

# Hepatic sexual dimorphism -implications for non-alcoholic fatty liver disease

Philippe Lefebvre, Bart Staels

## ▶ To cite this version:

Philippe Lefebvre, Bart Staels. Hepatic sexual dimorphism -implications for non-alcoholic fatty liver disease. Nature Reviews Endocrinology, 2021, 17 (11), pp.662-670. 10.1038/s41574-021-00538-6. inserm-03381913

## HAL Id: inserm-03381913 https://inserm.hal.science/inserm-03381913

Submitted on 18 Oct 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Hepatic sexual dimorphism — implications for non-alcoholic fatty liver disease

Philippe Lefebvre<sup>1,†</sup> and Bart Staels<sup>1,</sup>

<sup>1</sup>Univ. Lille, INSERM, CHU Lille, Institut Pasteur de Lille, U1011 - EGID, F-59000 Lille, France.

†philippe-claude.lefebvre@inserm.fr

#### **Abstract**

The liver is often thought of as a single functional unit, but both its structural and functional architecture make it highly multivalent and adaptable. In any given physiological situation, the liver can maintain metabolic homeostasis, conduct appropriate inflammatory responses, carry out endobiotic and xenobiotic transformation and synthesis reactions, as well as store and release multiple bioactive molecules. Moreover, the liver is a very resilient organ. This resilience means that chronic liver diseases can go unnoticed for decades, yet culminate in life-threatening clinical complications once the liver is no longer able to compensate for them. Non-alcoholic fatty liver disease (NAFLD) predisposes individuals to cirrhosis and increases liver-related and cardiovascular disease-related mortality. This Review discusses the accumulating evidence of sexual dimorphism in NAFLD, which is currently rarely considered in preclinical and clinical studies. Increased awareness of hepatic sexual dimorphism could lead to improved understanding of the biological processes that are dysregulated in NAFLD, to the identification of relevant therapeutic targets and to improved risk stratification of patients with NAFLD undergoing therapeutic intervention.

#### [H1] Introduction

The search for treatments for chronic metabolic diseases and other human ailments has often highlighted the difficulty of conducting drug trials in fully representative populations, as patient selection and randomization often introduces bias <sup>1</sup>. The ability to define the factors that drive a patient's response to treatment is the key concept of precision medicine, which aims to treat only sensitive subpopulations. Surprisingly, essential biological differences between male and female individuals are rarely considered during preclinical<sup>2</sup> and clinical<sup>3</sup> studies. This lack of consideration is a cause for concern because sex is a strong determinant of disease prevalence and outcomes in many fields, including cardiology<sup>4</sup>, diabetology<sup>5</sup>, oncology<sup>6</sup>, pulmonology<sup>7</sup> and, of specific interest to this Review, hepatology. Nevertheless, the vast majority of therapeutic guidelines are optimized for a standard 70 kg (white) male patient. The consequences of overlooking sex as a biological variable (reviewed in depth elsewhere<sup>8</sup>) are often deleterious for both sexes and can result in inadequate initiation and choice of treatments. Given the existence of physiological sexual dimorphism in many metabolic processes, it is not surprising that pathogenic events can affect metabolically active organs

differently in male and female individuals. As such, the liver responds differently in men and women during the progression of chronic liver diseases, including non-alcoholic fatty liver disease (NAFLD)<sup>9</sup>.

First described in the early 1980s, NAFLD is a condition defined by histological abnormalities in the liver similar to those encountered in patients with alcoholism<sup>10</sup>. The pathogenesis of NAFLD is driven by metabolic dysfunction in the context of a fatty liver and thus this disease has a high prevalence in obese and diabetic patients [. When fat accumulation becomes chronic, hepatic steatosis (also termed fatty liver or non-alcoholic fatty liver (NAFL)) progresses to non-alcoholic steatohepatitis (NASH) and fibrosis<sup>11</sup>. NASH was recognized as an important liver disease in the late 1990s<sup>12</sup> and the pathogenic events leading to NASH and end-stage liver fibrosis (cirrhosis) were subsequently shown to result from a 'multi-hit' mechanism in which steatosis paves the way for additional hepatic insults, such as inflammation (accompanied by the release of proinflammatory cytokines), oxidative stress (associated with increased levels of reactive oxygen species) and endoplasmic reticulum stress, which in turn propagate inflammatory and fibrogenic responses within the liver<sup>13</sup>. This initially simple model has evolved considerably in the past decade as a result of the recognition that factors originating in other tissues, such as adipokines, immune cells and metabolites produced by gut microbiota, make important contributions to the pathogenesis of NASH. These same factors are also implicated in the progression of NAFLD towards more severe stages <sup>14,15</sup> and act in concert with genetic markers of susceptibility to NAFLD<sup>16</sup>.

The global prevalence of NAFLD has now reached epidemic proportions; this condition is estimated to affect 15–31% of the total population when detected by indirect (non-histological) methods. The highest rates of NAFLD are reported in South America (30%) and the Middle East (32%), followed by Asia (27%), the USA (24%), Europe (23%) and Africa (14%) with ethnic disparities<sup>17</sup>. North Americans of Hispanic descent are more prone to NALFD (2 to 3-fold) when compared to the overall population. Moreover, ~20% of individuals with NAFLD probably also have features of NASH <sup>18</sup>. Although NASH is also detected in lean patients, in whom the condition shows a strong ethnic bias to vary between 10-15% in Asia, 15% in Europe and 37% in USA. While still elusive, reasons for these differences probably reflect genetic, social and dietary influences <sup>19,20</sup>. The prevalence of NASH is 2 to 3 fold higher in obese individuals than in lean ones<sup>21</sup> and in patients with type 2 diabetes mellitus (T2DM) than in the background population<sup>22,23</sup>. These two groups accordingly account for a high proportion of the patients with advanced fibrosis. Patients with fibrosis are more likely to develop liver-related adverse outcomes such as hepatocellular carcinoma (HCC) as well as liver-unrelated comorbidities such as cardiovascular disease (CVD)<sup>24,25</sup>. Thus, NASH accounts for an ever-increasing burden on health-care systems.

A wealth of studies of the various dysfunctional organs and cell types that contribute to progression of NAFLD has led to the identification of multiple signalling and metabolic pathways that

promote liver dysfunction and fibrosis, among which insulin resistance is a major initiating pathological event that drives de novo lipogenesis in the liver<sup>26-28</sup>. Despite tremendous efforts to develop drugs that interfere with these dysregulated metabolic, inflammatory and other biological processes, so far these therapeutic approaches have not been successful in either preventing or halting the progression toward NASH and fibrosis<sup>29</sup>. This failure is probably a reflection of the multifactorial and heterogeneous nature of this disease, as well as the lack of tools for its accurate diagnosis and for quantitative disease staging<sup>30,31</sup>. In addition, sex is rarely considered as a biological variable in these numerous and interesting studies, despite the reported sexual dimorphism of the gut microbiota<sup>32</sup>, the innate immune system<sup>33</sup> and the liver itself. As a result, in-depth studies conducted in relevant models are necessary to fully understand the natural history of NAFLD, and to identify sex-dependent determinants of NAFLD progression.

This Review highlights the evidence from preclinical and clinical studies indicating that sex is an important biological variable in hepatic physiology and NAFLD. This aspect becomes highly relevant as studies' accuracy reaches the single cell level, providing unique opportunities to provide a comprehensive atlas of cellular modifications in NAFLD.

#### [H1] Sexual dimorphism in the liver

Although some essential hepatic functions (gluconeogenesis, bilirubin detoxification, iron storage and production of clotting factors) are equally carried out by the liver in male and female individuals, others clearly differ according to sex. Indeed, sex-specific differences in hepatic function have long been observed in sexually mature oviparous animals and mammals<sup>34</sup>. Sex-specific production of hepatic proteins such as vitellogenin,  $\alpha$ 2-microglobulin, bile acids, xenobiotic transporters<sup>35</sup> and cytochrome P450 (CYP) enzymes involved in metabolism of sex steroids<sup>34</sup> has been described in various animal models and in humans. In most cases, sex-specific differences in levels of protein production are reflected in gene expression levels, and sex-biased expression of about 1,000 mRNA transcripts has been described in liver cells from male versus female individuals<sup>36</sup>. These features, which are partly a result of the exquisitely close coupling of metabolism to reproductive needs (a necessity for successful pregnancy in female individuals), orientate the female liver towards a highly efficient metabolic phenotype<sup>37</sup> characterized by increased mitochondrial biogenesis<sup>38</sup>.

In mice, hepatic sexual dimorphism is also observed at the metabolomic level<sup>39</sup> and this functional adaptation has been ascribed to the transcriptional control of gene expression by the pulsatile (in males) or constant (in females) secretion of growth hormone (GH)<sup>40</sup>. The GH–STAT5b pathway regulates CYP enzymes involved in metabolism of xenobiotic molecules and sex steroids<sup>41</sup> in a sex-specific and species-specific manner, such that sexual dimorphism is highly pronounced in rats

and attenuated but still substantial in humans<sup>42,43</sup>. This pathway is an important contributor to liver sexual dimorphism, in addition to the androgen and oestrogen signaling pathways<sup>44,45</sup>.

Over the years, additional regulatory mechanisms have been implicated in the control of sex-specific liver functions, including the circadian clock<sup>46,47</sup>, the bile acid-activated farnesoid X receptor (FXR) pathway<sup>48-50</sup> and the gut microbiome, which exacerbates sexual dimorphism in the mouse liver by controlling the secretion of GH and sex hormones<sup>39,48,51</sup>.

#### [H1] Molecular basis of liver sexual dimorphism

All the pathways that control sex-specific liver functions are deeply interconnected<sup>39,52</sup> but how they cooperate to establish molecular sexual dimorphism in the liver remains to be fully characterized. The mechanism through which GH determines hepatic sexual dimorphism is under intense investigation<sup>53</sup>. For instance, the level of expression of genes that regulate lipid, cholesterol and drug metabolism in the adult liver is determined by the GH secretion pattern<sup>43</sup>, which in rats is established by androgen exposure (which causes persistent epigenetic changes) during the neonatal period<sup>54,55</sup>. Importantly, gut microbiota-derived metabolites are also required to establish the male-specific, pulsatile GH secretion pattern in mice, and germ-free mice display a less pronounced transcriptional sexual dimorphism<sup>39</sup>. In female rats, by contrast, the absence of neonatal androgen exposure means that GH secretion by the pituitary gland is constant, which leads to a desensitized, low activity of the JAK2–STAT5b pathway. Of note, STAT5a participates in GH-mediated signaling to a lesser extent than does STAT5b, as STAT5a is expressed only in minute amounts in the liver<sup>56</sup>. Accordingly, transient phosphorylation and activation of STAT5b is considered to be a masculinization event that drives upregulation of male-specific genes as well as downregulation of female-specific genes<sup>57</sup> (Figure 1). GH dictates chromatin opening in a sex-dependent manner, which is accompanied by deposition of specific histone marks at gene-regulatory regions without altering the pericentral-to periportal zonal expression pattern of sex-specific genes<sup>58</sup>. Therefore, metabolic features of the liver are specified by brain hormones.

Sex hormones (notably  $17\beta$ -oestradiol, referred to herein as oestradiol) also play an important part in the regulation of metabolic genes with sex-biased expression. The results of hormone replacement therapy studies in humans as well as gene deletion experiments in mice concur that oestrogens suppress hepatic de novo lipogenesis, increase fatty acid oxidation, increase production of VLDL triglycerides, decrease the synthesis of cholesterol (via increased clearance of plasma apolipoprotein B particles) and increase the synthesis of cholesterol-derived bile acids<sup>59,60</sup>. Thus, oestradiol has a protective effect in female livers that is also observed [in male livers as a result of aromatase-mediated conversion of testosterone to oestradiol, which safeguards hepatic insulin sensitivity, modifies bile acid synthesis and limits the bile acid pool size<sup>61-64</sup>.

Gene deletion experiments and other studies in mice are mostly supportive of a contribution of nuclear estrogen receptor  $\alpha$  (ESR1) and to a lesser extent a contribution of G-protein coupled oestrogen receptor (GPER) to the protective effects of oestradiol, but neither ESR2 (ER $\beta$ ) nor nuclear androgen receptor (AR) contribute to this mechanism<sup>65-67</sup>. The role of androgens and AR in regulation of hepatic lipid metabolism (and hence their contribution to steatosis) has been studied because of the highly conserved pro-steatotic effect of orchidectomy in many species<sup>65</sup>. Both testosterone and dihydroxytestosterone can act as anti-steatotic agents, but whether aromatization to oestradiol plays a part in this effect is unclear. The findings of gene deletion studies in mice are inconsistent but generally point to a protective role of AR in males and no effect of AR in females<sup>65</sup>. Of note, other transcription factors, such as aryl hydrocarbon receptor (AHR), pregnane X receptor (PXR/NR1I2) and farnesoid X receptor (FXR/NR1H4) also participate in the establishment of liver sexual dimorphism by controlling male sexual maturation, a process under the control of gut microbiota-produced metabolites that regulate AHR, PXR and FXR transcriptional activities <sup>39</sup>. Importantly, the regulation of genes involved in metabolism is partly subject to circadian oscillation, which is itself controlled by sex. A few studies have found increased resilience of female individuals to circadian rhythm disruption<sup>68</sup>, which is a predisposing factor for cardiometabolic diseases including NAFLD<sup>69,70</sup>.

#### [H1] Sexual dimorphism in NAFLD and NASH

NAFLD, which in its most rapidly progressive form leads to NASH and fibrosis, is associated with an increased risk of liver-related and cardiovascular-related comorbidities in adults<sup>71,72</sup> (Figure 2). The early manifestations of NAFLD (that is, NAFL or steatosis), have a higher prevalence in men than in women irrespective of age<sup>73,74</sup>. In healthy men, the risk of NAFLD is inversely correlated with blood levels of oestradiol <sup>75</sup>. However, the prevalence of NAFLD is also steadily increasing in adolescents<sup>76</sup> and both the development of NAFLD and its progression to fibrosis show dose-dependent correlations with fructose intake <sup>77</sup>. Paediatric NAFLD is histologically characterized by portal inflammation that can progress to fibrosis in the absence of detectable hepatocyte degeneration or ballooning, in contrast to NAFLD in adults, which is characterized by mostly lobular inflammation and considerable ballooning<sup>78</sup>. Interestingly, levels of SHBG are inversely correlated with NAFL severity and although oestradiol levels do not correlate with NAFLD stage, testosterone levels are inversely correlated with fibrosis severity in boys<sup>79</sup>. Therefore, sex hormones are likely to also exert major effects in the paediatric population, but will not be further discussed in this Review owing to the scarcity of epidemiological and mechanistic studies<sup>80</sup>.

In general, men have more advanced grades of NASH than women<sup>81,82</sup>, and men are also more prone than women to develop fibrosis<sup>83</sup>. NASH-related HCC is diagnosed 2–4 times more often in men than women, and this sex bias is also observed in mouse studies, which suggests that the

increased susceptibility to HCC observed in men is unlikely to be due to increased exposure to risk factor(s)<sup>84</sup>. This protection from the development of HCC is similarly unlikely to reflect a globally higher resistance of the female liver to injury, as some other liver pathologies, such as alcoholic liver disease and primary biliary cirrhosis, develop more aggressively in women than in men<sup>85</sup>. As with the physiological aspects of liver sexual dimorphism, little is known of the intrinsic molecular mechanisms underlying the exacerbated development of NASH in males. Although genetic studies suggest that susceptibility to NASH and fibrosis (assessed by non-invasive methods) are heritable traits<sup>16,86</sup>, known risk alleles in the coding or noncoding regions of *PNPLA3*, *TM6SF2*, *GCKR*, *MBOAT7*, *SLC39A8*, *SLC30A10* and *HSD17B13* genes are not associated preferentially with either sex<sup>87,88</sup>. As the metabolic features of genetically driven NASH are distinct from those of metabolically driven NASH<sup>89,90</sup>, the genetic type will not be considered further in this Review.

Sex hormones (notably oestrogens) have a fundamental role in liver protection, as the prevalence of NASH in postmenopausal women (average age 51 years) in the USA is equivalent to that in men<sup>91</sup>, and premature menopause increases the risk of both NASH and fibrosis<sup>92</sup>. However, studies of the effects of hormone replacement therapy in humans have not yet formally established exogenous estrogens as liver-protective drugs, despite evidence of their positive effects in rodents<sup>93</sup>. Conversely, the results of a retrospective study suggest that the excess androgens present in women with polycystic ovary syndrome render them more likely to develop NAFLD<sup>94</sup>. Clinical trials of antiandrogen treatment in patients with HCC have failed to show any benefit, but were mostly carried out in patients with late-stage, virally induced tumours<sup>95</sup>.

#### [H1] Molecular explanations

As described in a number of recent comprehensive reviews, the natural history of NAFLD involves slow progression (over decades) from steatosis to steatohepatitis and ultimately to fibrosis, albeit with considerable inter-individual variation and spontaneous reversibility<sup>18,72,96-99</sup>. Progression through each stage increases the probability of worsening liver function, with advanced fibrosis being the most powerful predictor of liver-related or overall mortality<sup>25,100-102</sup>. Each stage is characterized by specific hepatic histological features<sup>18</sup> and is associated with prominent cellular (dys)functions that can be identified either by transcriptomic profiling of human bulk liver tissue or at the single-cell level<sup>103-109</sup>. Both clinical and experimental studies suggest a causative link between NASH and fibrogenesis<sup>26,110</sup>.

From this wealth of data consensus has emerged: that steatosis is linked to dysregulated expression of genes related to lipid storage, lipid and amino acid metabolism; that steatohepatitis is linked to dysregulation of genes involved in inflammation; and that fibrosis is linked to dyregulation of genes involved in extracellular matrix remodeling (Figure 2). Owing to considerable variations in

sample size, cohort constitution and technical factors, the results of these studies show little or no agreement as to whether the identified genes have a causative role in human disease. A core set of genes that are dysregulated in NASH or fibrosis has been identified through meta-analyses of mouse and human transcriptomic data, and the genes included in this core set mostly have functions involving the control of protein stability, inflammation or extracellular matrix architecture<sup>106,109,111</sup>. A random forest-based approach has identified male-specific and female-specific NASH gene signatures that had fairly high predictive capacity in independent cohorts<sup>112</sup>. However, these analyses could not establish whether physiologically sex-biased gene expression<sup>113</sup> is perturbed in individuals with NASH, as the initial cohort included morbidly obese patients but not lean controls.

Metabolomic approaches have also identified NAFLD-induced alterations in blood levels of lipidsand amino acid metabolites. The results of these studies are in general agreement that blood levels of lipid species increase during the progression of NAFLD<sup>114,115</sup> By contrast, some but not all studies reported altered levels of bile acids during progression of NAFLD, and this lack of consensus probably related to the use or not of cohort stratification criteria based on insulin resistance has led to controversial interpretations regarding the implication of bile acids in NAFLD pathogenic events. Interestingly, a multi-omics study of non-diabetic women reported, like others<sup>116-118</sup>, increased levels of branched chain amino acids in individuals with NAFLD. Synthesis of branched chain amino acids is catalyzed by the gut microbiota, which (in patients with steatosis) has low microbial gene richness and increased genetic potential for dietary lipid processing and endotoxin biosynthesis. In gut microbiota transplant experiments conducted in male mice, transfer of such low-richness gut microbiota triggered anaplerosis of the tricarboxylic acid cycle and ultimately steatosis<sup>119</sup>. In an independent study, increased levels of branched chain amino acids were specific to women, whereas men exhibited decreased levels <sup>120</sup>. This observation might provide a basis for the relative resistance of women to NAFLD, as in some studies supplementation with branched chain amino acids has been shown to be protective in preclinical models of NASH and also in patients with NASH. However, whether branched chain amino acids are beneficial or detrimental for liver health is still debated<sup>121</sup>.

Finally, HCC also occurs more frequently in men than in women<sup>122</sup>. HCC is characterized by a sex-biased transcriptional pattern<sup>123</sup> to which GH, sex hormones, their cognate nuclear receptors and associated pioneer transcription factors contribute<sup>122</sup>. The incidence of HCC is increased in patients with advanced NASH or cirrhosis<sup>124</sup>, but whether NAFLD-related HCC also exhibits sexual dimorphism is not clearly established<sup>125,126</sup>. In one rodent study, abrogated expression of the liver-specific P1 isoform of HNF4 $\alpha$ , which has an important role in the control of lipid, glucose and drug metabolism in the liver<sup>127,128</sup>, suggested that HNF4 $\alpha$  is critically involved in establishing the sex-biased development of NASH-induced HCC<sup>129</sup>. Whether the proposed mechanism, which involves IL-6 and STAT3, is also operative in humans is unknown.

#### [H1] Clinical considerations

Ample evidence supports the concept that sex is an important biological variable influencing the progression of NAFLD. However, the effect of sex continues to be overlooked even in the most recent publications relating to preclinical experiments, biomarker discovery and validation studies and even, surprisingly, in clinical trials in this field. This oversight is evident not only in the adult population, but also in paediatric and lean populations with NASH, which are much less thoroughly studied than the obese adult population. Although the need for improved stratification of patients with NASH is acknowledged both in clinical practice and in drug development, such stratification is rarely implemented in daily practice; therefore, insights might be gained from de-segregation of human transcriptomic data by sex<sup>112</sup>. Drug-based treatments options for NASH do not yet exist but the possibility of sex-based dosage optimization will need to be considered in the future. Early detection of individuals predisposed to develop NASH integrates parameters such as insulin resistance, but these parameters are also strongly influenced by sex<sup>130,131</sup> and interestingly, by GH, which is generally considered an antagonist of the actions of insulin. In addition to being sex-dependent, the secretion of GH has a circadian cycle that is affected by altered gut microbiota and sleep disorders, both of which contribute to the development of metabolic syndrome<sup>132</sup>.

#### [H1] Research needs and future perspectives

Complex multi-organ dysfunction contributes to the development of liver pathology in patients with NAFLD. Each of the affected components (liver, intestine, the gut microbiome, adipose tissue, immune cells and skeletal muscle) also has sex (hormone)-dependent physiology. Although most mechanistic data have been obtained from studies in male rodents, the sexual dimorphism of white adipose tissue depots is obvious from both histological and molecular viewpoints<sup>133,134</sup>. Crosstalk between the liver and white adipose tissue, exemplified by the occurrence of white adipose tissue inflammation preceding the development of NAFLD, has not yet been considered a sex-dependent process<sup>135</sup>. However, in mice, regulatory T cells (which regulate immune responses in visceral adipose tissue and exert tempering effects in NASH<sup>136</sup>) differ phenotypically and molecularly between the sexes<sup>137</sup>. In addition, resident and infiltrating macrophages in white adipose tissue and in the liver have important roles in NASH<sup>138</sup> and sex hormones strongly influence the phenotypes of these cells<sup>33</sup>.

Apart from gonadal sex, chromosomal sex (through X-escape genes — genes on the inactivated X chromosome that are nonetheless transcribed to produce RNA levels ≥10% of the expression of their counterparts on the active X chromosome) could also influence organ pathophysiology in NAFLD. The ability of white adipose tissue to expand, and specifically the ability of adipocytes to proliferate and differentiate in a metabolically challenging environment, also affects glucose homeostasis. Histone demethylase KDM5C, which is encoded on the X chromosome and

escapes X-chromosome inactivation, controls the proliferation of adipocytes<sup>139</sup> and contributes to the sex-specific expandability of white adipose tissue<sup>140</sup>. This circumstance raises the interesting possibility that X-linked gene expression levels might affect insulin resistance and hence progression of NAFLD, at least in its early stages. However, this hypothesis remains to be formally tested. Another parameter that is rarely taken into account in preclinical studies is ageing, mostly owing to cost and/or logistical constraints. Indeed, most preclinical studies of diet-induced NAFLD or NASH are conducted in young (usually male) animals and the disease barely reaches the fibrotic stage.

Finally, most knowledge of human NASH has been collected through transcriptomic data. Other causes of liver dysfunction, such as abnormal activation or inactivation of signalling pathways, might be overlooked. As an example, the GH–JAK2–STAT5 pathway strongly affects liver responses to metabolic and chemical insults in rodents<sup>141</sup>, yet the contributions of this and other signalling pathways to human NAFLD have not been directly tested using powerful approaches such as phosphoproteomics<sup>142</sup>. These knowledge gaps undoubtedly call for improved characterization of the animal models used in NASH research, the use of complementary techniques (such as integrative omics studies<sup>143,144</sup>) and for careful translation of observations made therein to human pathology<sup>145</sup>.

#### [H1] Conclusions

The liver is a sexually dimorphic organ and the pathological events leading to NAFLD progression in the two sexes are likely to differ, at least in part. This situation warrants a thorough investigation of sex-specific molecular mechanisms. In preclinical studies, the selection of appropriate experimental options (such as use of male or female animals) needs to go beyond the expectation of maximal and stable responses. Therefore, a full understanding of NAFLD progression and the identification of optimal therapeutic strategies requires the inclusion of sex as a critical parameter in driving pathological events.

## **Acknowledgements**

This work was supported by grants from Agence Nationale pour la Recherche (ANR-16-RHUS-0006-PreciNASH and ANR-10-LBEX-46), the European Union (FP6 Hepadip FP6-018734 and FP7 Resolve, FP7-305707), Fondation de France (Grant 2014 00047965), Fondation pour la Recherche Médicale (Equipe labellisée, DEQ20150331724). BS is a recipient of an Advanced ERC Grant (694717).

#### **Author contributions**

The authors contributed to all aspects of the article.

## **Competing interests**

The authors declare no competing interests.

#### **Peer review information**

Nature Reviews XXX thanks [Referee#1 name], [Referee#2 name] and the other, anonymous, reviewer(s) for their contribution to the peer review of this work.

### **Supplementary information**

Supplementary information is available for this paper at https://doi.org/10.1038/s415XX-XXXX-XXXX-X

## **Key points**

- Liver pathophysiology is sexually dimorphic
- Male individuals predominantly show more severe stages of NAFL, NASH and fibrosis
- Individual variations in humans require large cohorts to identify sex-specific features
- (Pre)clinical investigations of NAFLD and of fibrosis rarely consider sex as a biological variable
- The time (and technologies) are ripe for investigating the sexual dimorphism of liver diseases in time and space

**Figure 1: Main physiological pathways involved in liver sexual dimorphism.** The growth hormone (GH) and sex hormone signaling pathways are shown, as well as additional feedback loops controlling the release of GH from the pituitary gland. The principal biological outputs are also indicated.. GHRH, growth hormone-releasing hormone; HPA, hypothalamic–pituitary–adrenal axis; IR, insulin receptor; GHR, growth hormone receptor; NEFAs, non-esterified fatty acids; E2, 17β-oestradiol; *CYP19A1*, the gene encoding aromatase; JAK2, Janus kinase 2; STAT5, signal transducer and activator of transcription 5; ER, estrogen receptor (encoded by *ESR1*); IGF-I, insulin-like growth factor 1; *PPARG*, the gene encoding peroxisome proliferator activated receptor-γ; SREBP1c, sterol regulatory element-binding protein 1c (encoded by *SREBF1*).

**Figure 2: Progression of NAFLD is influenced by sexual dimorphism.** Various organ responses and dysfunctions influence the progression of nonalcoholic fatty liver disease (NAFLD). Only sexually dimorphic features that are established or at least partially described in the literature are indicated. NEFAs, non-esterified fatty acids; NAFL, non-alcoholic fatty liver; ECM, extracellular matrix; HCC: hepatocellular carcinoma.

#### **BIBLIOGRAPHY**

- 1 Schork, N. J. Personalized medicine: Time for one-person trials. *Nature* **520**, 609-611, (2015).
- Prendergast, B. J., Onishi, K. G. & Zucker, I. Female mice liberated for inclusion in neuroscience and biomedical research. *Neurosci Biobehav Rev* **40**, 1-5, (2014).
- Woitowich, N. C. & Woodruff, T. K. Opinion: Research community needs to better appreciate the value of sex-based research. *Proc Natl Acad Sci U S A* **116**, 7154-7156, (2019).
- 4 Lam, C. S. P. et al. Sex differences in heart failure. Eur Heart J 40, 3859-3868c, (2019).
- Gannon, M., Kulkarni, R. N., Tse, H. M. & Mauvais-Jarvis, F. Sex differences underlying pancreatic islet biology and its dysfunction. *Mol Metab* **15**, 82-91, (2018).
- Rubin, J. B. et al. Sex differences in cancer mechanisms. Biol Sex Differ 11, 17-46, (2020).
- Kokturk, N., Kilic, H., Baha, A., Lee, S. D. & Jones, P. W. Sex difference in chronic obstructive lung disease. Does it matter? A concise review. *COPD* **13**, 799-806, (2016).
- 8 Mauvais-Jarvis, F. *et al.* Sex and gender: modifiers of health, disease, and medicine. *The Lancet* **396**, 565-582, (2020).
- 9 Lonardo, A. & Suzuki, A. Sexual dimorphism of NAFLD in adults. Focus on clinical aspects and implications for practice and translational research. *J Clin Med* **9**, 1278-1307,(2020).
- Ludwig, J., Viggiano, T. R., McGill, D. B. & Oh, B. J. Nonalcoholic steatohepatitis: Mayo Clinic experiences with a hitherto unnamed disease. *Mayo Clinic proceedings* **55**, 434-438, (1980).
- James, O. F. & Day, C. P. Non-alcoholic steatohepatitis (NASH): a disease of emerging identity and importance. *J Hepatol* **29**, 495-501, (1998).
- James, O. & Day, C. Non-alcoholic steatohepatitis: another disease of affluence. *The Lancet* **353**, 1634-1636, (1999).
- Dowman, J. K., Tomlinson, J. W. & Newsome, P. N. Pathogenesis of non-alcoholic fatty liver disease. *QJM* **103**, 71-83, (2010).
- Tilg, H., Adolph, T. E. & Moschen, A. R. Multiple parallel hits hypothesis in NAFLD Revisited After a Decade. *Hepatology*, **73**, 833-842, (2021).
- 15 Kolodziejczyk, A. A., Zheng, D., Shibolet, O. & Elinav, E. The role of the microbiome in NAFLD and NASH. *EMBO Mol Med* **11**, e9302, (2019).
- Eslam, M., Valenti, L. & Romeo, S. Genetics and epigenetics of NAFLD and NASH: Clinical impact. *J Hepatol* **68**, 268-279, (2018).
- Younossi, Z. *et al.* Global burden of NAFLD and NASH: trends, predictions, risk factors and prevention. *Nat Rev Gastroenterol Hepatol* **15**, 11-20, (2018).
- 18 Sheka, A. C. et al. Nonalcoholic steatohepatitis: A Review. JAMA 323, 1175-1183, (2020).
- Ye, Q. *et al.* Global prevalence, incidence, and outcomes of non-obese or lean non-alcoholic fatty liver disease: a systematic review and meta-analysis. *The Lancet. Gastroenterology & hepatology*, (2020).
- 20 Chen, F. *et al.* Lean NAFLD: A distinct entity shaped by differential metabolic adaptation. *Hepatology*, **71**, 1213-1227 (2019).
- 21 Chang, Y. *et al.* Metabolically healthy obesity and the development of nonalcoholic fatty liver disease. *Am J Gastroenterol* **111**, 1133-1140, (2016).
- Younossi, Z. M. *et al.* The global epidemiology of NAFLD and NASH in patients with type 2 diabetes: A systematic review and meta-analysis. *J Hepatol* **71**, 793-801, (2019).
- Jarvis, H. *et al.* Metabolic risk factors and incident advanced liver disease in non-alcoholic fatty liver disease (NAFLD): A systematic review and meta-analysis of population-based observational studies. *PLoS Med* **17**, e1003100, (2020).
- Targher, G., Byrne, C. D. & Tilg, H. NAFLD and increased risk of cardiovascular disease: clinical associations, pathophysiological mechanisms and pharmacological implications. *Gut*, **69**, 1691-1705, (2020).
- Hagstrom, H. *et al.* Fibrosis stage but not NASH predicts mortality and time to development of severe liver disease in biopsy-proven NAFLD. *J Hepatol* **67**, 1265-1273, (2017).

- Haas, J. T., Francque, S. & Staels, B. Pathophysiology and mechanisms of nonalcoholic fatty liver disease. *Annu Rev Physiol* **78**, 181-205, (2016).
- Schwabe, R. F., Tabas, I. & Pajvani, U. B. Mechanisms of fibrosis development in nash. *Gastroenterology*, **158**, 1913–1928, (2020).
- Smith, G. I. *et al.* Insulin resistance drives hepatic de novo lipogenesis in nonalcoholic fatty liver disease. *J Clin Invest*, **130**, 1453-1460, (2019).
- Romero, F. A., Jones, C. T., Xu, Y., Fenaux, M. & Halcomb, R. L. The race to bash NASH: Emerging targets and drug development in a complex liver disease. *J Med Chem*, **63**, 5031-5073, (2020).
- Jensen, V. S. *et al.* Variation in diagnostic NAFLD/NASH read-outs in paired liver samples from rodent models. *J Pharmacol Toxicol Methods* **101**, 106651-106660, (2020).
- Rinella, M. E., Tacke, F., Sanyal, A. J., Anstee, Q. M. & participants of the, A. E. W. Report on the AASLD/EASL joint workshop on clinical trial endpoints in NAFLD. *J Hepatol* **71**, 823-833, (2019).
- Ren, C. & Sylvia, K.E. Sexual dimorphism in the gut microbiome, I U Journal of Undergraduate Research, **4**, 12–16, (2018).
- Jaillon, S., Berthenet, K. & Garlanda, C. Sexual dimorphism in innate immunity. *Clin Rev Allergy Immunol* **56**, 308-321, (2019).
- Roy, A. K. & Chatterjee, B. Sexual dimorphism in the liver. *Annu Rev Physiol* **45**, 37-50, (1983).
- 35 Klaassen, C. D. & Aleksunes, L. M. Xenobiotic, bile acid, and cholesterol transporters: function and regulation. *Pharmacol Rev* **62**, 1-96, (2010).
- Della Torre, S. & Maggi, A. Sex Differences: A resultant of an evolutionary pressure? *Cell Metab* **25**, 499-505, (2017).
- 37 Maggi, A. & Della Torre, S. Sex, metabolism and health. *Mol Metab* **15**, 3-7, (2018).
- Justo, R. *et al.* Gender dimorphism in rat liver mitochondrial oxidative metabolism and biogenesis. *Am J Physiol Cell Physiol* **289**, C372-378, (2005).
- Weger, B. D. *et al.* The mouse microbiome is required for sex-specific diurnal rhythms of gene expression and metabolism. *Cell Metab*, **29**, 1-21, (2018).
- Jansson, J. O., Edén, S. & Isaksson, O. Sexual dimorphism in the control of growth hormone secretion. *Endocr Rev* **6**, 128-150, (1985).
- Waxman, D. J. & Holloway, M. G. Sex differences in the expression of hepatic drug metabolizing enzymes. *Mol Pharmacol* **76**, 215-228, (2009).
- Davey, H. W., Wilkins, R. J. & Waxman, D. J. STAT5 signaling in sexually dimorphic gene expression and growth patterns. *Am J Hum Genet* **65**, 959-965, (1999).
- Waxman, D. J. & O'Connor, C. Growth hormone regulation of sex-dependent liver gene expression. *Mol Endocrinol* **20**, 2613-2629, (2006).
- Zheng, D., Wang, X., Antonson, P., Gustafsson, J. & Li, Z. Genomics of sex hormone receptor signaling in hepatic sexual dimorphism. *Mol Cell Endocrinol* **471**, 33-41, (2018).
- Rando, G. & Wahli, W. Sex differences in nuclear receptor-regulated liver metabolic pathways. *Biochim Biophys Acta* **1812**, 964-973, (2011).
- Yang, X., Zhang, Y. K., Esterly, N., Klaassen, C. D. & Wan, Y. J. Gender disparity of hepatic lipid homoeostasis regulated by the circadian clock. *J Biochem* **145**, 609-623, (2009).
- 47 Qian, J. *et al.* Sex differences in the circadian misalignment effects on energy regulation. *Proc Natl Acad Sci U S A* **116**, 23806-23812, (2019).
- Sheng, L. *et al.* Gender differences in bile acids and microbiota in relationship with gender dissimilarity in steatosis induced by diet and FXR inactivation. *Sci Rep* **7**, 1748, (2017).
- 49 Chiang, J. Y. L. Linking Sex Differences in non-alcoholic fatty liver disease to bile acid signaling, gut microbiota, and high fat diet. *Am J Pathol* **187**, 1658-1659, (2017).
- Jena, P. K. *et al.* Western diet-induced dysbiosis in Farnesoid X Receptor knockout mice causes persistent hepatic inflammation after antibiotic treatment. *Am J Pathol* **187**, 1800-1813, (2017).
- Giles, D. A. *et al.* Thermoneutral housing exacerbates nonalcoholic fatty liver disease in mice and allows for sex-independent disease modeling. *Nat Med* **23**, 829-838, (2017).

- Nobs, S. P., Tuganbaev, T. & Elinav, E. Microbiome diurnal rhythmicity and its impact on host physiology and disease risk. *EMBO Rep* **20**, e47129, (2019).
- Fernandez-Perez, L., de Mirecki-Garrido, M., Recio, C. & Guerra, B. Control of liver gene expression by sex steroids and growth hormone interplay in *Chemistry and Biological Activity of Steroids*, Chapter 4, (Intechopen, 2019).
- Jansson, J. O. & Frohman, L. A. Differential effects of neonatal and adult androgen exposure on the growth hormone secretory pattern in male rats. *Endocrinology* **120**, 1551-1557, (1987).
- Reizel, Y. *et al.* Gender-specific postnatal demethylation and establishment of epigenetic memory. *Genes Dev* **29**, 923-933, (2015).
- Hosui, A. & Hennighausen, L. Genomic dissection of the cytokine-controlled STAT5 signaling network in liver. *Physiol Genomics* **34**, 135-143, (2008).
- 57 Clodfelter, K. H. *et al.* Sex-dependent liver gene expression is extensive and largely dependent upon signal transducer and activator of transcription 5b (STAT5b): STAT5b-dependent activation of male genes and repression of female genes revealed by microarray analysis. *Mol Endocrinol* **20**, 1333-1351, (2006).
- Lau-Corona, D., Suvorov, A. & Waxman, D. J. Feminization of male mouse liver by persistent growth hormone stimulation: activation of sex-biased transcriptional networks and dynamic changes in chromatin states. *Mol Cell Biol* **37**, e00301-17, (2017).
- Palmisano, B. T., Zhu, L. & Stafford, J. M. Role of estrogens in the regulation of liver lipid metabolism. *Advances in experimental medicine and biology* **1043**, 227-256, (2017).
- 60 Mittendorfer, B. Sexual dimorphism in human lipid metabolism. *J Nutr* **135**, 681-686, (2005).
- Phelps, T., Snyder, E., Rodriguez, E., Child, H. & Harvey, P. The influence of biological sex and sex hormones on bile acid synthesis and cholesterol homeostasis. *Biol Sex Differ* **10**, 52-64, (2019).
- Koulouri, O., Ostberg, J. & Conway, G. S. Liver dysfunction in Turner's syndrome: prevalence, natural history and effect of exogenous oestrogen. *Clin Endocrinol (Oxf)* **69**, 306-310, (2008).
- Tramunt, B. *et al.* Sex differences in metabolic regulation and diabetes susceptibility. *Diabetologia* **63**, 453-461, (2020).
- Shen, M. & Shi, H. Sex hormones and their receptors regulate liver energy homeostasis. *Int J Endocrinol* **2015**, 294278, (2015).
- 65 Grossmann, M., Wierman, M. E., Angus, P. & Handelsman, D. J. Reproductive endocrinology of nonalcoholic fatty liver disease. *Endocr Rev* **40**, 417-446, (2019).
- Sharma, G. & Prossnitz, E. R. G-protein-coupled estrogen receptor (GPER) and sex-specific metabolic homeostasis. *Advances in experimental medicine and biology* **1043**, 427-453, (2017).
- 67 Meda, C. *et al.* Hepatic ERalpha accounts for sex differences in the ability to cope with an excess of dietary lipids. *Mol Metab* **32**, 97-108, (2020).
- Anderson, S. T. & FitzGerald, G. A. Sexual dimorphism in body clocks. *Science* **369**, 1164-1165, (2020).
- Saran, A. R., Dave, S. & Zarrinpar, A. Circadian Rhythms in the Pathogenesis and Treatment of Fatty Liver Disease. *Gastroenterology*, **158**, 1948–1966, (2020).
- Mukherji, A., Bailey, S. M., Staels, B. & Baumert, T. F. The circadian clock and liver function in health and disease. *J Hepatol*, **71**, 200-211, (2019).
- 5tols-Goncalves, D., Hovingh, G. K., Nieuwdorp, M. & Holleboom, A. G. NAFLD and atherosclerosis: two sides of the same dysmetabolic coin? *Trends Endocrinol Metab* **30**, 891-902, (2019).
- Anstee, Q. M., Reeves, H. L., Kotsiliti, E., Govaere, O. & Heikenwalder, M. From NASH to HCC: current concepts and future challenges. *Nat Rev Gastroenterol Hepatol* **16**, 411-428, (2019).
- Ballestri, S. *et al.* NAFLD as a Sexual Dimorphic Disease: Role of gender and reproductive status in the development and progression of nonalcoholic fatty liver disease and inherent cardiovascular risk. *Advances in therapy* **34**, 1291-1326, (2017).

- Beaudry, K. M. & Devries, M. C. Sex-based differences in hepatic and skeletal muscle triglyceride storage and metabolism (1). *Applied physiology, nutrition, and metabolism Physiologie appliquée, nutrition et métabolisme* **44**, 805-813, (2019).
- Tian, G. X. *et al.* Oestradiol is a protective factor for non-alcoholic fatty liver disease in healthy men. *Obesity reviews, Obesity* **13**, 381-387, (2012).
- Younossi, Z. M. *et al.* Global epidemiology of nonalcoholic fatty liver disease-Meta-analytic assessment of prevalence, incidence, and outcomes. *Hepatology* **64**, 73-84, (2016).
- Ribeiro, A., Igual-Perez, M. J., Santos Silva, E. & Sokal, E. M. Childhood fructoholism and fructoholic liver disease. *Hepatol Commun* **3**, 44-51, (2019).
- Nobili, V., Alisi, A., Newton, K. P. & Schwimmer, J. B. Comparison of the phenotype and approach to pediatric vs adult patients with nonalcoholic fatty liver disease. *Gastroenterology* **150**, 1798-1810, (2016).
- Mueller, N. T. *et al.* Sex hormone relations to histologic severity of pediatric nonalcoholic fatty liver disease. *J Clin Endocrinol Metab*, **105**, 3496-3504, (2020).
- Hatton, G., Alterio, T., Nobili, V. & Mann, J. P. Unmet needs in pediatric NAFLD research: what do we need to prioritize for the future? *Expert Rev Gastroenterol Hepatol* **12**, 961-967, (2018).
- Arun, J., Clements, R. H., Lazenby, A. J., Leeth, R. R. & Abrams, G. A. The prevalence of nonalcoholic steatohepatitis is greater in morbidly obese men compared to women. *Obes Surg* **16**, 1351-1358, (2006).
- Skubic, C., Drakulić, Ž. & Rozman, D. Personalized therapy when tackling nonalcoholic fatty liver disease: a focus on sex, genes, and drugs. *Expert opinion on drug metabolism & toxicology* **14**, 831-841, (2018).
- Lonardo, A. *et al.* Sex Differences in NAFLD: State of the art and identification of research gaps. *Hepatology*, **70**, 1457-1469, (2019).
- 84 El-Serag, H. B. & Rudolph, K. L. Hepatocellular carcinoma: epidemiology and molecular carcinogenesis. *Gastroenterology* **132**, 2557-2576, (2007).
- Durazzo, M. *et al.* Gender specific medicine in liver diseases: a point of view. *World J Gastroenterol* **20**, 2127-2135, (2014).
- Loomba, R. *et al.* Heritability of hepatic fibrosis and steatosis based on a prospective twin study. *Gastroenterology* **149**, 1784-1793, (2015).
- Parisinos, C. A. *et al.* Genome-wide and Mendelian randomisation studies of liver MRI yield insights into the pathogenesis of steatohepatitis. *J Hepatol*, **73**, 241-25, (2020).
- Verrijken, A. *et al.* A gene variant of PNPLA3, but not of APOC3, is associated with histological parameters of NAFLD in an obese population. *Obesity (Silver Spring)* **21**, 2138-2145, (2013).
- Petaja, E. M. & Yki-Jarvinen, H. Definitions of normal liver fat and the association of insulin sensitivity with acquired and genetic NAFLD-A Systematic Review. *Int J Mol Sci* **17**, 633-649, (2016).
- 90 Yki-Jarvinen, H. & Luukkonen, P. K. Heterogeneity of non-alcoholic fatty liver disease. *Liver Int* **35**, 2498-2500, (2015).
- Yang, J. D. *et al.* Gender and menopause impact severity of fibrosis among patients with nonalcoholic steatohepatitis. *Hepatology* **59**, 1406-1414, (2014).
- 92 Klair, J. S. *et al.* A longer duration of estrogen deficiency increases fibrosis risk among postmenopausal women with nonalcoholic fatty liver disease. *Hepatology* **64**, 85-91, (2016).
- Lee, C., Kim, J. & Jung, Y. Potential therapeutic application of estrogen in gender disparity of nonalcoholic fatty liver disease/nonalcoholic steatohepatitis. *Cells* **8**, 8(10):1259-1279, (2019).
- 94 Kumarendran, B. *et al.* Polycystic ovary syndrome, androgen excess, and the risk of nonalcoholic fatty liver disease in women: A longitudinal study based on a United Kingdom primary care database. *PLoS Med* **15**, e1002542- e1002562, (2018).
- Ali, M. A. *et al.* Nonalcoholic steatohepatitis-related hepatocellular carcinoma: is there a role for the androgen receptor pathway? *Onco Targets Ther* **10**, 1403-1412, (2017).
- 96 Negro, F. Natural history of NASH and HCC. Liver Int 40 Suppl 1, 72-76, (2020).

- 97 Altayar, O., Noureddin, N., Thanda Han, M. A., Murad, M. H. & Noureddin, M. Fibrosis Changes in the Placebo Arm of NASH Clinical Trials. *Clin Gastroenterol Hepatol* **17**, 2387, (2019).
- Singh, S. *et al.* Fibrosis progression in nonalcoholic fatty liver vs nonalcoholic steatohepatitis: a systematic review and meta-analysis of paired-biopsy studies. *Clin Gastroenterol Hepatol* **13**, 643-654, (2015).
- 99 Eslam, M., Sanyal, A. J., George, J. & an international consensus, p. MAFLD: A consensus-driven proposed nomenclature for metabolic associated fatty liver disease. *Gastroenterology*, **158**, 1999-2014 (2020).
- Angulo, P. et al. Liver fibrosis, but no other histologic features, is associated with long-term outcomes of patients with nonalcoholic fatty liver disease. *Gastroenterology* **149**, 389-397, (2015).
- Ekstedt, M. *et al.* Fibrosis stage is the strongest predictor for disease-specific mortality in NAFLD after up to 33 years of follow-up. *Hepatology* **61**, 1547-1554, (2015).
- Dulai, P. S. *et al.* Increased risk of mortality by fibrosis stage in nonalcoholic fatty liver disease: Systematic review and meta-analysis. *Hepatology* **65**, 1557-1565, (2017).
- Ramachandran, P. *et al.* Resolving the fibrotic niche of human liver cirrhosis at single-cell level. *Nature*, **575**, 512-518, (2019).
- Moylan, C. A. *et al.* Hepatic gene expression profiles differentiate presymptomatic patients with mild versus severe nonalcoholic fatty liver disease. *Hepatology* **59**, 471-482, (2014).
- Arendt, B. M. *et al.* Altered hepatic gene expression in nonalcoholic fatty liver disease is associated with lower hepatic n-3 and n-6 polyunsaturated fatty acids. *Hepatology* **61**, 1565-1578, (2015).
- Lefebvre, P. *et al.* Interspecies NASH disease activity whole-genome profiling identifies a fibrogenic role of PPARalpha-regulated dermatopontin. *JCI Insight* **2**, 92264-92281, (2017).
- Haas, J. T. *et al.* Transcriptional network analysis implicates altered hepatic immune function in NASH development and resolution. *Nature Metabolism* **1**, 604-614, (2019).
- Suppli, M. P. *et al.* Hepatic transcriptome signatures in patients with varying degrees of nonalcoholic fatty liver disease compared with healthy normal-weight individuals. *Am J Physiol Gastrointest Liver Physiol* **316**, G462-G472, (2019).
- Gerhard, G. S. *et al.* Transcriptomic profiling of obesity-related nonalcoholic steatohepatitis reveals a core set of fibrosis-specific genes. *J Endocr Soc* **2**, 710-726, (2018).
- 110 Ratziu, V. Back to Byzance: Querelles byzantines over NASH and fibrosis. *J Hepatol* **67**, 1134-1136, (2017).
- 111 Chen, W. *et al.* Multi-transcriptomes analyses reveal prioritized genes specifically associated with liver fibrosis progression independent of etiology. *Am J Physiol Gastrointest Liver Physiol*, **316**:G744-G754 (2019).
- Vandel, J. *et al.* Hepatic molecular signatures highlight the sexual dimorphism of Non-Alcoholic SteatoHepatitis (NASH). *Hepatology*, **73**, 920-936, (2020).
- 2hang, Y. *et al.* Transcriptional profiling of human liver identifies sex-biased genes associated with polygenic dyslipidemia and coronary artery disease. *PLoS One* **6**, e23506-e23522, (2011).
- Puri, P. *et al.* The plasma lipidomic signature of nonalcoholic steatohepatitis. *Hepatology* **50**, 1827-1838, (2009).
- Loomba, R., Quehenberger, O., Armando, A. & Dennis, E. A. Polyunsaturated fatty acid metabolites as novel lipidomic biomarkers for noninvasive diagnosis of nonalcoholic steatohepatitis. *J Lipid Res* **56**, 185-192, (2015).
- Gaggini, M. *et al.* Altered amino acid concentrations in NAFLD: Impact of obesity and insulin resistance. *Hepatology*, **67**, 145-158, (2017).
- van den Berg, E. H. *et al.* Non-alcoholic fatty liver disease and risk of incident type 2 diabetes: role of circulating branched-chain amino acids. *Nutrients* **11**, 705-720, (2019).
- Lake, A. D. *et al.* Branched chain amino acid metabolism profiles in progressive human nonalcoholic fatty liver disease. *Amino Acids* **47**, 603-615, (2015).

- Hoyles, L. *et al.* Molecular phenomics and metagenomics of hepatic steatosis in non-diabetic obese women. *Nat Med*, **24**, 1070-1080, (2018).
- Grzych, G. et al. Plasma BCAA changes in patients with NAFLD are sex dependent. J Clin Endocrinol Metab, 105, 2311–2321, (2020).
- 121 Canfora, E. E., Meex, R. C. R., Venema, K. & Blaak, E. E. Gut microbial metabolites in obesity, NAFLD and T2DM. *Nat Rev Endocrinol* **15**, 261-273, (2019).
- 122 Clocchiatti, A., Cora, E., Zhang, Y. & Dotto, G. P. Sexual dimorphism in cancer. *Nat Rev Cancer* **16**, 330-339, (2016).
- Natri, H. M., Wilson, M. A. & Buetow, K. H. Distinct molecular etiologies of male and female hepatocellular carcinoma. *BMC Cancer* **19**, 951-963, (2019).
- 124 Forner, A., Reig, M. & Bruix, J. Hepatocellular carcinoma. *The Lancet* **391**, 1301-1314, (2018).
- Reddy, S. K. *et al.* Outcomes of curative treatment for hepatocellular cancer in nonalcoholic steatohepatitis versus hepatitis C and alcoholic liver disease. *Hepatology* **55**, 1809-1819, (2012).
- Yasui, K. *et al.* Characteristics of patients with nonalcoholic steatohepatitis who develop hepatocellular carcinoma. *Clin Gastroenterol Hepatol* **9**, 428-433, (2011).
- Gupta, R. K. & Kaestner, K. H. HNF-4alpha: from MODY to late-onset type 2 diabetes. *Trends Mol Med* **10**, 521-524, (2004).
- Lau, H. H., Ng, N. H. J., Loo, L. S. W., Jasmen, J. B. & Teo, A. K. K. The molecular functions of hepatocyte nuclear factors In and beyond the liver. *J Hepatol* **68**, 1033-1048, (2018).
- Fekry, B. *et al.* HNF4 $\alpha$ -deficient fatty liver provides a permissive environment for sexindependent hepatocellular carcinoma. *Cancer Res* **79**, 5860-5873, (2019).
- Varlamov, O., Bethea, C. L. & Roberts, C. T., Jr. Sex-specific differences in lipid and glucose metabolism. *Front Endocrinol (Lausanne)* **5**, 241-248, (2014).
- Aldhoon-Hainerova, I. *et al.* Glucose homeostasis and insulin resistance: prevalence, gender differences and predictors in adolescents. *Diabetol Metab Syndr* **6**, 100-109, (2014).
- Weger, B. D., Rawashdeh, O. & Gachon, F. At the intersection of microbiota and circadian clock: are sexual dimorphism and growth hormones the missing link to pathology?: Circadian Clock and Microbiota: Potential Effect on Growth Hormone and Sexual Development. *Bioessays* **41**, e1900059, (2019).
- Bloor, I. D. & Symonds, M. E. Sexual dimorphism in white and brown adipose tissue with obesity and inflammation. *Horm Behav* **66**, 95-103, (2014).
- Moreira-Pais, A. *et al.* Sex differences on adipose tissue remodeling: from molecular mechanisms to therapeutic interventions. *J Mol Med (Berl)* **98**, 483-493, (2020).
- Korf, H., Boesch, M., Meelberghs, L. & van der Merwe, S. Macrophages as Key Players during Adipose Tissue-Liver Crosstalk in Nonalcoholic Fatty Liver Disease. *Semin Liver Dis* **39**, 291-300, (2019).
- Van Herck, M. A. *et al.* The Differential Roles of T Cells in Non-alcoholic Fatty Liver Disease and Obesity. *Front Immunol* **10**, 82-102, (2019).
- 137 Vasanthakumar, A. *et al.* Sex-specific adipose tissue imprinting of regulatory T cells. *Nature* **579**, 581-585, (2020).
- 138 Remmerie, A., Martens, L. & Scott, C. L. Macrophage Subsets in Obesity, Aligning the Liver and Adipose Tissue. *Front Endocrinol (Lausanne)* **11**, 259-264, (2020).
- Brier, A. B. *et al.* The KDM5 family is required for activation of pro-proliferative cell cycle genes during adipocyte differentiation. *Nucleic Acids Res*, **45**, 1743-1759, (2016).
- Link, J. C. *et al.* X chromosome dosage of histone demethylase KDM5C determines sex differences in adiposity. *J Clin Invest*, **130**, 5688-5701, (2020).
- 141 Kaltenecker, D. *et al.* Hepatic growth hormone JAK2 STAT5 signalling: Metabolic function, non-alcoholic fatty liver disease and hepatocellular carcinoma progression. *Cytokine* **124**, 154569-154581, (2019).
- 142 Wattacheril, J. *et al.* Non-alcoholic fatty liver disease phosphoproteomics: A functional piece of the precision puzzle. *Hepatol Res* **47**, 1469-1483, (2017).

- 143 Kurt, Z. *et al.* Tissue-specific pathways and networks underlying sexual dimorphism in non-alcoholic fatty liver disease. *Biol Sex Differ* **9**, 46-60, (2018).
- Cvitanović Tomaš, T., Urlep, Ž., Moškon, M., Mraz, M. & Rozman, D. LiverSex computational model: Sexual aspects in hepatic metabolism and abnormalities. *Front Physiol* **9**, 360-372, (2018).
- Haczeyni, F. *et al.* Mouse models of non-alcoholic steatohepatitis: A reflection on recent literature. *J Gastroenterol Hepatol* **33**, 1312-1320, (2018).



