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Review

Sphingolipids metabolism alteration in the central nervous system: Amyotrophic lateral sclerosis (ALS) and other neurodegenerative diseases

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ABSTRACT

Sphingolipids are complex lipids. They play a structural role in neurons, but are also involved in regulating cellular communication, and neuronal differentiation and maturation. There is increasing evidence to suggest that dysregulated metabolism of sphingolipids is linked to neurodegenerative processes in amyotrophic lateral sclerosis (ALS), Parkinson's disease and Gaucher's disease. In this review, we provide an overview of the role of sphingolipids in the development and maintenance of the nervous system. We describe the implications of altered metabolism of sphingolipids in the pathophysiology of certain neurodegenerative diseases, with a primary focus on ALS. Finally, we provide an update of potential treatments that could be used to target the metabolism of sphingolipids in neurodegenerative diseases.

1. Introduction

Lipids represent around 60% of the constituents of nervous tissue and are essential in the propagation of electrical and chemical signals [1]. The sphingolipids represent almost 20% of the lipids of the nervous system [2]. They belong to a major class of complex lipids that are involved in various physiological and developmental processes, thus ensuring the proper functioning of the nervous system. In this review, we describe the generalities of sphingolipids and their physiological roles in the nervous system, as well as the importance of sphingolipids in neurodegenerative diseases. Finally, we discuss therapeutic approaches associated with sphingolipids in animal models and clinical trials.

2. Biosynthesis of sphingolipids in the nervous system

The biosynthesis of sphingolipids begins at the cytosolic layer of the endoplasmic reticulum (ER) and progresses through several subcellular structures. The general structure of sphingolipids is defined by a sphingosine skeleton. Ceramide is the simplest sphingolipid. Made up of a sphingosine molecule and one fatty acid, it is the key precursor for the synthesis of many sphingolipids. Ceramide is obtained by the transformation of sphinganine into dihydroceramide by the ceramide synthase enzymes, which represent a large family of ER enzymes. Dihydroceramide is subsequently converted to ceramide by ceramidase [3]. During synthesis, the subclass of sphingolipids is determined through the addition of different chemical

Abbreviations: ALS, amyotrophic lateral sclerosis; ER, endoplasmic reticulum; UGT8, galactosyltransferase; FAPP2, four-phosphate adaptator protein 2; SM, sphingomyelin; CNS, central nervous system; UGCG, UDP-glucose ceramide glycosyltransferase; MAGs, myelin-associated glycoproteins; SMase, sphingomyelinase; Trk, Tropomyosin receptor kinase; AD, Alzheimer's disease; PD, Parkinson's disease; GBA, β -Glucocerebrosidase; S1P, sphingosine-1-phosphate; BBB, blood-brain barrier.

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groups (radical group "R") to the ceramide (Fig. 1). Among these subclasses are the glycosphingolipids which are characterized by the addition of one or more monosaccharides. Once formed in the ER, ceramide can be directly transformed into galactosylceramide through the addition of one galactose by the enzyme ceramide galactosyltransferase (UGT8), or transported to the Golgi apparatus to give rise to several glycosphingolipids and gangliosides (Fig. 2). For example, when ceramide is transported to the Cis-Golgi network, the addition of glucose leads to the formation of glucosylceramide. At this stage, glucosylceramide is transported to the trans-Golgi apparatus via two mechanisms: through vesicular transport or through the transport protein four-phosphate adaptator protein 2 (FAPP2) [4,5]. At the trans-Golgi network, glucosylceramide is transformed into different glycosphingolipids (e.g. lactosylceramide, gangliosides, etc.). Using the ceramide transferase protein, ceramide can be transported directly to the trans-Golgi network, where the addition of a phosphocholine produces sphingomyelin. At the trans-Golgi network, several glycosphingolipids are formed. These glycosphingolipids are transported to the plasma membrane where they are integrated into lipid rafts. Finally, the addition of glucose and galactose to the ceramide allows for the formation of lactosylceramide. Addition of sialic acid to lactosylceramide leads to the production of the first ganglioside, GM3. Various derivatives can then be generated through the addition of N-acetylgalactosamine and galactose, which gives rise to GM2, GM1, and so on (Fig. 1).

The degradation of glycosphingolipids occurs in the endosomes and lysosomes. Endosomal glycosphingolipids are recycled to subcellular compartments (ER and Golgi apparatus) while the glycosphingolipids that are redirected to the lysosomes are hydrolyzed to ceramide and then to sphingosine.

3. Sphingolipids have major roles in the nervous system

Several studies show that sphingolipids are widely distributed across the central nervous system (CNS). The CNS is mainly composed of glycosphingolipids and gangliosides [6,7]. Gangliosides are mainly located

in neurons, whereas galactosylceramide is mainly localized in the oligodendrocytes [8]. The distribution of sphingolipids in the CNS highlights their importance in the formation of different cell types, and in neurodevelopment.

3.1. Sphingolipids: driving the maturation of the nervous system

The gangliosides are the major class of sphingolipids that are found at high concentrations in the CNS. As such, they are indispensable for the development of the nervous system, axonal growth, and neuronal differentiation [9,10]. During embryonic development, ganglioside levels vary depending on the stage of maturation (Fig. 3). During the early stages of development, GM3 and GD3 are synthesized in large quantities during the formation of the neural tube, and they also participate in the proliferation and differentiation of neural stem cells [7,10,11]. Next, several derivatives of GM3 (like GD1a, GM1, GD1b and GT1b) participate in neuronal differentiation, synaptogenesis, and myelination until adulthood [12]. Sphingomyelin, galactosylceramide and sulfatide also participate in axonal arborization during development. Not surprisingly, an absence of GD3 synthase drastically delays axonal growth and myelination in mice [13]. By contrast, administration of GD3 improves the regeneration process after sciatic nerve injury in GD3 synthase knockout (KO) mice [14]. This age- and development-dependent expression of gangliosides highlights their crucial role in neurogenesis.

3.2. Implication of glycosphingolipids in neuronal differentiation and axonal growth

The other actors involved in the development of the nervous system are the glycosphingolipids. As gangliosides, glycosphingolipids are involved in establishing neuronal proliferation and maturation, as well as axonal growth [7,15,16]. In addition, many glycosphingolipid enzymes such as the synthetic enzymes of ceramide and glucosylceramide, ceramide synthase and the UDP-glucose ceramide glucosyltransferase

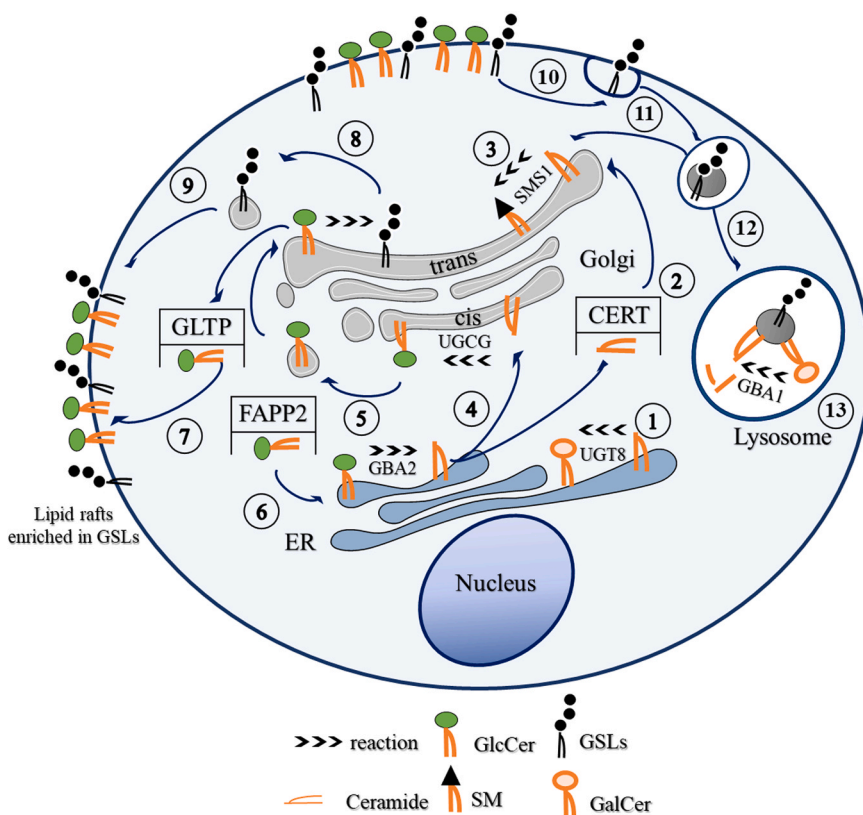


Fig. 1. Synthesis, degradation, and recycling of sphingolipids. The ceramide synthesized in the endoplasmic reticulum (ER) can be transformed into different metabolites. (1) Ceramide (Cer) is transformed to galactosylceramide (GalCer) in the ER. (2) Cer is transported from the ER to the trans-Golgi apparatus by the ceramide transfer protein (CERT) (3) where it is transformed into sphingomyelin (SM). (4) Ceramide can also be transported from the ER to the cis-Golgi network to be transformed into glucosylceramide (GlcCer). (5) GlcCer can be transported to the trans-Golgi apparatus through vesicular transport. At the trans-Golgi apparatus, GlcCer is transformed into different glycosphingolipids (GSLs). (6) Four-phosphate adaptator protein 2 (FAPP2) can transport GlcCer back to the ER where it can be degraded to Cer by GBA2. (7) GlcCer is transported to the membrane by the glycolipid transfer protein (GLTP) where it can integrate into lipid rafts. (8) GSLs are transported to the plasma membrane through vesicular transport, (9) where they can integrate into lipid rafts. (10) GSLs are recycled in the endosome (11) to the ER or Golgi apparatus (12) or degraded by lysosomal pathway. (13) Degradation of GlcCer at the lysosome level occurs by GBA1 (inspired by [6]).

Sphingolipids

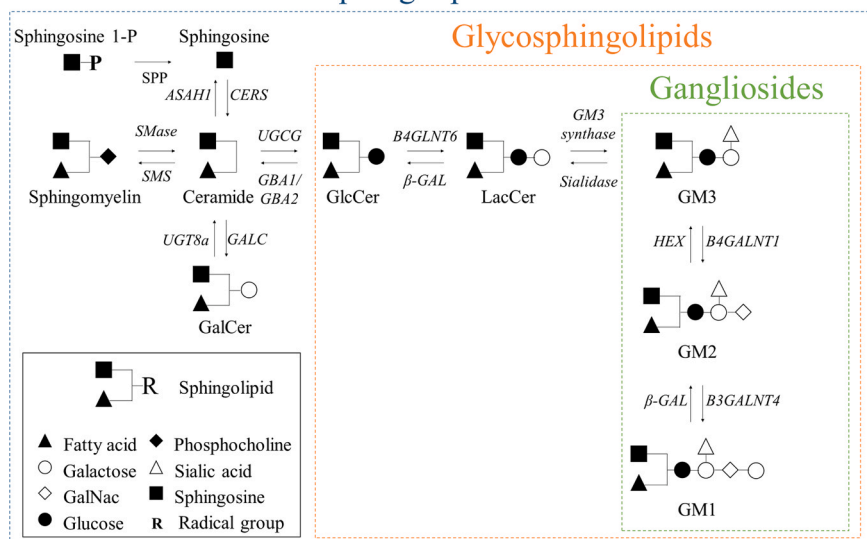


Fig. 2. Schematic representation of sphingolipid metabolism. Sphingolipids are characterized by the addition of a radical group to determine the subclass of sphingolipids; in this diagram the different classes of ceramide/dihydroceramide are not shown (e.g. hydrogen atom for ceramide is not shown). Sphingomyelin degradation by sphingomyelinase (SMase) generates ceramide (Cer). From the Cer, sphingosine and galactosylceramide (GalCer) are then synthesized by acid ceramidase (ASAHI) and ceramide galactosyltransferase (GALC) respectively. Glucosylceramide (GlcCer) is a glycosphingolipid which is obtained from Cer by glucocerebrosidase (GBA1/GBA2). The addition of galactose to GlcCer by galactosyltransferase (B4GLNT6) leads to the generation of lactosylceramide (LacCer). Then, the addition of sialic acid to LacCer forms GM3, a precursor of gangliosides. Ganglioside synthesis is continued through the successive addition of N-acetylgalactosamine (GalNAc) and galactose residues by GalNAc transferase 1 (GM2 synthase/ B4GLNT1 and B3GALNT4) respectively for GM2 and GM1.

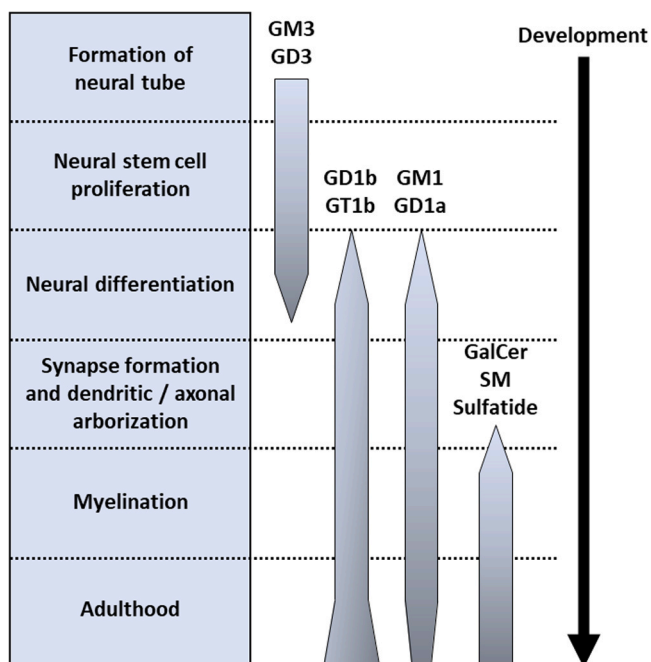


Fig. 3. Expression of gangliosides in the nervous system during development. Ganglioside composition changes during development. The gangliosides GD3 and GM3 are produced during the formation of the neural tube, and during neural stem cell proliferation. During neuronal differentiation, the expression of GD3 and GM3 decreases. In parallel, gangliosides necessary for neuronal differentiation, such as GD1a, GM1, GD1b and GT1b, are synthesized. In adulthood, the levels of GD1a and GM1 decrease while the levels of GD1b and GT1b continue to increase. Galactosylceramide (GalCer), sphingomyelin (SM) and sulfatide are synthesized during synaptogenesis. Their synthesis is maintained until adulthood (inspired by [8,11]).

(UGCG), have been shown to be essential for axonal and dendritic growth. In this regard, inhibition of ceramide synthase activity in murine neurons in vitro leads to a considerable decrease in levels of glycosphingolipid, thereby impairing neuronal growth [17,18]. Similarly, deletion of *UGCG* in the neural cells of mice causes axonal degeneration and demyelination of peripheral nerves, leading to the death [19]. In

2009, it was also shown that inhibition of *UGCG* with a shRNA led to a decrease in the expression of glycosphingolipids in mouse embryonic stem cells [20]. In sum, sphingolipids and the degradation of glucosylceramide into ceramide is crucial for the development of the nervous system.

3.3. Location and function in cell membranes of the nervous system

Cell membranes are composed of micro-domains, called lipid rafts, which are supplemented with phospholipids, cholesterol and sphingolipids. These lipid rafts play a structural role in cells but are also critical for signal transduction. Cholesterol is critical for the formation and maintenance of lipid rafts where they play a structural and anchoring role for membrane proteins involved in endocytosis, signal transduction, cell adhesion and rearrangement of the cytoskeleton [21–24]. Sphingolipids are an integral part of cell membranes where they act as modulators of signaling pathways. Ceramides, by their degree of saturation or chain length, can induce membrane gel domains, with far-reaching effects on signal transduction [25,26].

Several studies also suggest that lipid rafts, including sphingolipids, are directly responsible for the spatial organization of signaling molecules. These rafts group G proteins and neurotransmitter receptors to promote signal transmission [21]. Indeed, several sphingolipids (e.g. SM, GM1, etc.) interact with G protein coupled receptors [27–29]. Moreover, the glycosylated region of gangliosides, such as GM1, are positioned in the extracellular environment in a way that allows interaction with BDNF and NGF receptors [30–32]. Of note, GM1 is also found in the nuclear envelope of cells, where it regulates gene expression through its association with a sodium-calcium exchanger as during neuronal development [33–35]. To ensure the integrity of tissue in the nervous system, the gangliosides stabilize the myelin-associated glycoproteins (MAGs), thereby allowing strong architecture between the axon and the myelin sheath [7,36,37]. In 2005, Yamashita and collaborators reported disruptions in axon-myelin interactions in mice lacking GM3 and GM2 synthases, two enzymes that synthesize gangliosides [38]. Finally, sphingolipids are involved in cellular communication through the vesicles of endosomes and exosomes. Indeed, sphingomyelinase (SMase) for example, participates in the formation and secretion of exosome vesicles, and sphingosine kinase is involved in the regulation of endosomes [28]. All these data underline the importance of sphingolipids in cell integrity but also in the establishment of cellular communication.

3.4. GM1 and neurotrophic factors

It is widely accepted that GM1 interacts with neurotrophins, which themselves are essential for the growth and survival of neurons. GM1 is anchored to the cell membrane via its ceramide group, and 5 sugars (radical R) are exposed to the extracellular environment. The extracellular portion of GM1 can interact with growth factor receptors. Pioneering studies demonstrated that GM1 promoted the neurite growth in murine neuroblastomas [39–41]. Subsequently, a direct interaction between GM1 and the TrkA receptor, as well as the ability of GM1 to increase the activation of TrkA induced by NGF, was shown [30,42]. In the early 2000s, the enhanced effect of NGF on GM1-potentiated TrkA receptor activity was demonstrated in vivo in rats [32]. More recently, it has been proposed that this effect is due to the binding of NGF to the TrkA receptor, and activation of the Ras/Raf/MEK/Erk pathway [7]. Interestingly, cells deficient in GM1 do not express the TrkA receptor [43]. In line with these data, it has been shown that endogenous GM1 directly modulates the activity of the TrkA and TrkB receptors, as well as their associated signaling cascades [31,44,45]. GM1 also activates the various Trk receptors (A, B and C) by phosphorylation [32] and participates in the autophosphorylation of TrkC [46]. Critically, a 2002 study by Bachis and colleagues noted that GM1 appeared to prevent glutamate-related excitotoxicity by mimicking the action of BDNF on TrkB [47]. Collectively, these studies consolidate the close link between GM1 and Trk receptors in neuronal survival.

4. Sphingolipids in neurodegenerative diseases

Currently, more than 30 million people are affected by neurodegenerative diseases. Alzheimer's disease (AD) is the most common of these diseases and affects 25 million people worldwide, while 6.3 million people have Parkinson's disease (PD), and 220,000 have amyotrophic lateral sclerosis (ALS) [48]. The accumulation or dysregulation of sphingolipids is proposed to contribute to the pathogenesis of these neurodegenerative diseases by severely affecting lysosomal enzyme activity, which triggers an accumulation of lipids within organelles of the "endosomal-autophagic-lysosomal" system [49].

4.1. Amyotrophic lateral sclerosis

Amyotrophic lateral sclerosis is a fatal neurodegenerative disease that is characterized by the irrevocable degeneration of upper and lower motor neurons. The progressive loss of neurons results in debilitating motor weakness, increasing paralysis, and death usually occurs within 3–5 years from symptom onset [50]. It is becoming evident that ALS pathology extends beyond the motor system [51].

Several studies have identified metabolic alterations in ALS patients. These are mostly defined by hypermetabolism and dyslipidemia, which are clinically associated with the severity of symptoms [52–54]. Hypermetabolism is an early phenomenon that is present throughout the disease [55–57], and that appears to negatively impact prognosis [58–60]. By contrast, hyperlipidemia appears to be a protective factor in ALS, and it is known that a hyperlipidemic diet significantly improves survival of ALS mice (*Sod1^{G86R}*) [61] and slows progression of disease in ALS patients [62]. Congruent with this, high levels of total cholesterol [63] and elevated levels of serum triglyceride [64] appear to be beneficial in ALS patients. More recently, the beneficial effects of high-caloric nutrition on the survival of ALS patients with fast-progressing disease were shown [65]. While the cause for metabolic dysregulation in ALS remains unknown, consistent observations of the beneficial effects of lipids has spurred extensive research into the contribution of lipid metabolism to the disease.

An abnormal increase in sphingolipid levels (sphingomyelin, ceramide, cholesterol, etc.) has been observed in the spinal cord of ALS patients, and in the spinal cord of asymptomatic and symptomatic *SOD1^{G93A}* mice [66]. Subsequent studies confirming an increase in

metabolites of sphingolipids (glucosylceramide, ceramide, galactosylceramide, GM3, GM1, etc.) and their associated enzymes (GBA1, GBA2, etc.) in the spinal cord of *Sod1^{G86R}* mice [67], *SOD1^{G93A}* mice and ALS patients [68] indicate that altered metabolism of sphingolipids could be integral to the progression of ALS pathology [69]. Interestingly, the inhibition of glucosylceramide synthesis dramatically accelerates disease progression in *SOD1^{G93A}* mice while the intracerebroventricular perfusion of GM3 is able to significantly delay the onset of paralysis in *SOD1^{G93A}* mice [68]. While the clinical relevance of altered sphingolipid metabolism remains to be established, levels of sphingomyelin and long chain triglycerides in the CSF of ALS patients has been shown to correlate with the progression of ALS [70,71].

The detection of antibodies targeting gangliosides, in particular GM1, in serum [72,73], brain [74] and spinal cord [75] of some patients with ALS led to clinical trials of ganglioside administration. These trials have not been conclusive [76–78]. However, Xu et al. have shown that a single injection of human IgM (rHlgM12) binding gangliosides extends the survival of two ALS mouse models [79] thus emphasizing a role for glycosphingolipids in ALS.

An increase in the levels of glucosylceramide and GM1 have been observed in the CSF of ALS patients [80]. Transcriptomic studies of muscle biopsies from ALS patients have reported a significant increase in the expression of the *UGCG* gene, encoding the enzyme responsible for the synthesis of glucosylceramide. Similar to ALS patients, mouse models of ALS show accumulation of glucosylceramide in skeletal muscle. Lipidomic analysis revealed complete modulation of sphingolipids in skeletal muscles and spinal cords of *Sod1^{G86R}* mice prior to the onset of disease phenotypes. Interestingly, the levels of GM2 and GM3 were increased in *Sod1^{G86R}* mice, but also in non-transgenic mice that had undergone axotomy, suggesting that altered sphingolipid metabolic might occur as a consequence of denervation. In addition, inhibition of glucosylceramide synthesis by an *UGCG* inhibitor significantly delayed functional recovery after sciatic nerve injury [67]. Conversely, inhibition of the degradation of glucosylceramide by conduritol B epoxide, an irreversible inhibitor of GBA1 and GBA2, improved functional recovery in a model of sciatic nerve compression and slowed disease progression in *Sod1^{G86R}* mice [80]. Thus, current evidence indicates that sphingolipid metabolism is likely to be a key modulator of disease course in ALS.

4.2. Other neurological diseases

In recent years, there has been a growing body of evidence to indicate that the metabolism of sphingolipids contributes to the pathophysiology of other diseases such as Alzheimer's disease, Parkinson's disease and Gaucher disease. The gradual accumulation of protein aggregates, which leads to toxicity and neurodegeneration of cells of the nervous system, occurs in response to defects in the metabolism of sphingolipids, major players in the lysosomal degradation of cellular debris.

4.2.1. Alzheimer's disease

Alzheimer's disease (AD) is the most common neurodegenerative disease characterized by the accumulation of extracellular amyloid plaques and intraneuronal deposits of the Tau protein [81,82]. Although the molecular mechanism that underpins the development of the disease is not clear, numerous studies suggest that sphingolipids play a crucial role in the pathogenesis of AD [83–88]. Indeed, the levels of sphingolipids (e.g. sphingomyelin and ceramide) and their associated synthetic enzymes (sphingomyelin synthase and sphingomyelinase) are increased in the brains of AD mice and AD patients [89–91]. Several teams have also shown that elevated levels of serum ceramide in AD patients and AD model animals [92–96] may be predictive of AD and cognitive impairment [97–99]. Ceramide, produced by sphingomyelinases, stabilizes β -secretase, which leads to an increase in the cleavage of the amyloid precursor protein [100]. In this regard, increasing ceramide and sphingomyelinase levels promotes the accumulation of amyloid- β

(A β 42) in two mouse models of AD [90,91] and ceramide-enriched exosomes worsen disease pathology [91]. Conversely, genetic inhibition of sphingomyelinase enzyme in a mouse model of AD decreases amyloid- β deposits and cognitive impairment [91,101], and attenuates the progression of disease [91]. Furthermore, knocking out the *SMS1* gene in the hippocampus of an AD-like transgenic mouse appears to limit the formation of amyloid plaques and improve cognitive function [102].

4.2.2. Gaucher disease and parkinson's disease

Glucosylceramidase activity is critical in these two diseases, with the lysosomal enzyme (GBA1) playing a particularly important role. The loss of GBA1 activity strongly modulates lysosomal function, which is involved in the elimination of cellular material through autophagy. Mutations in the *GBA1* gene lead to non-functional enzymes, either by total loss of functions or by driving a defect in the targeting of GBA1 to the lysosome, which results in the accumulation of glucosylceramide in the lysosomes.

GBA1 mutations are associated with Gaucher disease [49,103,104]. Gaucher disease is a lysosomal overload disease characterized by a complete loss of function of the GBA1 protein. This leads to an accumulation of glucosylceramide, which can be the cause of neuropathic forms of the disease [49,105]. Although no correlation has been made between the severity of Gaucher disease and the *GBA2* gene, studies suggest that *GBA2* is involved in the pathophysiology of the disease [103,106].

Parkinson's disease is the second most common neurodegenerative disease that affects predominantly dopaminergic neurons of the substantia nigra. People with Parkinson's disease usually have motor symptoms, but the disease is also characterized by insidious cognitive decline, which increases with the duration of the disease. Mutations in the *GBA1* gene are considered a high-risk factor for Parkinson's disease. Indeed, *GBA1* mutations occur in 10–25% of patients with Parkinson's disease [104,107–112]. *GBA1* mutations cause a decrease in the levels of the GBA1 protein and its enzymatic activity, resulting in the toxic accumulation of α -synuclein and defecting autophagy [113,114]. Interestingly, activation of GBA1 induces clearance of α -synuclein and restores lysosomal function in dopaminergic neurons of Parkinson's patients [115]. Moreover, inhibition of GBA1 activity in a mouse model of synucleinopathy leads to accumulation of glucosylceramide and worsening of motor and cognitive phenotypes. By contrast, increasing the activity of GBA1 leads to a slowing of disease progression [116]. While studies suggest that the accumulation of glucosylceramide might alter the mechanisms that regulate α -synuclein [117,118], the molecular mechanisms that link GBA1 and α -synuclein are still poorly understood.

All of these data underline the importance of potential molecular targets (e.g. GBA1 and GBA2) in order to develop effective pharmacological strategies to limit the progression of these neurological diseases.

5. Therapeutic targeting of sphingolipids in neurodegeneration

With a growing body of evidence to show that sphingolipids actively participate in the pathophysiology of neurodegenerative disease [5,49,119], there is an increased focus on the targeting of the metabolism of sphingolipids for therapeutic development. Due to the complexity of the sphingolipid pathway, only the most common players in neurodegenerative diseases will be discussed.

5.1. Sphingolipids at the centre of new therapies in animal and cell models

Some molecules have proven to be promising candidates for attenuating various pathologies in *in vitro* and *in vivo* models of neurodegenerative disease.

5.1.1. Fingolimod (FTY720)

Fingolimod is an analogue of sphingosine. In the brain, it is converted to fingolimod -phosphate (FTY720-P). Fingolimod-P is a

sphingosine-1-phosphate (S1P) receptor agonist [120], which is reduced in Alzheimer's disease and Parkinson's disease [121–124]. Stimulation of S1P by FTY720 improves memory, synaptic plasticity, reduces β -amyloid production and enhances motor function in several distinct animal models of these diseases [125–128]. More recently, FTY720 was shown to significantly improve survival in the *SOD1^{G93A}* mouse model of ALS [129].

5.1.2. Isofagomine

In 2009, Liebermann and co-workers demonstrated that isofagomine by acting as a chaperone of GBA1 ensured good conformational stability and activity of human GBA1 while also increasing its enzymatic activity [130] (see Table 1). In support of this, isofagomine was shown to extend the lifespan in animal models of Gaucher disease and increase the activity and levels of GBA1 in the brain and visceral tissues, although its effects on the accumulation of glucosylceramide and glycosphingosine is yet to be determined [131]. Finally, isofagomine reduces aggregates of human wildtype α -synuclein in dopaminergic neurons in the substantia nigra of a mouse model of synucleinopathy [132]. In general, current data on the use of isofagomine in Gaucher disease and synucleinopathies is encouraging.

5.1.3. Ambroxol

Ambroxol is a molecule known for its mucolytic, antioxidant and anaesthetic properties [133]. It can cross the blood-brain barrier (BBB) and improve the activity of GBA1 in Gaucher disease patient cells *in vitro*, and in wildtype mice *in vivo* [134] (see Table 1). In 2009, Maegawa and collaborators demonstrated that ambroxol stabilizes the conformation of GBA1, thereby increasing its enzymatic activity, and reducing the accumulation of glucosylceramide in fibroblasts from Gaucher disease patients [135]. Ambroxol has also been shown to enhance the activity of GBA1 in fibroblasts of Parkinson's disease patients, and to reduce the aggregation of α -synuclein [114,136]. In wildtype and transgenic mice overexpressing human α -synuclein, treatment with ambroxol led to an increase in GBA1 mRNA and enzyme activity in the brain, and a concomitant reduction in levels of α -synuclein [137]. Finally, the activities of GBA1 and β -hexosaminidase are significantly improved in non-human primates after treatment with ambroxol [138]. In the context of ALS, our laboratory has demonstrated that ambroxol prevents the loss of muscle strength, delays disease progression, and extends survival of *Sod1^{G86R}* mice. Moreover, ambroxol promotes axonal growth and neuronal network complexity *in vitro* and *in vivo* [139,140]. Overall, ambroxol appears to be highly effective in modulating GBA1 activity to exert neuroprotective effects across a number of mouse models of neurodegeneration.

5.2. Sphingolipid modulation in humans

There is growing interest in targeting the metabolism of sphingolipids as a therapeutic strategy in human neurodegenerative diseases. In this section, we will only present clinical studies for Gaucher disease and Parkinson's disease.

5.2.1. Isofagomine

The defective activity of GBA1 in patients with Gaucher disease can be restored by isofagomine treatment. By stabilizing the binding of the GBA1 enzyme with its substrate [141], isofagomine increases GBA1 activity in Gaucher disease patient fibroblasts, thus preventing the toxic accumulation of glucosylceramide [130]. However, isofagomine has poor cell penetration and clinical trials for Gaucher disease have been inconclusive.

5.2.2. Ambroxol

In 2013, Zimran et al. showed that ambroxol was safe and had no adverse effects in patients with Gaucher disease [142]. In another pilot study, ambroxol was found to improve motor functions and neurological

Table 1
Summary of studies using treatments that modulate the metabolism of sphingolipids.

Drug	Disease	Model	Dose	Effects	References
Fingolimod (FTY720)	AD	Mouse	0.5 mg/kg	• Reduces amyloid- β production	[127]
	AD-like neurodegeneration	Rats	1 mg/kg	• Attenuates A β 42-induced learning and memory impairment	[125]
	AD-like neurodegeneration	Rats	1 mg/kg	• Reduces inflammatory markers	[126]
	–	Chemical model of neurodegeneration	10–1000 nM	• Attenuates excitotoxicity and neuroinflammation	[145]
	–	Rats (Focal cerebral ischemia)	0.5 mg/kg	• Improves synaptic plasticity and memory deficit	[143]
–	PD	Mouse	1 mg/kg	• Attenuates motor deficits	[146]
	ALS	Mouse	0.1 and 1 mg/kg	• Reduces the loss of dopaminergic neurons	[129]
	ALS	Mouse	0.1 and 1 mg/kg	• Improves neurological scores and survival	[129]
Isofagomine	GD	Patients lysosomal enzyme (GBA1)	–	• Ensure good conformation and stability	[130]
	GD	Mouse	20 or 600 mg/kg	• Increases GBA1 enzymatic activity	[147]
	α -synucleinopathy	Mouse	100 mg/kg	• Increases the activity and levels of GBA1	[132]
Ambroxol	GD	Patients fibroblasts	20–125 μ M	• Extends survival	[135]
	GD	Patients fibroblasts	0.3–10 mM	• Reduces aggregates in dopaminergic neurons	[134]
	GD	Patients	150 mg/d	• Ensure good conformation and stability	[142]
	GD	Patients fibroblasts	60 μ M	• Increases GBA1 enzymatic activity	[136]
	GD	Patients	1.3 g/d	• Reduces GlcCer storage	[143]
	GD	Patients	21 mg/kg/d	• Improves the activity of GBA1	[148]
	GD	Patients	27 mg/kg/d	• Neither toxicity in vivo	[148]
	PD	Mouse	4 mM	• Safe	[137]
	PD	Neural crest stem cells line from PD and GBA1 mutations patients	60 μ M	• No adverse effects	[114]
	PD	Non-human primate	100 mg/d	• Reduces ROS production	[138]
	PD	Patients	1.23 g/d	• Improves the activity of GBA1	[144]
	ALS	Mouse	3 mM	• Increases the activity of GBA1	[139]

AD: Alzheimer's disease, ALS: amyotrophic lateral sclerosis, BBB: blood brain barrier, GBA: glucocerebrosidase, GD: Gaucher disease, GlcCer: glucosylceramide, PD: Parkinson's disease, ROS: reactive oxygen species

symptoms by decreasing myoclonus in Gaucher disease patients [143]. Recently, a phase II clinical trial, which aimed to assess the therapeutic potential of ambroxol in Parkinson's disease patients with or without *GBA1* mutations was carried out [144] (NCT02941822). Interim results for this trial are positive. Seventeen patients (8 with *GBA1* mutations, 9 without) received increasing doses of ambroxol (up to 420 mg, 3 times a day at the end of the study). Ambroxol did not cause any specific side effects and was well tolerated by patients. Given that ambroxol crosses BBB [144], this molecule seems to be a promising therapeutic candidate for the treatment of diseases that are characterized by defective *GBA1* enzyme activity in parallel with the accumulation of misfolded proteins.

6. Conclusion

The pathophysiological mechanisms that underpin neurodegeneration are complex. The deregulation of sphingolipid metabolism appears to be common across a number of neurodegenerative diseases, yet the mechanisms by which the dysregulation in sphingolipid metabolism contributes to the degenerative process remains to be elucidated. While the targeting of sphingolipid metabolism has reached clinical stages for Parkinson's disease, this is yet to occur in Alzheimer's disease

or ALS. Nonetheless, results in mouse models of neurodegenerative diseases, and in particular ALS, highlight that the modulation of sphingolipid metabolism may be beneficial. Future studies that clarify the role of sphingolipids in neurodegeneration, with the view to translate this knowledge into clinical trials and effective drug treatments are desperately needed.

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Author contributions

Alexandra Bouscary and Cyril Quessada did the literature search and wrote the article. All authors contributed to the critical revision of the manuscript.

Patents

Henriques, Loeffler and Spedding hold a patent entitled “Inhibitors of Glucosylceramide Degradation in the Treatment of Diseases of the Motor Units”.

Conflicts of interest

The authors declare no conflict of interest.

References

- [1] J. Tamiji, D.A. Crawford, The neurobiology of lipid metabolism in autism spectrum disorders, *NeuroSignals* 18 (2011) 98–112, <https://doi.org/10.1159/000323189>.
- [2] C. Fasano, A. Hiol, J.P. Miolan, J.P. Niel, Les sphingolipides: vecteurs d'agents pathogènes et cause de maladies génétiques, *Med. Sci.* 22 (2006) 411–415, <https://doi.org/10.1051/medsci/2006224411>.
- [3] G. D'Angelo, S. Capasso, L. Sticco, D. Russo, Glycosphingolipids: synthesis and functions, *FEBS J.* 280 (2013) 6338–6353, <https://doi.org/10.1111/febs.12559>.
- [4] G. D'Angelo, T. Uemura, C.C. Chuang, E. Polishchuk, M. Santoro, H. Ohvo-Rekilä, T. Sato, G. Di Tullio, A. Varriale, S. D'Auria, T. Daniele, F. Capuani, L. Johannes, P. Mattjus, M. Monti, P. Pucci, R.L. Williams, J.E. Burke, F.M. Platt, A. Harada, M. A. De Matteis, Vesicular and non-vesicular transport feed distinct glycosylation pathways in the Golgi, *Nature* 501 (2013) 116–120, <https://doi.org/10.1038/nature12423>.
- [5] T.J. Tracey, F.J. Steyn, E.J. Wolvetang, S.T. Ngo, Neuronal lipid metabolism: multiple pathways driving functional outcomes in health and disease, *Front. Mol. Neurosci.* 11 (2018) 1–25, <https://doi.org/10.3389/fnmol.2018.00010>.
- [6] N. Baumann, D. Pham-Dinh, Biology of oligodendrocyte and myelin in the mammalian central nervous system, *Physiol. Rev.* 81 (2001) 871–927, <https://doi.org/10.1152/physrev.2001.81.2.871>.
- [7] A.S.B. Olsen, N.J. Færgeman, Sphingolipids: membrane microdomains in brain development, function and neurological diseases, *Open Biol.* 7 (2017), <https://doi.org/10.1098/rsob.170069>.
- [8] N. Schaeren-Wiemers, P. Van Der Bijl, M.E. Schwab, The UDP-galactose:ceramide galactosyltransferase: expression pattern in oligodendrocytes and schwann cells during myelination and substrate preference for hydroxyceramide, *J. Neurochem.* 65 (1995) 2267–2278, <https://doi.org/10.1046/j.1471-4159.1995.65052267.x>.
- [9] R.K. Yu, Y.T. Tsai, T. Ariga, Functional roles of gangliosides in Neurodevelopment: an overview of recent advances, *Neurochem. Res.* 37 (2012) 1230–1244, <https://doi.org/10.1007/s11064-012-0744-y>.
- [10] R.K. Yu, Y. Nakatani, M. Yanagisawa, The role of glycosphingolipid metabolism in the developing brain, *J. Lipid Res.* 50 (2009) 440–445, <https://doi.org/10.1194/jlr.R800028-JLR200>.
- [11] R.K. Yu, L.J. Macala, T. Taki, H.M. Weinfeld, F.S. Yu, Developmental changes in ganglioside composition and synthesis in embryonic rat brain, *J. Neurochem.* 50 (1988) 1825–1829, <https://doi.org/10.1111/j.1471-4159.1988.tb02484.x>.
- [12] K. Palmano, A. Rowan, R. Guillerimo, J. Guan, P. McJarrow, The role of gangliosides in neurodevelopment, *Nutrients* 7 (2015) 3891–3913, <https://doi.org/10.3390/nu7053891>.
- [13] J. Wang, A. Cheng, C. Wakade, R.K. Yu, Ganglioside GD3 is required for neurogenesis and long-term maintenance of neural stem cells in the postnatal mouse brain, *J. Neurosci.* 34 (2014) 13790–13800, <https://doi.org/10.1523/JNEUROSCI.2275-14.2014>.
- [14] V.T. Ribeiro-Resende, T.A. Gomes, S. De Lima, M. Nascimento-Lima, M. Bargas-Rega, M.F. Santiago, R.A. De Melo Reis, F.G. De Mello, Mice lacking GD3 synthase display morphological abnormalities in the sciatic nerve and neuronal disturbances during peripheral nerve regeneration, *PLoS One* 9 (2014) 1–12, <https://doi.org/10.1371/journal.pone.0108919>.
- [15] M. Aureli, M. Samarani, V. Mordica, L. Mauri, N. Loberto, R. Bassi, A. Prinetti, S. Sonnino, Gangliosides and cell surface ganglioside glycohydrolases in the nervous system, *Adv. Neurobiol.* (2014) 223–244, https://doi.org/10.1007/978-1-4939-1154-7_10.
- [16] M. Aureli, N. Loberto, P. Lanteri, V. Chigorno, A. Prinetti, S. Sonnino, Cell surface sphingolipid glycohydrolases in neuronal differentiation and aging in culture, *J. Neurochem.* 116 (2011) 891–899, <https://doi.org/10.1111/j.1471-4159.2010.07019.x>.
- [17] R. Harel, A.H. Futerman, Inhibition of sphingolipid synthesis affects axonal outgrowth in cultured hippocampal neurons, *J. Biol. Chem.* 268 (1993) 14476–14481.
- [18] S. Furuya, K. Ono, Y. Hirabayashi, Sphingolipid biosynthesis is necessary for dendrite growth and survival of cerebellar purkinje cells in culture, *J. Neurochem.* 65 (1995) 1551–1561, <https://doi.org/10.1046/j.1471-4159.1995.65041551.x>.
- [19] R. Jenemann, R. Sandhoff, S. Wang, E. Kiss, N. Gretz, C. Zuliani, A. Martin-Villalba, R. Jäger, H. Schorle, M. Kenzelmann, M. Bonrouhi, H. Wiegandt, H. J. Gröne, Cell-specific deletion of glucosylceramide synthase in brain leads to severe neural defects after birth, *Proc. Natl. Acad. Sci. U. S. A.* 102 (2005) 12459–12464, <https://doi.org/10.1073/pnas.0500893102>.
- [20] J.U. Jung, K. Ko, D.H. Lee, K. Ko, K.T. Chang, Y.K. Choo, The roles of glycosphingolipids in the proliferation and neural differentiation of mouse embryonic stem cells, *Exp. Mol. Med.* 41 (2009) 935–945, <https://doi.org/10.3858/emmm.2009.41.12.099>.
- [21] J.A. Allen, R.A. Halverson-Tamboli, M.M. Rasenick, Lipid raft microdomains and neurotransmitter signalling, *Nat. Rev. Neurosci.* 8 (2007) 128–140, <https://doi.org/10.1038/nrn2059>.
- [22] G. Van Meer, D.R. Voelker, G.W. Feigenson, Membrane lipids: where they are, *Nat. Rev. Mol. Cell Biol.* 101 (2009) 1–4, <https://doi.org/10.1038/nrm2330>.
- [23] A. Prinetti, N. Loberto, V. Chigorno, S. Sonnino, Glycosphingolipid behaviour in complex membranes, *Biochim. Biophys. Acta - Biomembr.* 1788 (2009) 184–193, <https://doi.org/10.1016/j.bbmem.2008.09.001>.
- [24] E. Posse de Chaves, S. Sipione, Sphingolipids and gangliosides of the nervous system in membrane function and dysfunction, *FEBS Lett.* 584 (2010) 1748–1759, <https://doi.org/10.1016/j.febslet.2009.12.010>.
- [25] S.N. Pinto, L.C. Silva, A.H. Futerman, M. Prieto, Effect of ceramide structure on membrane biophysical properties: the role of acyl chain length and unsaturation, *Biochim. Biophys. Acta Biomembr.* 1808 (2011) 2753–2760, <https://doi.org/10.1016/j.bbmem.2011.07.023>.
- [26] B.M. Castro, M. Prieto, L.C. Silva, Ceramide: a simple sphingolipid with unique biophysical properties, *Prog. Lipid Res.* 54 (2014) 53–67, <https://doi.org/10.1016/j.plipres.2014.01.004>.
- [27] E. Kiyokawa, T. Baba, N. Otsuka, A. Makino, S. Ohno, T. Kobayashi, Spatial and functional heterogeneity of sphingolipid-rich membrane domains, *J. Biol. Chem.* 280 (2005) 24072–24084, <https://doi.org/10.1074/jbc.M502244200>.
- [28] M. Trayssac, Y.A. Hannun, L.M. Obeid, Role of sphingolipids in senescence: Implication in aging and age-related diseases, *J. Clin. Invest.* 128 (2018) 2702–2712, <https://doi.org/10.1172/JCI97949>.
- [29] R. Ledeen, G. Wu, Gangliosides of the nervous system, 2018. https://doi.org/10.1007/978-1-4939-8552-4_2.
- [30] S.J. Rabin, I. Mocchetti, G.M.1 Ganglioside, Activates the high-affinity nerve growth factor receptor trkA, *J. Neurochem.* 65 (1995) 347–354, <https://doi.org/10.1046/j.1471-4159.1995.65010347.x>.
- [31] M. Pitto, T. Mutoh, M. Kuriyama, A. Ferraretto, P. Palestini, M. Masserini, Influence of endogenous GM1 ganglioside on TrkB activity, in cultured neurons, *FEBS Lett.* 439 (1998) 93–96, [https://doi.org/10.1016/S0014-5793\(98\)01344-1](https://doi.org/10.1016/S0014-5793(98)01344-1).
- [32] A.M. Duchemin, Q. Ren, L. Mo, N.H. Neff, M. Hadjiconstantinou, GM1 ganglioside induces phosphorylation and activation of Trk and Erk in brain, *J. Neurochem.* 81 (2002) 696–707, <https://doi.org/10.1046/j.1471-4159.2002.00831.x>.
- [33] G. Wu, X. Xie, Z.H. Lu, R.W. Ledeen, Sodium-calcium exchanger complexed with GM1 ganglioside in nuclear membrane transfers calcium from nucleoplasm to endoplasmic reticulum, *Proc. Natl. Acad. Sci. U. S. A.* 106 (2009) 10829–10834, <https://doi.org/10.1073/pnas.0903408106>.
- [34] Y.-T. Tsai, Y. Itokazu, R.K. Yu, GM1 ganglioside is involved in epigenetic activation loci of neuronal cells, *Neurochem. Res.* 41 (2016) 107–115, <https://doi.org/10.1007/s11064-015-1742-7>.
- [35] X. Xie, G. Wu, Z.H. Lu, R.W. Ledeen, Potentiation of a sodium-calcium exchanger in the nuclear envelope by nuclear GM1 ganglioside, *J. Neurochem.* 81 (2002) 1185–1195, <https://doi.org/10.1046/j.1471-4159.2002.00917.x>.
- [36] B. Pan, S.E. Fromholt, E.J. Hess, T.O. Crawford, J.W. Griffin, K.A. Sheikh, R. L. Schnaar, Myelin-associated glycoprotein and complementary axonal ligands, gangliosides, mediate axon stability in the CNS and PNS: neuropathology and behavioral deficits in single- and double-null mice, *Exp. Neurol.* 195 (2005) 208–217, <https://doi.org/10.1016/j.expneurol.2005.04.017>.
- [37] P.H. Lopez, R.L. Schnaar, Gangliosides in cell recognition and membrane protein regulation, *Curr. Opin. Struct. Biol.* 19 (2009) 549–557, <https://doi.org/10.1016/j.sbi.2009.06.001>.
- [38] T. Yamashita, Y.P. Wu, R. Sandhoff, N. Werth, H. Mizukami, J.M. Ellis, J. L. Dupreell, R. Geyer, K. Sandhoff, R.L. Proia, Interruption of ganglioside synthesis produces central nervous system degeneration and altered axon-glia interactions, *Proc. Natl. Acad. Sci. U. S. A.* 102 (2005) 2725–2730, <https://doi.org/10.1073/pnas.0407785102>.
- [39] F.J. Roisen, H. Bartfeld, R. Nagele, G. Yorke, Ganglioside stimulation of axonal sprouting in vitro, *Science* 214 (1981) 577–578, <https://doi.org/10.1126/science.7291999>.
- [40] M.C. Byrne, R.W. Ledeen, F.J. Roisen, G. Yorke, J.R. Sclafani, Ganglioside-induced neurogenesis: verification that gangliosides are the active agents, and comparison of molecular species, *J. Neurochem.* 41 (1983) 1214–1222, <https://doi.org/10.1111/j.1471-4159.1983.tb00814.x>.
- [41] A. Ferreira, J. Busciglio, C. Landa, A. Caceres, Ganglioside-enhanced neurite growth: evidence for a selective induction of high-molecular-weight MAP-2, *J. Neurosci.* 10 (1990) 293–302, <https://doi.org/10.1523/jneurosci.10-01-00293.1990>.

- [42] T. Mutoh, A. Tokuda, T. Miyadai, M. Hamaguchi, N. Fujiki, Ganglioside GM1 binds to the Trk protein and regulates receptor function, *Proc. Natl. Acad. Sci. U. S. A.* 92 (1995) 5087–5091, <https://doi.org/10.1073/pnas.92.11.5087>.
- [43] T. Mutoh, T. Hamano, S. Yano, H. Koga, H. Yamamoto, K. Furukawa, R. W. Ledeen, Stable transfection of GM1 synthase gene into GM1-deficient NG108-15 cells, CR-72 cells, rescues the responsiveness of Trk-neurotrophin receptor to its ligand, NGF, *Neurochem. Res* 27 (2002) 801–806, <https://doi.org/10.1023/A:1020209008169>.
- [44] S. Fukumoto, T. Mutoh, T. Hasegawa, H. Miyazaki, M. Okada, G. Goto, K. Furukawa, T. Urano, K. Furukawa, GD3 synthase gene expression in PC12 cells results in the continuous activation of TrkA and ERK1/2 and enhanced proliferation, *J. Biol. Chem.* 275 (2000) 5832–5838, <https://doi.org/10.1074/jbc.275.8.5832>.
- [45] K. Furukawa, Y. Ohmi, Y. Ohkawa, O. Tajima, K. Furukawa, Glycosphingolipids in the regulation of the nervous system, 2014, pp. 307–320. https://doi.org/10.1007/978-1-4939-1154-7_14.
- [46] S.J. Rabin, A. Bachis, I. Mocchi, Gangliosides activate Trk receptors by inducing the release of neurotrophins, *J. Biol. Chem.* 277 (2002) 49466–49472, <https://doi.org/10.1074/jbc.M203240200>.
- [47] A. Bachis, S.J. Rabin, M.D. Fiocco, I. Mocchi, Gangliosides prevent excitotoxicity through activation of TrkB receptor, *Neurotox. Res.* 4 (2002) 225–234, <https://doi.org/10.1080/10298420290015836>.
- [48] K.C. Arthur, A. Calvo, T.R. Price, J.T. Geiger, A. Chiò, B.J. Traynor, Projected increase in amyotrophic lateral sclerosis from 2015 to 2040, *Nat. Commun.* 7 (2016) 1–6, <https://doi.org/10.1038/ncomms12408>.
- [49] J.C. Dodge, Lipid involvement in neurodegenerative diseases of the motor system: insights from lysosomal storage diseases, *Front. Mol. Neurosci.* 10 (2017) 1–14, <https://doi.org/10.3389/fnmol.2017.00356>.
- [50] M.A. van Es, O. Hardiman, A. Chio, A. Al-Chalabi, R.J. Pasterkamp, J.H. Veldink, L.H. van den Berg, Amyotrophic lateral sclerosis, *Lancet* 390 (2017) 2084–2098, [https://doi.org/10.1016/S0140-6736\(17\)31287-4](https://doi.org/10.1016/S0140-6736(17)31287-4).
- [51] R.M. Ahmed, M. Irish, O. Piguet, G.M. Halliday, L.M. Ittner, S. Farooqi, J. R. Hodges, M.C. Kiernan, Amyotrophic lateral sclerosis and frontotemporal dementia: distinct and overlapping changes in eating behaviour and metabolism, *Lancet Neurol.* 15 (2016) 332–342, [https://doi.org/10.1016/S1474-4422\(15\)00380-4](https://doi.org/10.1016/S1474-4422(15)00380-4).
- [52] J.C. Desport, P.M. Preux, L. Magy, Y. Boirie, J.M. Vallat, B. Beaufrère, P. Couratier, Factors correlated with hypermetabolism in patients with amyotrophic lateral sclerosis, *Am. J. Clin. Nutr.* 74 (2001) 328–334, <https://doi.org/10.1093/ajcn/74.3.328>.
- [53] P.F. Pradat, G. Bruneteau, P.H. Gordon, L. Dupuis, D. Bonnefont-Rousselot, D. Simon, F. Salachas, P. Corcia, V. Frochet, J.M. Lacorte, C. Jardel, C. Coussieu, N. Le Forestier, L. Lacomblez, J.P. Loeffler, V. Meininger, Impaired glucose tolerance in patients with amyotrophic lateral sclerosis, *Amyotroph. Lateral Scler.* 11 (2010) 166–171, <https://doi.org/10.3109/17482960902822960>.
- [54] L. Dupuis, P.F. Pradat, A.C. Ludolph, J.P. Loeffler, Energy metabolism in amyotrophic lateral sclerosis, *Lancet Neurol.* 10 (2011) 75–82, [https://doi.org/10.1016/S1474-4422\(10\)70224-6](https://doi.org/10.1016/S1474-4422(10)70224-6).
- [55] C. Bouteloup, J.C. Desport, P. Clavelou, N. Guy, H. Derumeaux-Burel, A. Ferrier, P. Couratier, Hypermetabolism in ALS patients: an early and persistent phenomenon, *J. Neurol.* 256 (2009) 1236–1242, <https://doi.org/10.1007/s00415-009-5100-z>.
- [56] B. Funalot, J.C. Desport, F. Sturtz, W. Camu, P. Couratier, High metabolic level in patients with familial amyotrophic lateral sclerosis, *Amyotroph. Lateral Scler.* 10 (2009) 113–117, <https://doi.org/10.1080/17482960802295192>.
- [57] R.S. Peter, A. Rosenbohm, L. Dupuis, T. Brehme, J. Kassubek, D. Rothenbacher, G. Nagel, A.C. Ludolph, Life course body mass index and risk and prognosis of amyotrophic lateral sclerosis: results from the ALS registry Swabia, *Eur. J. Epidemiol.* 32 (2017) 901–908, <https://doi.org/10.1007/s10654-017-0318-z>.
- [58] L. Dupuis, P. Corcia, A. Fergani, J.-L. Gonzalez De Aguilar, D. Bonnefont-Rousselot, R. Bittar, D. Seilhean, J.-J. Hauw, L. Lacomblez, J.-P. Loeffler, V. Meininger, Dyslipidemia is a protective factor in amyotrophic lateral sclerosis, *Neurology* 70 (2008) 1004–1009, <https://doi.org/10.1212/01.wnl.0000285080.70324.27>.
- [59] P. Jésus, P. Fayemendy, M. Nicol, G. Lautrette, H. Sourisseau, P.M. Preux, J. C. Desport, B. Marin, P. Couratier, Hypermetabolism is a deleterious prognostic factor in patients with amyotrophic lateral sclerosis, *Eur. J. Neurol.* 25 (2018) 97–104, <https://doi.org/10.1111/ene.13468>.
- [60] F.J. Steyn, Z.A. Ioannides, R.P.A. Van Eijk, S. Heggie, K.A. Thorpe, A. Ceslis, S. Heshmat, A.K. Henders, N.R. Wray, L.H. Van Den Berg, R.D. Henderson, P. A. McCombe, S.T. Ngo, Hypermetabolism in ALS is associated with greater functional decline and shorter survival, *J. Neurol. Neurosurg. Psychiatry* 89 (2018) 1016–1023, <https://doi.org/10.1136/jnnp-2017-317887>.
- [61] L. Dupuis, H. Oudart, F. René, J.L. Gonzalez De Aguilar, J.P. Loeffler, Evidence for defective energy homeostasis in amyotrophic lateral sclerosis: benefit of a high-energy diet in a transgenic mouse model, *Proc. Natl. Acad. Sci. U. S. A.* 101 (2004) 11159–11164, <https://doi.org/10.1073/pnas.0402026101>.
- [62] J. Dorst, J. Cypionka, A.C. Ludolph, High-caloric food supplements in the treatment of amyotrophic lateral sclerosis: a prospective interventional study, *Amyotroph. Lateral Scler. Front. Degener.* 14 (2013) 533–536, <https://doi.org/10.3109/21678421.2013.823999>.
- [63] R.M. Ahmed, E. Highton-Williamson, J. Caga, N. Thornton, E. Ramsey, M. Zoing, W.S. Kim, G.M. Halliday, O. Piguet, J.R. Hodges, I.S. Farooqi, M.C. Kiernan, Lipid metabolism and survival across the frontotemporal dementia-amyotrophic lateral sclerosis spectrum: relationships to eating behavior and cognition, *J. Alzheimer's Dis.* 61 (2017) 773–783, <https://doi.org/10.3233/JAD-170660>.
- [64] R. Huang, X. Guo, X. Chen, Z. Zheng, Q. Wei, B. Cao, Y. Zeng, H. Shang, The serum lipid profiles of amyotrophic lateral sclerosis patients: a study from southwest China and a meta-analysis, *Amyotroph. Lateral Scler. Front. Degener.* 16 (2015) 359–365, <https://doi.org/10.3109/21678421.2015.1047454>.
- [65] A.C. Ludolph, J. Dorst, J. Dreyhaupt, J.H. Weishaupt, J. Kassubek, U. Weiland, T. Meyer, S. Petri, A. Hermann, A. Emmer, J. Grosskreutz, T. Grehl, D. Zeller, M. Boentert, B. Schrank, J. Prudlo, A.S. Winkler, S. Gorbulev, F. Roselli, J. Schuster, L. Dupuis, Effect of high-caloric nutrition on survival in amyotrophic lateral sclerosis, *Ann. Neurol.* 87 (2020) 206–216, <https://doi.org/10.1002/ana.25661>.
- [66] R.G. Cutler, W.A. Pedersen, S. Camandola, J.D. Rothstein, M.P. Mattson, Evidence that accumulation of ceramides and cholesterol esters mediates oxidative stress – induced death of motor neurons in amyotrophic lateral sclerosis, *Ann. Neurol.* 52 (2002) 448–457, <https://doi.org/10.1002/ana.10312>.
- [67] A. Henriques, V. Croixmarie, D.A. Priestman, A. Rosenbohm, S. Dirrig-Grosch, E. D'Ambra, M. Huebeker, G. Hussain, C. Boursier-Neyret, A. Echaniz-Laguna, A. C. Ludolph, F.M. Platt, B. Walther, M. Spedding, J.P. Loeffler, J.L.G. De Aguilar, Amyotrophic lateral sclerosis and denervation alter sphingolipids and up-regulate glucosylceramide synthase, *Hum. Mol. Genet.* 24 (2015) 7390–7405, <https://doi.org/10.1093/hmg/ddv439>.
- [68] J.C. Dodge, C.M. Treleaven, J. Pacheco, S. Cooper, C. Bao, M. Abraham, M. Cromwell, S.P. Sardi, W.L. Chuang, R.L. Sidman, S.H. Cheng, L. S. Shihabuddin, Glycosphingolipids are modulators of disease pathogenesis in amyotrophic lateral sclerosis, *Proc. Natl. Acad. Sci. U. S. A.* 112 (2015) 8100–8105, <https://doi.org/10.1073/pnas.1508767112>.
- [69] A. Henriques, V. Croixmarie, A. Bouscary, A. Mosbach, C. Keime, C. Boursier-Neyret, B. Walter, M. Spedding, J.P. Loeffler, Sphingolipid metabolism is dysregulated at transcriptomic and metabolic levels in the spinal cord of an animal model of amyotrophic lateral sclerosis, *Front. Mol. Neurosci.* 10 (2018) 1–14, <https://doi.org/10.3389/fnmol.2017.00433>.
- [70] H. Blasco, C. Veyrat-Durebex, C. Bocca, F. Patin, P. Vourc'h, J. Kouassi Nzoughet, G. Lenaers, C.R. Andres, G. Simard, P. Corcia, P. Reynier, Lipidomics reveals cerebrospinal-fluid signatures of ALS, *Sci. Rep.* 7 (2017) 1–10, <https://doi.org/10.1038/s41598-017-17389-9>.
- [71] H. Blasco, F. Patin, A. Descat, G. Garçon, P. Corcia, P. Gelé, T. Lenglet, P. Bede, V. Meininger, D. Devos, J.F. Gossens, P.F. Pradat, A pharmacometabonomics approach in a clinical trial of ALS: Identification of predictive markers of progression, *PLoS One* 13 (2018) 1–14, <https://doi.org/10.1371/journal.pone.0198116>.
- [72] A. Pestronk, R.N. Adams, L. Clawson, D. Cornblath, R.W. Kuncel, D. Griffin, D. B. Drachman, Serum antibodies to GM1 ganglioside in amyotrophic lateral sclerosis, *Neurology* 38 (1988), <https://doi.org/10.1212/WNL.38.9.1457>.
- [73] A. Pestronk, R.N. Adams, D. Cornblath, R.W. Kuncel, D.B. Drachman, L.C. Rn, Patterns of serum IgM antibodies to GM1 and GD1a gangliosides in amyotrophic lateral sclerosis, *Ann. Neurol.* 25 (1989) 98–102, <https://doi.org/10.1002/ana.410250118>.
- [74] M.M. Rapport, H. Donnenfeld, W. Brunner, B. Hungund, H. Bartfeld, Ganglioside patterns in amyotrophic lateral sclerosis brain regions, *Ann. Neurol.* 18 (1985) 60–67, <https://doi.org/10.1002/ana.410180111>.
- [75] G. Dawson, K. Stefansson, Gangliosides of human spinal cord: aberrant composition of cords from patients with amyotrophic lateral sclerosis, *J. Neurosci. Res.* 12 (1984) 213–220, <https://doi.org/10.1002/jnr.490120209>.
- [76] W.G. Bradley, Double-blind controlled trial of purified brain gangliosides in amyotrophic lateral sclerosis and experience with peripheral neuropathies, 1984, pp. 565–573. https://doi.org/10.1007/978-1-4684-1200-0_47.
- [77] M. Hallett, H. Harrington, H.R. Tyler, T. Flood, N. Slater, Trials of ganglioside therapy for amyotrophic lateral sclerosis and diabetic neuropathy, 1984, pp. 575–579. https://doi.org/10.1007/978-1-4684-1200-0_48.
- [78] H. Harrington, M. Hallett, H.R. Tyler, Ganglioside therapy for amyotrophic lateral sclerosis: a double-blind controlled trial, *Neurology* 34 (1984), <https://doi.org/10.1212/WNL.34.8.1083>.
- [79] X. Xu, A. Denic, L.R. Jordan, N.J. Wittenberg, A.E. Warrington, B. Woolta, L. M. Papke, L.J. Zocklein, D. Yoo, J. Shaver, S.H. Oh, L.R. Pease, M. Rodriguez, A natural human IgM that binds to gangliosides is therapeutic in murine models of amyotrophic lateral sclerosis, *DMM Dis. Model. Mech.* 8 (2015) 831–842, <https://doi.org/10.1242/dmm.020727>.
- [80] A. Henriques, M. Huebeker, H. Blasco, C. Keime, C.R. Andres, P. Corcia, D. A. Priestman, F.M. Platt, M. Spedding, J.P. Loeffler, Inhibition of β -glucocerebrosidase activity preserves motor unit integrity in a mouse model of amyotrophic lateral sclerosis, *Sci. Rep.* 7 (2017) 1–15, <https://doi.org/10.1038/s41598-017-05313-0>.
- [81] K. Blennow, H. Hampel, M. Weiner, H. Zetterberg, Cerebrospinal fluid and plasma biomarkers in Alzheimer disease, *Nat. Rev. Neurol.* 6 (2010) 131–144, <https://doi.org/10.1038/nrneuro.2010.4>.
- [82] A.M. Fagan, D. Head, A.R. Shah, D. Marcus, M. Mintun, J.C. Morris, D. M. Holtzman, Decreased cerebrospinal fluid A β 42 correlates with brain atrophy in cognitively normal elderly, *Ann. Neurol.* 65 (2009) 176–183, <https://doi.org/10.1002/ana.21559>.
- [83] T. Hartmann, J. Kuchenbecker, M.O.W. Grimm, Alzheimer's disease: the lipid connection, *J. Neurochem.* 103 (Suppl) (2007) 159–170, <https://doi.org/10.1111/j.1471-4159.2007.04715.x>.
- [84] K. Czubowicz, H. Jęsko, P. Wencel, W.J. Lukiw, R.P. Strosznajder, The role of ceramide and sphingosine-1-phosphate in Alzheimer's disease and other neurodegenerative disorders, *Mol. Neurobiol.* 56 (2019) 5436–5455, <https://doi.org/10.1007/s12035-018-1448-3>.

- [85] G. Van Echten-Deckert, J. Walter, Sphingolipids: critical players in Alzheimer's disease, *Prog. Lipid Res.* 51 (2012) 378–393, <https://doi.org/10.1016/j.plipres.2012.07.001>.
- [86] M. Trayssac, Y.A. Hannun, L.M. Obeid, Role of sphingolipids in senescence: Implication in aging and age-related diseases, *J. Clin. Invest.* 128 (2018) 2702–2712, <https://doi.org/10.1172/JCI97949>.
- [87] M. Kosicek, H. Zetterberg, N. Andreasen, J. Peter-Katalinic, S. Hecimovic, Elevated cerebrospinal fluid sphingomyelin levels in prodromal Alzheimer's disease, *Neurosci. Lett.* 516 (2012) 302–305, <https://doi.org/10.1016/j.neulet.2012.04.019>.
- [88] F. Parveen, D. Bender, S.-H. Law, V.K. Mishra, C.-C. Chen, L.-Y. Ke, Role of ceramidases in sphingolipid metabolism and human diseases, *Cells* 8 (2019) 1573, <https://doi.org/10.3390/cells8121573>.
- [89] A. Jana, K. Pahan, Fibrillar amyloid- β peptides kill human primary neurons via NADPH oxidase-mediated activation of neutral sphingomyelinase: implications for Alzheimer's disease, *J. Biol. Chem.* 279 (2004) 51451–51459, <https://doi.org/10.1074/jbc.M404635200>.
- [90] M.O.W. Grimm, T.L. Rothhaar, T. Hartmann, The role of APP proteolytic processing in lipid metabolism, *Exp. Brain Res.* 217 (2012) 365–375, <https://doi.org/10.1007/s00221-011-2975-6>.
- [91] M.B. Dinkins, J. Enasco, C. Hernandez, G. Wang, J. Kong, I. Helwa, Y. Liu, A. V. Terry, E. Bieberich, Neutral sphingomyelinase-2 deficiency ameliorates Alzheimer's disease pathology and improves cognition in the 5XFAD mouse, *J. Neurosci.* 36 (2016) 8653–8667, <https://doi.org/10.1523/JNEUROSCI.1429-16.2016>.
- [92] A.V. Alessenko, A.E. Bugrova, L.B. Dudnik, Connection of lipid peroxide oxidation with the sphingomyelin pathway in the development of Alzheimer's disease, *Biochem. Soc. Trans.* 32 (2004) 144–146, <https://doi.org/10.1042/BST0320144>.
- [93] H. Sato, H. Tomimoto, R. Ohtani, T. Kitano, T. Kondo, M. Watanabe, N. Oka, I. Akiguchi, S. Furuya, Y. Hirabayashi, T. Okazaki, Astroglial expression of ceramide in Alzheimer's disease brains: a role during neuronal apoptosis, *Neuroscience* 130 (2005) 657–666, <https://doi.org/10.1016/j.neuroscience.2004.08.056>.
- [94] N. Marks, M.J. Berg, M. Saito, M. Saito, Glucosylceramide synthase decrease in frontal cortex of Alzheimer brain correlates with abnormal increase in endogenous ceramides: consequences to morphology and viability on enzyme suppression in cultured primary neurons, *Brain Res.* 1191 (2008) 136–147, <https://doi.org/10.1016/j.brainres.2007.10.066>.
- [95] J. Wang, G.W. Farr, D.H. Hall, F. Li, K. Furtak, L. Dreier, A.L. Horwich, An ALS-linked mutant SOD1 produces a locomotor defect associated with aggregation and synaptic dysfunction when expressed in neurons of *Caenorhabditis elegans*, *PLoS Genet.* 5 (2009), e1000350, <https://doi.org/10.1371/journal.pgen.1000350>.
- [96] V. Filippov, M.A. Song, K. Zhang, H.V. Vinters, S. Tung, W.M. Kirsch, J. Yang, P. J. Duersen-Hughes, Increased ceramide in brains with Alzheimer's and other neurodegenerative diseases, *J. Alzheimer's Dis.* 29 (2012) 537–547, <https://doi.org/10.3233/JAD-2011-111202>.
- [97] M.M. Mielke, V.V.R. Bandaru, N.J. Haughey, P.V. Rabins, C.G. Lyketsos, M. C. Carlson, Serum sphingomyelin and ceramides are early predictors of memory impairment, *Neurobiol. Aging* 31 (2010) 17–24, <https://doi.org/10.1016/j.neurobiolaging.2008.03.011>.
- [98] M.M. Mielke, V.V.R. Bandaru, N.J. Haughey, J. Xia, L.P. Fried, S. Yasar, M. Albert, V. Varma, G. Harris, E.B. Schneider, P.V. Rabins, K. Bandeen-Roche, C. G. Lyketsos, M.C. Carlson, Serum ceramides increase the risk of Alzheimer disease: the Women's health and aging study II, *Neurology* 79 (2012) 633–641, <https://doi.org/10.1212/WNL.0b013e318264e380>.
- [99] Y. Xing, Y. Tang, L. Zhao, Q. Wang, W. Qin, X. Ji, J. Zhang, J. Jia, Associations between plasma ceramides and cognitive and neuropsychiatric manifestations in Parkinson's disease dementia, *J. Neurol. Sci.* 370 (2016) 82–87, <https://doi.org/10.1016/j.jns.2016.09.028>.
- [100] L. Puglielli, B.C. Ellis, A.J. Saunders, D.M. Kovacs, Ceramide stabilizes β -site amyloid precursor protein-cleaving enzyme 1 and promotes amyloid β -peptide biogenesis, *J. Biol. Chem.* 278 (2003) 19777–19783, <https://doi.org/10.1074/jbc.M300466200>.
- [101] J.K. Lee, H.K. Jin, M.H. Park, B.R. Kim, P.H. Lee, H. Nakauchi, J.E. Carter, X. He, E.H. Schuchman, J.S. Bae, Acid sphingomyelinase modulates the autophagic process by controlling lysosomal biogenesis in Alzheimer's disease, *J. Exp. Med.* 211 (2014) 1551–1570, <https://doi.org/10.1084/jem.20132451>.
- [102] M.H. Lu, W.L. Ji, D.E. Xu, P.P. Yao, X.Y. Zhao, Z.T. Wang, L.P. Fang, R. Huang, L. J. Lan, J.B. Chen, T.H. Wang, L.H. Cheng, R.X. Xu, C.F. Liu, L. Puglielli, Q.H. Ma, Inhibition of sphingomyelin synthase 1 ameliorates Alzheimer-like pathology in APP/PS1 transgenic mice through promoting lysosomal degradation of BACE1, *Exp. Neurol.* 311 (2019) 67–79, <https://doi.org/10.1016/j.expneurol.2018.09.012>.
- [103] M. Horowitz, D. Elstein, A. Zimran, O. Goker-Alpan, New directions in Gaucher disease, *Hum. Mutat.* 37 (2016) 1121–1136, <https://doi.org/10.1002/humu.23056>.
- [104] S. Mullin, D. Hughes, A. Mehta, A.H.V. Schapira, Neurological effects of glucocerebrosidase gene mutations, *Eur. J. Neurol.* 26 (2019) 388–e29, <https://doi.org/10.1111/ene.13837>.
- [105] T.M. Cox, J.P. Schofield, Gaucher's disease: clinical features and natural history, 1997. [https://doi.org/10.1016/S0950-3536\(97\)80033-9](https://doi.org/10.1016/S0950-3536(97)80033-9).
- [106] N. Dekker, T. Voorn-Brouwer, M. Verhoek, T. Wennekes, R.S. Narayan, D. Speijer, C.E.M. Hollak, H.S. Overkleeft, R.G. Boot, J.M.F.G. Aerts, The cytosolic β -glucosidase GBA3 does not influence type 1 Gaucher disease manifestation, *Blood Cells, Mol. Dis.* 46 (2011) 19–26, <https://doi.org/10.1016/j.bcmd.2010.07.009>.
- [107] M.J. Barrett, P. Giraldo, J.L. Capablo, P. Alfonso, P. Irun, B. Garcia-Rodriguez, M. Pocovi, G.M. Pastores, Greater risk of parkinsonism associated with non-N370S GBA1 mutations, *J. Inher. Metab. Dis.* 36 (2013) 575–580, <https://doi.org/10.1007/s10545-012-9527-5>.
- [108] J. Mitsui, I. Mizuta, A. Toyoda, R. Ashida, Y. Takahashi, J. Goto, Y. Fukuda, H. Date, A. Iwata, M. Yamamoto, N. Hattori, M. Murata, T. Toda, S. Tsuji, Mutations for Gaucher disease confer high susceptibility to Parkinson disease, *Arch. Neurol.* 66 (2009) 571–576, <https://doi.org/10.1001/archneurol.2009.72>.
- [109] G. O'Regan, R.M. Desouza, R. Balestrino, A.H. Schapira, Glucocerebrosidase mutations in parkinson disease, *J. Park. Dis.* 7 (2017) 411–422, <https://doi.org/10.3233/JPD-171092>.
- [110] E. Sidransky, M.A. Nalls, J.O. Aasly, G. Annesi, E.R. Barbosa, A. Bar-shira, D. Berg, J. Bras, A. Brice, L.N. Clark, C. Condroyer, A. Rolfs, H. Rosenbaum, R. Rozenberg, A. Samii, M. Spitz, E. Tan, N. Tayebi, T. Toda, Multicenter GBA mutations PD 361 (2010) 1651–1661, <https://doi.org/10.1056/NEJMoa0901281.Multi-center>.
- [111] A.H.V. Schapira, Glucocerebrosidase and Parkinson disease: recent advances, *Mol. Cell. Neurosci.* 66 (2015) 37–42, <https://doi.org/10.1016/j.mcn.2015.03.013>.
- [112] I. Stojkowska, D. Krainc, J.R. Mazzulli, Molecular mechanisms of α -synuclein and GBA1 in Parkinson's disease, *Cell Tissue Res.* 373 (2018) 51–60, <https://doi.org/10.1007/s00441-017-2704-y>.
- [113] T. Shachar, C. Lo Bianco, A. Recchia, C. Wiessner, A. Raas-Rothschild, A. H. Futerman, Lysosomal storage disorders and Parkinson's disease: Gaucher disease and beyond, *Mov. Disord.* 26 (2011) 1593–1604, <https://doi.org/10.1002/mds.23774>.
- [114] S.Y. Yang, M. Beavan, K.Y. Chau, J.W. Taanman, A.H.V. Schapira, A human neural crest stem cell-derived dopaminergic neuronal model recapitulates biochemical abnormalities in GBA1 mutation carriers, *Stem Cell Rep.* 8 (2017) 728–742, <https://doi.org/10.1016/j.stemcr.2017.01.011>.
- [115] J.R. Mazzulli, F. Zunke, T. Tsunemi, N.J. Tokar, S. Jeon, L.F. Burbulla, S. Patnaik, E. Sidransky, J.J. Marugan, C.M. Sue, D. Krainc, Activation of β -glucocerebrosidase reduces pathological α -synuclein and restores lysosomal function in Parkinson's patient midbrain neurons, *J. Neurosci.* 36 (2016) 7693–7706, <https://doi.org/10.1523/JNEUROSCI.0628-16.2016>.
- [116] E. Rockenstein, J. Clarke, C. Viel, N. Panarello, C.M. Treleaven, C. Kim, B. Spencer, A. Adame, H. Park, J.C. Dodge, S.H. Cheng, L.S. Shihabuddin, E. Masliah, S.P. Sardi, Glucocerebrosidase modulates cognitive and motor activities in murine models of Parkinson's disease, *Hum. Mol. Genet.* 25 (2016) 2645–2660, <https://doi.org/10.1093/hmg/ddw124>.
- [117] J. Blanz, P. Saftig, Parkinson's disease: acid-glucocerebrosidase activity and alpha-synuclein clearance, *J. Neurochem.* 139 (2016) 198–215, <https://doi.org/10.1111/jnc.13517>.
- [118] E. Sidransky, G. Lopez, The link between the GBA gene and parkinsonism, *Lancet Neurol.* 11 (2012) 986–998, [https://doi.org/10.1016/S1474-4422\(12\)70190-4](https://doi.org/10.1016/S1474-4422(12)70190-4).
- [119] F. Schmitt, G. Hussain, L. Dupuis, J.P. Loeffler, A. Henriques, A plural role for lipids in motor neuron diseases: energy, signaling and structure, *Front. Cell. Neurosci.* 8 (2014) 1–10, <https://doi.org/10.3389/fncel.2014.00025>.
- [120] M. Bigaud, D. Guerini, A. Billich, F. Bassilana, V. Brinkmann, Second generation S1P pathway modulators: Research strategies and clinical developments, *Biochim. Biophys. Acta Mol. Cell Biol. Lipids* 1841 (2014) 745–758, <https://doi.org/10.1016/j.bbalip.2013.11.001>.
- [121] J. Ceccom, N. Loukh, V. Lauwers-Cances, C. Touriol, Y. Nicaise, C. Gentil, E. Uro-Coste, S. Pitson, C.A. Maurage, C. Duyckaerts, O. Cuvillier, M.B. Delisle, Reduced sphingosine kinase-1 and enhanced sphingosine 1-phosphate lyase expression demonstrate deregulated sphingosine 1-phosphate signaling in Alzheimer's disease, *Acta Neuropathol. Commun.* 2 (2014) 1–10, <https://doi.org/10.1186/2051-5960-2-12>.
- [122] A. Di Pardo, V. Maglione, Sphingolipid metabolism: a new therapeutic opportunity for brain degenerative disorders, *Front. Neurosci.* 12 (2018) 1–8, <https://doi.org/10.3389/fnins.2018.00249>.
- [123] S. Grassi, L. Mauri, S. Prioni, L. Cabibbi, S. Sonnino, A. Prinetti, P. Giussani, Sphingosine 1-phosphate receptors and metabolic enzymes as druggable targets for brain diseases, *Front. Pharmacol.* 10 (2019) 1–20, <https://doi.org/10.3389/fphar.2019.00807>.
- [124] G. Vidal-Martinez, K. Najera, J.D. Miranda, C. Gil-Tommee, B. Yang, J. Vargas-Medrano, V. Diaz-Pacheco, R.G. Perez, FTY720 improves behavior, increases brain derived neurotrophic factor levels and reduces α -synuclein pathology in parkinsonian GM2 +/- mice, *Neuroscience* 411 (2019) 1–10, <https://doi.org/10.1016/j.neuroscience.2019.05.029>.
- [125] M. Asle-Rousta, Z. Kolahdooz, S. Oryan, A. Ahmadiani, L. Dargahi, FTY720 (Fingolimod) attenuates beta-amyloid peptide (A β 42)- induced impairment of spatial learning and memory in rats, *J. Mol. Neurosci.* 50 (2013) 524–532, <https://doi.org/10.1007/s12031-013-9979-6>.
- [126] M. Asle-Rousta, Z. Kolahdooz, L. Dargahi, A. Ahmadiani, S. Nasoohi, Prominence of central sphingosine-1-phosphate receptor-1 in attenuating A β -induced injury by fingolimod, *J. Mol. Neurosci.* 54 (2014) 698–703, <https://doi.org/10.1007/s12031-014-0423-3>.
- [127] N. Takasugi, T. Sasaki, I. Ebinuma, S. Osawa, H. Isshiki, K. Takeo, T. Tomita, T. Iwatsubo, FTY720/Fingolimod, a sphingosine analogue, reduces amyloid- β production in neurons, *PLoS One* 8 (2013) 1–8, <https://doi.org/10.1371/journal.pone.0064050>.
- [128] M. Nazari, S. Keshavarz, A. Rafati, M.R. Namavar, M. Haghani, Fingolimod (FTY720) improves hippocampal synaptic plasticity and memory deficit in rats

- following focal cerebral ischemia, *Brain Res. Bull.* 124 (2016) 95–102, <https://doi.org/10.1016/j.brainresbull.2016.04.004>.
- [129] R.L. Potenza, R. De Simone, M. Armida, V. Mazziotti, A. Pezzola, P. Popoli, L. Minghetti, Fingolimod: a disease-modifier drug in a mouse model of amyotrophic lateral sclerosis, *Neurotherapeutics* 13 (2016) 918–927, <https://doi.org/10.1007/s13311-016-0462-2>.
- [130] R.L. Lieberman, J.A. D'Aquino, D. Ringe, G.A. Petsko, Effects of pH and iminosugar pharmacological chaperones on lysosomal glycosidase structure and stability, *Biochemistry* 48 (2009) 4816–4827, <https://doi.org/10.1021/bi9002265>.
- [131] Y. Sun, H. Ran, B. Liou, B. Quinn, M. Zamzow, W. Zhang, J. Bielawski, K. Kitatani, K.D.R. Setchell, Y.A. Hannun, G.A. Grabowski, Isofagomine in vivo effects in a neuronopathic gaucher disease mouse, *PLoS One* 6 (2011), <https://doi.org/10.1371/journal.pone.0019037>.
- [132] F. Richter, S.M. Fleming, M. Watson, V. Lemesre, L. Pellegrino, B. Ranes, C. Zhu, F. Mortazavi, C.K. Mulligan, P.C. Sioshansi, S. Hean, K. De La Rosa, R. Khanna, J. Flanagan, D.J. Lockhart, B.A. Wustman, S.W. Clark, M.F. Chesselet, A GCCase chaperone improves motor function in a mouse model of synucleinopathy, *Neurotherapeutics* 11 (2014) 840–856, <https://doi.org/10.1007/s13311-014-0294-x>.
- [133] M. Malerba, B. Ragnoli, Ambroxol in the 21st century: pharmacological and clinical update, *Expert Opin. Drug Metab. Toxicol.* 4 (2008) 1119–1129, <https://doi.org/10.1517/17425255.4.8.1119>.
- [134] Z. Luan, L. Li, K. Higaki, E. Nanba, Y. Suzuki, K. Ohno, The chaperone activity and toxicity of ambroxol on Gaucher cells and normal mice, *Brain Dev.* 35 (2013) 317–322, <https://doi.org/10.1016/j.braindev.2012.05.008>.
- [135] G.H.B. Maegawa, M.B. Tropak, J.D. Buttner, B.A. Rigat, M. Fuller, D. Pandit, L. Tang, G.J. Kornhaber, Y. Hamuro, J.T.R. Clarke, D.J. Mahuran, Identification and characterization of ambroxol as an enzyme enhancement agent for Gaucher disease, *J. Biol. Chem.* 284 (2009) 23502–23516, <https://doi.org/10.1074/jbc.M109.012393>.
- [136] A. McNeill, J. Magalhaes, C. Shen, K.Y. Chau, D. Hughes, A. Mehta, T. Foltynie, J. M. Cooper, A.Y. Abramov, M. Gegg, A.H.V. Schapira, Ambroxol improves lysosomal biochemistry in glucocerebrosidase mutation-linked Parkinson disease cells, *Brain* 137 (2014) 1481–1495, <https://doi.org/10.1093/brain/awu020>.
- [137] A. Migdalska-Richards, L. Daly, E. Bezard, A.H.V. Schapira, Ambroxol effects in glucocerebrosidase and α -synuclein transgenic mice, *Ann. Neurol.* 80 (2016) 766–775, <https://doi.org/10.1002/ana.24790>.
- [138] A. Migdalska-Richards, W.K.D. Ko, Q. Li, E. Bezard, A.H.V. Schapira, Oral ambroxol increases brain glucocerebrosidase activity in a nonhuman primate, *Synapse* 71 (2017) 17–19, <https://doi.org/10.1002/syn.21967>.
- [139] A. Bouscary, C. Quessada, A. Mosbach, N. Callizot, M. Spedding, J. Loeffler-Philippe, A. Henriques, Ambroxol hydrochloride improves motor functions and extends survival in a mouse model of familial amyotrophic lateral sclerosis, *Front. Pharmacol.* 10 (2019), <https://doi.org/10.3389/fphar.2019.00883>.
- [140] A. Bouscary, C. Quessada, F. René, M. Spedding, A. Henriques, S. Ngo, J. P. Loeffler, Drug repositioning in neurodegeneration: an overview of the use of ambroxol in neurodegenerative diseases, *Eur. J. Pharmacol.* 884 (2020), 173446, <https://doi.org/10.1016/j.ejphar.2020.173446>.
- [141] R.L. Lieberman, B.A. Wustman, P. Huertas, A.C. Powe, C.W. Pine, R. Khanna, M. G. Schlossmacher, D. Ringe, G.A. Petsko, Structure of acid β -glucosidase with pharmacological chaperone provides insight into Gaucher disease, *Nat. Chem. Biol.* 3 (2007) 101–107, <https://doi.org/10.1038/nchembio850>.
- [142] A. Zimran, G. Altarescu, D. Elstein, Pilot study using ambroxol as a pharmacological chaperone in type 1 Gaucher disease, *Blood Cells Mol. Dis.* 50 (2013) 134–137, <https://doi.org/10.1016/j.bcmd.2012.09.006>.
- [143] A. Narita, K. Shirai, S. Itamura, A. Matsuda, A. Ishihara, K. Matsushita, C. Fukuda, N. Kubota, R. Takayama, H. Shigematsu, A. Hayashi, T. Kumada, K. Yuge, Y. Watanabe, S. Kosugi, H. Nishida, Y. Kimura, Y. Endo, K. Higaki, E. Nanba, Y. Nishimura, A. Tamasaki, M. Togawa, Y. Saito, Y. Maegaki, K. Ohno, Y. Suzuki, Ambroxol chaperone therapy for neuronopathic Gaucher disease: a pilot study, *Ann. Clin. Transl. Neurol.* 3 (2016) 200–215, <https://doi.org/10.1002/acn3.292>.
- [144] S. Mullin, L. Smith, K. Lee, G. D'Souza, P. Woodgate, J. Elflein, J. Hällqvist, M. Toffoli, A. Streeter, J. Hosking, W.E. Heywood, R. Khengar, P. Campbell, J. Hehir, S. Cable, K. Mills, H. Zetterberg, P. Limousin, V. Libri, T. Foltynie, A.H. V. Schapira, Ambroxol for the treatment of patients with Parkinson disease with and without glucocerebrosidase gene mutations: a nonrandomized, noncontrolled trial, *JAMA Neurol.* (2020) 1–8, <https://doi.org/10.1001/jamaneurol.2019.4611>.
- [145] R. Cipriani, J.C. Chara, A. Rodríguez-Antigüedad, C. Matute, FTY720 attenuates excitotoxicity and neuroinflammation, *J. Neuroinflamm.* 12 (2015) 1–14, <https://doi.org/10.1186/s12974-015-0308-6>.
- [146] P. Zhao, X. Yang, L. Yang, M. Li, K. Wood, Q. Liu, X. Zhu, Neuroprotective effects of fingolimod in mouse models of Parkinson's disease, *FASEB J.* 31 (2017) 172–179, <https://doi.org/10.1096/fj.201600751R>.
- [147] Y.H. Xu, Y. Sun, H. Ran, B. Quinn, D. Witte, G.A. Grabowski, Accumulation and distribution of β -synuclein and ubiquitin in the CNS of Gaucher disease mouse models, *Mol. Genet. Metab.* 102 (2011) 436–447, <https://doi.org/10.1016/j.ymgme.2010.12.014>.
- [148] Y.M. Kim, M.S. Yum, S.H. Heo, T. Kim, H.K. Jin, J.S. Bae, G.H. Seo, A. Oh, H. M. Yoon, H.T. Lim, H.W. Kim, T.S. Ko, H.S. Lim, M.J. Osborn, J. Tolar, C. Cozma, A. Rolf, A. Zimran, B.H. Lee, H.W. Yoo, Pharmacologic properties of high-dose ambroxol in four patients with Gaucher disease and myoclonic epilepsy, *J. Med. Genet.* 57 (2020) 124–131, <https://doi.org/10.1136/jmedgenet-2019-106132>.