

# Immune–metabolic interactions in homeostasis and the progression to NASH

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1 Title 2 Immune-metabolic interactions in homeostasis and the progression to NASH 3 4 **Authors** Joanne A. Hoogerland<sup>1</sup>, Bart Staels<sup>1\*</sup>, David Dombrowicz<sup>1\*</sup> 5 6 7 Affiliations 8 <sup>1</sup>Univ. Lille, Inserm, CHU Lille, Institut Pasteur de Lille, U1011-EGID, F-59000 Lille, France 9 \*Co-senior authors 10 11 Correspondence: david.dombrowicz@univ-lille.fr 12 13 Key words 14 Liver; immunology; NAFLD; NASH; metabolism 15 16 Abstract 17 The incidence of Non-Alcoholic Fatty Liver Disease (NAFLD) has tremendously increased over 18 the past 2 decades. NAFLD ranges from simple steatosis (NAFL) to Non-Alcoholic 19 Steatohepatitis (NASH) and predisposes to fibrosis and hepatocellular carcinoma. The 20 importance of the immune system in hepatic physiology and in the progression of NAFLD is 21 increasingly recognized. At homeostasis, the liver participates in the immune defense against 22 pathogens and in tolerance towards gut-derived microbial compounds. Hepatic immune cells

also respond to metabolic stimuli and play a role in NAFLD progression to NASH. We first

discuss how metabolic perturbations affect immune cell phenotype and function in NAFL

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- $\,\,25\,\,$   $\,$  and NASH. Then, we focus on the role of immune cells in liver homeostasis and in the
- development of NASH.

#### The liver as an immunological organ

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The liver plays a central role in glucose and lipid homeostasis as well as in cholesterol and bile acid synthesis. Hepatocytes account for about 70% of the total liver cell content and execute most hepatic metabolic processes [1]. The remaining liver cells are nonparenchymal cells such as cholangiocytes, liver sinusoidal endothelial cells (LSECs), hepatic stellate cells (HSCs), and resident innate and adaptive immune cells. The main hepatic blood supply (~80%) comes from the gut via the portal vein, passes through a network of liver sinusoids, and leaves the parenchyma via the central vein. The liver is therefore constantly exposed to metabolites and non-self antigens derived from nutrients and gut microbiota. When pathogens invade the liver, neutrophils are the first responders. They are mobilized from a large bone marrow pool, circulate via the blood stream, extravasate into tissue where they immobilize and kill pathogens. Neutrophils act through phagocytosis, production of reactive oxygen species (ROS), release of Neutrophil Extracellular Traps (NET), and secretion of a variety of cytokines. In addition, the antigen-rich portal blood is "probed" by antigenpresenting cells (APCs) and lymphocytes for the presence of non-self compounds as well as for pathogens. In the metabolically healthy liver, this process contributes to the establishment of (antigen-specific) tolerance. This immune tolerant state implies that the liver actively inhibits an overt immune response to harmless food-derived antigens and microbial products. However, to maintain homeostasis, the liver has also to allow efficient responses against infectious agents. APCs, such as dendritic cells (DCs) (represented as plasmacytoid DCs (pDCs) and classical DCs (cDCs)), and Kupffer cells (KCs), the resident hepatic macrophages, are predominantly located in the periportal and pericentral areas. They capture antigens that pass through the liver, process and present them through major histocompatibility complex class-I or II (MHC-I/II) molecules. Immunoglobulin-producing B,

and T lymphocytes are dispersed throughout the parenchyma being present in the portal tracts as well. Various subsets of T cells recognize presented antigens using highly variable T Cell antigen Receptors (TCRs), resulting - in non-inflammatory conditions - in tolerance induction rather than enhancement of T cell responses. In line, in these conditions, antigen presentation to naïve CD4<sup>+</sup> T cells induces differentiation of regulatory T cells (Tregs) rather than T helper (Th) 1 (Th1) or 17 (Th17) cells. In addition, CD8<sup>+</sup> T cells are highly effective in killing cells infected by intracellular pathogens and damaged cells. Next to conventional CD4<sup>+</sup> and CD8<sup>+</sup> T cells, non-conventional T cells (natural killer T (NKT) cells) are present in the liver and contribute to immune homeostasis by recognizing specific non-peptide antigens (Figure 1). In an altered metabolic environment, as occurring in NAFLD, the hepatic immune system can be triggered by 'metabolic overload', leading to an inappropriate immune response. The major risk factor for NAFLD is overnutrition, associated with obesity and type 2 diabetes. Metabolic overload already occurs in early stages of NAFLD characterized by benign steatosis (NAFL) with no evidence of hepatocellular injury [2]. Continuous metabolic overload by high caloric diets generates toxic lipids, such as free cholesterol, free fatty acids, diacylglycerols, and ceramides [3,4]. Nutrient overload causes oxidative and ER stress and hepatocyte apoptosis [5], thereby driving NAFLD progression. The strong association between NAFLD and type 2 diabetes indeed shows that persistent nutrient (glucose) overload affects hepatic cell function. Drastic reduction of caloric intake, such as after bariatric surgery, improves all stages of NAFLD, proving that metabolic overload is a main driver of NAFLD [6]. Metabolic overload first affects hepatic non-immune cells, which, at homeostasis, also exert immunemodulatory functions by participating in antigen presentation contributing to hepatic immune tolerance (Box 1 and 2). Inflammation, the central feature in the progression from

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NAFL to NASH is subsequently triggered by metabolic overload and ensuing hepatocyte injury [7]. Pathological NASH develops when fat accumulation in hepatocytes is accompanied by lobular inflammation and hepatocellular ballooning [8,9]. The final stages of NASH are advanced fibrosis/cirrhosis, hepatocellular carcinoma, and liver failure.

In the present review, we will discuss liver immunity at homeostasis and during development and progression of NASH. We will focus on 1) the effects of the altered metabolic environment, predisposing to NASH, on hepatic immune cells, and 2) the role of hepatic immune cells in the development and progression of NASH.

#### Triggers of the immune response in NAFLD

The altered metabolic environment in NAFLD affects immune cell phenotype and function, hence promoting NASH progression. Indeed, immune cells sense environmental and metabolic cues, such as nutrient and oxygen availability, and microbial metabolites (Box 3), and adapt their metabolic programs accordingly. This part of the review describes the current knowledge about the effects of NAFLD-related alterations in lipid-, glucose-, and cholesterol metabolism on hepatic immune cells.

#### Lipid metabolism

Hepatic steatosis results from an imbalance between hepatic triglyceride input and output. Hepatic free fatty acid influx rates and *de novo* lipogenesis are increased in NAFLD [10], affecting hepatocytes as well as immune cell function. Accumulation of (toxic) lipids in hepatocytes causes hepatocellular lipotoxicity, which leads to metabolic dysfunction and cellular oxidative and ER stress, and eventually cell death. Injured and dying hepatocytes release apoptotic bodies and extracellular vesicles (EVs) containing RNA, membrane,

cytosolic and nuclear proteins, lipids, and damage-associated molecular patterns (DAMPs) [11], thereby inducing monocyte recruitment and macrophage activation. For example, danger signals, such as histidine-rich glycoprotein (HRG), induce macrophage polarization towards a pro-inflammatory phenotype [12]. Next to hepatocyte-mediated effect of lipids, some studies show a direct effect of lipids on hepatic immune cells. Hepatic macrophages from mice fed a high fat diet display intracellular lipid accumulation which was associated with altered expression of genes involved in lipid synthesis, oxidation, uptake and secretion, and the increased production of inflammatory mediators [13]. Furthermore, in vitro assays showed that LPS-mediated activation of these lipid-loaden macrophages promote the recruitment of CD4<sup>+</sup> and CD8<sup>+</sup> T and B lymphocytes, which is a hallmark of NASH. MRS1, the key macrophage receptor for clearance of circulation lipoproteins, was recently suggested to be critical for the uptake of saturated fatty acids and the subsequent proinflammatory response [14]. These studies do not allow to discriminate between recruited and resident macrophages (or KCs), as the most specific markers for KCs, such as CLEC4F, were not used. However, it is likely that at least some hepatic macrophage subsets can be directly activated by lipid accumulation. More recent studies using single-cell RNAseq have identified distinct macrophage subtypes in livers mice fed a high-fat diet [15,16]. It was suggested that hepatic lipid accumulation promotes the recruitment of a subset of monocyte-derived lipid-associated macrophages (LAMs) – but not resident KCs [15]. Studies on activation of the newly identified macrophage subsets and the potential effects of lipids on their phenotype and function are still awaiting. Dendritic cells (DCs) are also sensitive to extracellular fatty acids during innate immune activation [17]. High fatty acid concentrations enhance the TLR-mediated innate activation of DCs by inhibiting the activity of hexokinase, the initial enzyme of glycolysis. Consequently,

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the glycolytic reprogramming of DCs is impaired and mitochondrial reactive oxygen species (ROS) production is increased. ROS affects the unfolded protein response and induces a specific inflammatory program, characterized by increased IL-23 production [17]. Notably, IL-23 expression is enhanced in livers of mice fed a NASH diet [18]. Finally, a few studies have investigated the effects of specific fatty acids on T cells. Generally, high concentrations of fatty acids are toxic to T cells, and non-toxic concentrations induce proliferation and cytokine production [19–21]. The mechanisms by which fatty acids affect T cell function, especially in NAFLD/NASH, is still unclear and deserves further research.

#### Glucose metabolism

Hepatic steatosis promotes liver insulin resistance ultimately leading to diabetes, itself a major risk factor for NAFLD. Glucose availability and uptake are critical determinants in directing macrophage differentiation. As DCs, macrophages integrate signals of overnutrition and accordingly reprogram their metabolism [22]. The transcription factor HIF1 $\alpha$  plays a key role in the induction of the glycolytic adaptation in inflammatory macrophages. HIF1 $\alpha$  expression is regulated both by inflammatory signals, such as pro-inflammatory cytokines, inducing the NF-kB signaling pathway [23], and growth factors, such as GM-CSF, triggering the AKT/mTOR pathway [24,25]. HIF1 $\alpha$ , in turn, regulates the expression of glycolytic enzymes and the glucose transporter GLUT1, thereby facilitating rapid glucose uptake. Glucose entry and its metabolization into lactate directly promotes inflammatory macrophage gene expression and activation [26]. Inflammatory macrophages further contribute to hepatic steatosis and inflammation by secretion of cytokines such as TNF- $\alpha$  and IL-1 $\beta$  [27].

Similar as macrophages, T cells are also dependent on the import of extracellular glucose to meet increased metabolic needs during activation. Naïve, resting T cells rely on oxidative phosphorylation to maintain housekeeping functions and cell survival, which is promoted by IL-4 and IL-7 [28]. Upon activation, T cells switch from a quiescent state to proliferative expansion, which requires uptake of essential metabolites to support anabolic growth [29]. T cell activation involves the costimulatory receptor CD28, which induces glucose uptake and glycolysis in T cells to a level that is sufficient for proliferation and function [30]. Glucose availability seems to (partly) control immune cell function since activated immune cells have high metabolic demands. However, more studies are needed to investigate the impact of a nutrient-rich NAFLD environment on immune cell function, not only at the single cell level, but also in cell-cell interactions.

#### Cholesterol metabolism

Elevated cholesterol levels in hepatocytes are associated with NAFLD risk and severity [4,31]. Cholesterol is synthesized *de novo* from acetyl-CoA or imported *via* lipoprotein uptake. Macrophages are important for cholesterol efflux and reverse cholesterol transport from peripheral tissues to the liver *via* high-density lipoprotein (HDL) particles. LXRα is a critical regulator of cholesterol and lipid metabolism in KCs, and links metabolism and inflammation by reducing the production of pro-inflammatory mediators upon stimulation [32,33]. Moreover, cholesterol is a precursor for bile acid synthesis, and (microbial) metabolites of bile acids were recently shown to control the adaptive immune response in the gut [34–36] (Box 3). However, studies about the effect of specific bile acids on hepatic immunity are lacking. Plasma bile acid levels and profiles are reported to be altered in NAFL and NASH [37–40], and a large inter-individual variation in human bile acid pool size and composition

exists [39]. It will be important to further investigate the signaling functions of bile acids and their metabolites in the control of immune cell phenotypes and functions, including interindividual differences in adaptive immunity.

#### Several liver immune cell types are involved in NAFLD onset and progression

NASH is characterized by hepatic inflammatory infiltrates and lobular inflammation. Immune cell profiling and transcriptomic analysis identified a hepatic immune- and inflammation-related gene signature in NASH patients, which reversibly associates with NAFL to NASH transition and is characterized by pathways related to antigen presentation and CD8<sup>+</sup> T cell activation [18]. Yet a wide range of hepatic immune cell populations have been associated with NAFL progression to NASH, including macrophages, DCs, neutrophils, natural killer cells, (non-conventional) T cells and B cells.

#### Macrophages are activated in the early phase of liver injury

KCs, the resident macrophages in the liver, are the most abundant hepatic immune cells accounting for ~35% of the non-parenchymal cells. These cells originate from yolk sacderived erythromyeloid progenitors in the fetal liver. KCs are enriched in mid regions in human and in periportal regions in mouse livers [41,42] and are located along LSECs in the sinusoids, with a significant part of the KC inside the space of Disse to ensure close contact with hepatocytes, HSCs and LSECs [43]. In the metabolically healthy liver, KCs clear common gut-derived antigens, including bacterial antigens and fungal pathogens, from blood by phagocytosing, processing, and presenting antigens to maintain immune homeostasis [44–47]. Both resident (KC) and infiltrating monocyte-derived macrophages play important roles in the development of hepatic steatosis and NASH. Analysis of liver biopsies of patients with

different stages of NAFLD show (portal) macrophage infiltration even at the stage of steatosis, before inflammation or fibrosis develop [48-50]. Macrophages are mainly observed around damaged hepatocytes, where they form hepatic crown-like structures (hCLS) [15,51-53]. In hCLS, activated macrophages aggregate and surround hepatocytes containing large lipid droplets and cholesterol crystals. The number of hCLS positively correlates with liver fibrosis [51], suggesting a pathophysiological role in NASH progression. Moreover, several studies in HFD-fed mice show that (selective) depletion of macrophages by gadolinium chloride (GdCl<sub>3</sub>) [27,54] or clodronate [55] improves hepatic steatosis, hepatocyte insulin resistance, and inflammation. However, as the employed depletion methods target all types of macrophages, firm conclusions about the exact role of KCs in NASH development cannot be drawn from these studies. Upon liver damage macrophages encompass several heterogeneous populations [56]. Resident KCs represent the largest macrophage population in the liver. When activated, KCs express high levels of inflammatory markers and are the major source of liver-derived MCP-1 (also known as CCL2), which plays an important role in the progression of simple steatosis to NASH) in HFD-fed mice [57]. The second most abundant population are monocyte-derived infiltrating macrophages which express high levels of LY6C and C-C motif chemokine receptor 2 (CCR2), and acquire macrophage characteristics over time [57]. Bonnardel et al. showed, in a KC-specific depletion model, that activated HSCs and LSECs induce the expression of monocyte-attracting chemokines and adhesion molecules [43]. Within a short time-frame – 24 h after KC depletion – monocytes colonize the liver and acquire LXRα and ID3 expression, which are an important part of the KC-specific identity program [58]. Expression of LXRa, via the DLL4-Notch pathway, and ID3 are respectively induced upon interaction with LSECs and hepatocytes, underlining the contribution of different hepatic cell

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types in shaping the KC phenotype. Another study, using a similar KC-specific depletion model, confirmed this rapid differentiation of recruited monocytes and showed that tissue environment signals, such as DLL4, TGF-B and desmosterol, induce differentiation by regulating the expression of KC identity transcription factors [59]. In NAFLD, monocytederived macrophages are recruited to the liver by the CCL2/CCR2 axis where they promote NASH development. Indeed, monocyte-derived infiltrating macrophages with a proinflammatory phenotype are abundant in HFD-fed mice, in ob/ob mice [60] and in humans with NAFLD [61]. Pharmacological inhibition of CCR2 decreased monocyte recruitment in the liver and ameliorated NASH in mice [62-64]. Three recent studies aimed to further identify resident and infiltrating macrophages in mice fed either Western-type (high fat and cholesterol) [65], long term NASH-inducing (high fat, high cholesterol, high fructose) [16], or methionine and choline-deficient (MCD) [66] diets (Box 4). Collectively, these studies showed that chronic steatohepatitis leads to permanent changes in the KC pool, with potential long-lasting impact on liver homeostasis and function, especially on the liver response to lipid overload. The use of different mouse models and protocols to induce steatosis and NASH (e.g. with or without additional fructose, choline-deficiency, study duration) with different phenotypic stages of hepatic NAFLD [67] complicates comparison of the identified populations. These studies also underscore the need for a consensus on the macrophage nomenclature. However, the heterogenous macrophage populations clearly play diverse roles in NASH. Functionally characterizing each subset will be of importance to develop more selective therapeutic (immune-based) strategies.

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The various subsets of dendritic cells (DCs) are differentially affected in NASH

NASH is associated with the accumulation of intrahepatic DCs and alterations in the proportions of DC subsets. DCs are the major APCs in the liver and in the metabolically healthy liver they control the induction of antigen-specific adaptive immune responses as well as the induction of central and peripheral tolerance to auto-antigens [68,69]. Hepatic DCs (~1% of the non-parenchymal cells) are represented by three populations: plasmacytoid DCs (pDCs), classical type 1 DCs (cDC1s, expressing XCR1 and CD103), priming the CD8<sup>+</sup> T cell response, and classical type 2 DCs (cDC2s, expressing CD11b and Sirpα), priming the CD4<sup>+</sup> T cell response [70-72]. At homeostasis, hepatic DCs are mostly immature and tolerogenic [73]. However, upon liver injury, activated DCs migrate to the periportal and pericentral areas and draining lymph nodes, secrete pro-inflammatory cytokines, such as IL-6, IL-12, and TNF-α, and express co-stimulatory molecules CD40, CD80 and CD86 [74,75]. The cDC1 (XCR1<sup>+</sup>)/cDC2 (CD172a<sup>+</sup>) ratio is decreased in livers of High Fat, High Cholesterol, High Sucrose diet fed mice, mainly due to a proportional (% of CD45<sup>+</sup> cells) increase in cDC2 and a slight reduction in cDC1 cells [18]. Moreover, cDC2 positively associate with NASH, while cDC1 negatively correlate with NASH. Surprisingly, the increase in cDC2 cells is paralleled by an increase in hepatic CD8<sup>+</sup> T cells, suggesting that changes in the cDC1/cDC2 ratio might fine tune CD8<sup>+</sup> T cell activation, likely through complex mechanisms. Indeed, T cell activation by cDC1 cells is classically MHC-I restricted. However, cDC1 cells also regulate the anti-tumor function of CD8<sup>+</sup> T cells through MHC-II-dependent regulation of CD4<sup>+</sup> helper T cells [76]. Another study reported elevated numbers of both cDC1 and cDC2 cells in mice fed Western, choline-deficient high fat, or MCD diets, while only the abundance of cDC2 cells was increased in high fat diet-fed or db/db and ob/ob obese mice [77]. The use of different diets and treatment durations to induce NASH-like pathologies, with variable relatedness to the

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human pathology [67,78], likely account for the observed differences in cDC1 and cDC2 abundance and ratio.

Immune cell profiling of blood and liver biopsies from NAFLD patients also revealed a NASH-dependent elevation of CD141<sup>+</sup> cells, corresponding to murine cDC1 cells [77]. Furthermore, an enrichment in (activated) migratory CCR7<sup>+</sup> DC (mDC) upon NASH induction was observed. In line, increased interactions between cDCs with NASH-associated CD8<sup>+</sup> T cells were observed in liver-draining lymph nodes of MCD-diet fed mice. A recent study, using *Batf3*<sup>-/-</sup> mice, which genetically lack cDC1 cells, identified murine CD103<sup>+</sup> cDC1 as a protective DC subtype during NASH development [79]. Adoptive transfer of these cells in *Batf3*<sup>-/-</sup> mice attenuated inflammation and reduced high sucrose diet-induced metabolic disturbances. Finally, specific cDC1 deletion, using XCR1-Cre<sup>DTA</sup> transgenic mice, protected mice for MCD-induced NASH [77]. The authors suggested that increased cDC1 numbers cause inflammation-related liver damage (NASH), in part *via* their interaction with T cells, while non-inflammatory accumulation of hepatic lipids (simple steatosis) is associated with increased numbers of cDC2 cells. However, lack of cDC2-specific depletion tools prevented so far the functional assessment of their role in NAFL and/or NASH development.

Infiltrating neutrophils facilitate NASH development

Bone-marrow-derived neutrophils mainly circulate via the blood stream for immune surveillance and are activated and attracted by inflammation or infection-activated endothelial cells [80]. Neutrophils infiltrate, as first responders, into tissues where they immobilize and kill pathogens acting through the production of reactive oxygen species (ROS), secretion of a variety of cytokines [81], such as IFN $\gamma$ , TNF- $\alpha$ , IL-4, and IL-10, and the formation of neutrophil extracellular traps (NETs) [82,83]. NETs are formed when

neutrophils actively release their nuclear DNA content, while still maintaining membrane integrity, chemotaxic responsiveness and phagocytosis [84].

Infiltration of neutrophils, mainly around steatotic hepatocytes and in the portal tract [50], is associated with the early stage of NAFL/NASH in patients and animal models [80,85-88]. Neutrophils contribute to NAFLD progression and the exacerbation of the inflammatory state by the production of neutrophil-derived myeloperoxidase (MPO), which accelerates ROS production and induces DNA damage [52,89]. NET formation (NETosis) also attracts monocyte-derived macrophages and subsequently induces steatosis-to-NASH progression [87]. In NASH patients, circulating neutrophils suppress the proliferation and activation of CD4<sup>+</sup> and CD8<sup>+</sup> T cells [90]. Over time, this may induce an inadequate immune-surveillance of liver damage, making patients more prone to the progression of liver damage. Although neutrophils are generally believed to promote NASH, they can also play a beneficial role. Indeed, neutrophils highly express microRNA-223, which upon transfer to hepatocytes elicits anti-inflammatory and anti-fibrotic responses [91]. However, antibody-mediated neutrophil depletion improved hepatic inflammation and fibrosis, and ameliorated HSC activation in high fat, high cholesterol-induced NASH in mice, indicating a detrimental role of neutrophils in NASH. Subsequent chow diet feeding, to study the role of neutrophils during (spontaneous) NASH resolution, showed that neutrophil depletion exacerbated fibrosis, HSC activation, and myeloid cell infiltration during resolution, demonstrating that neutrophils also play a pro-resolutive role in liver injury and fibrosis [92].

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The role of conventional  $CD4^{\dagger}$  Th1 and Th17 lymphocytes in the contribution to NASH development deserves further study

The main hepatic T cell populations in a healthy liver are conventional CD4 $^+$  and CD8 $^+$   $\alpha\beta$  T cells. Upon activation by APCs [93], they play a critical role in the hepatic immune response. Although the contribution of CD8<sup>+</sup> T cells to NAFL and NASH development has been recently extensively studied, data about the precise role of CD4<sup>+</sup> T cells are limited. CD4<sup>+</sup> T helper 1 (Th1) and T helper 17 (Th17) cells are increased in peripheral blood of NAFL [94] and NASH [18,95-98] patients. Th1 cells are important drivers of inflammation and contribute to NAFLD development by secreting IFNy [99]. Hepatic infiltration of and IL-17A production by Th17 cells increase steatosis, liver injury and fibrosis [98], while Th17 depletion in mice protects against NASH development [100]. The role of CD4<sup>+</sup> regulatory T cells (Tregs) in NASH development still remains controversial. Tregs are reported to be reduced in high-fat diet-fed mice [101], while choline-deficient high-fat and high-fat highcarbohydrate diets increase intrahepatic Tregs in mice [102,103]. Mechanistically, circulating IL-10<sup>+</sup> Tregs positively correlate with NASH features. However, as IL-10 is considered to be anti-inflammatory, this likely reflects a counter-adaptive immunoregulatory response [18,104]. A better understanding of the contribution of Tregs to NASH development will provide more insight in the hepatic immune regulation upon a metabolic challenge. Finally, currently no studies mechanistically addressed the role of CD4<sup>+</sup> T cells in NASH. Recently it was shown that Th2 cells negatively correlate with NASH activity [18], but the role of Th2 cells in NASH remains also to be investigated.

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CD8<sup>+</sup> cytotoxic T lymphocytes: the ultimate effectors in NASH?

Livers of high fat (and high cholesterol) and MCD diet fed mice are characterized by a hepatic infiltration of T cells, which is also observed in livers and peripheral blood of NAFL and NASH patients [18,50,105–108]. CD8<sup>+</sup> cytotoxic T cells play a key role in NASH progression, as

administration of neutralizing anti-CD8 antibodies decreased hepatic steatosis and inflammation [106,107]. Our previous work in NAFLD patients showed a strong correlation between circulating and hepatic cytotoxic (perforin and IFNγ-expressing) CD8<sup>+</sup> T cells and hepatic markers of inflammation in NASH [18]. Moreover, the cytotoxic potential of CD8<sup>+</sup> T cells in NASH was increased as illustrated by the increased expression of perforin, a poreforming protein, and the cytotoxic proteins granzyme A, a tryptase, and granzyme B, a serine protease, which are key for the effector function of cytotoxic T cells. Regression of NASHspecific features, such as lobular inflammation and ballooning, correlated with a decrease in cytotoxic CD8<sup>+</sup> T cells [18]. In line, Dudek et al. reported hepatic accumulation of tissue resident (CXCR6<sup>+</sup>) CD8<sup>+</sup> T cells in patients with NASH and in choline-deficient high-fat dietfed mice, which contribute to liver damage [109]. These short-lived tissue-resident CXCR6<sup>+</sup> CD8<sup>+</sup> T cells co-express programmed cell death protein 1 (PD1), which is associated with exhaustion. Furthermore, they express genes associated with apoptosis and with effector function, such as granzyme B, Tox, TNF- $\alpha$ , and IFN $\gamma$ . In the healthy liver, CXCR6 expression in CD8<sup>+</sup> T cells is inhibited by FOXO1. However, in NASH patients and mouse models of NASH, serum levels of the FOXO1-inhibitor IL-15 are increased, resulting in a higher expression of CXCR6 on T cells. In an environment with specific stimuli, such as TNF- $\alpha$  or acetate – a shortchain fatty acid which is elevated in NASH livers -, CXCR6<sup>+</sup> CD8<sup>+</sup> T cells acquire the potential of auto-aggression [109]. CXCR6<sup>+</sup> CD8<sup>+</sup> T cells were shown to contribute to liver damage by non-specific, MHC-I independent, killing of hepatocytes. This was triggered by ATP released from dying hepatocytes which, via the purinergic receptor P2X7, enhances calcium influx in CXCR6<sup>+</sup> CD8<sup>+</sup> T cells initiating auto-aggression. Tissue damage due to auto-aggression by CD8<sup>+</sup> T cells, and the upregulation of PD1 in these cells, can promote the development of liver cancer upon chronic liver damage [110–112]. Although the activation of auto-aggressive

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CXCR6<sup>+</sup> CD8<sup>+</sup> T cells is antigen-independent, the strong association between elevated hepatic CD8<sup>+</sup> T and cDC1 cells, as discussed above, also points toward a likely role of antigen presentation in CD8<sup>+</sup> T cell activation in NASH [18,77] (Figure 4). Future studies will be needed to further elucidate the respective contribution of antigen (in)dependent stimulated CD8<sup>+</sup> T cells in the progression of NAFL to NASH. Interestingly, the metabolic environment affects both CD8<sup>+</sup> T cell phenotype and function in NASH development. Hepatic CD8<sup>+</sup> T cells in Western diet-fed Low Density Lipoprotein Receptor-deficient mice, which display an obese/hyperlipidemic NASH phenotype, express IL-10, TNF-α, and IL-6, and induce hepatic inflammation, macrophage accumulation and HSC activation [113]. Although IL-10 has anti-inflammatory properties, it was hypothesized that IL-10 secretion from CD8<sup>+</sup> T cells promotes CD8<sup>+</sup> T cell survival and TIMP-1 expression in macrophages, hence stimulating NASH progression. Whereas CD8<sup>+</sup> T cells are the primary regulators of HSC activation selectively in an obese/hyperlipidemic environment, CD8<sup>+</sup> T cells play a minimal role in NASH development in non-obese animals [113]. Indeed, hepatic CD8<sup>+</sup> T cells in a lean NASH model, i.e. MCD diet-fed mice, exhibited a different phenotype and expressed low levels of IL-10, IFNγ, and granzyme B. Moreover, these CD8<sup>+</sup> T cells did not impact on NASH-associated inflammation nor HSC activation. These findings, indicating that metabolic conditions influence CD8<sup>+</sup> T cell phenotype and function, deserve further investigation in additional, more relevant (non-)obese NASH models.

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The role of non-conventional T lymphocytes in NASH is understudied

The healthy liver contains, next to innate lymphoid cells (Box 5), a substantial number of non-conventional T cells: natural killer T (NKT) cells, mucosal-associated invariant T (MAIT) cells and  $\gamma\delta$  T cells. NKT cells are the most abundant hepatic non-conventional T cells in mice

[114]. Two types of NKT cells are found: type I NKT cells expressing a semi-invariant TCR (invariant (i)NKT cells), and type II NKT cells, expressing a diverse TCR repertoire [115]. NKT cells recognize lipid antigens presented by CD1d and can rapidly respond by secreting multiple cytokines (IFNy, IL-4, IL-10 and IL-17A) [116]. CD1d is an MHC-I-like molecule that is reactive to self-derived or microbial lipid antigens, including glycolipids and phospholipids [114]. As CD1d is expressed by antigen presenting cells, such as DCs and KCs [117], NKT cells form a bridge between the innate and adaptive immune systems. NKT cell numbers change during the onset of NAFL and progression to NASH. A reduction in hepatic NKT cell numbers was reported in mouse models on short-term MCD, choline-deficient L-amino acid-deficient (CDAA), or high-fat diets [118,119]. Decreases in hepatic NKT cell numbers can be due to steatosis-induced KC-derived IL-12 and Tim-3/Gal-9 signaling, which promotes NKT cell apoptosis. By contrast, NKT cell numbers increase in advanced NASH. Mice fed a high fat, high cholesterol diet for 16 weeks showed increased NKT cells in liver [106]. NKT cell depletion protected against NASH progression, as these mice exhibited lower NAFLD activity scores (NAS) and AST/ALT levels, less hepatic infiltrating macrophages and decreased expression of genes related to fibrosis [106]. Similarly, a study combining choline-deficiency with a high fat diet for 6 months showed a significant increase in hepatic NKT cells [107]. A possible explanation for the increase in NKT cell numbers is the activation of the Hedgehog pathway in NASH. This results in higher levels of the NKT cell chemokine CXCL16, the adhesion molecule VCAM-1, and IL-15, all promoting NKT viability [120-122]. Several NKToperated mechanisms can contribute to NASH progression. Activated intrahepatic NKT cells enhance hepatocyte lipid uptake via secretion of LIGHT, thus promoting steatosis [107]. Moreover, NKT cells produce IL-17 and IL-22 at the beginning of the steatosis phase, becoming IFNy/IL-4/IL-13- secreting cells at a later stage of the disease [123]. The differential

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activation and cytokine secretion by NKT cells results in an evolving immune response during disease progression. Moreover, the synergistic action of NKT and CD8<sup>+</sup> T cells contributes to inflammation and fibrosis [106,123]. MAIT cells are a population of innate-like T lymphocytes that recognize a limited array of microbial-derived antigens and vitamin B metabolites presented by MHCI-I-related molecule 1 (MR1) [124]. In the metabolically healthy liver, they are important for antimicrobial immunity against bacteria and yeast, and exert cellular cytotoxicity against target cells by producing pro-inflammatory cytokines such as IFN $\gamma$  and TNF- $\alpha$  [125]. MAIT cells accelerate inflammation and fibrosis in models of CCL<sub>4</sub>- or bile duct ligation-induced liver injury and accumulate in livers of NASH patients [126]. By contrast, MAIT cells were found to be protective against liver inflammation in MCD diet-induced NASH models [127]. Beyond the fact that very different experimental models were studied, it is difficult to speculate about possible mechanisms accounting for these opposing results.  $\gamma\delta$  T cells express a TCR  $\gamma$ -chain and a  $\delta$ -chain and are a heterogenous population consisting of multiple subsets. They function as the first line of immune defense against infections and cancer via cytotoxic effects and production of IFNγ and IL-17 [128,129]. The liver contains γδ T cell precursors, which primarily develop into IFN $\gamma^+$   $\gamma\delta$  T cells, but their role in immune homeostasis and NAFL/NASH progression still needs to be determined [129].

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B lymphocytes and antibody production promotes NASH development

B cells are the largest population of hepatic lymphocytes (~20% of all CD45<sup>+</sup> cells in mice) [130] and are important for both humoral immunity, by producing antibodies, and cellular immunity, by functioning as APCs. They are located close to the bile ducts and are reported to function as APCs for MAIT cells by presenting gut-derived bacterial antigens *via* MR-1

[131,132]. Most hepatic B cells are mature follicular B cells, which survey for antigens. In the human liver, immature, "classical" B2, "non-conventional" B1, and antibody-producing plasma cells have also been detected, although their precise function at steady state is unclear [133,134]. Growing evidence suggests an essential role for B cells in NAFL and NASH development. B cells accumulate in the portal tract of livers of patients with NASH [50,135], and serum levels of IgA [136] and B cell-activating factor (BAFF), promoting B cell differentiation and survival, are increased in NASH patients [137]. Accumulated B cells in livers of HFD- or NASH-diet fed mice are pro-inflammatory and secrete cytokines such as IL-6 and TNF- $\alpha$  [137–139]. The (innate) TLR-associated Myeloid Differentiation primary response protein 88 (MyD88) pathway is required for B cell activation and maturation in NASH [139] and for their profibrotic role, as shown in a mouse model with CCl<sub>4</sub>-induced liver fibrosis [140]. Moreover, microbiota-derived metabolites draining into the liver as a result of increased gut permeability activate intrahepatic B cells during NASH [139]. B2 cells mature towards IgG producing plasma cells, that promote the presentation of oxidative stress-derived antigens to CD4<sup>+</sup> T cells and support NASH progression [135]. IgA producing plasma cells also contribute to NASH progression, as mice lacking IgA developed less liver injury and fibrosis upon a HFD [141]. Moreover, B cell-harboring, but antibodydeficient mice (IgMi mice) fed a high fat diet were protected from the development of hepatic steatosis and inflammation [142], suggesting a role for 'pathogenic' antibodies. Altogether, while hepatic B cell infiltration in NASH is established, the exact activation mechanisms and effector functions are yet to be determined.

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Concluding remarks; an integrated and perspective view of immune dysregulation in NASH

NAFL and its progression towards NASH are closely associated with environmental factors, such as overnutrition, and with dysregulation of energy metabolism, such as insulin resistance and obesity. Therefore, dietary and lifestyle interventions are the primary strategy to reduce the prevalence of this most common liver disease. Weight loss improves hepatic steatosis, and there is evidence that diets low in saturated and high in monounsaturated fats and dietary fiber (Mediterranean diet) are beneficial in NAFLD [10,143]. Lowering hepatic steatosis also decreases the risk for progression towards NASH. The fact that bariatric surgery and subsequent great and sustained weight loss can reverse pathological liver features in all stages of NAFLD, including fibrosis [6], proves that energy balance control is beneficial in NAFLD. Still, pharmacological therapies are eagerly awaited and several clinical trials are running to evaluate the effectiveness of different pharmacological targets on the reversal of NASH and/or fibrosis, assessing histological steatosis, hepatocyte ballooning, lobular inflammation and fibrosis stage. Inflammation is considered a key element of disease progression. The presence of a wide diversity of immune cells in NAFLD livers and their emerging roles in the progression to NASH makes the hepatic immune compartment an attractive target for therapeutic approaches. However, the complexity of different immune cell subsets in the sequential NAFL/NASH stages, and the knowledge gap about cell-cell interactions modifying disease progression, complicates the development of new therapies. An integration of the current knowledge will highlight unanswered questions on the role of immune cells in NASH, which need to be investigated in the future to develop specific immune-modulatory therapeutics for NAFLD (Figure 5, Outstanding questions). Accumulation of hepatic fat (steatosis), as occurs in energy substrate overload conditions such as diabetes, induces hepatocyte (mitochondrial, oxidative and ER) stress, which

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promotes the progression of NAFL towards NASH. Due to the altered metabolic environment, these damaged, necrotic or apoptotic hepatocytes initially induce the recruitment of monocytes, via the CCL2/CCR2 axis or other, yet unknown, metabolic and tissue environmental signals. The recruited monocyte-derived macrophages replace resident KCs, whose abundance decreases due to impaired self-renewal. Phenotypically diverse macrophage populations are present in NASH, but how dysregulation of the hepatic macrophage compartment impacts NASH development is yet unknown. Importantly, macrophages are the most abundant hepatic immune cells, but the impact of their altered phenotype on the immune response in NASH still remains to be investigated. It is tempting to speculate that alterations in macrophage subtypes affect antigen presentation, and/or DC and T cell function and phenotype, which are modified in NASH. DC abundance increases in NASH livers, whereas the cDC1/cDC2 ratio decreases. The exact mechanism behind the altered cDC1/cDC2 ratio, as well as its effect on CD8<sup>+</sup> T cell activation and the function of cDC2 in NASH are still unknown. However, cDC1 cells interact with CD8<sup>+</sup> T cells in NASH, enhancing their inflammatory signature. Moreover, antigen presentation by DCs contributes to the increase in (proinflammatory) Th1 and Th17 CD4<sup>+</sup> T cells. As DCs are important hepatic antigen presenting cells, the impact of the altered cDC1/cDC2 ratio in NASH development awaits further investigation. Alterations in CD4<sup>+</sup> T cell abundance may subsequently affect the activation B cells, whose abundance is increased in NASH livers. In turn, activated B cells secrete proinflammatory cytokines, such as TNF- $\alpha$  and IL-6, as well as IgA and IgG, and recruit T cells. The exact role of B cells in NASH development, the potential differential role of various B cell subsets, and the possible antigen specificity in NASH remain to be investigated. In a NASH environment, CD8<sup>+</sup> T cells acquire the ability to kill hepatocytes, and to activate HSC, hence likely enhancing fibrogenesis. Moreover, their

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synergistic interaction with NKT cells also promotes hepatic inflammation and fibrosis. Of note, NKT cell abundance is increased in NASH, probably due to IL-15 and CXCL16 secreted by macrophages. LIGHT secretion by NKT cells further enhances hepatocyte lipid accumulation, thus further promoting hepatic steatosis and contributing to NAFLD progression. The cellular interaction between NKT cells and hepatocytes shows integrated communication between parenchymal cells and the immune compartment. As described, major hepatic non-immune cell populations do play an important role in hepatic immunity. This role is fairly well documented at metabolic homeostasis, but information in a metabolically altered context is largely lacking. Further in depth understanding of the important, complex roles of both hepatic immune cells and parenchymal cells in NAFLD progression is necessary for a better understanding of the pathophysiology of this disease and the subsequent development of effective therapeutic strategies.

Targeting cDC-T cell interactions: an immune based therapeutic perspective in NASH.

Major recent publications [18,77,109,112] have highlighted the key association of cDCs and CD8<sup>+</sup> T cells upon progression of NAFL to NASH. While aberrant activation of CD8<sup>+</sup> T cells and development of an auto-aggressive exhausted phenotype appeared independent of antigen presentation, recent data from mice fed a MCD diet to induce NASH show that (mature or migratory) cDC1 and CD8<sup>+</sup> T cells physically interact in liver-draining lymph nodes [77]. Since physical interactions are a prerequisite for antigen presentation, it is likely that (conventional) antigen presentation to CD8<sup>+</sup> T cells takes place and plays a yet to be established role in NASH development. In line, cDC1 depletion significantly decreased NASH development, and immune cell infiltration, suggesting a functional role for these cells in NASH pathology. Finally, indoleamine 2,3-dioxygenase 1 (IDO1) expressing cDC1 cells

correlated with a prognostic liver signature of NAFLD progression. Inhibition of IDO1, the rate-limiting enzyme that converts L-tryptophan to L-kynurenine, in an *in vitro* system decreased the expression of several of the prognostic NAFLD liver signature genes [144]. While the molecular identification of (a) NASH-specific antigen(s) is awaiting, these data strongly suggest that blocking cDC-CD8<sup>+</sup> T cell interactions and thus antigen presentation might represent a promising therapeutic strategy in NASH. Moreover, identification of NASH-specific antigens could open new options for antigen-specific based therapies.

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### S40 References

- 541 1 Si-Tayeb, K. et al. (2010) Developmental Cell Organogenesis and Development of the
- 542 Liver. *DEVCEL* 18, 175–189
- 543 2 Haas, J. et al. (2016) Pathophysiology and Mechanisms of Nonalcoholic Fatty Liver
- 544 Disease. *Annu. Rev. Physiol.* 78, 181–205
- Neuschwander-Tetri, B. (2010) Hepatic lipotoxicity and the pathogenesis of
- 546 nonalcoholic steatohepatitis: the central role of nontriglyceride fatty acid metabolites.
- 547 *Hepatology* 52, 774–788
- 548 4 Horn, C.L. et al. (2022) Role of Cholesterol-Associated Steatohepatitis in the
- 549 Development of NASH. *Hepatol. Commun.* 6, 12–35
- 550 5 Bence, K.K. and Birnbaum, M.J. (2020) Metabolic drivers of non-alcoholic fatty liver
- 551 disease. *Mol. Metab.* 50, 101143
- 552 6 Laursen, T.L. et al. (2019) Bariatric surgery in patients with non-alcoholic fatty liver
- disease from pathophysiology to clinical effects. World J. Hepatol. 11, 138
- 554 7 Marra, F. and Svegliati-Baroni, G. (2018) Lipotoxicity and the gut-liver axis in NASH
- 555 pathogenesis. *J. Hepatol.* 68, 280–295
- 556 8 Kleiner, D.E. et al. (2019) Association of Histologic Disease Activity With Progression of
- Nonalcoholic Fatty Liver Disease. JAMA Netw. open 2, e1912565
- 558 9 Schuppan, D. et al. (2018) Determinants of fibrosis progression and regression in
- 559 NASH. *J. Hepatol.* 68, 238–250
- 560 10 Yki-Järvinen, H. et al. (2021) Dietary carbohydrates and fats in nonalcoholic fatty liver
- disease. *Nat. Rev. Gastroenterol. Hepatol.* 18, 770–786
- 11 Ibrahim, S.H. et al. (2018) Non-alcoholic steatohepatitis pathogenesis: sublethal
- hepatocyte injury as a driver of liver inflammation. *Gut* 67, 963

564 12 Bartneck, M. et al. (2016) Histidine-rich glycoprotein promotes macrophage activation 565 and inflammation in chronic liver disease. Hepatology 63, 1310-1324 566 13 Leroux, A. et al. (2012) Toxic lipids stored by Kupffer cells correlates with their proinflammatory phenotype at an early stage of steatohepatitis. J. Hepatol. 57, 141–149 567 568 14 Govaere, O. et al. (2022) Macrophage scavenger receptor 1 mediates lipid-induced 569 inflammation in non-alcoholic fatty liver disease. J. Hepatol. 76, 1001–1012 570 15 Daemen, S. et al. (2021) Dynamic Shifts in the Composition of Resident and Recruited 571 Macrophages Influence Tissue Remodeling in NASH. Cell Rep. 34, 108626 572 Seidman, J.S. et al. (2020) Niche-Specific Reprogramming of Epigenetic Landscapes 16 573 Drives Myeloid Cell Diversity in Nonalcoholic Steatohepatitis. Immunity 52, 1057-574 1074.e7 575 17 Mogilenko, D. et al. (2019) Metabolic and Innate Immune Cues Merge into a Specific 576 Inflammatory Response via the UPR. Cell 177, 1201-1216.e19 577 18 Haas, J.T. et al. (2019) Transcriptional network analysis implicates altered hepatic 578 immune function in NASH development and resolution. Nat. Metab. 1, 604-614 579 Stentz, F.B. and Kitabchi, A.E. (2006) Palmitic acid-induced activation of human T-19 580 lymphocytes and aortic endothelial cells with production of insulin receptors, reactive 581 oxygen species, cytokines, and lipid peroxidation. Biochem. Biophys. Res. Commun. 582 346, 721–726 583 20 Fernanda Cury-Boaventura, M. et al. (2006) Mechanisms involved in Jurkat cell death 584 induced by oleic and linoleic acids. Clin. Nutr. 25, 1004–1014 585 21 de Jong, A.J. et al. (2014) Fatty Acids, Lipid Mediators, and T-Cell Function. Front. 586 *Immunol.* 5, 1–7 587 Newsholme, P. et al. (1987) Rates of utilization and fates of glucose, glutamine, 22

588 pyruvate, fatty acids and ketone bodies by mouse macrophages. Biochem. J. 242, 589 631-636 590 23 van Uden, P. et al. (2008) Regulation of hypoxia-inducible factor-1α by NF-κB. 591 Biochem. J. 412, 477-484 592 24 Arranz, A. et al. (2012) Akt1 and Akt2 protein kinases differentially contribute to 593 macrophage polarization. Proc. Natl. Acad. Sci. 109, 9517-9522 594 25 Cheng, S.-C. et al. (2014) mTOR- and HIF- $1\alpha$ -mediated aerobic glycolysis as metabolic 595 basis for trained immunity. Science (80-. ). 345, 1250684 596 Viola, A. et al. (2019) The Metabolic Signature of Macrophage Responses. Front. 26 597 *Immunol.* 10, 1462 598 27 Huang, W. et al. (2010) Depletion of liver kupffer cells prevents the development of 599 diet-induced hepatic steatosis and insulin resistance. *Diabetes* 59, 347–357 600 Plas, D.R. et al. (2002) Homeostatic control of lymphocyte survival: potential origins 28 601 and implications. Nat. Immunol. 3, 515-521 602 29 Fox, C.J. et al. (2005) Fuel feeds function: energy metabolism and the T-cell response. 603 Nat. Rev. Immunol. 5, 844–852 604 30 Frauwirth, K.A. et al. (2002) The CD28 signaling pathway regulates glucose 605 metabolism. Immunity 16, 769-777 606 31 Enjoji, M. et al. (2012) Nutrition and nonalcoholic Fatty liver disease: the significance of cholesterol. Int. J. Hepatol. 2012, 1-6 607 608 Endo-Umeda, K. et al. (2018) Dysregulation of Kupffer Cells/Macrophages and Natural 32 609 Killer T Cells in Steatohepatitis in LXR α Knockout Male Mice. Endocrinology 159, 610 1419-1432 611 33 Endo-Umeda, K. and Makishima, M. (2019) Liver X Receptors Regulate Cholesterol

Metabolism and Immunity in Hepatic Nonparenchymal Cells. Int. J. Mol. Sci. 20, 5045 Campbell, C. et al. (2020) Bacterial metabolism of bile acids promotes generation of peripheral regulatory T cells. *Nature* 581, 475–479 Hang, S. et al. (2019) Bile acid metabolites control TH17 and Treg cell differentiation. *Nature* 576, 143–148 Song, X. et al. (2020) Microbial bile acid metabolites modulate gut RORy+ regulatory T cell homeostasis. Nature 577, 410-415 Chávez-Talavera, O. et al. (2019) Bile acid alterations in nonalcoholic fatty liver disease, obesity, insulin resistance and type 2 diabetes. Curr. Opin. Lipidol. 30, 244-Puri, P. et al. (2018) The presence and severity of nonalcoholic steatohepatitis is associated with specific changes in circulating bile acids. Hepatology 67, 534-548 Chen, L. et al. (2020) Genetic and Microbial Associations to Plasma and Fecal Bile Acids in Obesity Relate to Plasma Lipids and Liver Fat Content. Cell Rep. 33, 108212 Grzych, G. et al. (2021) NASH-related increases in plasma bile acid levels depend on insulin resistance. JHEP Reports 3, 100222 Gola, A. et al. (2021) Commensal-driven immune zonation of the liver promotes host defence. *Nature* 589, 131–136 Guilliams, M. et al. (2022) Spatial proteogenomics reveals distinct and evolutionarily conserved hepatic macrophage niches. Cell 185, 379-396.e38 Bonnardel, J. et al. (2019) Stellate Cells, Hepatocytes, and Endothelial Cells Imprint the Kupffer Cell Identity on Monocytes Colonizing the Liver Macrophage Niche. Immunity 51, 638-654.e9 Jenne, C.N. and Kubes, P. (2013) Immune surveillance by the liver. Nat. Immunol. 14,

| 536 |    | 996–1006  |
|-----|----|---|
| 537 | 45 | Sun, D. et al. (2019) Fungal dissemination is limited by liver macrophage filtration of   |
| 538 |    | the blood. Nat. Commun. 10, 4566  |
| 539 | 46 | Heymann, F. et al. (2015) Liver Inflammation Abrogates Immunological Tolerance            |
| 540 |    | Induced by Kupffer Cells. <i>Hepatology</i> 62, 279–291                                   |
| 541 | 47 | McDonald, B. et al. (2020) Programing of an Intravascular Immune Firewall by the Gut      |
| 542 |    | Microbiota Protects against Pathogen Dissemination during Infection. Cell Host            |
| 543 |    | Microbe 28, 660-668.e4  |
| 544 | 48 | Park, J.W. et al. (2007) Predictors reflecting the pathological severity of non-alcoholic |
| 545 |    | fatty liver disease: Comprehensive study of clinical and immunohistochemical findings     |
| 546 |    | in younger Asian patients. J. Gastroenterol. Hepatol. 22, 491–497                         |
| 547 | 49 | Lotowska, J.M. et al. (2013) The role of Kupffer cells in the morphogenesis of            |
| 548 |    | nonalcoholic steatohepatitis-ultrastructural findings. the first report in pediatric      |
| 549 |    | patients. Scand. J. Gastroenterol. 48, 352–357  |
| 550 | 50 | Gadd, V.L. et al. (2014) The portal inflammatory infiltrate and ductular reaction in      |
| 551 |    | human nonalcoholic fatty liver disease. Hepatology 59, 1393–1405                          |
| 552 | 51 | Itoh, M. et al. (2013) Hepatic Crown-Like Structure: A Unique Histological Feature in     |
| 553 |    | Non-Alcoholic Steatohepatitis in Mice and Humans. PLoS One 8, e82163                      |
| 554 | 52 | Rensen, S.S. et al. (2009) Increased hepatic myeloperoxidase activity in obese subjects   |
| 555 |    | with nonalcoholic steatohepatitis. Am. J. Pathol. 175, 1473–1482                          |
| 656 | 53 | Ioannou, G.N. et al. (2013) Hepatic cholesterol crystals and crown-like structures        |
| 557 |    | distinguish NASH from simple steatosis. J. Lipid Res. 54, 1326–1334                       |
| 558 | 54 | Neyrinck, A.M. et al. (2009) Critical role of Kupffer cells in the management of diet-    |
| 559 |    | induced diabetes and obesity. <i>Biochem. Biophys. Res. Commun.</i> 385, 351–356          |

660 55 Lanthier, N. et al. (2010) Kupffer cell activation is a causal factor for hepatic insulin 661 resistance. Am. J. Physiol. Liver Physiol. 298, G107-G116 662 56 Tacke, F. and Zimmermann, H.W. (2014) Macrophage heterogeneity in liver injury and 663 fibrosis. J. Hepatol. 60, 1090-1096 664 57 Morinaga, H. et al. (2015) Characterization of distinct subpopulations of hepatic 665 macrophages in HFD/obese mice. *Diabetes* 64, 1120–1130 666 58 Scott, C.L. et al. (2018) The Transcription Factor ZEB2 Is Required to Maintain the 667 Tissue-Specific Identities of Macrophages. *Immunity* 49, 312-325.e5 668 59 Sakai, M. et al. (2019) Liver-Derived Signals Sequentially Reprogram Myeloid 669 Enhancers to Initiate and Maintain Kupffer Cell Identity. Immunity 51, 655-670.e8 670 60 Deng, Z. Bin et al. (2009) Immature myeloid cells induced by a high-fat diet contribute 671 to liver inflammation. *Hepatology* 50, 1412–1420 672 61 Krenkel, O. et al. (2018) Therapeutic inhibition of inflammatory monocyte recruitment 673 reduces steatohepatitis and liver fibrosis. Hepatology 67, 1270-1283 674 62 Miura, K. et al. (2012) Hepatic recruitment of macrophages promotes nonalcoholic 675 steatohepatitis through CCR2. Am. J. Physiol. Liver Physiol. 302, G1310–G1321 676 63 Yang, S.J. et al. (2009) Inhibition of the chemokine (C-C motif) ligand 2/chemokine (C-677 C motif) receptor 2 pathway attenuates hyperglycaemia and inflammation in a mouse 678 model of hepatic steatosis and lipoatrophy. *Diabetologia* 52, 972–981 679 64 Baeck, C. et al. (2012) Pharmacological inhibition of the chemokine CCL2 (MCP-1) 680 diminishes liver macrophage infiltration and steatohepatitis in chronic hepatic injury. 681 Gut 61, 416-426 682 65 Remmerie, A. et al. (2020) Osteopontin Expression Identifies a Subset of Recruited 683 Macrophages Distinct from Kupffer Cells in the Fatty Liver. Immunity 53, 641-657.e14

Tran, S. et al. (2020) Impaired Kupffer Cell Self-Renewal Alters the Liver Response to Lipid Overload during Non-alcoholic Steatohepatitis. Immunity 53, 627-640.e5 Im, Y.R. et al. (2021) A Systematic Review of Animal Models of NAFLD Finds High-Fat, High-Fructose Diets Most Closely Resemble Human NAFLD. Hepatology 74, 1884-Hasegawa, H. and Matsumoto, T. (2018) Mechanisms of Tolerance Induction by Dendritic Cells In Vivo. Front. Immunol. 9, 1 Audiger, C. et al. (2017) The Importance of Dendritic Cells in Maintaining Immune Tolerance. J. Immunol. 198, 2223-2231 Miller, J.C. et al. (2012) Deciphering the transcriptional network of the dendritic cell lineage. Nat. Immunol. 13, 888-899 Guilliams, M. et al. (2014) Dendritic cells, monocytes and macrophages: a unified nomenclature based on ontogeny. Nat. Rev. Immunol. 14, 571-578 Mildner, A. and Jung, S. (2014) Development and Function of Dendritic Cell Subsets. *Immunity* 40, 642–656 Kelly, A. et al. (2014) CD141+ myeloid dendritic cells are enriched in healthy human liver. *J. Hepatol.* 60, 135–142 Shu, S.A. et al. (2007) The role of CD11c+ hepatic dendritic cells in the induction of innate immune responses. Clin. Exp. Immunol. 149, 335–343 Asselin-Paturel, C. et al. (2005) Type I interferon dependence of plasmacytoid dendritic cell activation and migration. J. Exp. Med. 201, 1157–1167 Ferris, S.T. et al. (2020) cDC1 prime and are licensed by CD4+ T cells to induce anti-tumour immunity. *Nature* 584, 624–629 Deczkowska, A. et al. (2021) XCR1+ type 1 conventional dendritic cells drive liver

708 pathology in non-alcoholic steatohepatitis. Nat. Med. 27, 1043-1054 709 78 Febbraio, M.A. et al. (2019) Preclinical Models for Studying NASH-Driven HCC: How 710 Useful Are They? Cell Metab. 29, 18–26 711 79 Heier, E.C. et al. (2017) Murine CD103+ dendritic cells protect against steatosis 712 progression towards steatohepatitis. J. Hepatol. 66, 1241–1250 713 80 Kolaczkowska, E. and Kubes, P. (2013) Neutrophil recruitment and function in health 714 and inflammation. Nat. Rev. Immunol. 13, 159-175 715 81 Tecchio, C. et al. (2014) Neutrophil-Derived Cytokines: Facts Beyond Expression. Front. 716 Immunol. 5, 717 82 Brinkmann, V. et al. (2004) Neutrophil Extracellular Traps Kill Bacteria. Science (80-. ). 718 303, 1532-1535 719 83 Takei, H. et al. (1996) Rapid killing of human neutrophils by the potent activator 720 phorbol 12-myristate 13-acetate (PMA) accompanied by changes different from 721 typical apoptosis or necrosis. J. Leukoc. Biol. 59, 229-240 722 84 Yipp, B.G. et al. (2012) Infection-induced NETosis is a dynamic process involving 723 neutrophil multitasking in vivo. *Nat. Med.* 18, 1386–1393 724 85 Bertola, A. et al. (2010) Hepatic Expression Patterns of Inflammatory and Immune 725 Response Genes Associated with Obesity and NASH in Morbidly Obese Patients. PLoS 726 One 5, e13577 727 86 Chang, B. et al. (2015) Short- or long-term high-fat diet feeding plus acute ethanol 728 binge synergistically induce acute liver injury in mice: An important role for CXCL1. 729 Hepatology 62, 1070–1085 730 87 van der Windt, D.J. et al. (2018) Neutrophil extracellular traps promote inflammation 731 and development of hepatocellular carcinoma in nonalcoholic steatohepatitis.

732 Hepatology 68, 1347–1360 733 88 Gao, B. and Tsukamoto, H. (2016) Inflammation in Alcoholic and Nonalcoholic Fatty 734 Liver Disease: Friend or Foe? Gastroenterology 150, 1704–1709 735 89 Hwang, S. et al. (2020) Interleukin-22 Ameliorates Neutrophil-Driven Nonalcoholic 736 Steatohepatitis Through Multiple Targets. Hepatology 72, 412–429 737 90 Antonucci, L. et al. (2020) Circulating Neutrophils of Nonalcoholic Steatohepatitis 738 Patients Show an Activated Phenotype and Suppress T Lymphocytes Activity. J. 739 Immunol. Res. 2020, 1-15 740 He, Y. et al. (2021) Neutrophil-to-hepatocyte communication via LDLR-dependent 91 741 miR-223—enriched extracellular vesicle transfer ameliorates nonalcoholic 742 steatohepatitis. J. Clin. Invest. 131, 743 92 Kim, A.D. et al. (2021) Dual role of neutrophils in modulating liver injury and fibrosis 744 during development and resolution of diet-induced murine steatohepatitis. Sci. Rep. 745 11, 1–12 746 93 Ficht, X. and Iannacone, M. (2020) Immune surveillance of the liver by T cells. Sci. 747 Immunol. 5, 748 94 Rau, M. et al. (2016) Progression from Nonalcoholic Fatty Liver to Nonalcoholic 749 Steatohepatitis Is Marked by a Higher Frequency of Th17 Cells in the Liver and an 750 Increased Th17/Resting Regulatory T Cell Ratio in Peripheral Blood and in the Liver. J. 751 *Immunol.* 196, 97–105 752 95 Pacifico, L. et al. (2006) Increased T-helper interferon-y-secreting cells in obese 753 children. Eur. J. Endocrinol. 154, 691–697 754 96 Ferreyra Solari, N.E. et al. (2012) The role of innate cells is coupled to a Th1-polarized 755 immune response in pediatric nonalcoholic steatohepatitis. J. Clin. Immunol. 32, 611Inzaugarat, M.E. et al. (2011) Altered phenotype and functionality of circulating immune cells characterize adult patients with nonalcoholic steatohepatitis. J. Clin. *Immunol.* 31, 1120–1130 Gomes, A.L. et al. (2016) Metabolic Inflammation-Associated IL-17A Causes Non-alcoholic Steatohepatitis and Hepatocellular Carcinoma. Cancer Cell 30, 161–175 Her, Z. et al. (2020) CD4+ T Cells Mediate the Development of Liver Fibrosis in High Fat Diet-Induced NAFLD in Humanized Mice. Front. Immunol. 11, Rolla, S. et al. (2016) The balance between IL-17 and IL-22 produced by liver-infiltrating T-helper cells critically controls NASH development in mice. Clin. Sci. 130, 193-203 Ma, X. et al. (2007) A high-fat diet and regulatory T cells influence susceptibility to endotoxin-induced liver injury. *Hepatology* 46, 1519–1529 Wang, H. et al. (2021) Regulatory T-cell and neutrophil extracellular trap interaction contributes to carcinogenesis in non-alcoholic steatohepatitis. J. Hepatol. 75, 1271-Dywicki, J. et al. (2022) The Detrimental Role of Regulatory T Cells in Nonalcoholic Steatohepatitis. *Hepatol. Commun.* 6, 320–333 Vonghia, L. et al. (2015) CD4+ROR γ t++ and Tregs in a Mouse Model of Diet-Induced Nonalcoholic Steatohepatitis. *Mediators Inflamm.* 2015, 1–10 Van Herck, M.A. et al. (2019) The Differential Roles of T Cells in Non-alcoholic Fatty Liver Disease and Obesity. Front. Immunol. 10, 82 Bhattacharjee, J. et al. (2017) Hepatic natural killer T-cell and CD8+ T-cell signatures in mice with nonalcoholic steatohepatitis. *Hepatol. Commun.* 1, 299–310

780 107 Wolf, M.J. et al. (2014) Metabolic activation of intrahepatic CD8+ T cells and NKT cells 781 causes nonalcoholic steatohepatitis and liver cancer via cross-talk with hepatocytes. 782 Cancer Cell 26, 549-564 783 Alegre, N.S. et al. (2020) Limited expression of TLR9 on T cells and its functional 108 784 consequences in patients with nonalcoholic fatty liver disease. Clin. Mol. Hepatol. 26, 785 216-226 786 109 Dudek, M. et al. (2021) Auto-aggressive CXCR6+ CD8 T cells cause liver immune 787 pathology in NASH. Nature 592, 444-449 788 Ma, C. et al. (2016) NAFLD causes selective CD4+ T lymphocyte loss and promotes 110 789 hepatocarcinogenesis. Nature 531, 253–257 790 111 Hansel, C. et al. (2019) The Inhibitory T Cell Receptors PD1 and 2B4 Are Differentially 791 Regulated on CD4 and CD8 T Cells in a Mouse Model of Non-alcoholic Steatohepatitis. 792 Front. Pharmacol. 10, 793 Pfister, D. et al. (2021) NASH limits anti-tumour surveillance in immunotherapy-112 794 treated HCC. *Nature* 592, 450–456 795 113 Breuer, D.A. et al. (2020) CD8 + T cells regulate liver injury in obesity-related 796 nonalcoholic fatty liver disease. Am. J. Physiol. Liver Physiol. 318, G211-G224 797 114 Chen, Y. and Tian, Z. (2021) Innate lymphocytes: pathogenesis and therapeutic targets 798 of liver diseases and cancer. Cell. Mol. Immunol. 18, 57–72 799 115 Shen, H. et al. (2020) Unveiling the heterogeneity of NKT cells in the liver through 800 single cell RNA sequencing. Sci. Rep. 10, 19453 801 116 Crosby, C.M. and Kronenberg, M. (2018) Tissue-specific functions of invariant natural 802 killer T cells. Nat. Rev. Immunol. 18, 559-574 803 117 Geissmann, F. et al. (2005) Intravascular immune surveillance by CXCR6+ NKT cells

| 804 |     | patrolling liver sinusoids. <i>PLoS Biol.</i> 3, 0650–0661   |
|-----|-----|--|
| 805 | 118 | Tang, ZH. et al. (2013) Tim-3/Galectin-9 Regulate the Homeostasis of Hepatic NKT                         |
| 806 |     | Cells in a Murine Model of Nonalcoholic Fatty Liver Disease. J. Immunol. 190, 1788–                      |
| 807 |     | 1796   |
| 808 | 119 | Kremer, M. et al. (2010) Kupffer cell and interleukin-12-dependent loss of natural                       |
| 809 |     | killer T cells in hepatosteatosis. <i>Hepatology</i> 51, 130–141   |
| 810 | 120 | Wehr, A. et al. (2013) Chemokine Receptor CXCR6-Dependent Hepatic NK T Cell                              |
| 811 |     | Accumulation Promotes Inflammation and Liver Fibrosis. J. Immunol. 190, 5226–5236                        |
| 812 | 121 | Locatelli, I. et al. (2013) NF-кВ1 deficiency stimulates the progression of non-alcoholic                |
| 813 |     | steatohepatitis (NASH) in mice by promoting NKT-cell-mediated responses. Clin. Sci.                      |
| 814 |     | 124, 279–287   |
| 815 | 122 | Syn, W.K. et al. (2010) Accumulation of natural killer T cells in progressive                            |
| 816 |     | nonalcoholic fatty liver disease. <i>Hepatology</i> 51, 1998–2007  |
| 817 | 123 | Maricic, I. et al. (2018) Differential Activation of Hepatic Invariant NKT Cell Subsets                  |
| 818 |     | Plays a Key Role in Progression of Nonalcoholic Steatohepatitis. J. Immunol. 201,                        |
| 819 |     | 3017–3035  |
| 820 | 124 | Treiner, E. et al. (2003) Selection of evolutionarily conserved mucosal-associated                       |
| 821 |     | invariant T cells by MR1. <i>Nature</i> 422, 164–169   |
| 822 | 125 | Legoux, F. $\it{et~al.}$ (2017) Unconventional or Preset $\alpha\beta$ T Cells: Evolutionarily Conserved |
| 823 |     | Tissue-Resident T Cells Recognizing Nonpeptidic Ligands. Annu. Rev. Cell Dev. Biol. 33,                  |
| 824 |     | 511–535  |
| 825 | 126 | Hegde, P. et al. (2018) Mucosal-associated invariant T cells are a profibrogenic                         |
| 826 |     | immune cell population in the liver. Nat. Commun. 9, 2146  |
| 827 | 127 | Li, Y. et al. (2018) Mucosal-Associated Invariant T Cells Improve Nonalcoholic Fatty                     |

| 828 |     | Liver Disease Through Regulating Macrophage Polarization. Front. Immunol. 9,                  |
|-----|-----|---|
| 829 | 128 | Chien, Y. et al. (2014) $\gamma\delta$ T Cells: First Line of Defense and Beyond. Annu. Rev.  |
| 830 |     | Immunol. 32, 121–155  |
| 831 | 129 | Hu, Y. et al. (2021) Single-cell analysis reveals the origins and intrahepatic                |
| 832 |     | development of liver-resident IFN-γ-producing γδ T cells. <i>Cell. Mol. Immunol.</i> 18, 954- |
| 833 |     | 968   |
| 834 | 130 | Nakagaki, B.N. et al. (2018) Immune and metabolic shifts during neonatal                      |
| 835 |     | development reprogram liver identity and function. J. Hepatol. 69, 1294–1307                  |
| 836 | 131 | Jeffery, H.C. et al. (2016) Biliary epithelium and liver B cells exposed to bacteria          |
| 837 |     | activate intrahepatic MAIT cells through MR1. J. Hepatol. 64, 1118–1127                       |
| 838 | 132 | Salerno-Goncalves, R. et al. (2014) B Cells Modulate Mucosal Associated Invariant T           |
| 839 |     | Cell Immune Responses. Front. Immunol. 4,   |
| 840 | 133 | Aizarani, N. et al. (2019) A human liver cell atlas reveals heterogeneity and epithelial      |
| 841 |     | progenitors. <i>Nature</i> 572, 199–204   |
| 842 | 134 | MacParland, S.A. et al. (2018) Single cell RNA sequencing of human liver reveals              |
| 843 |     | distinct intrahepatic macrophage populations. Nat. Commun. 9, 4383                            |
| 844 | 135 | Bruzzì, S. et al. (2018) B2-Lymphocyte responses to oxidative stress-derived antigens         |
| 845 |     | contribute to the evolution of nonalcoholic fatty liver disease (NAFLD). Free Radic.          |
| 846 |     | Biol. Med. 124, 249–259   |
| 847 | 136 | McPherson, S. et al. (2014) Serum immunoglobulin levels predict fibrosis in patients          |
| 848 |     | with non-alcoholic fatty liver disease. J. Hepatol. 60, 1055–1062                             |
| 849 | 137 | Miyake, T. et al. (2013) B cell-activating factor is associated with the histological         |
| 850 |     | severity of nonalcoholic fatty liver disease. <i>Hepatol. Int.</i> 7, 539–547                 |
| 851 | 138 | Zhang, F. et al. (2016) Role of intrahepatic B cells in non-alcoholic fatty liver disease by  |

| 852 |     | secreting pro-inflammatory cytokines and regulating intrahepatic T cells. J. Dig. Dis.      |
|-----|-----|---|
| 853 |     | 17, 464–474   |
| 854 | 139 | Barrow, F. et al. (2021) Microbiota-Driven Activation of Intrahepatic B Cells               |
| 855 |     | Aggravates NASH Through Innate and Adaptive Signaling. Hepatology 74, 704–722               |
| 856 | 140 | Thapa, M. et al. (2015) Liver fibrosis occurs through dysregulation of MyD88-               |
| 857 |     | dependent innate B-cell activity. Hepatology 61, 2067–2079                                  |
| 858 | 141 | Shalapour, S. et al. (2017) Inflammation-induced IgA+ cells dismantle anti-liver cancer     |
| 859 |     | immunity. <i>Nature</i> 551, 340–345  |
| 860 | 142 | Karl, M. et al. (2022) Dual roles of B lymphocytes in mouse models of diet-induced          |
| 861 |     | nonalcoholic fatty liver disease. <i>Hepatology</i> 00, 1–15                                |
| 862 | 143 | El-Agroudy, N.N. et al. (2019) Are Lifestyle Therapies Effective for NAFLD Treatment?       |
| 863 |     | Trends Endocrinol. Metab. 30, 701–709   |
| 864 | 144 | Fujiwara, N. et al. (2022) Molecular signatures of long-term hepatocellular carcinoma       |
| 865 |     | risk in nonalcoholic fatty liver disease. Sci. Transl. Med. 14, eabo4474                    |
| 866 | 145 | Warren, A. et al. (2006) T lymphocytes interact with hepatocytes through                    |
| 867 |     | fenestrations in murine liver sinusoidal endothelial cells. <i>Hepatology</i> 44, 1182–1190 |
| 868 | 146 | Guidotti, L.G. et al. (2015) Immunosurveillance of the liver by intravascular effector      |
| 869 |     | CD8+ T cells. <i>Cell</i> 161, 486–500  |
| 870 | 147 | Dolina, J.S. et al. (2017) Cross-Presentation of Soluble and Cell-Associated Antigen by     |
| 871 |     | Murine Hepatocytes Is Enhanced by Collectrin Expression. J. Immunol. 198, 2341–             |
| 872 |     | 2351  |
| 873 | 148 | Holz, L.E. et al. (2008) Intrahepatic Murine CD8 T-Cell Activation Associates With a        |
| 874 |     | Distinct Phenotype Leading to Bim-Dependent Death. Gastroenterology 135, 989–997            |
| 875 | 149 | Crispe, I.N. (2011) Liver Antigen-Presenting Cells. J. Hepatol. 54, 357                     |

Poisson, J. et al. (2017) Liver sinusoidal endothelial cells: Physiology and role in liver diseases. J. Hepatol. 66, 212–227 Ganesan, L.P. et al. (2011) Rapid and efficient clearance of blood-borne virus by liver sinusoidal endothelium. PLoS Pathog. 7, e1002281 Øie, C.I. et al. (2020) Liver sinusoidal endothelial cells contribute to the uptake and degradation of entero bacterial viruses. Sci. Rep. 10, 1–9 Øie, C.I. et al. (2011) Rat liver sinusoidal endothelial cells (LSECs) express functional low density lipoprotein receptor-related protein-1 (LRP-1). J. Hepatol. 55, 1346–1352 Thomson, A.W. and Knolle, P.A. (2010) Antigen-presenting cell function in the tolerogenic liver environment. Nat. Rev. Immunol. 10, 753-766 Carambia, A. et al. (2014) TGF-β-dependent induction of CD4+CD25+Foxp3 + Tregs by liver sinusoidal endothelial cells. J. Hepatol. 61, 594–599 Schurich, A. et al. (2010) Dynamic regulation of CD8 T cell tolerance induction by liver sinusoidal endothelial cells. J. Immunol. 184, 4107-4114 Schurich, A. et al. (2009) Distinct kinetics and dynamics of cross-presentation in liver sinusoidal endothelial cells compared to dendritic cells. Hepatology 50, 909-919 Schildberg, F.A. et al. (2011) Prominent regulatory but weak antigen-presenting cell function of hepatic stellate cells. Hepatology 54, 1108-1108 Winau, F. et al. (2007) Ito Cells Are Liver-Resident Antigen-Presenting Cells for Activating T Cell Responses. *Immunity* 26, 117–129 Bomble, M. et al. (2010) Analysis of antigen-presenting functionality of cultured rat hepatic stellate cells and transdifferentiated myofibroblasts. Biochem. Biophys. Res. Commun. 396, 342-347 Gupta, G. et al. (2019) Role of hepatic stellate cell (HSC)-derived cytokines in hepatic

| 900 |     | inflammation and immunity. Cytokine 124, 154542  |
|-----|-----|--|
| 901 | 162 | Shang, L. et al. (2018) Human hepatic stellate cell isolation and characterization. J.     |
| 902 |     | Gastroenterol. 53, 6–17  |
| 903 | 163 | Payen, V.L. et al. (2021) Single-cell RNA sequencing of human liver reveals hepatic        |
| 904 |     | stellate cell heterogeneity. JHEP Reports 3, 100278  |
| 905 | 164 | Charles, R. et al. (2013) Human hepatic stellate cells inhibit t-cell response through B7- |
| 906 |     | H1 pathway. Transplantation 96, 17–24  |
| 907 | 165 | Ichikawa, S. et al. (2011) Hepatic Stellate Cells Function as Regulatory Bystanders. J.    |
| 908 |     | Immunol. 186, 5549–5555  |
| 909 | 166 | Dunham, R.M. et al. (2013) Hepatic Stellate Cells Preferentially Induce Foxp3 +            |
| 910 |     | Regulatory T Cells by Production of Retinoic Acid. J. Immunol. 190, 2009–2016              |
| 911 | 167 | Mucida, D. et al. (2007) Reciprocal TH17 and regulatory T cell differentiation mediated    |
| 912 |     | by retinoic acid. Science 317, 256–260   |
| 913 | 168 | Broomé, U. et al. (1993) Induced expression of heat-shock protein on biliary               |
| 914 |     | epithelium in patients with primary sclerosing cholangitis and primary biliary cirrhosis.  |
| 915 |     | Hepatology 18, 298–303   |
| 916 | 169 | Ayres, R.C.S. et al. (1993) Intercellular adhesion molecule- i and MHC antigens on         |
| 917 |     | human intrahepatic bile duct cells: Effect of proinflammatory cytokines. Gut 34, 1245–     |
| 918 |     | 1249   |
| 919 | 170 | Leon, M.P. et al. (1996) Immunogenicity of biliary epithelium: Investigation of antigen    |
| 920 |     | presentation to CD4+ T cells. <i>Hepatology</i> 24, 561–567                                |
| 921 | 171 | Auth, M.K.H. et al. (1993) Establishment and immunological characterization of             |
| 922 |     | cultured human gallbladder epithelial cells. Hepatology 18, 546–555                        |
| 923 | 172 | Schrumpf, E. et al. (2015) The biliary epithelium presents antigens to and activates       |

| 924 |     | natural killer T cells. <i>Hepatology</i> 62, 1249–1259                                    |
|-----|-----|--|
| 925 | 173 | Lee, G. et al. (2020) Distinct signatures of gut microbiome and metabolites associated     |
| 926 |     | with significant fibrosis in non-obese NAFLD. Nat. Commun. 11, 4982                        |
| 927 | 174 | Sharpton, S. et al. (2021) Current Concepts, Opportunities, and Challenges of Gut          |
| 928 |     | Microbiome-Based Personalized Medicine in Nonalcoholic Fatty Liver Disease. Cell           |
| 929 |     | Metab. 33, 21–32   |
| 930 | 175 | Mouries, J. et al. (2019) Microbiota-driven gut vascular barrier disruption is a           |
| 931 |     | prerequisite for non-alcoholic steatohepatitis development. J. Hepatol. 71, 1216–1228      |
| 932 | 176 | Rahman, K. et al. (2016) Loss of Junctional Adhesion Molecule A Promotes Severe            |
| 933 |     | Steatohepatitis in Mice on a Diet High in Saturated Fat, Fructose, and Cholesterol.        |
| 934 |     | Gastroenterology 151, 733-746.e12  |
| 935 | 177 | Sharpton, S.R. et al. (2019) Gut Microbial Metabolism and Nonalcoholic Fatty Liver         |
| 936 |     | Disease. Hepatol. Commun. 3, 29–43   |
| 937 | 178 | Huh, J.R. et al. (2011) Digoxin and its derivatives suppress T H17 cell differentiation by |
| 938 |     | antagonizing RORγ3t activity. Nature 472, 486–490  |
| 939 | 179 | Paik, D. et al. (2022) Human gut bacteria produce TH17-modulating bile acid                |
| 940 |     | metabolites. <i>Nature</i>   |
| 941 | 180 | Puchalska, P. et al. (2018) Isotope Tracing Untargeted Metabolomics Reveals                |
| 942 |     | Macrophage Polarization-State-Specific Metabolic Coordination across Intracellular         |
| 943 |     | Compartments. iScience 9, 298–313  |
| 944 | 181 | Glass, O. et al. (2018) Serum Interleukin-8, Osteopontin, and Monocyte                     |
| 945 |     | Chemoattractant Protein 1 Are Associated With Hepatic Fibrosis in Patients With            |
| 946 |     | Nonalcoholic Fatty Liver Disease. <i>Hepatol. Commun.</i> 2, 1344–1355                     |
| 947 | 182 | Honda, M. et al. (2020) Neutralizing antibody against osteopontin attenuates non-          |

948 alcoholic steatohepatitis in mice. J. Cell Commun. Signal. 14, 223–232 949 183 McFarland, A.P. et al. (2021) Multi-tissue single-cell analysis deconstructs the complex 950 programs of mouse natural killer and type 1 innate lymphoid cells in tissues and 951 circulation. *Immunity* 54, 1320-1337.e4 952 Mikulak, J. et al. (2019) Hepatic Natural Killer Cells: Organ-Specific Sentinels of Liver 184 953 Immune Homeostasis and Physiopathology. Front. Immunol. 10, 946 954 185 Vivier, E. et al. (2011) Innate or Adaptive Immunity? The Example of Natural Killer 955 Cells. Science 331, 44 956 Bai, L. et al. (2021) Liver type 1 innate lymphoid cells develop locally via an interferon-186 957 y-dependent loop. Science (80-.). 371, 958 187 Di Censo, C. et al. (2021) Granzyme A and CD160 expression delineates ILC1 with 959 graded functions in the mouse liver. Eur. J. Immunol. 51, 2568–2575 960 Tang, L. et al. (2016) Differential phenotypic and functional properties of liver-resident 188 961 NK cells and mucosal ILC1s. J. Autoimmun. 67, 29-35 962 189 Zhou, J. et al. (2019) Liver-Resident NK Cells Control Antiviral Activity of Hepatic T Cells 963 via the PD-1-PD-L1 Axis. *Immunity* 50, 403-417.e4 964 190 Martínez-Chantar, M.L. et al. (2021) Revisiting the Role of Natural Killer Cells in Non-965 Alcoholic Fatty Liver Disease. Front. Immunol. 12, 640869 966 191 Diedrich, T. et al. (2020) Characterization of the immune cell landscape of patients 967 with NAFLD. PLoS One 15, e0230307 968 192 Stiglund, N. et al. (2019) Retained NK cell phenotype and functionality in non-alcoholic 969 fatty liver disease. Front. Immunol. 10, 1255 970 193 Fan, Y. et al. (2020) Hepatic NK cells attenuate fibrosis progression of non-alcoholic 971 steatohepatitis in dependent of CXCL10-mediated recruitment. Liver Int. 40, 598-608

| 972 | 194 | Cuff, A.O. et al. (2019) The Obese Liver Environment Mediates Conversion of NK Cells         |
|-----|-----|--|
| 973 |     | to a Less Cytotoxic ILC1-Like Phenotype. Front. Immunol. 10, 2180                            |
| 974 | 195 | Tosello-Trampont, A.C. et al. (2016) NKp46+ natural killer cells attenuate metabolism-       |
| 975 |     | induced hepatic fibrosis by regulating macrophage activation in mice. Hepatology 63,         |
| 976 |     | 799–812  |
| 977 | 196 | Luci, C. et al. (2019) Natural Killer Cells and Type 1 Innate Lymphoid Cells Are New         |
| 978 |     | Actors in Non-alcoholic Fatty Liver Disease. Front. Immunol. 10, 1192                        |
| 979 | 197 | Gomez-Santos, L. et al. (2012) Inhibition of natural killer cells protects the liver against |
| 980 |     | acute injury in the absence of glycine N-methyltransferase. <i>Hepatology</i> 56, 747–759    |
| 981 | 198 | Gur, C. et al. (2012) NKp46-mediated killing of human and mouse hepatic stellate cells       |
| 982 |     | attenuates liver fibrosis. Gut 61, 885–893   |
| 983 |     |  |

### 984 Text boxes

## Box 1 – Hepatocytes exert an immune function at homeostasis

While most antigens in the liver are taken up and processed by non-parenchymal APCs, parenchymal cells also possess antigen presentation capacities. Hepatocytes are lined by LSECs, which provide an hepatic barrier to protect the hepatocytes against portal-delivered gut-derived pathogens. However, hepatocytes are accessible for naïve CD8<sup>+</sup> T lymphocytes *via* T cell cytoplasmic extensions penetrating the endothelial fenestrations that perforate the LSECs [145,146]. This is of importance for immune surveillance and, for example, for the detection of Hepatitis B Virus-infected hepatocytes by virus-specific CD8<sup>+</sup> T cells [146]. Hepatocytes constitutively express MHC-I molecules allowing them to cross-present antigens and to prime naïve CD8<sup>+</sup> T cells directly [147], or *via* LSEC-mediated cross-presentation [148]. In a healthy liver, hepatocyte activation of CD8<sup>+</sup> T cells generally leads to an abortive immune response and early T cell apoptosis [149], maintaining liver tolerance.

# Box 2 – LSEC, cholangiocytes, and HSC act as antigen presenting cells at homeostasis

Liver sinusoidal endothelial cells (LSECs) are, contrary to hepatocytes, in direct contact with the blood. Their numerous fenestrae and absence of basal membrane allow efficient filtration from the sinusoidal lumen to the space of Disse [150]. LSECs play an important role in the clearance of circulating microorganisms [151,152] and molecules [153], as well as in mediating immune tolerance [154] under physiological conditions. Antigens that are taken up and processed by LSECs can be presented *via* MHC-II to CD4<sup>+</sup> T cells, and *via* MHC-I to naïve CD8<sup>+</sup> T cells. Priming of naïve CD4<sup>+</sup> T cells mainly induces the development of CD4<sup>+</sup>CD25<sup>+</sup>Foxp3<sup>+</sup> Tregs [155], which is an important mechanism for maintaining hepatic tolerance. Antigen presentation by LSECs to CD8<sup>+</sup> T cells results – at low antigen

1008 concentrations – in the downregulation of IFNγ and IL-2 cytokine production by CD8<sup>+</sup> T cells, 1009 leading to antigen-specific T cell tolerance [156,157]. 1010 Hepatic stellate cells (HSCs) constitute 8-10% of total liver cells. HSCs are located at the 1011 space of Disse, between the hepatocytes and the LSECs. In a healthy liver, HSCs are 1012 quiescent, serve as the primary storage site for vitamin A, and are essential in the regulation 1013 of retinoic acid homeostasis. Moreover, they are the main collagen- and extracellular matrix 1014 producing cells in liver fibrosis. Recent studies have also identified a role of HSCs in hepatic 1015 immunology. Antigen uptake and presentation by HSCs in mice is low [158], although they 1016 express MHC-II, costimulatory molecules such as CD40 and CD80 [159,160], and they 1017 produce various growth factors, cytokines [161] and chemokines [162]. A recent study using 1018 single-cell RNA sequencing identified two HSC subpopulations in human liver with one of 1019 them displaying a gene signature that is reminiscent of antigen-presenting cells [163]. 1020 Although it has been suggested that HSCs can present antigens to T cells [159,164], this 1021 function is still controversial. However, HSCs do contribute to hepatic immune tolerance via 1022 indirect priming of DC-activated Tregs [165,166]. Treg induction and Th17 inhibition is 1023 mediated through the release of retinoic acid [167] during priming of T cells by DCs [165]. At 1024 homeostasis HSCs thus regulate hepatic immune reactivity mainly indirectly by modulating 1025 the outcome of T cell activation by other APCs. 1026 Cholangiocytes, or biliary epithelial cells (BECs), line the bile duct and are constantly exposed 1027 to bile content. Apart from their function in the regulation of bile flow, composition, and pH, 1028 they also play a role in antigen presentation. On the apical side, BECs are exposed to bile and 1029 (lipid) antigens present in bile. On the basolateral side, BECs are in close contact with hepatic 1030 immune cells. Disorders associated with immune damage of the bile ducts, such as primary biliary cholangitis (PBC) [168,169], are associated with increased basolateral MHC-II 1031

expression by BECs, although *in vitro* assays failed to demonstrate CD4<sup>+</sup> T cell-activation [170]. BECs also constitutively express MHC-I molecules [171] and are capable of presenting antigens *via* MR1 [131] and CD1d [172] to unconventional T cells such as MAIT and NKT cells. NKT cells react to lipid antigens presented by CD1d expressed on the basolateral side, and *in vitro* studies have shown that murine and human cholangiocyte cell lines are able to activate NKT cells upon exposure to lipids and endogenous antigens [172]. Interestingly, CD1d expression was reported to be a characteristic of healthy BECs, being downregulated in PBC and alcoholic cirrhosis. This suggests that CD1d-dependent antigen presentation and the potential ensuing activation of NKT cells is important to maintain immune tolerance in a healthy liver. However, more studies are needed to fully investigate the role of BECs in hepatic immune homeostasis.

## Box 3 – Microbial metabolites trigger the immune response

The gut microbiota is increasingly recognized to contribute to NAFLD development and progression towards fibrosis [173,174]. Diet-induced microbial dysbiosis damages the gut-vasculature barrier function [175]. Bacterial translocation into the blood and subsequently in the liver leads to hepatic inflammation and NASH development [175,176]. Moreover, microbial-derived compounds, such as short-chain fatty acids, branched-chain amino acids, and secondary bile acids, affect the pathogenesis of NAFLD [177]. Interestingly, three recent studies show that (microbial) metabolites of bile acids, which are produced from hepatic cholesterol, control the adaptive immune response in the gut [34–36] (Figure 2). A bile acid metabolite library screen for T cell modulatory effects identified two derivatives of lithocholic acid (LCA) which affect Th17 and Treg differentiation [35]. 3-oxoLCA, on the one hand, inhibits Th17 differentiation by physically interacting with the transcription factor

retinoic acid receptor-related orphan receptor yt (RORyt). RORyt is a key transcription factor required for induction of IL-17 transcription [178]. 3-oxoLCA inhibits its transcriptional activity and thus Th17 differentiation. Conversely, isoalloLCA promotes Treg differentiation by enhancing mitochondrial ROS production and increasing histone acetylation at the FoxP3 promoter region. Overall, the LCA metabolites 3-oxoLCA and isoalloLCA exert antiinflammatory actions by directly affecting CD4<sup>+</sup> T cell differentiation [179]. Similarly, Song et al. reported that bile acids induce (immunosuppressive) RORyt-expressing Tregs residing in the lamina propria of the colon [36]. The authors speculate that the most abundant intestinal primary bile acid species, such as cholic acid and chenodeoxycholic acid, act via the vitamin D receptor (VDR) to maintain a healthy colonic RORyt<sup>+</sup> Treg pool (Figure 2). Recently, the secondary bile acid isoDCA was found to indirectly induce peripheral Tregs in the colon by inhibiting FXR activity in DCs [34]. Collectively, these studies identify an immune suppressive role for bile acid metabolites in the control of the adaptive immune response in the colon. Although these studies are not performed in a NASH context and do not directly relate to the hepatic immune system, they underline the important immune regulatory function of bile acids (and metabolites). NASH is associated with changes in bile acid metabolism and pool composition and altered plasma bile acid levels and profiles [37]. However, so far studies about the effect of altered bile acid profiles or specific bile acid species on hepatic immunity and NASH development are lacking.

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### Box 4 - Macrophages in NASH: beyond the M1/M2 paradigm

As for macrophages in other tissues, in the past, KCs were subdivided into M1 (classically activated, immunogenic) and M2 (alternatively activated, tolerogenic) cells. However, it is increasingly recognized that KCs exhibit a high plasticity, are highly dynamic and can

differentiate into a wide range of polarization or activation states. Depending on the context, KCs exhibit specific metabolic signatures [180] and M1 and M2 KCs only represent the extreme phenotypes of a wide spectrum. Upon Western diet feeding, 4 abundant macrophage populations were identified in the liver, including resident KCs and 3 subsets of recruited macrophages: monocyte-derived KCs, macrophages becoming monocyte-derived KCs (termed pre-moKCs), and hepatic lipidassociated macrophages (LAMs). In agreement with other studies [15], resident KCs were gradually lost during NAFLD progression and replaced by monocyte-derived KCs. Interestingly, monocyte-derived KCs resembled resident KCs, while the transcriptome of hepatic LAMs largely differed from that of resident KCs. LAMs expressed lower levels of immune activation-associated genes, such as II18, Fpr2, Tlr4 and CD38, and had the distinct ability to metabolize lipids. LAMs were only identified in later stages of NAFLD and express Spp1 (next to Trem2, Cd9, Cd63, and Gpnmb), encoding the chemokine osteopontin. Osteopontin was found to be a good biomarker of NASH [181] and is thought to contribute to fibrosis [182]. Moreover, Daemen et al. showed that LAMs preferentially localize to HSC expansion regions, where they aggregate into hepatic crown-like structures (hCLS) [15], suggesting a role for hepatic LAMs in NASH development and liver fibrosis. Another recent study identified low numbers of LAMs already in periportal regions of healthy livers, and showed accumulation of more mature LAMs in pericentral regions in steatotic mouse livers [42]. The exact function of LAMs in the NASH liver is as yet unclear. A more detailed sc-RNAseq analysis based on transcriptional differences identified 5 macrophage clusters in mice fed control or NASH diets [16]: healthy KCs, NASH KCs, recruited macrophages, LY6Chi recruited macrophages and LY6Clo recruited macrophages (Figure 3). NASH KCs and recruited macrophages were predominantly found in NASH livers,

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while healthy livers consisted almost entirely of healthy KCs. Both LY6C<sup>hi&lo</sup> recruited macrophages were also predominantly found in NASH livers and differed in gene expression and zonation from recruited macrophages. While recruited macrophages were mainly located in the liver sinusoid, in contact with the LSECs, LY6C<sup>hi&lo</sup> recruited macrophages reside proximal to the large portal and central vein vessels in contact with vascular endothelium. These observations underline that disease-promoting metabolic and tissue environmental signals can instruct macrophages to acquire distinct gene expression programs. Mechanistically, MCD diet-induced NASH was associated with impaired KC self-renewal and disease progression with increased recruitment of monocyte-derived KCs [66]. Moreover, death of NASH KCs, possibly induced by increased ER stress, modifies the KC niche and enables repopulation by monocyte-derived KCs. Monocyte-derived KCs are enriched in genes involved in cellular stress and activation, limited hepatic triglyceride storage and increased inflammation.

## Box 5 – Hepatic innate lymphoid cells in NAFLD: a controversial role

Liver innate lymphoid cells mostly include natural killer (NK) cells and closely-related group 1 lymphoid cells (ILC1s). They both lack antigen-specific receptors and produce the signature cytokine IFN $\gamma$  [114,183] in response to IL-12, IL-15, or IL-18 secreted by DCs, macrophages and T lymphocytes [184,185]. Mouse hepatic NK, unlike ILC1, require the transcription factor EOMES for their development, while ILC1 express the transcription factor Zpf683. Moreover, the development of hepatic ILC1s from its liver-resident progenitor cells is promoted by IFN $\gamma$  production by ILC1s themselves [186].

In the metabolically healthy liver, NK cells rapidly recognize and kill viral-infected or tumor-transformed cells in the absence of specific immunization by releasing perforin and

granzymes [185]. Hepatic ILC1s exhibit cytotoxicity mediated through granzyme A [187] and B, perforin, TRAIL, and FasL [188]. By expressing inhibitory receptors, such as PD-1 (programmed cell death-1) and lymphocyte-activation protein-3 (LAG-3), ILC1s regulate the anti-viral activity of hepatic T cells [189], thereby preventing hepatic virus infection.

Conflicting data exist on changes in ILC1 and NK cell numbers and proportions in blood and livers of NAFL/NASH patients [190–192] and in livers of mouse NAFLD models [193–196], probably due to differences in disease stage and the use of different NASH-inducing protocols. On the one hand, NK cells have a detrimental role in NAFL to NASH progression as TRAIL-producing NK cells were shown to promote a pro-inflammatory environment at an early NAFLD stage [197]. On the other hand, NK cells were reported to exert anti-fibrotic effects [193,195] and to actively contribute to limiting NASH-associated tissue damage and fibrosis by killing activated HSCs, the main collagen producing cells in fibrosis [198]. A better characterization of (tissue resident) NK cells and the seemingly controversial role of NK cells as contributors or modulators of NASH progression should be further investigated.

1142 Figure legends

Figure 1. Hepatic immunity in healthy liver.

At homeostasis the liver exhibits an immune tolerant state, while it also allows efficient responses against infectious agents. LSECs prime naïve CD4<sup>+</sup> T cells and induce Treg differentiation. KCs are important to maintain immune homeostasis by clearing common gut-derived antigens. CD8<sup>+</sup> effector T cells scan and recognize MHC-I/peptide complexes presented by hepatocytes to respond to intracellular pathogens and damaged cells.

BEC; biliary epithelial cells, MHC-I/II; major histocompatibility complex, class I/II, TCR; T cell receptor, HSC; hepatic stellate cell, KC; Kupffer cell, neutro; neutrophils, Treg; regulatory T cell, CD4; CD4<sup>+</sup> T cell, CD8; CD8<sup>+</sup> T cell, NKT; natural killer T cell, pDC; plasmacytoid dendritic cell, cDC; conventional dendritic cell.

peripheral Treg.

Figure 2. The immune suppressive role of bile acids and their metabolites affect T cell homeostasis in the colonic lamina propria and control the adaptive immune response.

The most abundant primary bile acids (CA and CDCA) maintain a healthy colonic RORyt Treg pool *via* VDR signaling. 3-oxo-LCA, which is formed by the gut microbiota, inhibits Th17 differentiation by physical interaction with RORyt. IsoalloLCA promotes Treg differentiation by enhancing mitoROS and FoxP3 acetylation. isoDCA indirectly induces pTreg generation by inhibiting FXR transcriptional activity in DCs.

CA; cholic acid, CDCA; chenodeoxycholic acid, 3-oxo-LCA; 3-oxo-litocholic acid, isoalloLCA; isoallolitocholic acid, isoDCA; isodeoxycholic acid, mitoROS; mitochondrial reactive oxygen species, VDR; vitamin D receptor, RORγt; retinoid-related orphan receptor γt, FoxP3; forkhead box P3, FXR; farnesoid X receptor, Treg; regulatory T cell, Th17; T helper 17, pTreg;

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1167 Figure 3. Kupffer cell populations in healthy and NASH livers, according to Seidman et al. [16]

and Bonnardel et al. [43]

KC-H cells are enriched in healthy liver. KC-H loss upon NASH-induced liver injury, most

probably due to apoptosis, induces TNFα and IL-1-dependent receptor activation of HSCs

and LSECs. VCAM, SEL-E, and CCL2 upregulation results in monocyte recruitment. LSEC DLL4

expression drives monocyte differentiation and supports niche specialization via the DLL4-

Notch pathway. KC-hepatocyte interactions induce ID3 expression, LSECs and HSCs induce

LXRα expression. 4 macrophage populations are enriched in NASH liver: KC-N, KN-RM (both

localized in liver sinusoid), Ly6Chi-RM, and Ly6Clo-RM (both localized around portal and

1176 central vein vasculature).

HSC; hepatic stellate cell, TNF $\alpha$ ; tumor necrosis factor  $\alpha$ , IL-1; interleukin-1, VCAM; vascular

cell adhesion molecule, SEL-E; selectin-E, DLL4; delta like canonical notch ligand 4, LXRα; liver

X receptor alpha, ID3; inhibitor of differentiation 3, KC-H; healthy Kupffer cells, KC-N; NASH-

Kupffer cells, KN-RM; recruited macrophages, Ly6Chi-RM; Ly6Chi recruited macrophages,

Ly6C<sup>lo</sup>-RM; Ly6C<sup>lo</sup> recruited macrophages.

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1183 Figure 4. Cytotoxic CD8<sup>+</sup> T cells in NASH.

(a) IL-15-driven transcriptional (re)programming and metabolic stimuli promote auto-

aggressive CXCR6<sup>+</sup> CD8+ T cells displaying enhanced expression of PD1 and GZMB. Auto-

aggression is MHC-I-independent, as the purinergic receptor P2X7 responds to ATP released

from dying hepatocytes. Calcium influx is enhanced to initiate auto-aggression. (b) GZMA

and perforin expressing CD8<sup>+</sup> T cells associate with NASH. The link between the altered

cDC1/cDC2 ratio and antigen dependent CD8<sup>+</sup> T cell activation remains to be investigated.

GZMA; granzyme A, GZMB; granzyme B, PD1; programmed cell death protein 1, CXCR6; C-X-C motif chemokine receptor 6, IL-15; interleukin 15, FOXO1; forkhead box O1, P2X7; purinergic receptor P2X 7, MHC-I; major histocompatibility complex, class I, TCR; T cell receptor, cDC1; conventional dendritic cells 1, cDC2; conventional dendritic cells 2.

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Figure 5. Immune dysregulation in NASH liver.

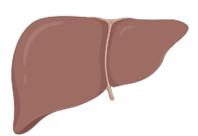
Macrophages are recruited via the CCL2/CCR2 axis or by other (environmental) signals and replace resident KCs. DC abundance increases in NASH livers and cDC1 cells enhance the inflammatory signature of CD8<sup>+</sup> T cells. The cDC1/cDC2 ratio decreases, but its effect on CD4<sup>+</sup> and CD8<sup>+</sup> T cell activation and the function of cDC2 in NASH is unknown. B cells secrete proinflammatory cytokines, such as TNFa and IL-6, as well as IgA, although the exact role of B cells in NASH remains to be investigated. CD8<sup>+</sup> T cells in a NASH environment acquire the ability to kill hepatocytes, and to induce HSC activation, thereby enhancing the development of fibrosis. The synergistic action of CD8<sup>+</sup> T cells with NKT cells might lead to NASH progression. LIGHT secretion by NKT cells enhances hepatocyte lipid accumulation, thus promoting hepatic steatosis, while the secretion of proinflammatory cytokines, such as IFNy, may recruit and activate other immune cell types, enhancing immune dysregulation and NASH progression. NASH-related metabolites, such as excess glucose and fatty acids and increase in plasma bile acid levels also affects phenotype and function of the hepatic immune compartment. HSC; hepatic stellate cells, CCL2; C-C motif chemokine ligand 2, CCR2; C-C motif chemokine receptor 2, ATP; adenosinetriphosphate, NKT; natural killer T cells, IFNγ; interferon γ, IL; interleukin, MPO; myeloperoxidase, NETosis; NET (neutrophil extracellular traps) formation, CXCR6; C-X-C motif chemokine receptor 6, cDC1; classical type 1 DCs, cDC2; classical type 2 DCs, DCs; dendritic cells, moKC; monocyte-derived Kupffer cells, Th1; T helper 1, Th17; T helper 17, IgA; immunoglobuline A, TNFα; tumor necrosis factor α, ER stress; endoplasmic reticulum stress, KC; Kupffer cells, FA; fatty acids, chol; cholesterol, BA; bile acids.

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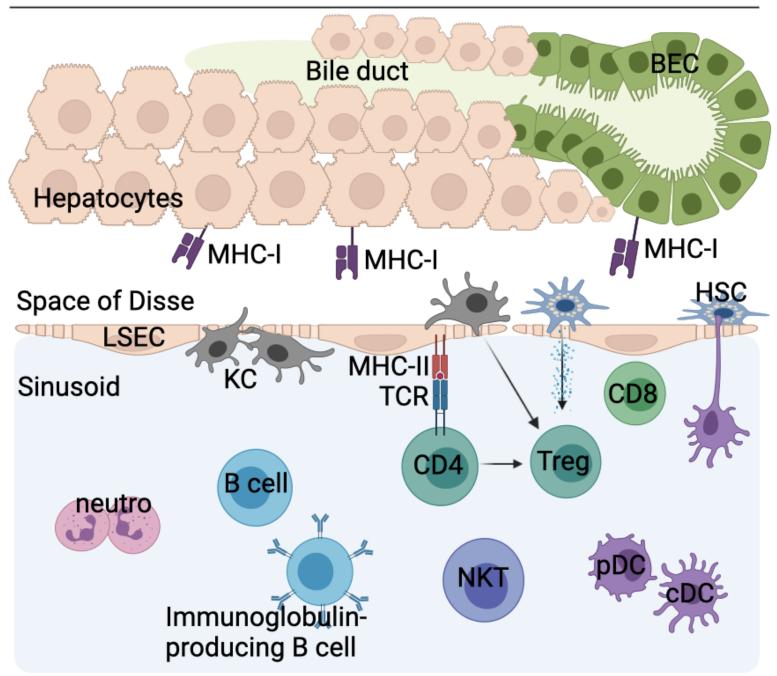
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Figures are created with BioRender.com

Figure 1



- Immune tolerant state
- Efficient response against infectious agents
- T cell tolerance induction
- Regulatory T cell development



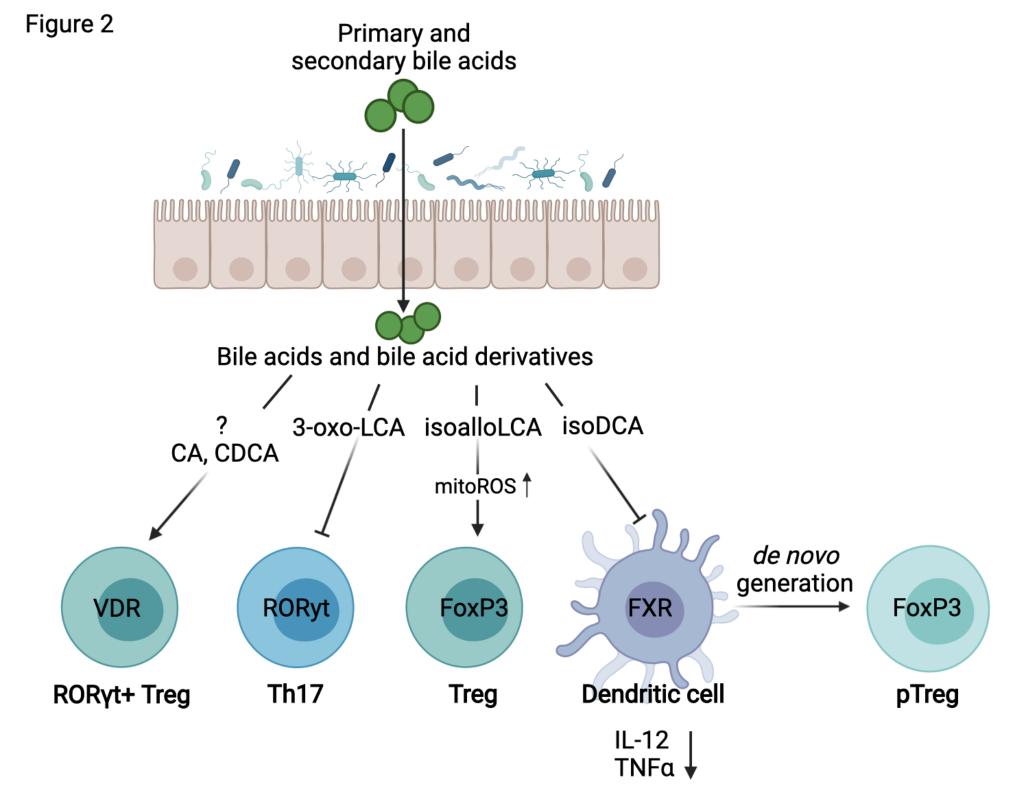
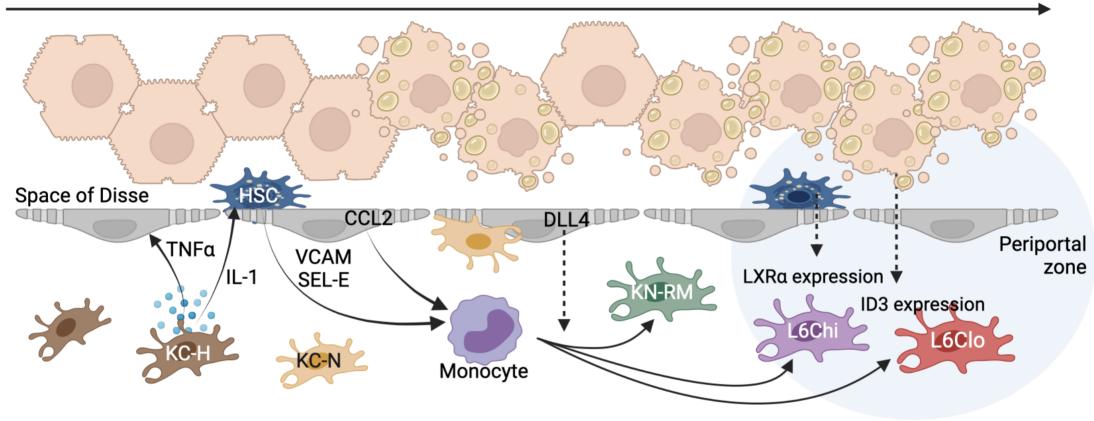


Figure 3

# Homeostasis



|                 | 302                                 | 366                                       | 200  | 3  | 300  |
|-----------------|-------------------------------------|---|--|--|--|
| Cell population | KC-H (healthy Kupffer cells)        | KC-N (NASH-Kupffer cells)                 | KN-RM (recruited macrophage occupying the KC-N nice) | Ly6Chi-RM (Ly6Chi<br>recruited macrophage) | Ly6C <sup>lo</sup> -RM (Ly6C <sup>lo</sup> recruited macrophage) |
| Enriched in     | Healthy liver                       | NASH liver                                | NASH liver   | NASH liver                                 | NASH liver   |
| Localization    | Liver sinusoid                      | Liver sinusoid                            | Liver sinusoid                                       | Around portal and central vein vasculature | Around portal and central vein vasculature                       |
| Identity genes  | F4/80, Maf, Cd163,<br>Clec4f, Timd4 | F4/80, Maf, Clec4f,<br>Cd5l, Fabp7, Ccl24 | F4/80, Maf, Clec4f<br>Cd63, Cdh5                     | Ly6c2, Ccr2, Itgam                         | Cd209a, Cd7, Itgam   |

Figure 4

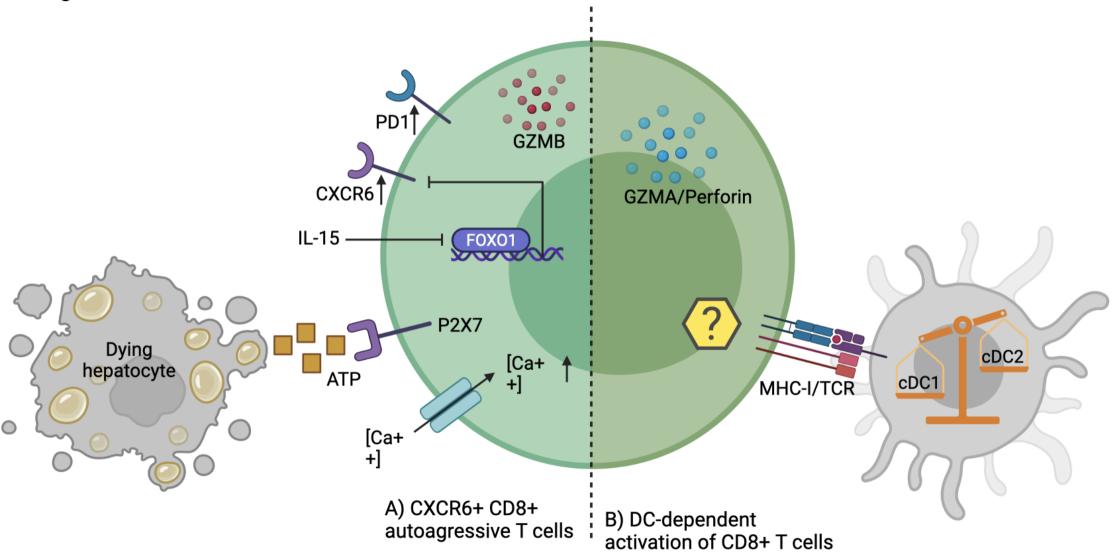
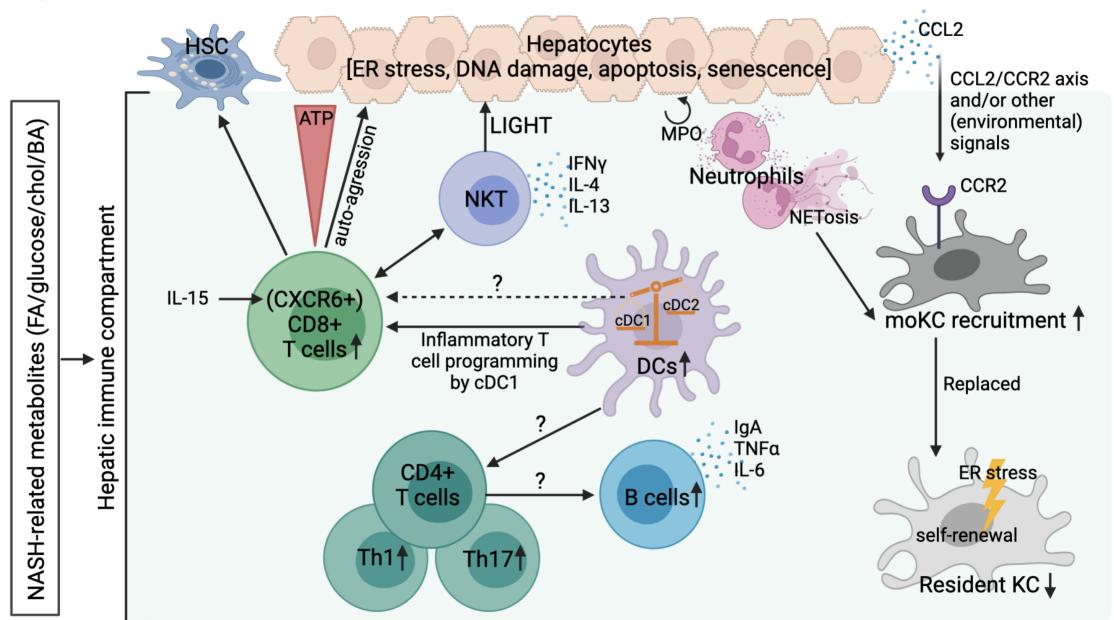


Figure 5



1 Highlights

- The altered metabolic environment in non-alcoholic fatty liver disease (NAFLD) promotes
- 4 hepatocyte metabolic dysfunction and cellular stress, affecting the highly diverse hepatic
- 5 immune compartment.
- Resident macrophages are activated in the early phase of liver injury and disease-
- 7 promoting metabolic and tissue environmental signals alter their gene expression
- 8 programs resulting in an identity loss.
- In a NASH environment, cDC and CD8<sup>+</sup> T cell populations are altered. CD8 cells with an
- 10 "exhausted phenotype" are activated and kill hepatocytes, hence promoting tissue
- 11 damage.
- The phenotype and function of hepatic immune cells is reshaped during the progression
- of NAFL to non-alcoholic steatohepatitis (NASH), but the role of the cellular interactions
- in disease progression is only starting to be uncovered.

1 Outstanding questions

- 3 1. How does the dysregulation of the macrophage compartment impact NASH
- 4 development?
- 5 2. Does an altered cDC1/cDC2 ratio rather than absolute numbers play a role in NASH?
- 6 3. Is antigen-dependent activation of CD8 T lymphocytes playing a role in NASH?
- 7 4. Do various B cell subsets play a differential role in NASH development?