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Review

Canonical and Non-Canonical Autophagy in HIV-1 Replication Cycle

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Abstract: Autophagy is a lysosomal-dependent degradative process essential for maintaining cellular homeostasis, and is a key player in innate and adaptive immune responses to intracellular pathogens such as human immunodeficiency virus type 1 (HIV-1). In HIV-1 target cells, autophagy mechanisms can (i) selectively direct viral proteins and viruses for degradation; (ii) participate in the processing and presentation of viral-derived antigens through major histocompatibility complexes; and (iii) contribute to interferon production in response to HIV-1 infection. As a consequence, HIV-1 has evolved different strategies to finely regulate the autophagy pathway to favor its replication and dissemination. HIV-1 notably encodes accessory genes encoding Tat, Nef and Vpu proteins, which are able to perturb and hijack canonical and non-canonical autophagy mechanisms. This review outlines the current knowledge on the complex interplay between autophagy and HIV-1 replication cycle, providing an overview of the autophagy-mediated molecular processes deployed both by infected cells to combat the virus and by HIV-1 to evade antiviral response.

Keywords: HIV-1; autophagy; LC3-associated phagocytosis; Env; Tat; Nef; Vpu; LC3

1. Introduction

Macroautophagy (herein called autophagy) is a lysosomal degradation pathway-dependent mechanism, initially described as being involved in the turnover of cellular components (proteins, lipids and organelles), either to maintain cell homeostasis or as a source of energy in response of metabolic stress. In the early 2000s, it became obvious that autophagy is also involved in immune response as a key element of innate defense against bacterial infection [1–3] or adaptive immunity by participating in major histocompatibility complex class II (MHC-II) processing of viral antigens [4]. Since then, numerous studies have provided evidence that autophagy plays a crucial role in host response, especially by selectively degrading intracellular pathogens or targeting cytosolic exogenous antigens to MHC-II containing compartment. The autophagic process also plays a part in the recognition of pathogen-associated molecular patterns (PAMPs) through Toll-like receptors (TLRs) by delivering cytosolic microbial genetic material to endosomal TLRs. Furthermore, autophagy acts as a downstream effector of pattern-recognition receptors (PRRs), signaling to eliminate intracellular pathogens or to regulate inflammatory response.

Human immunodeficiency virus (HIV), the etiologic agent responsible for AIDS, as an obligate intracellular pathogen, needs to circumvent cells' intrinsic defenses and evade immune surveillance in order to efficiently propagate in infected patients. Interestingly, HIV infects CD4-expressing cells, such as CD4+ T lymphocytes (CD4+ T cells), macrophages and dendritic cells (DCs), leading to the failure of the immune system and resulting in a positive feedback loop for virus replication and spread.

In addition, HIV can persist for a long time in these target cells. Thus, CD4+ T cells, macrophages and DCs form viral reservoirs, despite the fact that these cellular populations are among the most specialized host cells to detect and mount an immune response to eliminate pathogens. Since it is clear that the autophagy mechanism participates in numerous antiviral activities deployed by CD4-expressing immune cells, many studies have been carried out to evaluate the impact of autophagy on the HIV life cycle. This review discusses the intricate interplay between HIV and autophagy, highlighting how HIV exercises fine control over the autophagy pathway in its target cells.

2. An Overview of the Autophagy Mechanism

In mammalian cells, canonical autophagy is regulated by approximately 30 autophagy-related proteins (ATG) and several non-ATG proteins, which coordinate the formation of intracellular vesicles, sequestering portions of cytoplasmic material in a three-step process: initiation, elongation and maturation (Figure 1). The initiation step starts with the isolation of a cellular membrane (called phagophore) at the endoplasmic reticulum (ER) surface by the formation of an omega-like shape protrusion [5–7]. This structure is induced by a preinitiation complex composed of UNC-51-like kinase (ULK) 1 and ULK2 proteins, a focal adhesion kinase family-interacting protein of 200 kDa (FIP200), and ATG101 and ATG13 proteins [8–12]. In the absence of stimuli, the preinitiation complex is inhibited by a key regulator of the autophagy machinery, the serine/threonine kinase mammalian target of rapamycin (mTOR) protein. mTOR, in complex with different cellular cofactors to form mTOR complex 1 (mTORC1), maintains the preinitiation complex in an inactivated state by phosphorylating ULK1 and ATG13 [10–12]. After stimulation by metabolic stress (e.g., starvation, oxidative stress or hypoxia) or chemical autophagy inducers such as rapamycin [13], mTORC1 is inhibited, allowing the release and the translocation of the preinitiation complex to the site of phagophore formation [14,15]. Subsequently, ULK1 activates the class III phosphatidylinositol 3-kinase (PI3K) complex formed by the PI3K vacuolar protein sorting 34 (VPS34), VPS15 (also called p150), Beclin 1/ATG6, ATG14L and the activating molecule in Beclin 1-related autophagy 1 (AMBRA1) protein. The activated PI3K complex is targeted to the ER-mitochondria contact sites, where it produces phosphatidylinositol-3-phosphate (PI(3)P), triggering phagophore formation [16–19]. The elongation step, corresponding to phagophore expansion, is regulated by the ATG5-ATG12 conjugate and lipidated forms of ATG8 family members [20–23]. Production of these components requires two ATG7-dependent ubiquitin-like conjugation systems. Indeed, the E1-like enzyme ATG7, in association with E2-like enzymes ATG10 or ATG3, promotes the formation of ATG5-ATG12 conjugate [24–26] or the lipidation of ATG8s with phosphatidylethanolamine (mostly designated ATG8-II or LC3-II) [27,28], respectively. ATG8 members, comprising LC3-like (LC3A, LC3B and LC3C) and GABARAP-like (GABARAP, GABARAPL1 and GABARAPL2) subfamilies, are key markers used to monitor the autophagy process [29]. At the steady state, ATG8 proteins exist as free cytosolic forms, usually referred to as ATG8-I or LC3-I. Once autophagy is induced, ATG8 proteins are translocated to the nascent phagophore, where the lipidation reaction tightly binds ATG8s to the membrane. The ATG5-ATG12 conjugate, in interaction with ATG16L, controls ATG8s lipidation by specifying the membrane localization where the reaction takes place [30]. ATG8 proteins drive membrane expansion [31] and phagophore closure [32]. The resulting double-layered membrane vesicle is called autophagosome. The maturation step corresponds to the fusion of autophagosomes with lysosomes to generate autolysosomes, where the autophagosomal content is degraded. This step is regulated by cellular machineries involved in vesicle transport and fusion. In particular, several studies have shown that members of the Rab GTPase family (RAB7, RAB8B, RAB9, RAB11, RAB23 or RAB24) [1,33–36] and the soluble N-ethylmaleimide attachment protein receptor (SNARE) superfamily (VAMP3, VAMP7, VAMP8, VTI1B or STX17) [36–39] are essential for the late stage of autophagy mechanism.

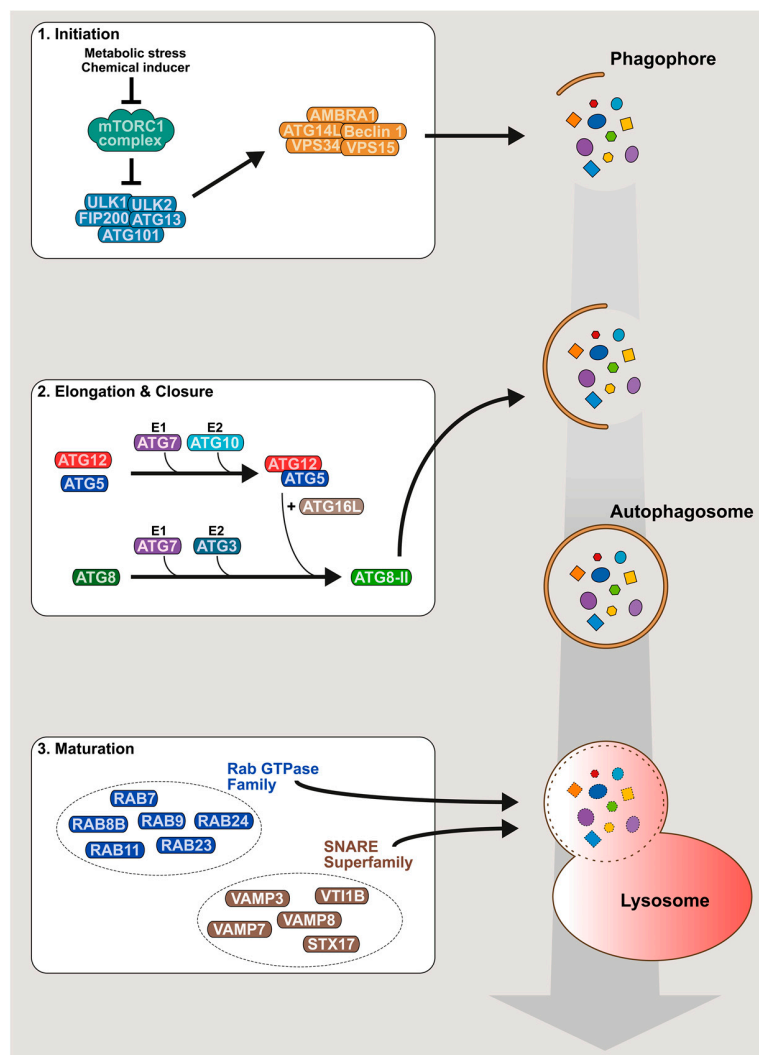


Figure 1. Schematic model of canonical autophagy. Canonical autophagy, which induces formation of double-layered membrane vesicles (autophagosomes) sequestering cytoplasmic material, can be divided into three main stages: (1) initiation; (2) elongation and closure; and (3) maturation. Briefly, in the absence of pro-autophagy signals, mTORC1 complex maintains the preinitiation complex of autophagy (ULK1, ULK2, FIP200, ATG101 and ATG13) in an inactivated state by phosphorylating ULK1 and ATG13. Upon autophagy stimulation (metabolic stress or chemical inducers), mTORC1 complex is inhibited, allowing the preinitiation complex to activate the PI3K complex (VPS34, VPS15, Beclin 1, ATG14L and AMBRA1). This latter triggers the isolation of a cellular membrane (phagophore) at the endoplasmic reticulum (ER) surface by producing phosphatidylinositol-3-phosphate. Elongation and closure of phagophore are driven by lipidated forms of ATG8 family members. ATG8 orthologs exist in two forms, depending on the autophagy state: a non-lipidated cytosolic form (ATG8; inactive autophagy state) or a lipidated membrane-bound form (ATG8-II; active autophagy state). ATG8 lipidation reaction is controlled by two ubiquitine-like systems composed of the E1-like enzyme ATG7 and E2-like enzymes ATG10 or ATG3. The ATG7/ATG10 system generates the ATG5-ATG12 conjugate which, in interaction with ATG16L, is essential for specifying the site of ATG8-II production at the phagophore membrane. The maturation step, corresponding to the fusion between autophagosomes and lysosomes, is regulated by members of the Rab GTPase family and the SNARE superfamily. This process results in degradation of the autophagosomal content. ULK: UNC-51-like kinase; FIP200: focal adhesion kinase family-interacting protein of 200 kDa; mTORC1: mTOR complex 1; PI3K: phosphatidylinositol 3-kinase; VPS: vacuolar protein sorting proteins; AMBRA1: activating molecule in Beclin 1-related autophagy 1; ATG: autophagy-related proteins; SNARE: soluble N-ethylmaleimide attachment protein receptor.

3. Cell Type Specific Modulation of Autophagy during HIV-1 Infection

Among HIV-infected patients, a few (around 5%) can control disease progression without antiretroviral therapy, remaining clinically asymptomatic and maintaining a normal CD4+ T cell count for at least 8 to 10 years [40,41]. These patients are divided into two groups, according to the viremia level: the viremic controllers (50–2000 HIV RNA copies/mL; VC) and the elite controllers (<50 HIV RNA copies/mL; EC) [42–44]. These two groups have been extensively studied to identify the cellular factors allowing viral control. A comparative analysis of peripheral blood mononuclear cells (PBMC) isolated from VC-, EC- and HIV-1-infected normal progressor donors underlined differences in autophagy pathway activity [45]. In particular, this study reports that VC and EC donors have a greater number of autophagic vesicles associated with an increased expression of autophagic markers with respect to normal progressors. In addition, rapamycin treatment of PBMC from VC donors induces an increased average number of autophagosomes per cell, where viral particles are trapped, an enhanced autophagic flux and a reduced virus production in comparison to untreated condition, while no modification is observed with normal progressors' PBMC. HIV-1 could thus block the autophagy process even upon rapamycin treatment in normal progressors' PBMC. Conversely, HIV-1 controllers seem to be able to bypass this block and sustain a fully functional autophagy mechanism engaged in viral particle elimination. However, this global analysis of autophagic response in infected PBMC does not take into account the cell-type-specific modulation of autophagy during HIV-1 infection. A more detailed examination of the heterogeneous population composing PBMC (lymphocytes, monocytes/macrophages and DC) reveals significant differences in the autophagic response to HIV-1 infection depending on the target cell (Figure 2).

In primary monocyte-derived macrophages (MDM), HIV infection induces the activation of the early steps of the autophagy mechanism and inhibition of the late stage of this process. This latter one is necessary for an optimal HIV-1 replication [46,47]. Interestingly, a transmission electron microscopy analysis carried out by Espert et al. highlights the fact that viral particles are only detected in weakly autophagic MDM [46]. These data suggest that HIV-1 maintains a delicate balance between activation and inhibition of the autophagy pathway to favor its replication in MDM.

In infected CD4+ T cells, the role of autophagy on HIV-1 production remains controversial. On the one hand, two studies report that productive HIV-1 infection of CD4+ T cells (primary cells or MOLT-X4 cell line) downregulates the autophagy pathway [46,48]. On the other hand, data presented by Laforge et al. indicate an increased expression of Beclin 1, ATG5 and LC3-II in HIV-1-infected primary CD4+ T cells compared to uninfected cells [49]. The use of different HIV-1 strains in these studies could explain those discrepancies. Interestingly, a more recent study showed that HIV-1 glycoprotein envelope (Env) triggered autophagy within the first 4 h following CD4+ T cell infection, and that autophagy is inhibited by HIV-1 48 h post-infection [50]. Accordingly, one may assume that, depending on the strain, HIV-1 cannot always invert the Env-mediated induction of autophagy in CD4+ T cells which could lead to an impaired HIV-1 replication in these cells. With regard to this, Laforge et al. [49] reveal that the induction of autophagy in response to the HIV-1 infection of CD4+ T cells is mediated by the damage-regulated autophagy modulator (DRAM), a lysosomal protein previously identified as a regulator of p53-induced autophagy [51]. In particular, they notice that depletion of DRAM increases the HIV-1 viral capsid (p24) present in the supernatant of infected CD4+ T cells. However, DRAM depletion also prevents HIV-1-mediated cell death, which does not allow a determination as to whether the p24 increase observed is due to a greater number of HIV-1 producer cells, or enhanced HIV-1 production. Nevertheless, the study by Sagnier et al. [50] demonstrated that treatment of primary CD4+ T cells or chronically-infected MOLT-X4 cells with Torin1, an mTOR inhibitor, impairs HIV-1 production. Consequently, this result suggests that autophagy activation has an antiviral effect in CD4+ T cells.

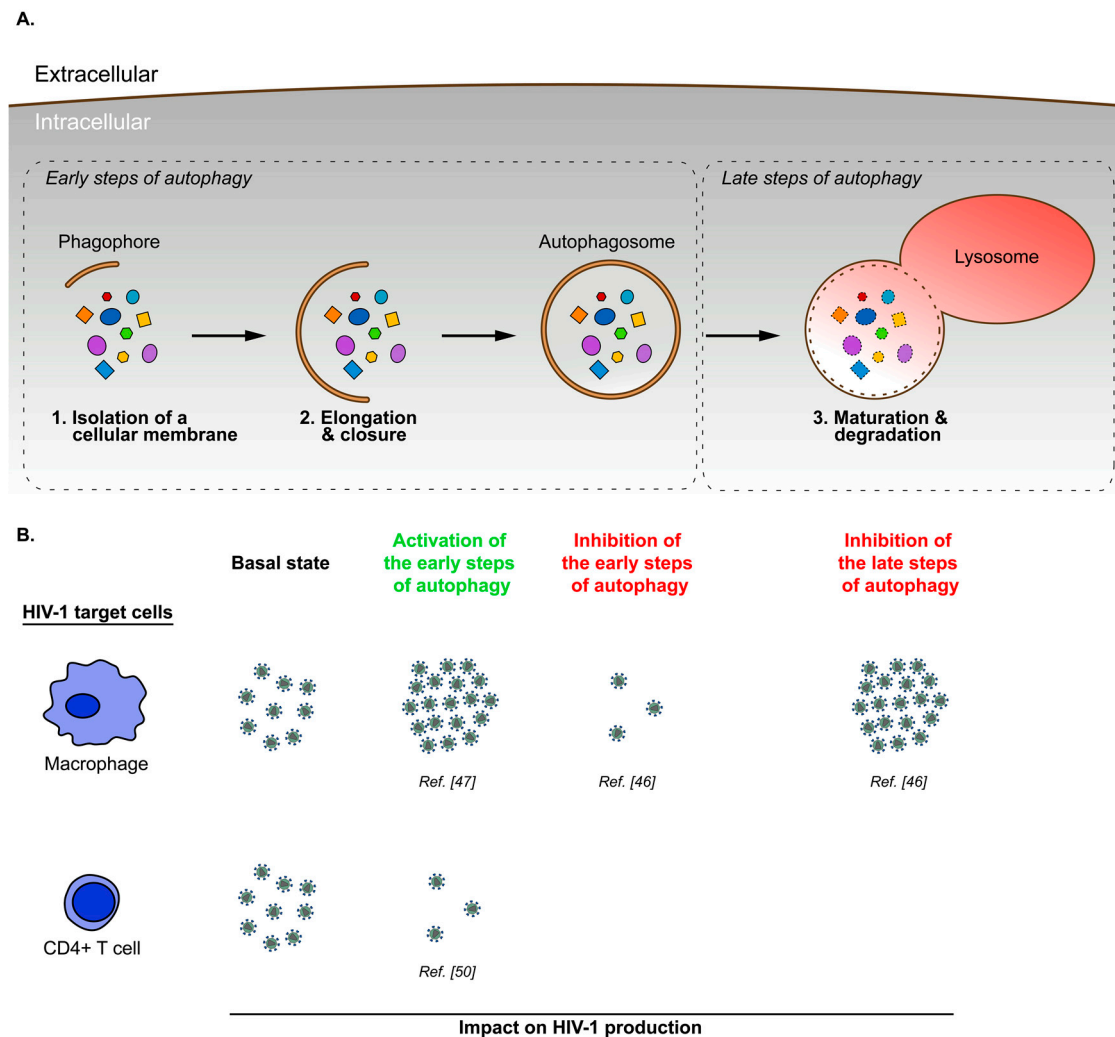


Figure 2. The interplay between autophagy and HIV-1 production in cell type specific manner. (A) Canonical autophagy mechanism can be decomposed into early steps, corresponding to the isolation, elongation and closure of a cellular membrane (phagophore) to form a double-layered membrane vesicle enclosing cytoplasmic materials (autophagosome); and late steps, corresponding to the autophagosome-lysosome fusion/maturation and the degradation of autophagosomal content; (B) In HIV-1 infected cells, activation or inhibition of early and late steps of autophagy process modulate virus production in a cell-type-specific manner. While induction of autophagy dramatically impairs HIV-1 production in CD4+ T cells, activation of early steps of autophagy is indispensable for the optimal production of viruses in macrophages. Conversely, blockage of late steps of autophagy favors an efficient HIV-1 production in macrophages.

4. Autophagy-Mediated Viral Restriction in HIV-1 Infected Cells

To face viral infection, host cells possess a large array of “self-defense” mechanisms that constitute the intrinsic and innate immunity [52]. Intrinsic immunity is mediated by cellular pathways such as autophagy, RNA interference or may be conferred by a set of proteins, called restriction factors, which provide an immediate and direct antiviral activity. Conversely, innate immunity is an inducible mechanism, which is deployed less rapidly than intrinsic immunity, but results in a broader antiviral response. This mechanism is mediated by PRRs, such as TLRs, which detect virus-associated molecular patterns and trigger proinflammatory signaling cascades. In this section, we will highlight how autophagy pathway intersects with intracellular defense mechanisms to prevent HIV-1 infection. On the one hand, we will discuss the role of autophagy in the selective targeting to lysosomal

degradation of viral components by proteins p62, the tripartite motif containing 5 α (TRIM5 α) and histone deacetylase 6 (HDAC6). On the other hand, we will examine the implication of autophagy in the induction of TLR-mediated proinflammatory response.

4.1. Degradation of the HIV-1 Tat Protein by the Selective Autophagy Mechanism

Although autophagy can be a non-specific degradation process, it can also specifically deliver cytosolic components to lysosomal degradation through the so-called selective autophagy mechanism. This mechanism requires autophagy receptors, which bridge selected cargo to nascent autophagosomes by interacting with the LC3-II proteins associated with the inner membrane of autophagosomes. Notably, interaction with LC3-II is mediated by LC3-interaction region (LIR) motifs present on autophagy receptors. One of the most extensively studied autophagy receptors is the p62 protein (also named sequestosome 1, SQSTM1), which is known to bind ubiquitinated substrates and to direct them for degradation [53]. Recently, p62 was reported to be involved in an autophagy-dependent degradation of the HIV-1 transactivator of transcription (Tat) protein [50]. Tat is an essential regulator of HIV-1 transcription by recruiting cellular transcription factors to the transactivation response element (TAR) presented within the 5' end of viral transcripts [54]. In the study by Sagnier et al. [50], the authors revealed that p62 directly interacts with Tat in an ubiquitin-independent manner and mediates Tat degradation via the autophagic process. In particular, they showed that the transient Env-mediated induction of autophagy triggers this mechanism in CD4+ T cells, leading to impaired HIV-1 replication. In addition, their results indicate that secreted Tat proteins, known to affect gene expression and function in bystander cells, are similarly degraded in neighboring cells via selective autophagy.

4.2. Contribution of Autophagy in the TRIM5 α Restriction Factor Activities against HIV-1

The p62 protein is also reported to participate in the anti-retroviral activity of the TRIM5 α restriction factor. TRIM5 α is a member of the TRIM family of proteins, which controls a large number of cellular processes (such as autophagy, innate immunity, apoptosis, or intracellular signaling). This restriction factor is also known to block HIV-1 infection by inducing a pro-inflammatory innate response and accelerating the uncoating of the viral core through binding to viral capsid [55–57]. More recent evidence highlights that TRIM5 α can act as an autophagic receptor for HIV-1 p24, as well [58,59]. In particular, one study revealed that autophagy activation in HIV-1-infected rhesus monkey CD4+ T cells induced HIV-1 p24 degradation in a process depending on TRIM5 α , p62, Beclin 1 and ATG7 [58]. The authors notably showed that mutating two LIR motifs in rhesus monkey TRIM5 α (rhTRIM5 α) prevents autophagy-mediated degradation of p24. Furthermore, on the basis that rhTRIM5 α cannot bind simian immunodeficiency virus (SIV) capsid [57,60], Mandell et al. [58] demonstrated that autophagy induction hampers HIV-1 but not SIV infection, supporting the role of TRIM5 α as an autophagy receptor for HIV-1 viral capsid. Some of the first investigations into human TRIM5 α (huTRIM5 α) found that this variant was much less efficient at blocking HIV-1 infection than rhTRIM5 α [57,60–62]. However, recent work on Langerhans cells has suggested that huTRIM5 α restriction efficacy may depend on autophagy process and the route of HIV-1 entry into host cells [59]. Langerhans cells form a subset of DC that is found in epidermis and mucosa. These cells are known to be the first line of defense against HIV-1 by capturing viruses via their cell surface C-type lectin receptor Langerin and degrading viral particles [63]. The study by Ribeiro et al. [59] revealed that depletion of Langerin, huTRIM5 α , ATG5 or ATG16L in HIV-1 infected Langerhans cells leads to increased integration of viral DNA into the host genome. Conversely, HIV-1 integration is dramatically reduced after rapamycin treatment. Moreover, ectopic expression of Langerin in CD4+/CCR5+ U87 cells, a human glioblastoma cell line, also strongly inhibits HIV-1 infection in a huTRIM5 α -, ATG16L-dependent manner. Interestingly, HIV-1 restriction activity of Langerin is not shared with the other C-type lectin receptor DC-SIGN (dendritic cell-specific ICAM-3-grabbing non-integrin). Indeed, depletion of huTRIM5 α in DC-SIGN+ DC, a subset of DC known to be easily infected by HIV-1 [64], does not modify HIV integration and production. Finally, the authors showed that TRIM5 α and

ATG16L form a complex with Langerin through the binding of both TRIM5 α and Langerin with leukocyte-specific protein 1 (LSP1). This led Ribeiro et al. to propose that, following binding to HIV-1, the Langerin-LSP1-TRIM5 α -ATG16L complex is internalized into Birbeck granules, a specific compartment of Langerhans cells connected to the endosomal recycling pathway [65], which are subsequently delivered to nascent autophagosomes for degradation.

4.3. Modulation of Vif-Mediated Countermeasure against APOBEC3G Restriction by Autophagy

During the course of its evolution, HIV-1 has acquired accessory genes, encoding for proteins able to block cellular restriction factor activity. Among HIV-1 accessory proteins, the viral infectivity factor (Vif) binds and induces a proteasome-dependent degradation of the restriction factor apolipoprotein B mRNA editing enzyme catalytic polypeptide-like 3G (APOBEC3G) [66]. This cellular factor was identified for both its ability to produce G to A hypermutations in the HIV-1 genome and to inhibit HIV-1 reverse transcription and proviral DNA integration [67–71]. Interestingly, it has been shown that HDAC6 is able to protect APOBEC3G against Vif-mediated degradation by interacting with APOBEC3G and/or inducing autophagy-dependent degradation of Vif, impairing HIV-1 infectivity [72]. This study has notably revealed that HDAC6 directly interacts with APOBEC3G and Vif, forming the ternary complex APOBEC3G-HDAC6-Vif or the two binary complexes HDAC6-APOBEC3G and HDAC6-Vif. By forming the HDAC6-APOBEC3G complex, HDAC6 prevents the association of APOBEC3G with Vif and, thus, inhibits the Vif-mediated ubiquitination and delivery of APOBEC3G to the proteasome. In addition, the HDAC6-Vif complex favors the autophagy-mediated degradation of Vif in a HDAC6 deacetylase activity and an ubiquitin-binding domain (BUZ domain)-dependent manner. HDAC6 deacetylase activity and BUZ domain have been linked to the clearance of polyubiquitin-enriched aggresome, a subcellular structure formed by the accumulation of polyubiquitin-positive misfolded proteins [73], by promoting fusion between lysosomes and aggresome-containing autophagosomes [74,75]. Thus, it can be hypothesized that HDAC6 promotes Vif degradation by favoring the association of Vif to the nascent aggresome-containing autophagosomes. In contrast with data by Valera et al. [72], a study indicates that inhibition of HDAC proteins, via HDAC chemical inhibitors (HDACi), promotes autophagy-mediated HIV-1 degradation in MDM by inhibiting the mTOR protein [76]. Although these studies might seem contradictory at first sight, HDAC family proteins differ widely in terms of sequence homology and cellular functions [77,78]. In addition, Campbell et al. [76] have not determined whether HDAC proteins directly act on cellular factors involved in mTOR regulation, or if HDAC proteins act on the transcription of autophagic factors related to their role as histone regulators. Thus, it is likely that HDAC family members differently regulate autophagy mechanism during HIV-1 infection.

4.4. Participation of Autophagy in the Induction of Innate Immunity to HIV-1 Infection

Recognition of HIV-1 by TLRs is also involved in the implementation of an efficient innate response to block HIV-1 spread. Particularly, it has been highlighted that the treatment of macrophages with two TLR8 agonists, ssRNA40 (a 20-mer phosphorothioate protected single-stranded RNA oligonucleotide) or the imidazoquinoline CL097, inhibits HIV-1 replication in an autophagy dependent manner in these cells [79]. This mechanism depends on the vitamin D signaling pathway, known to upregulate transcription of *Beclin 1* and *Atg5* in macrophages through the human cathelicidin antimicrobial peptide (CAMP)-dependent activation of the transcription factor CCAAT-enhancer-binding protein β (C/EBP- β) and the p38 mitogen-activated protein kinase (p38-MAPK) [80]. More precisely, in the presence of the active form of vitamin D, 1 α ,25-dihydroxycholecalciferol (1,25D3), and upon recognition of PAMPs, TLR8 induces *CAMP* gene upregulation, leading to an autophagy-mediated lysosomal-dependent inhibition of HIV-1 replication [79]. Autophagy is also implicated in the induction of interferon α (IFN α) by TLR7 in response to infectious or noninfectious HIV-1 or ssRNA40, in plasmacytoid DC (pDC) [81]. Indeed, ATG7 depletion by small interfering RNA (siRNA) or treatment with the autophagy inhibitor 3-methyladenine (3-MA) dramatically impedes IFN α production in pDC

in the presence of HIV-1 particles. Interestingly, the authors revealed that 3-MA treatment during HIV-1 infection reduces IFN regulatory transcription factor 7 (IRF7) phosphorylation, which is necessary for the translocation of IRF7 to the nucleus and subsequent stimulation of IFN α expression.

5. Disruption of Autophagy-Associated Adaptive Immune Response by HIV-1

The establishment of a specific adaptive immune response to HIV-1 necessitates, in part, the processing and presentation of exogenous or endogenous viral-derived antigens through the MHC pathway in DC for an ensuing T cell activation and clonal expansion [82]. However, it has been shown that HIV-1 infection impedes DC maturation, leading to an impaired T-cell response [83–85]. In recent years, there has been growing evidence supporting the role of autophagy in the establishment of an effective adaptive immune response by participating in antigen processing and presentation [86]. Notably, autophagy can enhance antigen presentation by MHC molecules (class I or II) in antigen-presenting cells (APC) like DC or macrophages in response to microbial infection [87–91]. Interestingly, the study by Blanchet et al. [92] revealed that HIV-1 affects the presentation of exogenous antigens through MHC-II by downregulating autophagy flux initiation. Indeed, as early as 10 hours after exposure of monocyte-derived DC to viral particles, HIV-1 promotes mTOR activation, leading to a strong reduction of the DC-mediated CD4⁺ T cell response against HIV-1 antigens. This effect is not associated with any defects in DC maturation and cell surface MHC-II expression. Conversely, induction of autophagy with rapamycin in DC induced a robust CD4⁺ T cell response. These findings suggest that HIV-1-mediated inhibition of autophagy in DC might be a mechanism for evading adaptive immune response. In some circumstances, APC can also present endogenous viral antigens through MHC-II molecules. However, it has been highlighted that the role of autophagy in the MHC-II pathway is limited to exogenous antigen presentation. A recent study revealed that induction or inhibition of autophagy in HIV-1-infected DC do not alter endogenous viral antigen processing and presentation by MHC-II, and subsequent CD4⁺ T cell activation [83]. In addition, Blanchet et al. [92] reported that autophagy does not contribute to the cross-presentation process in DC, a mechanism allowing the processing and presentation of exogenous HIV-1 antigens by MHC-I molecules to CD8⁺ T cells.

The crucial role of autophagy in an effective MHC-II-mediated CD4⁺ T cell response was also confirmed in DC. Indeed, transduction of DC with a recombinant construct expressing a fusion protein formed by LC3B and the HIV-1 Gag precursor protein allows the specific targeting of this viral component to autophagosomes [83]. More interestingly, in an *in vivo* mouse model, expression of a SIV Gag protein fused to LC3B induces a stronger CD4⁺ T cell response-associated cytokine production (IFN γ , tumor necrosis factor α (TNF α) and interleukin 2 (IL-2)) and a higher number of SIV Gag-specific IFN γ -secreting CD4⁺ T cells in comparison with transduction of the SIV Gag protein alone [93]. Intradermal injection of mice with constructs expressing either the HIV-1 p24 alone or fused to the autophagy receptor p62 (p24/p62) led to very similar results. Thus, p24/p62-immunized mice displayed a higher number of p24-responding T cells, recognizing a more diverse p24 peptide repertoire, than p24-immunized mice [94]. Finally, the study by Yin et al. [93] revealed that the SIV-Gag-LC3B antigen induced a greater level of SIV Gag specific antibodies in mice compared to SIV Gag alone. This observation indicates that the contribution of autophagy in MHC-II-mediated CD4⁺ T cell activation also has important consequences for the establishment of an efficient humoral response against HIV-1.

6. Hijacking of Canonical Autophagy by HIV-1 Viral Proteins

Several HIV-1 proteins have been identified for their ability to alter the canonical autophagy pathway to promote viral production and propagation, evade the innate and adaptive immune system, or even favor pro-survival mechanisms to delay cell death. This section highlights the most documented HIV-1 proteins, demonstrating connections with canonical autophagy (Table 1).

Table 1. Non-autophagic and autophagic functions of HIV-1 Env, Tat, Nef and Vpu proteins.

HIV-1 Proteins	Non-Autophagic Functions	Autophagic Functions	References (Associated with Autophagy)
Env (gp120 and gp41)	Binds to HIV-1 receptor and coreceptors; mediates fusion between virus envelope and plasma membrane.	Induces apoptosis in CD4+ T cell; impairs dendritic cells-mediated CD4+ T cell response; increases HIV-1 spreading from dendritic cells to CD4+ T cells; reduces TLR signaling response; upregulates autophagy in neurons.	[92,95–100]
Tat	Regulates HIV-1 and cellular genes transcription; upregulates cell surface CXCR4 expression in uninfected CD4+ T cells; mediates T cell activation; impedes CD4+ and CD8+ maturation; alters mature CD8+ activity; modulates apoptosis.	Blocks IFN γ -mediated autophagy in macrophages/monocytes; inhibits autophagy in uninfected macrophages/monocytes by stimulating Src-Akt signaling pathway; alters autophagosome-lysosomes fusion in neurons.	[101–107]
Nef	Downregulates cell surface expression of CD4, MHC-I, MHC-II, CD28, T cell receptor/CD3 complex; prevents incorporation of restriction factors SERINC3/5 in budding viruses; modulates CD4+ T cell activation.	Prevents autophagy-mediated HIV-1 degradation by blocking autophagosome maturation in macrophages and astrocytes; blocks nuclear translocation of the pro-autophagy transcription factor TFEB in macrophages.	[47,108–110]
Vpr	Induces apoptosis in infected and/or uninfected T cells, DCs, monocytes and neurons; blocks the cell cycle at G2 phase; reactivates viral expression in latently infected cells; influences reverse transcription accuracy; regulates nuclear import of the HIV-1 pre-integration complex.	Prevents apoptosis in macrophages by diverting early steps of autophagy.	[111]
Vpu	Induces CD4 degradation; downregulates cell surface expression of MHC-I, CD1d, CCR7, CD40, CD81, SLAMF6; induces degradation of the restriction factor BST2; induces mistrafficking of BST2; drives displacing of BST2 from virus budding sites; depolarizes membrane potential by forming ion channel.	Drives displacing of BST2 from virus budding sites via a non-canonical autophagy mechanism (LC3-associated phagocytosis).	[112]

MHC: major histocompatibility complex; SERINC3/5: restriction factors serine incorporator 3 and 5; TFEB: transcription factor EB; CCR7: chemokine receptor 7; SLAMF6: SLAM family member 6; BST2: bone marrow stromal cell antigen 2; CXCR4: CXC chemokine receptor 4.

6.1. Envelope Proteins

HIV-1 Env, which promotes viral entry by binding to cell surface receptor CD4 and co-receptor CXCR4 (CXC chemokine receptor 4) or CCR5 and fusing viral and plasma membranes, was the first HIV-1 viral protein identified for its ability to subvert the autophagy mechanism. Indeed, Env has been shown to induce autophagy-dependent apoptosis of uninfected bystander CD4+ T cells, likely significantly contributing to AIDS pathogenesis by depleting the CD4+ T cell population [95,96]. Env is a trimer of the heterodimer composed of non-covalently associated subunits gp120 and gp41, which are dedicated to viral receptor interaction and membrane fusion, respectively. In these studies [95,96], the authors revealed that the binding of gp120 with the CXCR4 co-receptor, but not CD4, is essential for induction of autophagy-mediated apoptosis. However, this process is not directly triggered by the gp120-CXCR4 interaction, but rather by the fusogenic activity of gp41. Indeed, a point mutation in the fusion domain of gp41 is sufficient to inhibit autophagy and the ensuing cell death of CD4+ T cells [96]. How Env-mediated autophagy induces apoptosis in uninfected CD4+ T cells has not been determined yet. Nevertheless, it appears that this mechanism requires a fully functional autophagy pathway since upon treatment with early or late autophagy inhibitors, such as 3-MA and bafilomycin A1 (a specific inhibitor of autophagosome-lysosome fusion) respectively, or following depletion of Beclin 1 and ATG7, uninfected CD4+ T cells do not undergo cell death following contact of Env [95]. Surprisingly, Env induces increased caspase-3 (a protease involved in cell death execution) activity in uninfected CD4+ T cells, treatment with the pan-caspase inhibitor z-VAD does not prevent Env-mediated apoptosis in these cells [95]. This result suggests that both caspase-dependent and caspase-independent apoptosis are involved in this mechanism.

In contrast to what is observed in CD4+ T cells, it has been shown that gp120 downregulates autophagy in DC [92]. This mechanism depends in part on the engagement of CD4 receptor with gp120, which results in Erk-mTOR signaling pathway activation and subsequent autophagy inhibition. Interestingly, this study revealed that HIV-1-mediated downregulation of autophagy in DC leads to a reduced DC-mediated CD4+ T cell response (as mentioned above), an impaired TLR signaling and an enhanced HIV-1 transmission to CD4+ T cells. Therefore, these observations suggest that gp120-mediated inhibition of autophagy in DC could promote viral spreading.

HIV-1 can infect macrophages and microglia in the brain, resulting in a progressive neurological injury of the central nervous system (CNS) which is due in part to the neurotoxic effect of gp120 [113]. Indeed, although neurons cannot be productively infected by HIV-1, virions or free-gp120 released from infected macrophages and microglia impact viability and biological functions of neurons [114]. Dysregulation of autophagy has been reported to contribute to various neurodegenerative diseases [115]. Interestingly, a higher expression of autophagy proteins (Beclin 1, ATG5, ATG7, LC3B-II) and a greater number of autophagosomes per neurons are found in post-mortem brain tissues from HIV-1-infected patients developing HIV-associated encephalitis (HIVE) than those with no HIVE [97]. Furthermore, several studies have demonstrated that treatment of neuroblastoma cells or astrocytes with free-gp120 induces autophagy [97–100]. Consequently, gp120-mediated autophagy induction in the CNS may be associated with neuronal damage. In addition, gp120 seems to have a mild to strong effect on neuroblastoma cell apoptosis, whereas no cell death induction is observed in gp120-treated astrocytes [98–100]. Two cellular factors have been identified to play a critical role in gp120-mediated autophagy in neuroblastoma cells, the apoptosis-stimulating protein of p53-2 (ASPP2) and proline oxidase (POX) proteins [98,99]. ASPP2 is induced in the presence of gp120, and positively or negatively regulates autophagy and apoptosis in a gp120 concentration-dependent manner [99]. Thus, while ASPP2 overexpression inhibits autophagy and apoptosis at low doses of gp120, it promotes activation of these cellular pathways at high doses of gp120. gp120 has also been shown to trigger autophagy by upregulating *POX* gene expression in a p53-dependent manner following its interaction with cell surface CXCR4 [98]. More specifically, engagement of CXCR4 with gp120 results in upregulation and nuclear translocation of p53. In turn, p53 recognizes and activates the *POX* gene promoter. Thereafter, POX, by catalyzing its substrate, generates reactive oxygen species

(ROS), which induces autophagy in neuroblastoma cells. Why autophagy is induced in the presence of gp120 in neuronal cells remains to be determined. Nevertheless, it has been suggested that autophagy induction could function as a prosurvival response in these cells. This mechanism could be necessary to remove toxic stress and delay cell death, although contributing to neurological disorders [97,98].

6.2. Tat Protein

HIV-1 Tat has been shown to cause neuronal toxicity and dysfunction, and therefore seems to be a critical factor in HIV-1-associated neurocognitive disorder (HAND) [116–119]. In the brain, Tat is secreted by infected macrophages, astrocytes and microglia cells, but can also circulate in the bloodstream and cross the blood-brain barrier [120]. Tat can enter in neurons through endocytosis pathways which are mediated by numerous receptors such as CXCR4, CD26, heparin sulfate proteoglycans or low-density lipoprotein receptor-related proteins [121–125]. In the last few years, an increasing number of studies has found that Tat can modulate autophagy in brain tissues [101–105]. Interestingly, Tat regulates autophagy in a cell-type-dependent manner. It has been reported that Tat induces autophagy in human astrocyte cell lines and embryonic rat hippocampal neurons [103,105], and represses this mechanism in primary mouse neuron cells [104]. More surprisingly, Tat appears to have a dual role in autophagy in neuroblastoma cell lines depending on its concentration [101,102]. Indeed, autophagy is upregulated or downregulated at low or high doses of Tat, respectively. Moreover, while Tat reduces the number of autophagosomes and increases their size at low concentrations [102], it induces autophagosome accumulation and elicits the formation of misshapen autophagosomes at high concentration [101]. Interestingly, Fields and collaborators identified an interaction between Tat and lysosome-associated membrane protein 2 (LAMP2) [101], a lysosomal protein essential for chaperone-mediated autophagy and canonical autophagy [126,127]. Their work also revealed that Tat colocalizes with autophagosome and lysosomal markers and enhances autophagosome-lysosome fusion in neuroblastoma cells [101]. Therefore, the authors proposed that Tat, by interacting with LAMP2, may promote rapid and aberrant autophagosome-lysosome fusion, resulting in an accumulation of autophagolysosomes with altered morphology. Fields and collaborators have confirmed in vitro results by using an inducible Tat transgenic mouse model. Remarkably, they showed that Tat-mediated induction or repression of autophagy in neurons promote neurodegeneration [101,102]. In addition, they have demonstrated that Tat-mediated autophagy alteration can be reversed by treatment with autophagy inducers, reducing neuronal injury and improving locomotor activity in mice.

Tat also dysregulates autophagy mechanisms in macrophage/monocyte populations. In infected macrophages, Tat suppresses IFN γ -mediated autophagy by inhibiting phosphorylation of signal transducer and activator of transcription-1 (STAT1) [106], a crucial signaling molecule downstream of IFN γ receptor. This block impairs autophagosome-mediated clearance of intracellular pathogens in macrophages, as illustrated by the loss of colocalization between IFN γ -induced autophagosomes and *Bacillus Calmette Guerin* in the presence of Tat. In a study by Van Grol et al. [107], Tat has been shown to inhibit autophagy in uninfected macrophages/monocytes by stimulating the Src-Akt signaling pathway, known to negatively regulate this mechanism [128–130]. More specifically, Tat activates Src-Akt signaling by interacting with the cell surface receptors CXCR4, vascular endothelial growth factor receptor (VEGFR) and β -integrins which are upstream of this pathway. This process also requires the STAT3 protein and concomitant activation of the IL-10 signaling pathway. Considering that Tat is secreted by infected macrophages, further investigation is needed in order to determine whether Tat modulates autophagy mechanisms in an autocrine manner as well.

6.3. Nef Protein

As mentioned above, autophagy is tightly regulated in macrophages by HIV-1. While early stages of this mechanism are essential for viral production, the degradation process is deleterious. In this respect, it has been highlighted that the HIV-1 negative regulatory factor (Nef) protein plays a

pivotal role in the control of the late steps of autophagy. Nef is a multifunctional protein regulating several proteins with various functions in the cell [131]. Notably, Nef is well known for its ability to downregulate cell surface expression of CD4 and MHC complexes [132–135], and to antagonize the restriction factors serine incorporator 3 and 5 (SERINC 3/5) [136–138]. In 2009, Kyei et al. [47] identified an autophagy-inhibitory activity for Nef in macrophages. More specifically, they showed that Nef prevents autophagosome maturation and subsequent degradation of HIV-1. This effect is mediated by a direct or an indirect interaction of Nef with Beclin 1, and requires the Nef diacidic motif $_{174}DD_{175}$ [47] and the amino acid residues 267–284 in the evolutionarily conserved domain (ECD) of Beclin 1 [110]. Moreover, Nef is observed in intracellular structures that are positive for Beclin 1, ATG7 and ATG12. Interestingly, Nef-induced inhibition of autophagy appears to occur late in the course of macrophage infection; between days 3 and 5, post-infection [108]. Therefore, the authors proposed that Nef blocks autophagy once a productive infection of macrophages is established. They focused on the transcription factor EB (TFEB), a cellular protein that positively regulates the expression of several autophagy genes [139,140]. The authors particularly showed that, between 3 and 5 days post-infection, Nef suppresses the dephosphorylation of TFEB, preventing the ensuing nuclear translocation of this transcription factor, and hence inhibiting autophagy activity. In addition, Campbell et al. [108] revealed that, in the absence of Nef, HIV-1-induced TFEB translocation depends on Beclin 1, which is consistent with the Nef-Beclin 1 interaction identified previously. Note that, similarly to macrophages, Nef has also been shown to inhibit autophagosomes maturation in astrocytes [109], suggesting that Nef-mediated autophagy inhibition occurs in various cell types.

6.4. Vpr Protein

Very recently, a pro-autophagy activity was identified for the HIV-1 viral protein R (Vpr) [111]. More particularly, data revealed an increased level of LC3B and Beclin 1 in monocytic cell line-derived macrophages transiently expressing Vpr. Surprisingly, this increased level of autophagic markers is not accompanied by a greater number of autophagosomes in the cell, suggesting that only the early steps of autophagy are induced by Vpr. Interestingly, treatment with an early autophagy inhibitor promotes apoptosis in Vpr expressing macrophages. On the contrary, in the absence of Vpr, no increase in cell death is observed in early autophagy inhibitor-treated macrophages. Previous works in the literature noticed a dual role of Vpr in modulation of apoptosis in a cell type-dependent manner. Indeed, it has been shown that Vpr induces apoptosis in infected and/or uninfected lymphocytes, DCs, monocytes and neurons, but not in macrophages [141–148]. HIV-1-infected macrophages are known to be highly resistant to viral cytopathic effect and to have a prolonged lifespan [149–153], making these cells an HIV-1 reservoir. Thus, the role played by autophagy in the apoptotic properties of Vpr could be an important element in the disease progression. In this regard, it seems essential to further explore the interplay between Vpr, autophagy and apoptosis both in infected macrophages and in the other HIV-1 target cells.

7. Diversion of a Non-Canonical Autophagy Pathway, the LC3-Associated Phagocytosis, by HIV-1 Viral Protein Vpu

Several non-canonical autophagy mechanisms have been identified in the past decade [154]. Among these, the LC3-Associated Phagocytosis (LAP) process was highlighted in 2007 by Sanjuan et al. [155]. The authors observed that TLR2 stimulation with zymosan, a yeast cell wall glucan, results in the engulfment of the TLR2-zymosan complex in a phagosome decorated with LC3-II molecules in macrophages (Figure 3A). This process depends notably on ATG5 and ATG7 proteins. In addition, the authors noticed a rapid Beclin 1 association with phagosomes, and an increased PI3K complex activity preceding LC3 recruitment. However, contrary to canonical autophagy, it was shown that LAP does not require all autophagy machinery components, since this process is independent of the autophagy preinitiation complex [156,157]. In a major advance in 2015, Martinez et al. [158] defined more precisely the molecular steps controlling the LAP mechanism, and identified the Rubicon

protein as a LAP-specific marker. Thus, they revealed that, following zymosan internalization by phagocytosis, Rubicon is recruited at the delimiting membrane of the phagosome, resulting in local production of PI(3)P. Indeed, Rubicon interacts with a PI3K complex containing the UV radiation resistance-associated gene (UVRAG) protein, Beclin 1, VPS34, but not ATG14L or AMBRA1. Subsequently, PI(3)P mediates the recruitment of the NADPH oxidase-2 (NOX2) complex composed of NOX2 and the subunits p22PHOX, p40PHOX, p47PHOX and p67PHOX. This complex is stabilized at the phagosome membrane by a direct interaction between Rubicon and p22PHOX, inducing ROS production in the phagosome. ROS is demonstrated to be indispensable for LC3-II translocation at the phagosome membrane, although the precise mechanism has not been determined yet. The authors also demonstrated that components of the autophagy conjugation system (ATG5, ATG7, ATG12, ATG16L, ATG3 and ATG4B) are required for LC3-II recruitment. Finally, Martinez et al. [158] noticed that, by associating with phagosomes, LC3-II assists fusion between these structures and lysosomes, enhancing the maturation process.

Numerous studies have described the involvement of LAP in antibacterial [159–166], antifungus [167–169] and antiparasite [170–172] responses. Recently, our laboratory showed for the first time that a virus can hijack the LAP mechanism to promote viral spreading. Indeed, we demonstrated that the HIV-1 Vpu activity to counteract the cellular restriction factor bone marrow stromal cell antigen 2 (BST2) requires, in part, the LAP mechanism (Figure 3B) [112]. BST2 is a cell surface glycoprotein that is double-linked to cellular membranes through a C-terminus α -helical transmembrane domain and an extracellular N-terminus glycosylphosphatidylinositol anchor. This unusual protein structure allows BST2 to tether nascent viral particles to cellular membranes by inserting its extremities both into viral envelope and plasma membrane [173–176]. Therefore, BST2 prevents HIV-1 release, resulting in virus accumulation at the cell surface.

Prior to our recent study, some Vpu activities required to counterattack BST2 mediated restriction on HIV-1 budding had been highlighted. First, Vpu accelerates the targeting of BST2 to the lysosomal degradation pathway by promoting ubiquitination of the newly synthesized and/or recycling BST2 molecules trafficking through endosomes [177–180]. This mechanism leads to reduced BST2 recycling to the cell surface and BST2 cell surface downregulation. Independently, Vpu also favors HIV-1 release by interfering with BST2 trafficking via interaction of Vpu with clathrin adaptor protein complex-1 (AP-1) and AP-2 [181–183], or by directly removing BST2 from virus budding sites at the plasma membrane [184–186]. Our recent work reveals that Vpu can also counteract BST2 restriction by hijacking a non-canonical autophagy process, reminiscent of a LAP process [112]. Indeed, in HeLa cells infected with VSV-G (vesicular stomatitis virus glycoprotein)-pseudotyped HIV-1 viruses, we observed that depletion of Beclin 1, ATG5 and LC3C, but not of preinitiation complex components (ULK1 and FIP200), leads to reduced HIV-1 release in a BST2 dependent manner. We describe a direct interaction between Vpu and LC3C mediated by the Vpu₆₃LVEM₆₆ motif. Mutation in this motif also results in decreased HIV-1 release. Furthermore, LC3C depletion results in virus sequestration in intracellular single membrane-bound compartments very similar to LC3-associated phagosomes. Interestingly, we noticed that LC3C is involved in Vpu-mediated removal of BST2 from virus budding site without affecting the BST2 turnover at the cell surface. This observation indicates that the LAP-dependent Vpu-mediated removing of BST2 is independent of Vpu-mediated BST2 degradation.

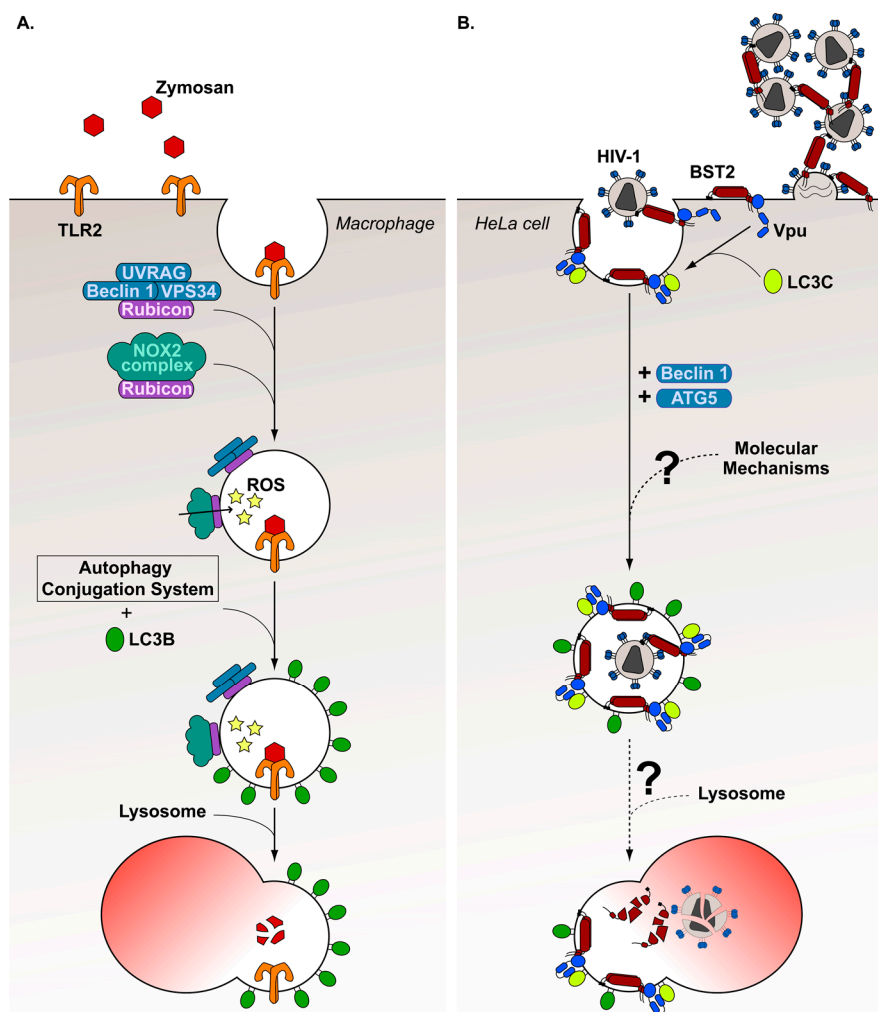


Figure 3. Comparative overview of LC3-Associated Phagocytosis (LAP)- and LC3C-dependent Vpu-mediated BST2 antagonism mechanisms. **(A)** Following its engagement with Toll-like receptor 2 (TLR2), zymosan is internalized by phagocytosis in macrophages. Subsequently, a PI3K complex composed of Rubicon, VPS34, Beclin 1 and UV radiation resistance-associated gene (UVRAG) proteins is recruited to the delimiting membrane of the phagosome resulting in a local production of phosphatidylinositol-3-phosphate (PI(3)P). This latter promotes recruitment of the NADPH oxidase-2 (NOX2) complex, which is responsible for reactive oxygen species (ROS) production into the phagosome. ROS, as along with the autophagy conjugation system, leads to decoration of the phagosome with LC3B, accelerating the phagosome-lysosome fusion process. **(B)** Following its interaction with the BST2 transmembrane domain at plasma membrane, Vpu recruits the LC3C protein to enhance internalization of BST2 into intracellular compartments, which are decorated with LC3 orthologs. This process requires Beclin 1 and ATG5 but not components of the preinitiation complex of autophagy (ULK1 and FIP200), which is reminiscent of the LAP mechanism, and results in the removal of BST2 from virus budding sites. However, the precise mechanisms by which this intracellular compartment is formed and the fate of this compartment within the cell are not yet determined.

8. Concluding Remarks

In the past decade, it has emerged that the autophagy pathway intersects with the HIV-1 life cycle. This review highlights the fact that autophagy acts as a double-edged sword in HIV-1 infection. Autophagy provides not only an integral part of intracellular defenses developed by infected cells to eliminate HIV-1 and mount a pro-inflammatory response, but also serves an essential role in the proper production and dissemination of this virus. HIV-1 finely regulates the autophagy pathway,

thereby making the analysis of this intricate interplay challenging. Furthermore, it appears that the role of autophagy in HIV-1 infection is very closely related to the cell type, given that the early steps of autophagy are required for viral replication in macrophages, but are detrimental in CD4+ T cells. In line with this, the HIV-1 genome encodes both pro- and anti-autophagy proteins, some of which display a dual role in a cell type-dependent manner. The molecular mechanisms in which early and late steps of autophagy are engaged in these different cell types remains largely unclear, and thus requires further investigation in the future. Moreover, the time course of viral infection should also be taken into consideration in future studies, in order to better understand the interplay between HIV-1 and autophagy and to resolve some discrepancies observed in previous studies. Finally, the recent observation that HIV-1 hijacks a non-canonical autophagy mechanism to counteract an innate barrier to viral infection could notably provide some clues to understand the benefit of autophagy proteins for HIV-1 production. Further investigation will be needed to precisely decipher the engagement of the LAP process in viral production in HIV-1 target cells. Addressing all these questions should provide the basis for the development of new drug design strategies to combat the virus.

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References

1. Gutierrez, M.G.; Master, S.S.; Singh, S.B.; Taylor, G.A.; Colombo, M.I.; Deretic, V. Autophagy is a defense mechanism inhibiting BCG and *Mycobacterium tuberculosis* survival in infected macrophages. *Cell* **2004**, *119*, 753–766. [[CrossRef](#)] [[PubMed](#)]
2. Nakagawa, I.; Amano, A.; Mizushima, N.; Yamamoto, A.; Yamaguchi, H.; Kamimoto, T.; Nara, A.; Funao, J.; Nakata, M.; Tsuda, K.; et al. Autophagy defends cells against invading group A Streptococcus. *Science* **2004**, *306*, 1037–1040. [[CrossRef](#)] [[PubMed](#)]
3. Ogawa, M.; Yoshimori, T.; Suzuki, T.; Sagara, H.; Mizushima, N.; Sasakawa, C. Escape of intracellular Shigella from autophagy. *Science* **2005**, *307*, 727–731. [[CrossRef](#)] [[PubMed](#)]
4. Paludan, C.; Schmid, D.; Landthaler, M.; Vockerodt, M.; Kube, D.; Tuschl, T.; Münz, C. Endogenous MHC class II processing of a viral nuclear antigen after autophagy. *Science* **2005**, *307*, 593–596. [[CrossRef](#)] [[PubMed](#)]
5. Wesselborg, S.; Stork, B. Autophagy signal transduction by ATG proteins: From hierarchies to networks. *Cell. Mol. Life Sci.* **2015**, *72*, 4721–4757. [[CrossRef](#)] [[PubMed](#)]
6. Ylä-Anttila, P.; Vihinen, H.; Jokitalo, E.; Eskelinen, E.-L. 3D tomography reveals connections between the phagophore and endoplasmic reticulum. *Autophagy* **2009**, *5*, 1180–1185. [[CrossRef](#)] [[PubMed](#)]
7. Hayashi-Nishino, M.; Fujita, N.; Noda, T.; Yamaguchi, A.; Yoshimori, T.; Yamamoto, A. A subdomain of the endoplasmic reticulum forms a cradle for autophagosome formation. *Nat. Cell Biol.* **2009**, *11*, 1433–1437. [[CrossRef](#)] [[PubMed](#)]
8. Hosokawa, N.; Sasaki, T.; Iemura, S.I.; Natsume, T.; Hara, T.; Mizushima, N. Atg101, a novel mammalian autophagy protein interacting with Atg13. *Autophagy* **2009**, *5*, 973–979. [[CrossRef](#)] [[PubMed](#)]
9. Mercer, C.A.; Kaliappan, A.; Dennis, P.B. A novel, human Atg13 binding protein, Atg101, interacts with ULK1 and is essential for macroautophagy. *Autophagy* **2009**, *5*, 649–662. [[CrossRef](#)] [[PubMed](#)]
10. Jung, C.H.; Jun, C.B.; Ro, S.-H.; Kim, Y.-M.; Otto, N.M.; Cao, J.; Kundu, M.; Kim, D.-H. ULK-Atg13-FIP200 Complexes Mediate mTOR Signaling to the Autophagy Machinery. *Mol. Biol. Cell* **2009**, *20*, 1992–2003. [[CrossRef](#)] [[PubMed](#)]
11. Hosokawa, N.; Hara, T.; Kaizuka, T.; Kishi, C.; Takamura, A.; Miura, Y.; Iemura, S.-I.; Natsume, T.; Takehana, K.; Yamada, N.; et al. Nutrient-dependent mTORC1 Association with the ULK1-Atg13-FIP200 Complex Required for Autophagy. *Mol. Biol. Cell* **2009**, *20*, 1981–1991. [[CrossRef](#)] [[PubMed](#)]
12. Ganley, I.G.; Lam, D.H.; Wang, J.; Ding, X.; Chen, S.; Jiang, X. ULK1.ATG13.FIP200 Complex Mediates mTOR Signaling and Is Essential for Autophagy. *J. Biol. Chem.* **2009**, *284*, 12297–12305. [[CrossRef](#)] [[PubMed](#)]
13. Shigemitsu, K.; Tsujishita, Y.; Hara, K.; Nanahoshi, M.; Avruch, J.; Yonezawa, K. Regulation of translational effectors by amino acid and mammalian target of rapamycin signaling pathways. Possible involvement of autophagy in cultured hepatoma cells. *J. Biol. Chem.* **1999**, *274*, 1058–1065. [[CrossRef](#)] [[PubMed](#)]

14. Itakura, E.; Mizushima, N. Characterization of autophagosome formation site by a hierarchical analysis of mammalian Atg proteins. *Autophagy* **2010**, *6*, 764–776. [[CrossRef](#)] [[PubMed](#)]
15. Karanasiou, E.; Stapleton, E.; Manifava, M.; Kaizuka, T.; Mizushima, N.; Walker, S.A.; Ktistakis, N.T. Dynamic association of the ULK1 complex with omegasomes during autophagy induction. *J. Cell Sci.* **2013**, *126*, 5224–5238. [[CrossRef](#)] [[PubMed](#)]
16. Axe, E.L.; Walker, S.A.; Manifava, M.; Chandra, P.; Roderick, H.L.; Habermann, A.; Griffiths, G.; Ktistakis, N.T. Autophagosome formation from membrane compartments enriched in phosphatidylinositol 3-phosphate and dynamically connected to the endoplasmic reticulum. *J. Cell Biol.* **2008**, *182*, 685–701. [[CrossRef](#)] [[PubMed](#)]
17. Petiot, A.; Ogier-Denis, E.; Blommaert, E.F.; Meijer, A.J.; Codogno, P. Distinct classes of phosphatidylinositol 3'-kinases are involved in signaling pathways that control macroautophagy in HT-29 cells. *J. Biol. Chem.* **2000**, *275*, 992–998. [[CrossRef](#)] [[PubMed](#)]
18. Blommaert, E.F.; Krause, U.; Schellens, J.P.; Vreeling-Sindelárová, H.; Meijer, A.J. The phosphatidylinositol 3-kinase inhibitors wortmannin and LY294002 inhibit autophagy in isolated rat hepatocytes. *Eur. J. Biochem.* **1997**, *243*, 240–246. [[CrossRef](#)] [[PubMed](#)]
19. Hamasaki, M.; Furuta, N.; Matsuda, A.; Nezu, A.; Yamamoto, A.; Fujita, N.; Oomori, H.; Noda, T.; Haraguchi, T.; Hiraoka, Y.; et al. Autophagosomes form at ER-mitochondria contact sites. *Nature* **2013**, *495*, 389–393. [[CrossRef](#)] [[PubMed](#)]
20. Kirisako, T.; Baba, M.; Ishihara, N.; Miyazawa, K.; Ohsumi, M.; Yoshimori, T.; Noda, T.; Ohsumi, Y. Formation process of autophagosome is traced with Apg8/Aut7p in yeast. *J. Cell Biol.* **1999**, *147*, 435–446. [[CrossRef](#)] [[PubMed](#)]
21. Mizushima, N.; Kuma, A.; Kobayashi, Y.; Yamamoto, A.; Matsubae, M.; Takao, T.; Natsume, T.; Ohsumi, Y.; Yoshimori, T. Mouse Apg16L, a novel WD-repeat protein, targets to the autophagic isolation membrane with the Apg12-Apg5 conjugate. *J. Cell Sci.* **2003**, *116*, 1679–1688. [[CrossRef](#)] [[PubMed](#)]
22. Kabeya, Y.; Mizushima, N.; Ueno, T.; Yamamoto, A.; Kirisako, T.; Noda, T.; Kominami, E.; Ohsumi, Y.; Yoshimori, T. LC3, a mammalian homologue of yeast Apg8p, is localized in autophagosome membranes after processing. *EMBO J.* **2000**, *19*, 5720–5728. [[CrossRef](#)] [[PubMed](#)]
23. Mizushima, N.; Yamamoto, A.; Hatano, M.; Kobayashi, Y.; Kabeya, Y.; Suzuki, K.; Tokuhisa, T.; Ohsumi, Y.; Yoshimori, T. Dissection of autophagosome formation using Apg5-deficient mouse embryonic stem cells. *J. Cell Biol.* **2001**, *152*, 657–668. [[CrossRef](#)] [[PubMed](#)]
24. Ohsumi, Y.; Mizushima, N.; Noda, T.; Yoshimori, T.; Tanaka, Y.; Ishii, T.; George, M.D.; Klionsky, D.J.; Ohsumi, M. A protein conjugation system essential for autophagy. *Nature* **1998**, *395*, 395–398. [[CrossRef](#)] [[PubMed](#)]
25. Shintani, T.; Mizushima, N.; Ogawa, Y.; Matsuura, A.; Noda, T.; Ohsumi, Y. Apg10p, a novel protein-conjugating enzyme essential for autophagy in yeast. *EMBO J.* **1999**, *18*, 5234–5241. [[CrossRef](#)] [[PubMed](#)]
26. Mizushima, N.; Sugita, H.; Yoshimori, T.; Ohsumi, Y. A new protein conjugation system in human. The counterpart of the yeast Apg12p conjugation system essential for autophagy. *J. Biol. Chem.* **1998**, *273*, 33889–33892. [[CrossRef](#)] [[PubMed](#)]
27. Ohsumi, Y.; Ichimura, Y.; Kirisako, T.; Takao, T.; Satomi, Y.; Shimonishi, Y.; Ishihara, N.; Mizushima, N.; Tanida, I.; Kominami, E.; et al. A ubiquitin-like system mediates protein lipidation. *Nature* **2000**, *408*, 488–492. [[CrossRef](#)] [[PubMed](#)]
28. Tanida, I.; Tanida-Miyake, E.; Ueno, T.; Kominami, E. The human homolog of *Saccharomyces cerevisiae* Apg7p is a Protein-activating enzyme for multiple substrates including human Apg12p, GATE-16, GABARAP, and MAP-LC3. *J. Biol. Chem.* **2001**, *276*, 1701–1706. [[CrossRef](#)] [[PubMed](#)]
29. Mizushima, N.; Yoshimori, T.; Levine, B. Methods in Mammalian Autophagy Research. *Cell* **2010**, *140*, 313–326. [[CrossRef](#)] [[PubMed](#)]
30. Fujita, N.; Itoh, T.; Oomori, H.; Fukuda, M.; Noda, T.; Yoshimori, T. The Atg16L complex specifies the site of LC3 lipidation for membrane biogenesis in autophagy. *Mol. Biol. Cell* **2008**, *19*, 2092–2100. [[CrossRef](#)] [[PubMed](#)]
31. Xie, Z.; Nair, U.; Klionsky, D.J. Atg8 Controls Phagophore Expansion during Autophagosome Formation. *Mol. Biol. Cell* **2008**, *19*, 3290–3298. [[CrossRef](#)] [[PubMed](#)]

32. Fujita, N.; Hayashi-Nishino, M.; Fukumoto, H.; Omori, H.; Yamamoto, A.; Noda, T.; Yoshimori, T. An Atg4B mutant hampers the lipidation of LC3 paralogues and causes defects in autophagosome closure. *Mol. Biol. Cell* **2008**, *19*, 4651–4659. [[CrossRef](#)] [[PubMed](#)]
33. Nozawa, T.; Aikawa, C.; Goda, A.; Maruyama, F.; Hamada, S.; Nakagawa, I. The small GTPases Rab9A and Rab23 function at distinct steps in autophagy during Group A Streptococcus infection. *Cell. Microbiol.* **2012**, *14*, 1149–1165. [[CrossRef](#)] [[PubMed](#)]
34. Munafó, D.B.; Colombo, M.I. Induction of autophagy causes dramatic changes in the subcellular distribution of GFP-Rab24. *Traffic* **2002**, *3*, 472–482. [[CrossRef](#)] [[PubMed](#)]
35. Pilli, M.; Arko-Mensah, J.; Ponpuak, M.; Roberts, E.; Master, S.; Mandell, M.A.; Dupont, N.; Ornatowski, W.; Jiang, S.; Bradfute, S.B.; et al. TBK-1 promotes autophagy-mediated antimicrobial defense by controlling autophagosome maturation. *Immunity* **2012**, *37*, 223–234. [[CrossRef](#)] [[PubMed](#)]
36. Fader, C.M.; Sánchez, D.; Furlán, M.; Colombo, M.I. Induction of autophagy promotes fusion of multivesicular bodies with autophagic vacuoles in k562 cells. *Traffic* **2008**, *9*, 230–250. [[CrossRef](#)] [[PubMed](#)]
37. Itakura, E.; Kishi-Itakura, C.; Mizushima, N. The hairpin-type tail-anchored SNARE syntaxin 17 targets to autophagosomes for fusion with endosomes/lysosomes. *Cell* **2012**, *151*, 1256–1269. [[CrossRef](#)] [[PubMed](#)]
38. Furuta, N.; Fujita, N.; Noda, T.; Yoshimori, T.; Amano, A. Combinational Soluble N-Ethylmaleimide-sensitive Factor Attachment Protein Receptor Proteins VAMP8 and Vti1b Mediate Fusion of Antimicrobial and Canonical Autophagosomes with Lysosomes. *Mol. Biol. Cell* **2010**, *21*, 1001–1010. [[CrossRef](#)] [[PubMed](#)]
39. Fader, C.M.; Sánchez, D.G.; Mestre, M.B.; Colombo, M.I. TI-VAMP/VAMP7 and VAMP3/cellubrevin: Two v-SNARE proteins involved in specific steps of the autophagy/multivesicular body pathways. *Biochim. Biophys. Acta-Mol. Cell Res.* **2009**, *1793*, 1901–1916. [[CrossRef](#)] [[PubMed](#)]
40. Kumar, P. Long term non-progressor (LTNP) HIV infection. *Indian J. Med. Res.* **2013**, *138*, 291–293. [[PubMed](#)]
41. Gaardbo, J.C.; Hartling, H.J.; Gerstoft, J.; Nielsen, S.D. Thirty Years with HIV Infection-Nonprogression Is Still Puzzling: Lessons to Be Learned from Controllers and Long-Term Nonprogressors. *AIDS Res. Treat.* **2012**, *2012*, 161584. [[CrossRef](#)] [[PubMed](#)]
42. Okulicz, J.F.; Marconi, V.C.; Landrum, M.L.; Wegner, S.; Weintrob, A.; Ganesan, A.; Hale, B.; Crum-Cianflone, N.; Delmar, J.; Barthel, V.; et al. Infectious Disease Clinical Research Program (IDCRP) HIV Working Group Clinical outcomes of elite controllers, viremic controllers, and long-term nonprogressors in the US Department of Defense HIV natural history study. *J. Infect. Dis.* **2009**, *200*, 1714–1723. [[CrossRef](#)] [[PubMed](#)]
43. Lambotte, O.; Boufassa, F.; Madec, Y.; Nguyen, A.; Goujard, C.; Meyer, L.; Rouzioux, C.; Venet, A.; Delfraissy, J.-F. SEROCO-HEMOCO Study Group HIV Controllers: A Homogeneous Group of HIV-1-Infected Patients with Spontaneous Control of Viral Replication. *Clin. Infect. Dis.* **2005**, *41*, 1053–1056. [[CrossRef](#)] [[PubMed](#)]
44. Grabar, S.; Selinger-Leneman, H.; Abgrall, S.; Pialoux, G.; Weiss, L.; Costagliola, D. Prevalence and comparative characteristics of long-term nonprogressors and HIV controller patients in the French Hospital Database on HIV. *AIDS* **2009**, *23*, 1163–1169. [[CrossRef](#)] [[PubMed](#)]
45. Nardacci, R.; Amendola, A.; Ciccocanti, F.; Corazzari, M.; Esposito, V.; Vlassi, C.; Taibi, C.; Fimia, G.M.; del Nonno, F.; Ippolito, G.; et al. Autophagy plays an important role in the containment of HIV-1 in nonprogressor-infected patients. *Autophagy* **2014**, *10*, 1167–1178. [[CrossRef](#)] [[PubMed](#)]
46. Espert, L.; Varbanov, M.; Robert-Hebmann, V.; Sagnier, S.; Robbins, I.; Sanchez, F.; Lafont, V.; Biard-Piechaczyk, M. Differential role of autophagy in CD4 T cells and macrophages during X4 and R5 HIV-1 infection. *PLoS ONE* **2009**, *4*, e5787. [[CrossRef](#)] [[PubMed](#)]
47. Kyei, G.B.; Dinkins, C.; Davis, A.S.; Roberts, E.; Singh, S.B.; Dong, C.; Wu, L.; Kominami, E.; Ueno, T.; Yamamoto, A.; et al. Autophagy pathway intersects with HIV-1 biosynthesis and regulates viral yields in macrophages. *J. Cell Biol.* **2009**, *186*, 255–268. [[CrossRef](#)] [[PubMed](#)]
48. Zhou, D.; Spector, S.A. Human immunodeficiency virus type-1 infection inhibits autophagy. *AIDS* **2008**, *22*, 695. [[CrossRef](#)] [[PubMed](#)]
49. Laforge, M.; Limou, S.; Harper, F.; Casartelli, N.; Rodrigues, V.; Silvestre, R.; Haloui, H.; Zagury, J.F.; Senik, A.; Estaquier, J. DRAM Triggers Lysosomal Membrane Permeabilization and Cell Death in CD4+ T Cells Infected with HIV. *PLoS Pathog.* **2013**, *9*. [[CrossRef](#)] [[PubMed](#)]

50. Sagnier, S.; Daussy, C.F.; Borel, S.; Robert-Hebmann, V.; Faure, M.; Blanchet, F.P.; Beaumelle, B.; Biard-Piechaczyk, M.; Espert, L. Autophagy Restricts HIV-1 Infection by Selectively Degrading Tat in CD4⁺ T Lymphocytes. *J. Virol.* **2015**, *89*, 615–625. [[CrossRef](#)] [[PubMed](#)]
51. Crichton, D.; Wilkinson, S.; O'Prey, J.; Syed, N.; Smith, P.; Harrison, P.R.; Gasco, M.; Garrone, O.; Crook, T.; Ryan, K.M. DRAM, a p53-Induced Modulator of Autophagy, Is Critical for Apoptosis. *Cell* **2006**, *126*, 121–134. [[CrossRef](#)] [[PubMed](#)]
52. Bieniasz, P.D. Intrinsic immunity: A front-line defense against viral attack. *Nat. Immunol.* **2004**, *5*, 1109–1115. [[CrossRef](#)] [[PubMed](#)]
53. Pankiv, S.; Clausen, T.H.; Lamark, T.; Brech, A.; Bruun, J.-A.; Outzen, H.; Øvervatn, A.; Bjørkøy, G.; Johansen, T. p62/SQSTM1 binds directly to Atg8/LC3 to facilitate degradation of ubiquitinated protein aggregates by autophagy. *J. Biol. Chem.* **2007**, *282*, 24131–24145. [[CrossRef](#)] [[PubMed](#)]
54. Clark, E.; Nava, B.; Caputi, M. Tat is a multifunctional viral protein that modulates cellular gene expression and functions. *Oncotarget* **2017**, *8*, 27569–27581. [[CrossRef](#)] [[PubMed](#)]
55. Pertel, T.; Hausmann, S.; Morger, D.; Züger, S.; Guerra, J.; Lascano, J.; Reinhard, C.; Santoni, F.A.; Uchil, P.D.; Chatel, L.; et al. TRIM5 is an innate immune sensor for the retrovirus capsid lattice. *Nature* **2011**, *472*, 361–365. [[CrossRef](#)] [[PubMed](#)]
56. Ganser-Pornillos, B.K.; Chandrasekaran, V.; Pornillos, O.; Sodroski, J.G.; Sundquist, W.I.; Yeager, M. Hexagonal assembly of a restricting TRIM5 α protein. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 534–539. [[CrossRef](#)] [[PubMed](#)]
57. Stremlau, M.; Perron, M.; Lee, M.; Li, Y.; Song, B.; Javanbakht, H.; Diaz-Griffero, F.; Anderson, D.J.; Sundquist, W.I.; Sodroski, J. Specific recognition and accelerated uncoating of retroviral capsids by the TRIM5 α restriction factor. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 5514–5519. [[CrossRef](#)] [[PubMed](#)]
58. Mandell, M.A.; Jain, A.; Arko-Mensah, J.; Chauhan, S.; Kimura, T.; Dinkins, C.; Silvestri, G.; Münch, J.; Kirchhoff, F.; Simonsen, A.; et al. TRIM proteins regulate autophagy and can target autophagic substrates by direct recognition. *Dev. Cell* **2014**, *30*, 394–409. [[CrossRef](#)] [[PubMed](#)]
59. Ribeiro, C.M.S.; Sarrami-Forooshani, R.; Setiawan, L.C.; Zijlstra-Willems, E.M.; van Hamme, J.L.; Tigchelaar, W.; van der Wel, N.N.; Kootstra, N.A.; Gringhuis, S.I.; Geijtenbeek, T.B.H. Receptor usage dictates HIV-1 restriction by human TRIM5 α in dendritic cell subsets. *Nature* **2016**, *1*, 448–452. [[CrossRef](#)] [[PubMed](#)]
60. Stremlau, M.; Owens, C.M.; Perron, M.J.; Kiessling, M.; Autissier, P.; Sodroski, J. The cytoplasmic body component TRIM5 α restricts HIV-1 infection in Old World monkeys. *Nature* **2004**, *427*, 848–853. [[CrossRef](#)] [[PubMed](#)]
61. Yap, M.W.; Nisole, S.; Stoye, J.P. A single amino acid change in the SPRY domain of human TRIM5 α leads to HIV-1 restriction. *Curr. Biol.* **2005**, *15*, 73–78. [[CrossRef](#)] [[PubMed](#)]
62. Sawyer, S.L.; Wu, L.I.; Emerman, M.; Malik, H.S. Positive selection of primate TRIM5 α identifies a critical species-specific retroviral restriction domain. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 2832–2837. [[CrossRef](#)] [[PubMed](#)]
63. de Witte, L.; Nabatov, A.; Pion, M.; Fluitsma, D.; de Jong, M.A.W.P.; de Gruijl, T.; Piguet, V.; van Kooyk, Y.; Geijtenbeek, T.B.H. Langerin is a natural barrier to HIV-1 transmission by Langerhans cells. *Nat. Med.* **2007**, *13*, 367–371. [[CrossRef](#)] [[PubMed](#)]
64. Gringhuis, S.I.; van der Vlist, M.; van den Berg, L.M.; den Dunnen, J.; Litjens, M.; Geijtenbeek, T.B.H. HIV-1 exploits innate signaling by TLR8 and DC-SIGN for productive infection of dendritic cells. *Nat. Immunol.* **2010**, *11*, 419–426. [[CrossRef](#)] [[PubMed](#)]
65. Mc Dermott, R.; Ziylan, U.; Spehner, D.; Bausinger, H.; Lipsker, D.; Mommaas, M.; Cazenave, J.-P.; Raposo, G.; Goud, B.; de la Salle, H.; et al. Birbeck granules are subdomains of endosomal recycling compartment in human epidermal Langerhans cells, which form where Langerin accumulates. *Mol. Biol. Cell* **2002**, *13*, 317–335. [[CrossRef](#)] [[PubMed](#)]
66. Mehle, A.; Strack, B.; Ancuta, P.; Zhang, C.; McPike, M.; Gabuzda, D. Vif overcomes the innate antiviral activity of APOBEC3G by promoting its degradation in the ubiquitin-proteasome pathway. *J. Biol. Chem.* **2004**, *279*, 7792–7798. [[CrossRef](#)] [[PubMed](#)]
67. Mangeat, B.; Turelli, P.; Caron, G.; Friedli, M.; Perrin, L.; Trono, D. Broad antiretroviral defence by human APOBEC3G through lethal editing of nascent reverse transcripts. *Nature* **2003**, *424*, 99–103. [[CrossRef](#)] [[PubMed](#)]

68. Iwatani, Y.; Chan, D.S.B.; Wang, F.; Maynard, K.S.; Sugiura, W.; Gronenborn, A.M.; Rouzina, I.; Williams, M.C.; Musier-Forsyth, K.; et al. Deaminase-independent inhibition of HIV-1 reverse transcription by APOBEC3G. *Nucleic Acids Res.* **2007**, *35*, 7096–7108. [[CrossRef](#)] [[PubMed](#)]
69. Mbisa, J.L.; Barr, R.; Thomas, J.A.; Vandegraaff, N.; Dorweiler, I.J.; Svarovskaia, E.S.; Brown, W.L.; Mansky, L.M.; Gorelick, R.J.; Harris, R.S.; et al. Human immunodeficiency virus type 1 cDNAs produced in the presence of APOBEC3G exhibit defects in plus-strand DNA transfer and integration. *J. Virol.* **2007**, *81*, 7099–7110. [[CrossRef](#)] [[PubMed](#)]
70. Guo, F.; Cen, S.; Niu, M.; Saadatmand, J.; Kleiman, L. Inhibition of tRNA₃(Lys)-primed reverse transcription by human APOBEC3G during human immunodeficiency virus type 1 replication. *J. Virol.* **2006**, *80*, 11710–11722. [[CrossRef](#)] [[PubMed](#)]
71. Mbisa, J.L.; Bu, W.; Pathak, V.K. APOBEC3F and APOBEC3G inhibit HIV-1 DNA integration by different mechanisms. *J. Virol.* **2010**, *84*, 5250–5259. [[CrossRef](#)] [[PubMed](#)]
72. Valera, M.-S.; de Armas-Rillo, L.; Barroso-González, J.; Ziglio, S.; Batisse, J.; Dubois, N.; Marrero-Hernández, S.; Borel, S.; García-Expósito, L.; Biard-Piechaczyk, M.; et al. The HDAC6/APOBEC3G complex regulates HIV-1 infectiveness by inducing Vif autophagic degradation. *Retrovirology* **2015**, *12*, 53. [[CrossRef](#)] [[PubMed](#)]
73. Kopito, R.R. Aggresomes, inclusion bodies and protein aggregation. *Trends Cell Biol.* **2000**, *10*, 524–530. [[CrossRef](#)]
74. Kawaguchi, Y.; Kovacs, J.J.; McLaurin, A.; Vance, J.M.; Ito, A.; Yao, T.P. The deacetylase HDAC6 regulates aggresome formation and cell viability in response to misfolded protein stress. *Cell* **2003**, *115*, 727–738. [[CrossRef](#)]
75. Lee, J.-Y.; Koga, H.; Kawaguchi, Y.; Tang, W.; Wong, E.; Gao, Y.-S.; Pandey, U.B.; Kaushik, S.; Tresse, E.; Lu, J.; et al. HDAC6 controls autophagosome maturation essential for ubiquitin-selective quality-control autophagy. *EMBO J.* **2010**, *29*, 969–980. [[CrossRef](#)] [[PubMed](#)]
76. Campbell, G.R.; Bruckman, R.S.; Chu, Y.L.; Spector, S.A. Autophagy induction by histone deacetylase inhibitors inhibits HIV type 1. *J. Biol. Chem.* **2015**, *290*, 5028–5040. [[CrossRef](#)] [[PubMed](#)]
77. Delcuve, G.P.; Khan, D.H.; Davie, J.R. Roles of histone deacetylases in epigenetic regulation: emerging paradigms from studies with inhibitors. *Clin. Epigenet.* **2012**, *4*, 5. [[CrossRef](#)] [[PubMed](#)]
78. Witt, O.; Deubzer, H.E.; Milde, T.; Oehme, I. HDAC family: What are the cancer relevant targets? *Cancer Lett.* **2009**, *277*, 8–21. [[CrossRef](#)] [[PubMed](#)]
79. Campbell, G.R.; Spector, S.A. Toll-Like Receptor 8 Ligands Activate a Vitamin D Mediated Autophagic Response that Inhibits Human Immunodeficiency Virus Type 1. *PLoS Pathog.* **2012**, *8*, 1003017. [[CrossRef](#)] [[PubMed](#)]
80. Yuk, J.-M.; Shin, D.-M.; Lee, H.-M.; Yang, C.-S.; Jin, H.S.; Kim, K.-K.; Lee, Z.-W.; Lee, S.-H.; Kim, J.-M.; Jo, E.-K. Vitamin D3 induces autophagy in human monocytes/macrophages via cathelicidin. *Cell Host Microbe* **2009**, *6*, 231–243. [[CrossRef](#)] [[PubMed](#)]
81. Zhou, D.; Kang, K.H.; Spector, S.A. Production of Interferon by Human Immunodeficiency Virus Type 1 in Human Plasmacytoid Dendritic Cells Is Dependent on Induction of Autophagy. *J. Infect. Dis.* **2012**, *205*, 1258–1267. [[CrossRef](#)] [[PubMed](#)]
82. O'Brien, M.; Manches, O.; Bhardwaj, N. Plasmacytoid dendritic cells in HIV infection. *Adv. Exp. Med. Biol.* **2013**, *762*, 71–107. [[PubMed](#)]
83. Coulon, P.-G.; Richetta, C.; Rouers, A.; Blanchet, F.P.; Urrutia, A.; Guerbois, M.; Piguat, V.; Theodorou, I.; Bet, A.; Schwartz, O.; et al. HIV-Infected Dendritic Cells Present Endogenous MHC Class II-Restricted Antigens to HIV-Specific CD4+ T Cells. *J. Immunol.* **2016**, *197*, 517–532. [[CrossRef](#)] [[PubMed](#)]
84. Granelli-Piperno, A.; Golebiowska, A.; Trumpfheller, C.; Siegal, F.P.; Steinman, R.M. HIV-1-infected monocyte-derived dendritic cells do not undergo maturation but can elicit IL-10 production and T cell regulation. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 7669–7674. [[CrossRef](#)] [[PubMed](#)]
85. Macatonia, S.E.; Lau, R.; Patterson, S.; Pinching, A.J.; Knight, S.C. Dendritic cell infection, depletion and dysfunction in HIV-infected individuals. *Immunology* **1990**, *71*, 38–45. [[PubMed](#)]
86. Münz, C. Autophagy Beyond Intracellular MHC Class II Antigen Presentation. *Trends Immunol.* **2016**, *37*, 755–763. [[CrossRef](#)] [[PubMed](#)]

87. Lee, H.K.; Mattei, L.M.; Steinberg, B.E.; Alberts, P.; Lee, Y.H.; Chervonsky, A.; Mizushima, N.; Grinstein, S.; Iwasaki, A. In Vivo Requirement for Atg5 in Antigen Presentation by Dendritic Cells. *Immunity* **2010**, *32*, 227–239. [[CrossRef](#)] [[PubMed](#)]
88. Schmid, D.; Pypaert, M.; Münz, C. Antigen-loading compartments for major histocompatibility complex class II molecules continuously receive input from autophagosomes. *Immunity* **2007**, *26*, 79–92. [[CrossRef](#)] [[PubMed](#)]
89. Leib, D.A.; Alexander, D.E.; Cox, D.; Yin, J.; Ferguson, T.A. Interaction of ICP34.5 with Beclin 1 modulates herpes simplex virus type 1 pathogenesis through control of CD4+ T-cell responses. *J. Virol.* **2009**, *83*, 12164–12171. [[CrossRef](#)] [[PubMed](#)]
90. Jagannath, C.; Lindsey, D.R.; Dhandayuthapani, S.; Xu, Y.; Hunter, R.L.; Eissa, N.T. Autophagy enhances the efficacy of BCG vaccine by increasing peptide presentation in mouse dendritic cells. *Nat. Med.* **2009**, *15*, 267–276. [[CrossRef](#)] [[PubMed](#)]
91. English, L.; Chemali, M.; Duron, J.; Rondeau, C.; Laplante, A.; Gingras, D.; Alexander, D.; Leib, D.; Norbury, C.; Lippé, R.; et al. Autophagy enhances the presentation of endogenous viral antigens on MHC class I molecules during HSV-1 infection. *Nat. Immunol.* **2009**, *10*, 480–487. [[CrossRef](#)] [[PubMed](#)]
92. Blanchet, F.P.; Moris, A.; Nikolic, D.S.; Lehmann, M.; Cardinaud, S.; Stalder, R.; Garcia, E.; Dinkins, C.; Leuba, F.; Wu, L.; et al. Human Immunodeficiency Virus-1 Inhibition of Immunoamphisomes in Dendritic Cells Impairs Early Innate and Adaptive Immune Responses. *Immunity* **2010**, *32*, 654–669. [[CrossRef](#)] [[PubMed](#)]
93. Jin, Y.; Sun, C.; Feng, L.; Li, P.; Xiao, L.; Ren, Y.; Wang, D.; Li, C.; Chen, L. Regulation of SIV Antigen-Specific CD4+ T Cellular Immunity via Autophagosome-Mediated MHC II Molecule-Targeting Antigen Presentation in Mice. *PLoS ONE* **2014**, *9*, e93143. [[CrossRef](#)] [[PubMed](#)]
94. Andersen, A.N.; Landsverk, O.J.; Simonsen, A.; Bogen, B.; Corthay, A.; Øynebråten, I. Coupling of HIV-1 Antigen to the Selective Autophagy Receptor SQSTM1/p62 Promotes T-Cell-Mediated Immunity. *Front. Immunol.* **2016**, *7*, 167. [[CrossRef](#)] [[PubMed](#)]
95. Espert, L.; Denizot, M.; Grimaldi, M.; Robert-Hebmann, V.; Gay, B.; Varbanov, M.; Codogno, P.; Biard-Piechaczyk, M. Autophagy is involved in T cell death after binding of HIV-1 envelope proteins to CXCR4. *J. Clin. Investig.* **2006**, *116*, 2161–2172. [[CrossRef](#)] [[PubMed](#)]
96. Denizot, M.; Varbanov, M.; Espert, L.; Robert-Hebmann, V.; Sagnier, S.; Garcia, E.; Curriu, M.; Mamoun, R.; Blanco, J.; Biard-Piechaczyk, M. HIV-1 gp41 fusogenic function triggers autophagy in uninfected cells. *Autophagy* **2008**, *4*, 998–1008. [[CrossRef](#)] [[PubMed](#)]
97. Zhou, D.; Masliah, E.; Spector, S.A. Autophagy is increased in postmortem brains of persons with HIV-1-associated encephalitis. *J. Infect. Dis.* **2011**, *203*, 1647–1657. [[CrossRef](#)] [[PubMed](#)]
98. Pandhare, J.; Dash, S.; Jones, B.; Villalta, F.; Dash, C. A novel role of proline oxidase in HIV-1 envelope glycoprotein-induced neuronal autophagy. *J. Biol. Chem.* **2015**, *290*, 25439–25451. [[CrossRef](#)] [[PubMed](#)]
99. Liu, Z.; Qiao, L.; Zhang, Y.; Zang, Y.; Shi, Y.; Liu, K.; Zhang, X.; Lu, X.; Yuan, L.; Su, B.; et al. ASPP2 plays a dual role in gp120-induced autophagy and apoptosis of neuroblastoma cells. *Front. Neurosci.* **2017**, *11*, 1–8. [[CrossRef](#)] [[PubMed](#)]
100. Cao, L.; Fu, M.; Kumar, S.; Kumar, A. Methamphetamine potentiates HIV-1 gp120-mediated autophagy via Beclin-1 and Atg5/7 as a pro-survival response in astrocytes. *Cell Death Dis.* **2016**, *7*, e2425. [[CrossRef](#)] [[PubMed](#)]
101. Fields, J.; Dumaop, W.; Eleuteri, S.; Elueteri, S.; Campos, S.; Serger, E.; Trejo, M.; Kosberg, K.; Adame, A.; Spencer, B.; et al. HIV-1 Tat alters neuronal autophagy by modulating autophagosome fusion to the lysosome: implications for HIV-associated neurocognitive disorders. *J. Neurosci.* **2015**, *35*, 1921–1938. [[CrossRef](#)] [[PubMed](#)]
102. Fields, J.A.; Metcalf, J.; Overk, C.; Adame, A.; Spencer, B.; Wrasidlo, W.; Florio, J.; Rockenstein, E.; He, J.J.; Masliah, E. The anticancer drug sunitinib promotes autophagy and protects from neurotoxicity in an HIV-1 Tat model of neurodegeneration. *J. Neurovirol.* **2017**, *23*, 290–303. [[CrossRef](#)] [[PubMed](#)]
103. Bruno, A.P.; de Simone, F.I.; Iorio, V.; De Marco, M.; Khalili, K.; Sariyer, I.K.; Capunzo, M.; Nori, S.L.; Rosati, A. HIV-1 Tat protein induces glial cell autophagy through enhancement of BAG3 protein levels. *Cell Cycle* **2014**, *13*, 3640–3644. [[CrossRef](#)] [[PubMed](#)]
104. Hui, L.; Chen, X.; Haughey, N.J.; Geiger, J.D. Role of endolysosomes in HIV-1 Tat-induced neurotoxicity. *ASN Neuro* **2012**, *4*, 243–252. [[CrossRef](#)] [[PubMed](#)]

105. De Simone, F.I.; Darbinian, N.; Amini, S.; Muniswamy, M.; White, M.K.; Elrod, J.W.; Datta, P.K.; Langford, D.; Khalili, K. HIV-1 Tat and Cocaine Impair Survival of Cultured Primary Neuronal Cells via a Mitochondrial Pathway. *J. Neuroimmune Pharmacol.* **2016**, *11*, 358–368. [[CrossRef](#)] [[PubMed](#)]
106. Li, J.C.; Au, K.; Fang, J.; Yim, H.C.; Chow, K.; Ho, P.; Lau, A.S. HIV-1 trans-activator protein dysregulates IFN- γ signaling and contributes to the suppression of autophagy induction. *AIDS* **2011**, *25*, 15–25. [[CrossRef](#)] [[PubMed](#)]
107. Van Grol, J.; Subauste, C.; Andrade, R.M.; Fujinaga, K.; Nelson, J.; Subauste, C.S. HIV-1 inhibits autophagy in bystander macrophage/monocytic cells through Src-Akt and STAT3. *PLoS ONE* **2010**, *5*, e11733. [[CrossRef](#)] [[PubMed](#)]
108. Campbell, G.R.; Rawat, P.; Bruckman, R.S.; Spector, S.A. Human Immunodeficiency Virus Type 1 Nef Inhibits Autophagy through Transcription Factor EB Sequestration. *PLoS Pathog.* **2015**, *11*, 1–24. [[CrossRef](#)] [[PubMed](#)]
109. Saribas, A.S.; Khalili, K.; Sariyer, I.K. Dysregulation of autophagy by HIV-1 Nef in human astrocytes. *Cell Cycle* **2015**, *14*, 2899–2904. [[CrossRef](#)] [[PubMed](#)]
110. Shoji-Kawata, S.; Sumpster, R.; Leveno, M.; Campbell, G.R.; Zou, Z.; Kinch, L.; Wilkins, A.D.; Sun, Q.; Pallauf, K.; MacDuff, D.; et al. Identification of a candidate therapeutic autophagy-inducing peptide. *Nature* **2013**, *494*, 201–206. [[CrossRef](#)] [[PubMed](#)]
111. Zhou, H.; Zheng, Y.; He, Y.; Chen, Z.; He, B. The role of autophagy in THP-1 macrophages resistance to HIV-Vpr-induced apoptosis. *Exp. Cell Res.* **2017**, *351*, 68–73. [[CrossRef](#)] [[PubMed](#)]
112. Madjo, U.; Leymarie, O.; Frémont, S.; Kuster, A.; Nehlich, M.; Gallois-Montbrun, S.; Janvier, K.; Berlioz-Torrent, C. LC3C Contributes to Vpu-Mediated Antagonism of BST2/Tetherin Restriction on HIV-1 Release through a Non-canonical Autophagy Pathway. *Cell Rep.* **2016**, *17*, 2221–2233. [[CrossRef](#)] [[PubMed](#)]
113. Nath, A. Human immunodeficiency virus (HIV) proteins in neuropathogenesis of HIV dementia. *J. Infect. Dis.* **2002**, *186*, S193–S198. [[CrossRef](#)] [[PubMed](#)]
114. Rao, V.R.; Ruiz, A.P.; Prasad, V.R. Viral and cellular factors underlying neuropathogenesis in HIV associated neurocognitive disorders (HAND). *AIDS Res. Ther.* **2014**, *11*, 13. [[CrossRef](#)] [[PubMed](#)]
115. Cherra, S.J.; Chu, C.T. Autophagy in neuroprotection and neurodegeneration: A question of balance. *Future Neurol.* **2008**, *3*, 309–323. [[PubMed](#)]
116. Pérez, A.; Probert, A.W.; Wang, K.K.; Sharmeen, L. Evaluation of HIV-1 Tat induced neurotoxicity in rat cortical cell culture. *J. Neurovirol.* **2001**, *7*, 1–10. [[CrossRef](#)] [[PubMed](#)]
117. King, J.E.; Eugenin, E.A.; Buckner, C.M.; Berman, J.W. HIV tat and neurotoxicity. *Microbes Infect.* **2006**, *8*, 1347–1357. [[CrossRef](#)] [[PubMed](#)]
118. Buscemi, L.; Ramonet, D.; Geiger, J.D. Human immunodeficiency virus type-1 protein Tat induces tumor necrosis factor- α -mediated neurotoxicity. *Neurobiol. Dis.* **2007**, *26*, 661–670. [[CrossRef](#)] [[PubMed](#)]
119. Agrawal, L.; Louboutin, J.-P.; Reyes, B.A.S.; Van Bockstaele, E.J.; Strayer, D.S. HIV-1 Tat neurotoxicity: A model of acute and chronic exposure, and neuroprotection by gene delivery of antioxidant enzymes. *Neurobiol. Dis.* **2012**, *45*, 657–670. [[CrossRef](#)] [[PubMed](#)]
120. Banks, W.A.; Robinson, S.M.; Nath, A. Permeability of the blood–brain barrier to HIV-1 Tat. *Exp. Neurol.* **2005**, *193*, 218–227. [[CrossRef](#)] [[PubMed](#)]
121. He, J.J.; Liu, Y.; Jones, M.; Hingtgen, C.M.; Bu, G.; Larabee, N.; Tanzi, R.E.; Moir, R.D.; Nath, A. Uptake of HIV-1 Tat protein mediated by low-density lipoprotein receptor-related protein disrupts the neuronal metabolic balance of the receptor ligands. *Nat. Med.* **2000**, *6*, 1380–1387. [[CrossRef](#)] [[PubMed](#)]
122. Gutheil, W.G.; Subramanyam, M.; Flentke, G.R.; Sanford, D.G.; Munoz, E.; Huber, B.T.; Bachovchin, W.W. Human immunodeficiency virus 1 Tat binds to dipeptidyl aminopeptidase IV (CD26): A possible mechanism for Tat's immunosuppressive activity. *Proc. Natl. Acad. Sci. USA* **1994**, *91*, 6594–6598. [[CrossRef](#)] [[PubMed](#)]
123. Xiao, H.; Neuveut, C.; Tiffany, H.L.; Benkirane, M.; Rich, E.A.; Murphy, P.M.; Jeang, K.T. Selective CXCR4 antagonism by Tat: Implications for in vivo expansion of coreceptor use by HIV-1. *Proc. Natl. Acad. Sci. USA* **2000**, *97*, 11466–11471. [[CrossRef](#)] [[PubMed](#)]
124. Vendeville, A.; Rayne, F.; Bonhoure, A.; Bettache, N.; Montcourrier, P.; Beaumelle, B. HIV-1 Tat enters T cells using coated pits before translocating from acidified endosomes and eliciting biological responses. *Mol. Biol. Cell* **2004**, *15*, 2347–2360. [[CrossRef](#)] [[PubMed](#)]
125. Tyagi, M.; Rusnati, M.; Presta, M.; Giacca, M. Internalization of HIV-1 tat requires cell surface heparan sulfate proteoglycans. *J. Biol. Chem.* **2001**, *276*, 3254–3261. [[CrossRef](#)] [[PubMed](#)]

126. Cuervo, A.M.; Wong, E. Chaperone-mediated autophagy: roles in disease and aging. *Cell Res.* **2014**, *24*, 92–104. [[CrossRef](#)] [[PubMed](#)]
127. Saftig, P.; Tanaka, Y.; Guhde, G.; Suter, A.; Eskelinen, E.-L.; Hartmann, D.; Lüllmann-Rauch, R.; Janssen, P.M.L.; Blanz, J.; von Figura, K. Accumulation of autophagic vacuoles and cardiomyopathy in LAMP-2-deficient mice. *Nature* **2000**, *406*, 902–906. [[CrossRef](#)] [[PubMed](#)]
128. Farrow, J.M.; Yang, J.C.; Evans, C.P. Autophagy as a modulator and target in prostate cancer. *Nat. Rev. Urol.* **2014**, *11*, 508–516. [[CrossRef](#)] [[PubMed](#)]
129. Pal, R.; Palmieri, M.; Loehr, J.A.; Li, S.; Abo-Zahrah, R.; Monroe, T.O.; Thakur, P.B.; Sardiello, M.; Rodney, G.G. Src-dependent impairment of autophagy by oxidative stress in a mouse model of Duchenne muscular dystrophy. *Nat. Commun.* **2014**, *5*, 4425. [[CrossRef](#)] [[PubMed](#)]
130. Lum, J.J.; DeBerardinis, R.J.; Thompson, C.B. Autophagy in metazoans: Cell survival in the land of plenty. *Nat. Rev. Mol. Cell Biol.* **2005**, *6*, 439–448. [[CrossRef](#)] [[PubMed](#)]
131. Basmaciogullari, S.; Pizzato, M. The activity of Nef on HIV-1 infectivity. *Front. Microbiol.* **2014**, *5*, 232. [[CrossRef](#)] [[PubMed](#)]
132. Schindler, M.; Würfl, S.; Benaroch, P.; Greenough, T.C.; Daniels, R.; Easterbrook, P.; Brenner, M.; Münch, J.; Kirchhoff, F. Down-modulation of mature major histocompatibility complex class II and up-regulation of invariant chain cell surface expression are well-conserved functions of human and simian immunodeficiency virus nef alleles. *J. Virol.* **2003**, *77*, 10548–10556. [[CrossRef](#)] [[PubMed](#)]
133. Schwartz, O.; Maréchal, V.; Le Gall, S.; Lemonnier, F.; Heard, J.M. Endocytosis of major histocompatibility complex class I molecules is induced by the HIV-1 Nef protein. *Nat. Med.* **1996**, *2*, 338–342. [[CrossRef](#)] [[PubMed](#)]
134. Aiken, C.; Konner, J.; Landau, N.R.; Lenburg, M.E.; Trono, D. Nef induces CD4 endocytosis: requirement for a critical dileucine motif in the membrane-proximal CD4 cytoplasmic domain. *Cell* **1994**, *76*, 853–864. [[CrossRef](#)]
135. Garcia, J.V.; Miller, A.D. Serine phosphorylation-independent downregulation of cell-surface CD4 by nef. *Nature* **1991**, *350*, 508–511. [[CrossRef](#)] [[PubMed](#)]
136. Matheson, N.J.; Sumner, J.; Wals, K.; Rapiteanu, R.; Weekes, M.P.; Vigan, R.; Weinelt, J.; Schindler, M.; Antrobus, R.; Costa, A.S.H.; et al. Cell Surface Proteomic Map of HIV Infection Reveals Antagonism of Amino Acid Metabolism by Vpu and Nef. *Cell Host Microbe* **2015**, *18*, 409–423. [[CrossRef](#)] [[PubMed](#)]
137. Usami, Y.; Wu, Y.; Göttliger, H.G. SERINC3 and SERINC5 restrict HIV-1 infectivity and are counteracted by Nef. *Nature* **2015**, *526*, 218–223. [[CrossRef](#)] [[PubMed](#)]
138. Rosa, A.; Chande, A.; Ziglio, S.; de Sanctis, V.; Bertorelli, R.; Goh, S.L.; McCauley, S.M.; Nowosielska, A.; Antonarakis, S.E.; Luban, J.; et al. HIV-1 Nef promotes infection by excluding SERINC5 from virion incorporation. *Nature* **2015**, *526*, 212–227. [[CrossRef](#)] [[PubMed](#)]
139. Settembre, C.; di Malta, C.; Polito, V.A.; Aencibia, M.G.; Vetrini, F.; Erdin, S.; Erdin, S.U.; Huynh, T.; Medina, D.; Colella, P.; et al. TFEB Links Autophagy to Lysosomal Biogenesis. *Science* **2011**, *332*, 1429–1433. [[CrossRef](#)] [[PubMed](#)]
140. Palmieri, M.; Impey, S.; Kang, H.; di Ronza, A.; Pelz, C.; Sardiello, M.; Ballabio, A. Characterization of the CLEAR network reveals an integrated control of cellular clearance pathways. *Hum. Mol. Genet.* **2011**, *20*, 3852–3866. [[CrossRef](#)] [[PubMed](#)]
141. Busca, A.; Saxena, M.; Kumar, A. Critical role for antiapoptotic Bcl-xL and Mcl-1 in human macrophage survival and cellular IAP1/2 (cIAP1/2) in resistance to HIV-Vpr-induced apoptosis. *J. Biol. Chem.* **2012**, *287*, 15118–15133. [[CrossRef](#)] [[PubMed](#)]
142. Godet, A.N.; Guernon, J.; Croset, A.; Cayla, X.; Falanga, P.B.; Colle, J.-H.; Garcia, A. PP2A1 binding, cell transducing and apoptotic properties of Vpr(77–92): A new functional domain of HIV-1 Vpr proteins. *PLoS ONE* **2010**, *5*, e13760. [[CrossRef](#)] [[PubMed](#)]
143. Mukerjee, R.; Chang, J.R.; del Valle, L.; Bagashev, A.; Gayed, M.M.; Lyde, R.B.; Hawkins, B.J.; Brailoiu, E.; Cohen, E.; Power, C.; et al. Deregulation of microRNAs by HIV-1 Vpr protein leads to the development of neurocognitive disorders. *J. Biol. Chem.* **2011**, *286*, 34976–34985. [[CrossRef](#)] [[PubMed](#)]
144. Patel, C.A.; Mukhtar, M.; Pomerantz, R.J. Human immunodeficiency virus type 1 Vpr induces apoptosis in human neuronal cells. *J. Virol.* **2000**, *74*, 9717–9726. [[CrossRef](#)] [[PubMed](#)]

145. Saxena, M.; Busca, A.; Pandey, S.; Kryworuchko, M.; Kumar, A. CpG protects human monocytic cells against HIV-Vpr-induced apoptosis by cellular inhibitor of apoptosis-2 through the calcium-activated JNK pathway in a TLR9-independent manner. *J. Immunol.* **2011**, *187*, 5865–5878. [[CrossRef](#)] [[PubMed](#)]
146. Arokium, H.; Kamata, M.; Chen, I. Virion-associated Vpr of human immunodeficiency virus type 1 triggers activation of apoptotic events and enhances fas-induced apoptosis in human T cells. *J. Virol.* **2009**, *83*, 11283–11297. [[CrossRef](#)] [[PubMed](#)]
147. Muthumani, K.; Hwang, D.S.; Desai, B.M.; Zhang, D.; Dayes, N.; Green, D.R.; Weiner, D.B. HIV-1 Vpr induces apoptosis through caspase 9 in T cells and peripheral blood mononuclear cells. *J. Biol. Chem.* **2002**, *277*, 37820–37831. [[CrossRef](#)] [[PubMed](#)]
148. Ferrucci, A.; Nonnemacher, M.; Wigdahl, B. Cellular phenotype impacts human immunodeficiency virus type 1 viral protein R subcellular localization. *Virol. J.* **2011**, *8*, 397. [[CrossRef](#)] [[PubMed](#)]
149. Carter, C.A.; Ehrlich, L.S. Cell Biology of HIV-1 Infection of Macrophages. *Annu. Rev. Microbiol.* **2008**, *62*, 425–443. [[CrossRef](#)] [[PubMed](#)]
150. Ho, D.D.; Rota, T.R.; Hirsch, M.S. Infection of monocyte/macrophages by human T lymphotropic virus type III. *J. Clin. Investig.* **1986**, *77*, 1712–1715. [[CrossRef](#)] [[PubMed](#)]
151. Coleman, C.M.; Wu, L. HIV interactions with monocytes and dendritic cells: Viral latency and reservoirs. *Retrovirology* **2009**, *6*, 51. [[CrossRef](#)] [[PubMed](#)]
152. Reynoso, R.; Wieser, M.; Ojeda, D.; Bönisch, M.; Kühnel, H.; Bolcic, F.; Quendler, H.; Grillari, J.; Grillari-Voglauer, R.; Quarleri, J. HIV-1 induces telomerase activity in monocyte-derived macrophages, possibly safeguarding one of its reservoirs. *J. Virol.* **2012**, *86*, 10327–10337. [[CrossRef](#)] [[PubMed](#)]
153. Swingler, S.; Mann, A.M.; Zhou, J.; Swingler, C.; Stevenson, M. Apoptotic killing of HIV-1-infected macrophages is subverted by the viral envelope glycoprotein. *PLoS Pathog.* **2007**, *3*, 1281–1290. [[CrossRef](#)] [[PubMed](#)]
154. Codogno, P.; Mehrpour, M.; Proikas-Cezanne, T. Canonical and non-canonical autophagy: variations on a common theme of self-eating? *Nat. Rev. Mol. Cell Biol.* **2012**, *13*, 7–12. [[CrossRef](#)] [[PubMed](#)]
155. Sanjuan, M.A.; Dillon, C.P.; Tait, S.W.G.; Moshiah, S.; Dorsey, F.; Connell, S.; Komatsu, M.; Tanaka, K.; Cleveland, J.L.; Withoff, S.; et al. Toll-like receptor signalling in macrophages links the autophagy pathway to phagocytosis. *Nature* **2007**, *450*, 1253–1257. [[CrossRef](#)] [[PubMed](#)]
156. Florey, O.; Kim, S.E.; Sandoval, C.P.; Haynes, C.M.; Overholtzer, M. Autophagy machinery mediates macroendocytic processing and entotic cell death by targeting single membranes. *Nat. Cell Biol.* **2011**, *13*, 1335–1343. [[CrossRef](#)] [[PubMed](#)]
157. Martinez, J.; Almendinger, J.; Oberst, A.; Ness, R.; Dillon, C.P.; Fitzgerald, P.; Hengartner, M.O.; Green, D.R. Microtubule-associated protein 1 light chain 3 α (LC3)-associated phagocytosis is required for the efficient clearance of dead cells. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 17396–17401. [[CrossRef](#)] [[PubMed](#)]
158. Martinez, J.; Malireddi, R.K.S.; Lu, Q.; Cunha, L.D.; Pelletier, S.; Gingras, S.; Orchard, R.; Guan, J.-L.; Tan, H.; Peng, J.; et al. Molecular characterization of LC3-associated phagocytosis reveals distinct roles for Rubicon, NOX2 and autophagy proteins. *Nat. Cell Biol.* **2015**, *17*, 893–906. [[CrossRef](#)] [[PubMed](#)]
159. Gong, L.; Cullinane, M.; Treerat, P.; Ramm, G.; Prescott, M.; Adler, B.; Boyce, J.D.; Devenish, R.J. The *Burkholderia pseudomallei* Type III Secretion System and BopA Are Required for Evasion of LC3-Associated Phagocytosis. *PLoS ONE* **2011**, *6*, e17852. [[CrossRef](#)] [[PubMed](#)]
160. Lam, G.Y.; Cemma, M.; Muise, A.M.; Higgins, D.E.; Brumell, J.H. Host and bacterial factors that regulate LC3 recruitment to *Listeria monocytogenes* during the early stages of macrophage infection. *Autophagy* **2013**, *9*, 985–995. [[CrossRef](#)] [[PubMed](#)]
161. Baxt, L.A.; Goldberg, M.B. Host and bacterial proteins that repress recruitment of LC3 to Shigella early during infection. *PLoS ONE* **2014**, *9*, e94653. [[CrossRef](#)] [[PubMed](#)]
162. Ligeon, L.-A.; Moreau, K.; Barois, N.; Bongiovanni, A.; Lacorre, D.-A.; Werkmeister, E.; Proux-Gillardeaux, V.; Galli, T.; Lafont, F. Role of VAMP3 and VAMP7 in the commitment of *Yersinia pseudotuberculosis* to LC3-associated pathways involving single- or double-membrane vacuoles. *Autophagy* **2014**, *10*, 1588–1602. [[CrossRef](#)] [[PubMed](#)]
163. Abnave, P.; Mottola, G.; Gimenez, G.; Boucherit, N.; Trouplin, V.; Torre, C.; Conti, F.; Ben Amara, A.; Lepolard, C.; Djian, B.; et al. Screening in planarians identifies MORN2 as a key component in LC3-associated phagocytosis and resistance to bacterial infection. *Cell Host Microbe* **2014**, *16*, 338–350. [[CrossRef](#)] [[PubMed](#)]

164. Mimouna, S.; Bazin, M.; Mograbi, B.; Darfeuille-Michaud, A.; Brest, P.; Hofman, P.; Vouret-Craviari, V. HIF1A regulates xenophagic degradation of adherent and invasive *Escherichia coli* (AIEC). *Autophagy* **2014**, *10*, 2333–2345. [[CrossRef](#)] [[PubMed](#)]
165. Hubber, A.; Kubori, T.; Coban, C.; Matsuzawa, T.; Ogawa, M.; Kawabata, T.; Yoshimori, T.; Nagai, H. Bacterial secretion system skews the fate of Legionella-containing vacuoles towards LC3-associated phagocytosis. *Sci. Rep.* **2017**, *7*, 44795. [[CrossRef](#)] [[PubMed](#)]
166. Deen, N.S.; Gong, L.; Naderer, T.; Devenish, R.J.; Kwok, T. Analysis of the Relative Contribution of Phagocytosis, LC3-Associated Phagocytosis, and Canonical Autophagy During *Helicobacter pylori* Infection of Macrophages. *Helicobacter* **2015**, *20*, 449–459. [[CrossRef](#)] [[PubMed](#)]
167. Tam, J.M.; Mansour, M.K.; Khan, N.S.; Seward, M.; Puranam, S.; Tanne, A.; Sokolovska, A.; Becker, C.E.; Acharya, M.; Baird, M.A.; et al. Dectin-1–Dependent LC3 Recruitment to Phagosomes Enhances Fungicidal Activity in Macrophages. *J. Infect. Dis.* **2014**, *210*, 1844–1854. [[CrossRef](#)] [[PubMed](#)]
168. Oikonomou, V.; Moretti, S.; Renga, G.; Galosi, C.; Borghi, M.; Pariano, M.; Puccetti, M.; Palmerini, C.A.; Amico, L.; Carotti, A.; et al. Noncanonical Fungal Autophagy Inhibits Inflammation in Response to IFN- γ via DAPK1. *Cell Host Microbe* **2016**, *20*, 744–757. [[CrossRef](#)] [[PubMed](#)]
169. Akoumianaki, T.; Kyrmizi, I.; Valsecchi, I.; Gresnigt, M.S.; Samonis, G.; Drakos, E.; Boumpas, D.; Muszkieta, L.; Prevost, M.-C.; Kontoyiannis, D.P.; et al. Aspergillus Cell Wall Melanin Blocks LC3-Associated Phagocytosis to Promote Pathogenicity. *Cell Host Microbe* **2016**, *19*, 79–90. [[CrossRef](#)] [[PubMed](#)]
170. Boonhok, R.; Rachaphaew, N.; Duangmanee, A.; Chobson, P.; Pattaradilokrat, S.; Utaisinchaoen, P.; Sattabongkot, J.; Ponpuak, M. LAP-like process as an immune mechanism downstream of IFN- γ in control of the human malaria *Plasmodium vivax* liver stage. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, e3519–e3528. [[CrossRef](#)] [[PubMed](#)]
171. Crauwels, P.; Bohn, R.; Thomas, M.; Gottwalt, S.; Jäckel, F.; Krämer, S.; Bank, E.; Tenzer, S.; Walther, P.; Bastian, M.; et al. Apoptotic-like Leishmania exploit the host's autophagy machinery to reduce T-cell-mediated parasite elimination. *Autophagy* **2015**, *11*, 285–297. [[CrossRef](#)] [[PubMed](#)]
172. Matte, C.; Casgrain, P.-A.; Séguin, O.; Moradin, N.; Hong, W.J.; Descoteaux, A. *Leishmania major* Promastigotes Evade LC3-Associated Phagocytosis through the Action of GP63. *PLoS Pathog.* **2016**, *12*, e1005690. [[CrossRef](#)] [[PubMed](#)]
173. Campbell, S.M.; Crowe, S.M.; Mak, J. Lipid rafts and HIV-1: From viral entry to assembly of progeny virions. *J. Clin. Virol.* **2001**, *22*, 217–227. [[CrossRef](#)]
174. Rollason, R.; Korolchuk, V.; Hamilton, C.; Jepson, M.; Banting, G. A CD317/tetherin-RICH2 complex plays a critical role in the organization of the subapical actin cytoskeleton in polarized epithelial cells. *J. Cell Biol.* **2009**, *184*, 721–736. [[CrossRef](#)] [[PubMed](#)]
175. Billcliff, P.G.; Rollason, R.; Prior, I.; Owen, D.M.; Gaus, K.; Banting, G. CD317/tetherin is an organiser of membrane microdomains. *J. Cell Sci.* **2013**, *126*, 1553–1564. [[CrossRef](#)] [[PubMed](#)]
176. Rollason, R.; Dunstan, K.; Billcliff, P.G.; Bishop, P.; Gleeson, P.; Wise, H.; Digard, P.; Banting, G. Expression of HIV-1 Vpu Leads to Loss of the Viral Restriction Factor CD317/Tetherin from Lipid Rafts and Its Enhanced Lysosomal Degradation. *PLoS ONE* **2013**, *8*, e75680. [[CrossRef](#)] [[PubMed](#)]
177. Dubé, M.; Bhusan Roy, B.; Guiot-Guillain, P.; Binette, J.; Mercier, J.; Chiasson, A.; Cohen, É.A. Antagonism of Tetherin Restriction of HIV-1 Release by Vpu Involves Binding and Sequestration of the Restriction Factor in a Perinuclear Compartment. *PLoS Pathog.* **2010**, *6*, e1000856. [[CrossRef](#)] [[PubMed](#)]
178. Mitchell, R.S.; Katsura, C.; Skasko, M.A.; Fitzpatrick, K.; Lau, D.; Ruiz, A.; Stephens, E.B.; Margottin-Gouget, F.; Benarous, R.; Guatelli, J.C. Vpu antagonizes BST-2-mediated restriction of HIV-1 release via beta-TrCP and endo-lysosomal trafficking. *PLoS Pathog.* **2009**, *5*, e1000450. [[CrossRef](#)] [[PubMed](#)]
179. Janvier, K.; Pelchen-Matthews, A.; Renaud, J.-B.; Caillet, M.; Marsh, M.; Berlioz-Torrent, C. The ESCRT-0 component HRS is required for HIV-1 Vpu-mediated BST-2/tetherin down-regulation. *PLoS Pathog.* **2011**, *7*, e1001265. [[CrossRef](#)] [[PubMed](#)]
180. Roy, N.; Pacini, G.; Berlioz-Torrent, C.; Janvier, K. Characterization of E3 ligases involved in lysosomal sorting of the HIV-1 restriction factor BST2. *J. Cell Sci.* **2017**, *130*, 1596–1611. [[CrossRef](#)] [[PubMed](#)]
181. Kueck, T.; Neil, S.J.D. A cytoplasmic tail determinant in HIV-1 Vpu mediates targeting of tetherin for endosomal degradation and counteracts interferon-induced restriction. *PLoS Pathog.* **2012**, *8*, e1002609. [[CrossRef](#)] [[PubMed](#)]

182. Kueck, T.; Foster, T.L.; Weinelt, J.; Sumner, J.C.; Pickering, S.; Neil, S.J.D. Serine Phosphorylation of HIV-1 Vpu and Its Binding to Tetherin Regulates Interaction with Clathrin Adaptors. *PLoS Pathog.* **2015**, *11*, e1005141. [[CrossRef](#)] [[PubMed](#)]
183. Jia, X.; Weber, E.; Tokarev, A.; Lewinski, M.; Rizk, M.; Suarez, M.; Guatelli, J.; Xiong, Y. Structural basis of HIV-1 Vpu-mediated BST2 antagonism via hijacking of the clathrin adaptor protein complex 1. *eLife* **2014**, *3*, e02362. [[CrossRef](#)] [[PubMed](#)]
184. McNatt, M.W.; Zang, T.; Bieniasz, P.D. Vpu Binds Directly to Tetherin and Displaces It from Nascent Virions. *PLoS Pathog.* **2013**, *9*, e1003299. [[CrossRef](#)] [[PubMed](#)]
185. Lewinski, M.K.; Jafari, M.; Zhang, H.; Opella, S.J.; Guatelli, J. Membrane anchoring by a C-terminal tryptophan enables HIV-1 Vpu to displace bone marrow stromal antigen 2 (BST2) from sites of viral assembly. *J. Biol. Chem.* **2015**, *290*, 10919–10933. [[CrossRef](#)] [[PubMed](#)]
186. Jafari, M.; Guatelli, J.; Lewinski, M.K. Activities of transmitted/founder and chronic clade B HIV-1 Vpu and a C-terminal polymorphism specifically affecting virion release. *J. Virol.* **2014**, *88*, 5062–5078. [[CrossRef](#)] [[PubMed](#)]



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