 627 forms of activation likely identified by common or selective transcriptional programs (Plemel et al., 2020). Similarly, a specific ApoE-dependent molecular signature was identified in 628 microglia/macrophages from different models of neurodegenerative diseases including EAE 629 630 (Krasemann et al., 2017). Strikingly, one of these subsets displayed an expression profile comparable to the one found in MS microglia (Zrzavy et al., 2017). Real-time in vivo imaging 631 632 of microglia/macrophages in EAE mice visualized the expected switch from pro-inflammatory to immuno-regulatory markers mostly in initial lesions with an increase during lesion resolution 633 634 (Locatelli et al., 2018). In a consistent manner, in actively demyelinating lesions from MS patients, some microglia/macrophages express pro-inflammatory markers while active lesions 635 prone to remyelinate comprise microglia/macrophages expressing anti-inflammatory markers 636 at a higher level than chronic inactive lesions (Miron et al., 2013;Zrzavy et al., 2017). 637

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Factors controling the pro-regenerative microglia/macrophages

Recently, the subpopulation of CD11c-expressing microglia previously reported to be transiently expanded soon after birth and critical for primary myelination, was found to be remarkably increased in EAE and cuprizone models of demyelination. The expansion of this subpopulation is induced by the stimulation of CSF1R, the key transducer of CSF1 and interleukin (IL)-34 signals, essential in myeloid cell development. CSF1R activation induces the expression of the chemokine CCL2, itself leading to a dramatic increase of CD11c⁺

microglia, finally resulting in improvement of EAE symptoms and lower level of demyelination 645 (Wlodarczyk et al., 2018). Among factors implicated in promotion of the remyelinating 646 phenotype of microglia, the homeobox gene Msh-like homeobox-3 (MSX3) is a pivotal 647 regulator for microglial polarization that is induced and repressed in M2 and M1 cells, 648 respectively. This finding is supported by the dynamic regulation of MSX3 in the EAE model 649 as well as by the beneficial and deleterious effects observed after MSX3 gain- and loss-of 650 function experiments in different demyelination models. Pparg, Stat6 and Jak3 are key genes 651 regulated by MSX3 (Yu et al., 2015). In the same line, pro-regenerative microglia express a 652 high level of the purinergic receptor P2X4R at the peak of recovery in the EAE model. P2X4R 653 activation increases and decreases pro-regenerative and pro-inflammatory genes, respectively, 654 whereas its blockade prevents myelin debris clearance (Zabala et al., 2018). 655

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The critical role of microglia/macrophages in myelin phagocytosis

The ability of microglia/macrophages to phagocyte myelin debris is critical for 657 promoting OPC recruitment and differentiation into myelinating cells (Triarhou and Herndon, 658 1985;Kotter et al., 2001;Kotter et al., 2005;Kotter et al., 2006;Neumann et al., 2009). The main 659 phagocytic receptor in microglia/macrophages is the complement receptor-3 (CR3). Binding of 660 myelin to CR3 initiates structural changes characteristic of phagocytosis mostly reflected by 661 filopodia-like membrane protrusions that engulf myelin as they extend and then pull myelin 662 into phagocytes as they retract. Filopodia production depends on F-actin remodeling, which is 663 promoted by active unphosphorylated cofilin and impeded by inactive phosphorylated cofilin. 664 Spleen tyrosine kinase (Syk), the non-receptor tyrosine kinase known to be recruited by 665 phagocytic receptors including CR3, downregulates CR3-mediated myelin phagocytosis by 666 increasing the inactive state of cofilin. This self-negative control of phagocytosis is useful in 667 protecting phagocytes from excessive phagocytosis during prolonged exposure to particles that 668 are fated to ingestion (Hadas et al., 2012). Remarkably, myelin debris are able to inhibit their 669 own clearance by using a protective mechanism whereby CD47 protein expressed by myelin 670 debris binds the immune inhibitory receptor signal regulatory protein- α (SIRP α) on the surface 671 of phagocytes (Fig. 3B). The mechanism supporting such detrimental protection involves the 672 regulators of cytoskeleton function paxilin and cofilin, which are both either activated by 673 phagocytic receptors or inactivated by the immune inhibitory SIRPa (Gitik et al., 2014). The 674 finding that expression of Galectin-3 correlates with myelin-debris phagocytosis in microglia 675 led to show its implication in phagocytosis activation first by advancing cofilin activation, 676 causing filopodia/lamellipodia to extend/engulf myelin-debris and, second, by advancing 677 678 actin/myosin-based contraction through K-Ras.GTP/PI3K signaling, causing 679 filopodia/lamellipodia to retract and thus internalize myelin debris (Reichert and Rotshenker, 680 2019).

The retinoid X receptor (RXR) plays an important role in monocyte/macrophage 681 phagocytosis of myelin as suggested by pioglitazone-induced activation of peroxisome 682 proliferator-activated receptor c, one of the permissive binding partners of RXR, shown to 683 684 inhibit pro-inflammatory differentiation of MS patient-derived monocytes/macrophages and to 685 enhance myelin phagocytosis (Natrajan et al., 2015b). Moreover, the blockade of myelin debris clearance by microglia/macrophages in cuprizone-demyelinated CX3CR1-deficient mice, 686 revealed the critical role played by CX3CR1, the receptor controlling microglial physiology 687 and orchestrating the crosstalk between microglia and neurons (Lampron et al., 2015). 688 689 Moreover, macrophages lacking miR-223 expression have impaired ability to polarize towards

the M2 phenotype thus altering myelin debris clearance and remyelination in the lysolecithin-690 demyelinated corpus callosum (Galloway et al., 2019). Interestingly, the use of human 691 monocyte-derived macrophages and microglia led to demonstrate that myelin phagocytosis is 692 significantly enhanced in cells exposed to TGF- β compared with resting basal conditions 693 whereas it is markedly reduced in classically activated polarized cells. The transcriptional 694 analysis of TGF-β-treated microglia revealed the tyrosine kinase receptor MerTK as one of the 695 most upregulated among differentially expressed genes. In contrast, the enzyme and its ligands 696 (growth arrest-specific 6 and Protein S) are down-regulated in classically activated cells (Healy 697 698 et al., 2016).

Recent publications identified novel signaling pathways aimed at restoring homeostatic 699 700 microglial phagocytosis in aging CNS. Indeed, old mice fail to resolve the inflammatory response initiated after myelin damage. This observation was related to accumulation of 701 excessive amounts of myelin debris by aged phagocytes, which trigger cholesterol crystal 702 formation and phagolysosomal membrane rupture, thus stimulating inflammasomes. However, 703 stimulation of cholesterol efflux and solubility by administering 2-hydroxypropyl-β-704 cyclodextrine in old demyelinated mice knockout for the major cholesterol transporter ApoE 705 706 was sufficient to restore the capacity to remyelinate lesioned tissue (Cantuti-Castelvetri et al., 707 2018).

Combined CRISPR-Cas9 knockout screens together with RNA-seq led to discover age-708 related genetic modifiers of microglial phagocytosis such as the canonical B-cell receptor 709 CD22, a negative regulator of phagocytosis that is upregulated on aged microglia. CD22 710 mediates the anti-phagocytic effect of $\alpha 2$ -6-linked sialic acid. Its inhibition promotes the 711 clearance of myelin debris in vivo and reprograms microglia towards a homeostatic 712 transcriptional state leading to improvement of cognitive function in aged mice (Pluvinage et 713 al., 2019). By up-regulating the expression of the scavenger receptor CD36, niacin (also called 714 vitamin B3) induces myelin phagocytosis through binding its receptor, hydroxycarboxylic acid 715 receptor 2, both in vitro and in vivo (Rawji et al., 2020). 716

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Pro-regenerative microglia/macrophage-derived factors

Many cytokines, growth factors and soluble factors are known to be secreted by 718 microglia/macrophages (Fig. 3C). Although induced by IL1ß characterized as a pro-719 inflammatory cytokine (Mason et al., 2001), microglia/macrophage-derived IGF-1 enhances 720 remyelination as suggested by EAE rats receiving IGF-1 subcutaneously (Yao et al., 1996) and 721 in lysolecithin-demyelinated spinal cord of young and aged rats intrathecally infused with the 722 growth factor (Hlavica et al., 2017). TNFα through its TNFR2 receptor promotes remyelination 723 by increasing OPC proliferation as shown by using mouse strains knockout for the ligand or its 724 receptor as well as by their up-regulation during the remyelination stage in the cuprizone model 725 (Arnett et al., 2001). T helper cell-derived TGF- β induces the secretion of microglial hepatocyte 726 growth factor (HGF), which drives OPC chemotaxis and differentiation. In agreement with this 727 finding, spinal cord lesions from relapsing-remitting EAE mice contain both OPC and HGF-728 producing microglia/macrophages in the recovery-, but not in the acute phase (Lalive et al., 729 730 2005). Highly expressed in microglia, the β -galactoside-binding lectin, galectin-3, has a critical role in driving OPC differentiation and myelination in agreement with the oligodendrocyte 731 maturation effects displayed by supernatants derived from galectin-3-expressing- but not 732 733 galectin-3-deficient microglia (Pasquini et al., 2011).

Microglia/macrophages also express activin-A at onset of remyelination of focally-734 induced demyelination. This growth factor regulates oligodendrocyte differentiation and 735 myelin compaction via the activin receptor subtype Acvr2a up-regulated during efficient myelin 736 regeneration, in contrast to the other receptor subtype Acvr2b, which has to be down-regulated 737 for allowing oligodendrocyte differentiation. In actively remyelinating areas of MS tissue, 738 oligodendrocyte lineage cells expressing Acvr2a consistently outnumber cells expressing 739 Acvr2b suggesting that increase in Acvr2b expression may impair oligodendrocyte 740 differentiation and myelin formation induced by Acvr2a (Dillenburg et al., 2018). 741

Finally, microglia is a main iron source in the form of ferritin. The latter is required for 742 OPC proliferation and differentiation and stands as an essential cofactor for enzymes involved 743 744 in myelin cholesterol and fatty acid synthesis (Schonberg et al., 2012). The cellular iron level in microglia/macrophages is controlled by hepcidin and ferroportin. The transcription of the 745 former is regulated by the BMP/Smad and IL-6/Jak-STAT3 signaling pathways during 746 inflammation resulting in the degradation of the only known iron exporter ferroportin, 747 subsequently increasing intracellular iron level. Recently, the infusion of the BMP antagonist 748 noggin in a model of ischemic stroke decreased the induction of hepcidin and ferritin proteins 749 whereas it increased the number of myelinated axons and myelin thickness (Shin et al., 2018). 750

751 Several microglial enzymes are also clearly implicated in remyelination. As shown in development, microglia-derived TG2 signals to GPR56 on OPCs in the presence of the ECM 752 protein laminin, promotes OPC proliferation and improves remyelination in vivo in the 753 cuprizone model and *ex vivo* in cerebellar slices exposed to lysolecithin (Giera et al., 2015). 754 Lipoprotein-lipase involved in lipid-processing, which plays an important role during initiation 755 of remyelination and is considered as a feature of alternatively-activated microglia, is 756 significantly increased in the EAE model when clinical symptoms start to decrease (Bruce et 757 al., 2018;Kamermans et al., 2019). Finally, CD38 that catalyzes the synthesis of cyclic 758 adenosine diphosphate-ribose (cADPR) from nicotinamide adenine dinucleotide (NAD⁺), is 759 increased in the cuprizone model of demyelination in both astrocytes and microglia and may be 760 required for myelin clearance and oligodendrocyte repopulation (Roboon et al., 2019). 761

762 Recently, extracellular vesicles produced in vitro by either pro-inflammatory- or proregenerative microglia were proposed to be one of the mechanisms used to promote ou block 763 myelin repair. Remarkably, exposure of cultured OPCs to inflammatory vesicles blocked OPC 764 maturation only in the presence of astrocytes, thus implicating the latter in remyelination failure 765 via a mechanism involving astrocyte conversion into deleterious cells. Moreover, biochemical 766 fractionation revealed that the inflammatory cargo of the pro-inflammatory vesicles mainly 767 contribute to the blockade of OPC maturation whereas surface lipid components may be mostly 768 involved in extracellular vesicle-mediated migration and/or differentiation of OPCs likely 769 implicating sphingosine 1 phosphate at least for OPC migration (Lombardi et al., 2019). 770

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Therapeutic modulation of microglia/macrophages

Major improvement of our knowledge of the role of microglia/macrophages during remyelination recently led to new therapeutic perspectives. One of the first papers providing evidences that increased recruitment of microglia/macrophages can support remyelination, regarded the anti-fungal amphotericin B and its association with macrophage colonystimulating factor (M-CSF). Although using amphotericin B is precluded by its ability to stimulate TNF secretion and its toxic properties (Doring et al., 2015), the increased uptake of

myelin debris induced by M-CSF in the cuprizone model remains interesting (Laflamme et al., 778 2018). The adoptive transfer of bone-marrow-derived M2 macrophages results in a shift of the 779 immunological response from helper T1 to helper T2 lymphocytes through the production of 780 anti-inflammatory cytokines, which in turn induces the polarization of microglia/macrophages 781 to the M2 phenotype in a model of spinal cord injury (Ma et al., 2015). Preventive IL-13 gene 782 therapy in the cuprizone model directs the polarization of microglia and infiltrating 783 macrophages towards an alternatively activated phenotype, thereby limiting lesion severity and 784 improving disease outcome (Guglielmetti et al., 2016). 785

Several molecules modulating microglia/macrophages are presently trialled in patients. 786 For instance, the anti-muscarinic molecule Clemastine, modulates macrophage inflammatory 787 788 responses in vitro and microglia/macrophage activation in vivo (Mei et al., 2014). Bexarotene, an agonist of the retinoid X receptor pathway, has been identified as a positive regulator of 789 myelin debris clearance and a key player in the age-related decline in remyelination acting via 790 a vet undetermined direct or indirect effect (Natrajan et al., 2015a). The anti-psychotic 791 quetiapine fumarate promotes remyelination in vivo together with attenuating microglial 792 responses to inflammatory stimuli by decreasing TNF and nitric oxide secretion and preventing 793 activation of NF-KB (Wang et al., 2015a). 794

795 New molecules are continously being identified. Among them, a recombinant version of a naturally occurring human IgM, rHIgM22, has been shown to promote remyelination in 796 Theiler's virus infection and cuprizone animal models and to stimulate myelin phagocytosis in 797 a dose-dependent manner (Zorina et al., 2018). The CSF1R kinase inhibitor BLZ945 798 prophylactically and therapeutically prevents demyelination in the cuprizone model, namely in 799 800 the corpus callosum, but however increase myelin debris and axonal damage in other fiber tracts 801 reminding the phenotype observed in cuprizone-treated TREM2 knock-out mice (Beckmann et al., 2018; Wies Mancini et al., 2019). Although TNF is a master proinflammatory product of 802 activated microglia/macrophages implicated in CNS demyelination, the blockade of its soluble 803 form did not prevent cuprizone-induced oligodendrocyte loss and demyelination, but led to 804 efficient early remyelination due to improved phagocytosis of myelin debris and resolution of 805 microglia/macrophage activation (Karamita et al., 2017). Finally, the allosteric modulator of 806 P2X4R, Ivermectin, mimics P2X4R activation and improved motor function, electrical nerve 807 conductance, myelin debris phagocytosis and remyelination in the EAE model (Zabala et al., 808 809 2018).

810 Key studies supporting detrimental roles of microglia/macrophages on 811 oligodendrocytes and myelin

Besides the growing number of data supporting the beneficial consequences of 812 promoting the pro-regenerative phenotype of microglia/macrophages, several critical works 813 support the deleterious effects of microglia activated through the classical way. First, activated 814 815 microglia synthesize a multitude of cytokines, chemokines, cell adhesion glycoproteins or reactive oxygen radicals able to damage axons, myelin, oligodendrocytes (Minghetti and Levi, 816 1998; Raivich and Banati, 2004) and therefore involved in the initiation and propagation of the 817 818 inflammatory cascade, which promotes demyelination in neural disorders (Perry et al., 2010). In a consistent manner, LPS-activated microglia, polarized to pro-inflammatory status secrete 819 tumor necrosis factor alpha (TNF α) and interleukin-1beta (IL-1 β), both known to be cytotoxic 820 821 for oligodendrocytes (Selmaj and Raine, 1988). TNFα and cuprizone supplementation to rat primary cultures of oligodendrocytes consistently decreased cell viability (Pasquini et al., 2007). Moreover, minocycline-mediated blockade of microglial activation in cuprizone-treated
mice prevented demyelination while a positive correlation was shown between the production of nitric oxide and oligodendrocyte death (Merrill et al., 1993). Finally, LPS-induced secretion
of the stress chaperone protein, heat shock protein 60 (HSP60), was found to initiate OPC apoptosis (Li et al., 2017).

828 As competent presenters of antigen, microglia activated in the classical way express molecules for antigen presentation such as MHC II and its co-stimulatory factors CD40 and 829 CD86, characterized as classical markers of microglia activation in MS and Alzheimer's disease 830 patients (Gobin et al., 2001;Lehmann et al., 2001;Hoftberger et al., 2004). Remarkably, 831 832 activated microglia/macrophage is also highly proliferative in both MS and animal models of demyelination mostly in actively demyelinating areas at the early stages of the disease but not 833 in remyelinating lesions again attesting the major role of classically activated 834 microglia/macrophages during the early stages of demyelinating diseases (Matsumoto et al., 835 1992;Schonrock et al., 1998;Ponomarev et al., 2005). Early activation of microglia is similarly 836 observed in ischemic dementia models as shown by the increased expression of MHC-I/II or 837 matrix metalloprotease-2 (MMP-2) 3 days after hypoperfusion whereas administration of the 838 natural dipeptide carnosine (*β*-alanyl-L-histidine) able to decrease microglial activation 839 improved cognitive function and white matter lesions (Wakita et al., 1994; Ihara et al., 2001). 840 Finally, as previously mentionned, extracellular vesicles released by pro-inflammatory 841 microglia block remyelination via a mechanism depending on the presence of astrocytes 842 apparently converted into harmful cells by the inflammatory extracellular vesicle cargo 843 (Lombardi et al., 2019). 844

845 In summary, the majority of experimental evidence reported in this review, points to astrocytes and microglia/macrophages as being crucial in both developmental and repairing 846 oligodendrogenesis and myelination. During development, astrocytes and a unique phenotype 847 of neonatal microglia secrete different patterns of molecules controling oligodendrocyte 848 development and myelination. Although both cell types shape the number of OPCs with 849 consequences for subsequent myelinogenesis, astrocytes are also implicated in a tight crosstalk 850 between OPCs and axons making them indispensable for myelination. In addition, by providing 851 a substantial fraction of lipids incorporated into CNS myelin, astrocytes are also required for 852 the proper generation of the myelin sheath. Under pathological and demyelinating conditions, 853 reactive astrocytes and activated microglia/macrophages both exhibit a dual activity inducing 854 855 detrimental or beneficial effects, the balance of which appears to be critical for tissue regeneration. Besides the secretion of a wide range of molecules, both cell types are able to 856 phagocyte myelin debris. However, in contrast to the beneficial role of microglia/macrophages-857 858 induced phagocytosis, myelin uptake occurring as an early response of astrocytes results in 859 recruitment of immune cells possibly increasing tissue damage in demyelinating conditions 860 driven by inflammation. As a whole, the review indicate that beyond the secretion of a multitude directly controling diverse facets of OPC biology, 861 of cues astrocytes and microglia/macrophages strongly modify the lesion microenvironment. Further delineation of 862 863 this crosstalk should open the way towards novel therapeutic approaches aimed at recovering proper developmental myelination and relieving the obstacles on failure of regeneration of 864 damaged myelin characterizing CNS demyelinating diseases. 865

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1499 Figure legends

1500 Figure 1: Major roles of astrocytes and microglia during developmental oligodendrogenesis and myelination. The different steps of oligodendroglial cell production 1501 (purple) are shown and include OPC migration, proliferation, differentiation as well as the 1502 maturation of immature oligodendrocytes into cells able to myelinate neuronal axons. 1503 Astrocyte- (A) and microglia- (B) derived molecules are indicated. The green arrows indicate 1504 molecules exhibiting a positive activity on oligodendrocyte lineage and myelin production (top 1505 in each panel) whereas the red arrows and blocking symbols indicate molecules displaying an 1506 inhibiting activity on oligodendroglial lineage production (bottom in each panel). 1507

Figure 2 : Reactive astrocyte-derived secretion products during myelin repair. The oligodendroglial lineage is shown in purple. Reactive astrocytes generate molecules promoting (top, green arrows) or blocking (bottom, red arrows and blocking symbols) myelin regeneration. Besides reactive astrocytes, two additional forms of astrocytes are indicated including myelinladen astrocytes endowed with a deleterious effect under inflammatory conditions and transplanted immature astrocytes which promote OPC proliferation / differentiation and additionally prevent glial scar formation.

Figure 3 : Key effects of microglia/macrophages during myelin repair. (A) Illustration of 1515 morphological modifications of ramified microglia/macrophages, which adopt a rhomboid 1516 shape in the presence of any insult and become activated into a classical or alternative manner 1517 giving rise to a range of cell phenotypes expressing mostly pro- (red) or anti-inflammatory 1518 (green) molecules, respectively. (B) Phagocytosis of myelin debris is a critical activity of 1519 microglia/macrophages for promoting myelin regeneration. Phagocytic receptors such as CR3 1520 advance phagocytosis by promoting the activation of paxillin and cofilin whereas the immune 1521 inhibitory receptor SIRPa, inhibits phagocytosis by promoting the inactivation of the two. 1522 Recruited by CR3, Syk downregulates CR3-mediated myelin phagocytosis protecting 1523 phagocytes from excessive phagocytosis during prolonged exposure to debris. (C) Pro-1524 regenerative microglia secrete a range of molecules intervening at various steps of myelin 1525 repair. 1526

1527







1534 Figure 2.





1537 Figure 3.