



Relevance of Polymorphic KIR and HLA Class I Genes in NK-Cell-Based Immunotherapies for Adult Leukemic Patients

Léa Dubreuil, Patrice Chevallier, Christelle Retiere, Katia Gagne

► To cite this version:

Léa Dubreuil, Patrice Chevallier, Christelle Retiere, Katia Gagne. Relevance of Polymorphic KIR and HLA Class I Genes in NK-Cell-Based Immunotherapies for Adult Leukemic Patients. *Cancers*, 2021, 13 (15), pp.3767. 10.3390/cancers13153767 . inserm-03317983

HAL Id: inserm-03317983

<https://inserm.hal.science/inserm-03317983>

Submitted on 9 Aug 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Review

Relevance of Polymorphic KIR and HLA Class I Genes in NK-Cell-Based Immunotherapies for Adult Leukemic Patients

Léa Dubreuil ^{1,2,3}, Patrice Chevallier ^{2,3,4}, Christelle Retière ^{1,2,3}  and Katia Gagne ^{1,2,3,5,*} 

¹ Department of Research, Etablissement Français du Sang, 44011 Nantes, France; lea.dubreuil@efs.sante.fr (L.D.); christelle.retiere@efs.sante.fr (C.R.)

² University de Nantes, INSERM U1232 CNRS, CRCINA, F-44000 Nantes, France; patrice.chevallier@chu-nantes.fr

³ LabEx IGO “Immunotherapy, Graft, Oncology”, F-44000 Nantes, France

⁴ Hematology Clinic, CHU, 44093 Nantes, France

⁵ LabEx Transplantex, Université de Strasbourg, 67000 Strasbourg, France

* Correspondence: katia.gagne@efs.sante.fr; Tel.: +33-2-40-12-34-72

Simple Summary: Immunotherapies are promising approaches to curing different acute leukemias. Natural killer (NK) cells are lymphocytes that are efficient in the elimination of leukemic cells. NK-cell-based immunotherapies are particularly attractive, but the landscape of the heterogeneity of NK cells must be deciphered. This review provides an overview of the polymorphic KIR and HLA class I genes that modulate the NK cell repertoire and how these markers can improve the outcomes of patients with acute leukemia. A better knowledge of these genetic markers that are linked to NK cell subsets that are efficient against hematological diseases will optimize hematopoietic stem-cell donor selection and NK immunotherapy design.



Citation: Dubreuil, L.; Chevallier, P.; Retière, C.; Gagne, K. Relevance of Polymorphic KIR and HLA Class I Genes in NK-Cell-Based Immunotherapies for Adult Leukemic Patients. *Cancers* **2021**, *13*, 3767. <https://doi.org/10.3390/cancers13153767>

Academic Editor: Antonio Curti

Received: 21 June 2021

Accepted: 23 July 2021

Published: 27 July 2021

Publisher’s Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Since the mid-1990s, the biology and functions of natural killer (NK) cells have been deeply investigated in healthy individuals and in people with diseases. These effector cells play a particularly crucial role after allogeneic hematopoietic stem-cell transplantation (HSCT) through their graft-versus-leukemia (GvL) effect, which is mainly mediated through polymorphic killer-cell immunoglobulin-like receptors (KIRs) and their cognates, HLA class I ligands. In this review, we present how KIRs and HLA class I ligands modulate the structural formation and the functional education of NK cells. In particular, we decipher the current knowledge about the extent of KIR and HLA class I gene polymorphisms, as well as their expression, interaction, and functional impact on the KIR⁺ NK cell repertoire in a physiological context and in a leukemic context. In addition, we present the impact of NK cell alloreactivity on the outcomes of HSCT in adult patients with acute leukemia, as well as a description of genetic models of KIRs and NK cell reconstitution, with a focus on emergent T-cell-repleted haplo-identical HSCT using cyclophosphamide post-grafting (haplo-PTCy). Then, we document how the immunogenetics of KIR/HLA and the immunobiology of NK cells could improve the relapse incidence after haplo-PTCy. Ultimately, we review the emerging NK-cell-based immunotherapies for leukemic patients in addition to HSCT.

Keywords: natural killer cells; killer-cell immunoglobulin-like receptors; HLA class I; polymorphism; acute leukemia; hematopoietic stem-cell transplantations; NK-cell-based immunotherapy

1. Introduction

Natural killer (NK) cells, which develop from common lymphoid progenitors in the bone marrow, correspond to large granular lymphocytes and are usually identified as CD3[−] CD56⁺, representing about 5–20% of peripheral blood mononuclear cells (PBMCs) [1]. These innate immune effectors [2] proliferate, produce cytokines, and are cytotoxic against virus-infected cells or tumor cells. According to the level of CD56 expression, two NK cell subsets with distinct repertoire signatures have been defined; CD56^{dim} NK cells (90%) are

mainly cytotoxic, while CD56^{bright} NK cells (10%) produce high levels of cytokines, but have a low cytotoxicity [3]. NK cells are able to lyse target cells by using antibody-dependent cellular cytotoxicity (ADCC) driven by the activating CD16 receptor. They express a wide range of activating receptors including NKG2D, DNAM-1, 2B4, and NCRs such as NKp30, NKp44, and NKp46 that recognize induced or upregulated molecules on tumor and virus-infected cells. NK cells recognize healthy cells that express normal levels of HLA class I molecules and sense the absence of HLA class I molecules on allogeneic cells or their decreased expression on tumor or virus-infected cells, as demonstrated by the “missing-self” hypothesis [4,5]. Killer-cell immunoglobulin-like receptors (KIRs) and the CD94/NKG2A heterodimer represent the two main inhibitory NK receptors that are engaged in this missing-self recognition. Overall, NK cell functions are regulated by a balance of synergistic signals from combinations of inhibitory and activating receptors engaged with cognate ligands on target cells. HLA class I molecules play a central role in the regulation of immune responses with respect to their interactions with T- or NK-cell-specific receptors [6]. KIRs and their HLA class I ligands constitute the main polymorphic and polygenic pair in humans for the regulation of functions and the licensing of NK cells [6,7]. KIRs are clonally expressed on the surface of mature NK cells [8,9]. Functional KIR/HLA interactions drive the tremendous diversity of NK cells with a limited number of germline-encoded genes [10]. In addition to KIRs, the CD94/NKG2A heterodimer recognizes the nonclassical HLA-E molecule. This receptor/ligand is expressed in all individuals and does not present the broad genetic diversity that characterizes the KIR/HLA pair. The inhibitory CD94/NKG2A receptor and its activating NKG2C counterpart are involved in missing-self recognition, and CD94/NKG2A is also involved in NK cell licensing [11]. Moreover, the expression of inhibitory and activating receptors is strongly associated with NK cell differentiation and maturation. In particular, NK cell subsets can be defined on the basis of differentiation markers. During their development, NK cells first acquire NKG2A expression and, at a later stage, KIR expression, followed by the terminal differentiation CD57 receptor [11]. The cytomegalovirus (CMV) also modulates the NK cell repertoire with an expansion of KIR2D⁺ NKG2C⁺ NK cells [12,13]. The model of NK cell differentiation established by Björsktorm et al. [11], therefore, permits the investigation of the heterogeneity of NK cells.

In cancer immunotherapies, NK cells represent key components especially for patients with hematological malignancies. In these cases, NK cells can easily access tumor sites without the barrier represented by the tumor microenvironment, unlike in the case of solid tumors [14]. Although they are diverse, hematological malignancies include acute myeloid leukemia (AML) and acute lymphoid leukemia (ALL), which are frequent in adult patients. For patients with acute leukemia, the relapse of the initial disease remains the major cause of death. The relapse may be due to a failure of conventional chemotherapies but also due to the escape of tumor cells from immune responses [15]. The lost or decreased expression of HLA molecules and/or upregulation of ligands on leukemic cells that are recognized by activating NK cell receptors can sustain the crucial role of NK cells. For leukemic patients for whom conventional chemotherapies have failed, allogeneic hematopoietic stem-cell transplantation (HSCT) is usually suggested in order to replace both the defective hematopoiesis and the immune system of the patient with healthy HSCs from donors. In that case, NK cells play a crucial role through their graft-versus-leukemia (GvL) effect, as they are the first lymphocytes to participate in the reconstitution of the immune system of the patient even before T cells after HSCT [16,17]. However, the NK cell functions in leukemic patients can be altered by the tumor microenvironment. The efficiency of NK-cell-based immunotherapies can be enhanced by cytokines, inhibitory specific monoclonal antibodies (mAbs), and adoptive transfer of in vitro activated NK cells [18,19]. Promising recent advances have shown the use of NK cells armed with chimeric antigen receptors (CARs) to enhance their antileukemic function [20]. The CRISPR technology also permits the optimization of some cellular pathways to boost the antileukemic efficacy of NK cell immunotherapies [21]. Numerous approaches to NK cell immunotherapy have been developed by using mainly autologous or allogeneic NK cells in bulk without considering

all genetic and allelic KIR and HLA polymorphisms known to govern the structural and functional heterogeneity of NK cells.

In the present review, we present the characteristics of KIR and HLA class I gene polymorphisms, their expression, the KIR/HLA interactions that regulate the NK cell repertoire in healthy individuals, and the relevance of KIR/KIR ligands to NK-cell-based immunotherapies for adult patients with hematological malignancies, especially in the context of allogeneic HSCT. The roles of NK cells in physiological, allogeneic, infectious, and leukemic contexts are summarized in Figure 1.

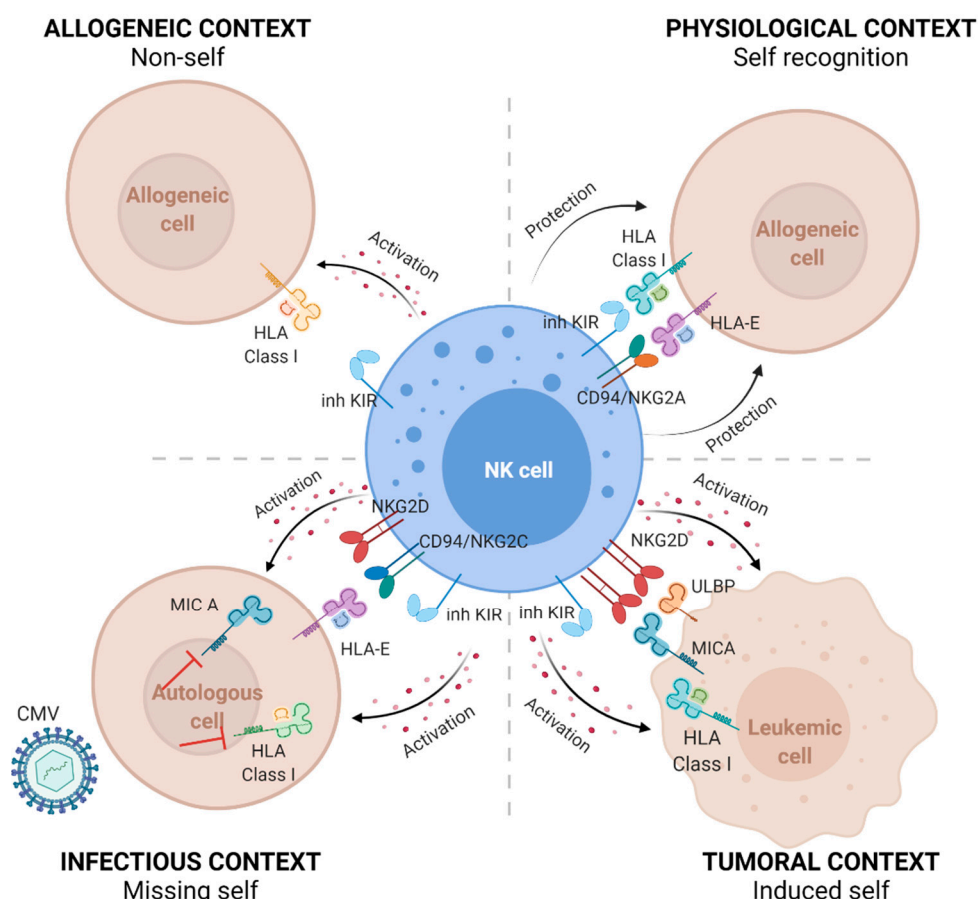


Figure 1. The natural cytotoxicity of NK cells is driven by the missing-self recognition. NK cells recognize healthy cells that express normal levels of HLA class I molecules and sense the absence of HLA class I molecules on allogeneic cells (non-self) or decreased expression of HLA class I molecules (missing-self) on tumor cells or virus-infected cells, as reported for the cytomegalovirus (CMV). Recognition of “self” mainly involves interactions between polymorphic inhibitory killer-cell immunoglobulin-like receptors (inh KIRs) and classical HLA class I molecules, as well as interactions between the CD94/NKG2A heterodimer and the nonclassical HLA-E molecule. The lack of interaction of inh KIRs with their HLA class I ligands or CD94-NKG2A with HLA-E induces the activation of NK cell lysis. In addition to KIR/KIR ligands, interactions of the induced MHC class I-like MICA ligand, ULBP ligand expressed on tumor cells, or the HLA-E molecule by respectively activating NKG2D or CD94/NKG2C receptors could activate NK cell lysis.

2. Relevance of the KIR and HLA Class I Genes to the NK Cell Repertoire

In healthy individuals, the formation of the NK cell repertoire is influenced by various factors including the variability of the KIR and HLA class I genes, the CMV status, and the frequency of NK cell subsets. KIRs represent a family of receptors belonging to the immunoglobulin (Ig)-like superfamily. The KIR family is composed of receptors with two or three (2D, 3D) extracellular domains. The intracytoplasmic tail can be long (L) with two

ITIMs (immune tyrosine-based inhibitory motifs) or short (S) and coupled with the DAP12 molecule, which bears an ITAM (immune tyrosine-based activating motif) discriminating inhibitory and activating KIRs, respectively [22].

2.1. KIR Genes

The genetics of KIRs are complex because KIR genes vary in structure, exon/intron organization, and relative size. Currently, the KIR gene family consists of 15 different loci (2DL1, 2DL2/2DL3, 2DL4, 2DL5A, 2DL5B, 2DS1, 2DS2, 2DS3, 2DS4, 2DS5, 3DL1/3DS1, 3DL2, 3DL3, 2DP1, 3DP1) encoded in a 100–200 kb region in the leucocyte region complex (LRC) located on chromosome 19. The LRC constitutes a dense cluster of immune-related genes that encode, in addition to KIR, other NK cell receptors [23]. KIR genes have a head-to-tail organization and are arranged within 2.4 kb of each other, except between 3DP1 and 2DL4, where there is a gap of 14 kb [24]. KIR genes have four to nine exons with an entire genomic size ranging from 4 kb for 3DP1 to 16 kb for 3DL2 and long introns [25]. Three KIR exon/intron gene organizations have been described. The first one corresponds to the genes encoding KIR3DL, such as 3DL1/2. These genes consist of nine exons: exons 1 and 2 encode the signal peptide; exons 3, 4, and 5 encode each Ig-like domain (D0, D1, and D2 respectively); exon 6 encodes the stem connecting the D2 domain to the transmembrane part, which is encoded by exon 7; exons 8 and 9 encode the intracytoplasmic tail. KIR3DL3 has a similar structure, but with the absence of exon 6 and a stop codon in exon 9. KIR3DS1 has the same exon/intron organization as that of 3DL1, but a shorter exon 9 encoding for the short tail. Interestingly, KIR3DL1 and KIR3DS1 are defined as alleles of the same gene, although they have different functions [26]. The second and third exon/intron gene organization corresponds to KIR2D: type I KIR2D includes genes that encode 2DL1/2/3/S1/2/3/4/5. These genes consist of eight exons with exons 4 and 5 encoding the D1 and D2 domains respectively, and a region homologous to exon 3 encoding the D0 domain of KIR3D, which is called “pseudo-exon 3” [27,28]. The KIR2DS4 gene is found in two forms, with one common 1D variant that is characterized by a 22 bp deletion in exon 5, which block transcription of the corresponding receptor [24]; the type II KIR2D genes include 2DL4/5, which exhibits eight exons and a 22 kb deletion in exon 4. Exons 3 and 5 of these genes encode the D0 and D2 domains, respectively.

The number and nature of KIR genes differ between individuals, allowing one to distinguish two major KIR haplotypes (A, B) [29]. The KIR A and B haplotypes both have four framework genes (3DL3, 3DP1, 2DL4, and 3DL2), and are present in all individuals with some rare exceptions. KIR3DL3 and 3DL2 are located at the centromeric and telomeric ends, respectively, and delimit the LRC, while 3DP1 and 2DL4 are located between both. The KIR A haplotype is defined by a fixed set of nine KIR genes (3DL3, 2DL3, 2DP1, 2DL1, 3DP1, 2DL4, 3DL1, 2DS4, and 3DL2) with only 2DS4 as an activating gene. In contrast, the B haplotypes are diverse with a variable number of KIR genes and are characterized by the presence of specific inhibitory KIRs, such as 2DL2, and more than one activating KIR gene. Based on the combination of the KIR A and B haplotypes, 660 KIR genotypes have been described worldwide (<http://www.allelefreqencies.net/>) (accessed on 26 July 2021). Conventionally, the gene frequencies of KIR are close to 100% for inhibitory 2DL1/2/3, whereas the frequencies of the activating KIR genes are low, except for 2DS4. Different commercial or in-house methods including the use of specific KIR primers or probes, permit the assessment of the presence or absence of KIR genes. Various studies have shown that frequencies of KIR genes, haplotypes and/or genotypes vary between populations [30–32]. Interestingly, a majority (>50%) of individuals harbors the KIR AA genotype in Asian populations [30,33], whereas only 35% of Europeans have this genotype [34,35].

In addition to the definition of the KIR A and B haplotypes, framework KIR genes allow for the delineation of two chromosomal parts: centromeric (cen) (2DS2, 2DL2/3, 2DL5B, 2DS3, 2DP1, 2DL1) and telomeric (tel) KIR genes (3DL1/S1, 2DL5A, 2DS5, 2DS1) [36]. Depending on the KIR2DL2/3/S2 genes, the cenAA (2DL3 only), cenAB (2DL3 with 2DS2 and/or 2DL2), and cenBB (2DL2 and/or 2DS2, no 2DL3) motifs are defined. Depending on

KIR2DS1/S4/3DL1/S1 genes, the telAA (3DL1 and 2DS4 only), telAB (3DL1 and 2DS4 with 3DS1 and/or 2DS1), and telBB (lacking 3DL1 and/or 2DS4) motifs are defined. This subdivision into gene content motifs can increase the diversity of KIR genes. Thus, individuals with a KIR AB genotype could present different cen and tel motifs, whereas individuals with the KIR AA genotype exhibit only the cenAA and telAA motifs. Another parameter of this diversity is linked to the copy number variation (CNV) [37]. The arrangement of KIR genes in a head-to-tail orientation and their high sequence homology allow for a gain or loss of KIR genes with the generation of expanded or contracted KIR haplotypes, as well as the formation of fusion genes [38]. It has been shown that some KIR genes are more affected by CNV than others [33].

Lastly, KIR genes exhibit an allelic diversity; currently, 1110 KIR alleles have been referenced (<https://www.ebi.ac.uk/ipd/kir/>) (accessed on 26 July 2021). The allelic polymorphism of KIRs is particularly high for inhibitory KIRs (the strongest being for 3DL3 with 228 alleles) and is limited for activating KIRs (the lowest being for 2DS1 with 33 alleles). To name KIR alleles, a specific nomenclature [39] has been established, which uses an asterisk as a separator after the gene name, before a numerical allele designation (2DL1*). The next three digits are used to name the different alleles of the same gene that differ in coding sequences (2DL1*003). The following two digits are used to distinguish alleles that only differ in synonymous (noncoding) substitutions within the coding sequence (2DL1*00302), and the last two digits are used to distinguish alleles that only differ in substitutions in noncoding regions (2DL1*0030202). For more than 15 years, specific KIR primers, probes or sequence-based-typing (SBT) methods have been used to assign KIR alleles of limited KIR genes [40]. More recently, next-generation sequencing (NGS) technologies have permitted the assignment of the alleles of all KIR genes [41–43]. As mentioned for KIR genes, KIR allele frequencies vary among populations. Interestingly, although 68 KIR2DL1 alleles have been reported worldwide, only five predominant alleles (*001, *002, *003, *004, and *007) are encountered in Europeans [34,44]. Moreover, the non-expressed KIR3DL1*004 allele [45], which is frequent in Caucasians [46], is absent in the Japanese population [30]. Interestingly, the group of P. Parham was the first to describe that allelic polymorphism of KIRs synergizes with variable gene content, thus making it possible to individualize KIR genotypes [47]. Recently, the KIR A haplotype and cenAA motif were reported to be associated with the 2DL1*001, *002, and *003 alleles, while the B haplotype and cenB+ motif were associated with the 2DL1*004 and *007 alleles in European populations [34,48–50]. In addition, different linkage disequilibria (LD) between KIR genes have been described [24,48]. Overall, these population-based genetic studies could help in the prediction of KIR alleles from KIR gene content, including pseudogenes [51].

2.2. HLA Class I Genes

The main characteristics of the HLA system correspond to an exhaustive polymorphism and the inheritance of HLA genes from the parental haplotypes in blocks. The HLA system represents the most polymorphic gene locus in humans [52], with 28,938 HLA alleles reported so far [53]. The HLA locus contains more than 200 genes, many of which have immune-related functions [54]. HLA class I genes encode both classical and nonclassical HLA class I molecules. Classical HLA class I genes (HLA-A, -B, -C) are located on the short arm of chromosome 6 (6p21.3) and are composed of eight exons and seven introns. Most polymorphic exons correspond to exons 2 and 3, which respectively, encode the $\alpha 1$ and $\alpha 2$ domains which form the peptide-binding groove of HLA class I molecules that are ubiquitously expressed on nucleated cells. The length of the HLA-A, -B, and -C genes is closed to 3 kb. In terms of allelic polymorphism, HLA-A, -B, and -C are divergent genes with numerous alleles, and all positions in the $\alpha 1$ and $\alpha 2$ domains exhibit variations [55]. The polymorphism of HLA-C genes, although extremely diverse, is more limited in terms of the alleles described in comparison with the polymorphism of HLA-B genes. The allele frequencies of HLA-C and the LD of HLA-C/HLA-B are frequently updated through genetic population studies [56–58]. From its phylogeny, HLA-C appears to be more recent

compared to the HLA-A and -B loci, and regulatory elements of transcription differ between the HLA-C and HLA-A/B genes [59]. Today, HLA class I alleles are assigned by using NGS methods [57,60–63], which allow detailed mapping of all polymorphic bases encountered in the exons, introns, and untranslated regions (UTR). Identification of regulatory variants located in UTRs is important because they can influence the expression of HLA class I molecules, as detailed later. In addition to the classical HLA class I genes, the HLA-E and MICA genes, despite their limited polymorphism are of interest in NK cell studies, as they encode for the CD94/NKG2C and NKG2D ligands, respectively [64,65].

2.3. KIR/HLA Interactions and the KIR⁺ NK Cell Repertoire

The distribution of KIRs on the surface of NK cells is clonal due to the stochastic expression of KIRs [66,67]. Therefore, a heterogeneous repertoire of NK cell subpopulations expressing different combinations of KIRs or a lack thereof is observed. Usually, the expression of KIRs on NK cells is assessed by using commercial (which is limited to some KIRs) and/or in-house anti-KIR mAbs, which make it possible to distinguish the main inhibitory and activating KIR2D [68]. The KIR gene content influences the KIR⁺ NK cell repertoire. Thus, KIR AA genotyped individuals show a less diverse NK cell repertoire with higher frequencies of the corresponding KIR⁺ NK subpopulations compared to KIR B+ genotyped individuals, who have a broader KIR gene content and lower frequencies of KIR⁺ NK subpopulations [69]. In addition to KIR gene content, the expression of KIR allele encoded products (allotypes) on NK cells may differ depending on the allelic polymorphism, as deeply documented for 3DL1 by P. Parham's group [70,71]. We also reported high (3DL1*001, *002, *015, *008), low (*005, *007), or no (*004, *019) expression of KIR3DL1 on NK cells depending on the 3DL1 alleles or the 3DL1/S1 allele combinations [46]. The differential expression of KIR3DL1 allotypes could be partially explained by the polymorphisms encountered in the KIR3DL1 promoter [72]. Similarly, three single-nucleotide polymorphisms (SNPs) located in the recruiting sites of transcriptional factors in the KIR2DL1 promoter induce a higher transcriptional activity of 2DL1 compared to 2DL3 [73]. Similarly to KIR3DL1, the expression level of KIR2DL1 is variable depending on the 2DL1 alleles. In particular, the KIR2DL1*004 allotype, which carries a cysteine in the intracytoplasmic tail at position 245 (Cys²⁴⁵), is poorly expressed on NK cells, with a low frequency of KIR2DL1⁺ NK cells compared to the 2DL1*002 and *003 (Arg²⁴⁵) allotypes [34,74,75].

KIRs interact with HLA class I molecules. Inhibitory KIR2DL1/2/3 all have HLA-Cw molecules as ligands, which are initially divided into two groups as a function of the 77 and 80 amino acids located in the $\alpha 2$ domain. KIR2DL1 specifically recognizes the HLA-Cw molecules of group 2 (C2) with Lys⁸⁰ [76]. The peptide presented by the HLA-Cw molecule seems to play a role in the binding affinity of KIR2DL1/C2 because a substitution at the P8 position with an acidic residue results in a loss of ligand binding [77,78]. KIR2DL2/3 recognize the HLA-Cw molecules of group C1 (C1) with Asp⁸⁰ [79]. However, the recognition of KIR2DL2/3 for C1 is less stringent than the recognition of 2DL1 for C2 because some C2 ligands, such as HLA-Cw4, are also recognized by KIR2DL2/3 [34,80,81]. In addition, the avidity of KIR2DL2 for the C1 ligands is stronger than for 2DL3 [80,82]. Interestingly, the peptide could increase the affinity of KIR2DL2/3 for the C1 ligands, as reported in the context of HCV [83]. KIR2DL4, which is an activating receptor, has the HLA-G molecule as a ligand, which is a nonclassical HLA class I molecule expressed on trophoblast cells during pregnancy [84]. KIR2DS1 recognizes C2 ligands as its inhibitory 2DL1 counterpart, but with a lower affinity [85,86]. KIR2DS2 recognizes C1 ligands and the HLA-A11 molecule [87,88]. As reported for KIR2DL2/3, the recognition of KIR2DS2 of HLA-C molecules is peptide-dependent [88,89]. KIR2DS4 recognizes some HLA-C molecules and the HLA-A11 molecule [90]. To date, the ligands of KIR2DL5 and 2DS3 remain unknown. KIR3DL1 recognizes HLA-B and some HLA-A molecules that bear Bw4 motifs [91,92], while its activating 3DS1 counterpart probably interact with the nonclassical HLA-F molecule [93], although a Bw4 specificity was reported in the context of HIV [94]. KIR3DL2 interacts with HLA-A3/A11 molecules, and this interaction is peptide-dependent,

which allows a modulation of receptor ligand interactions [95,96]. In addition to HLA class I ligands, the HERV-H LTR-associating 2 (HHLA2) molecule, a member of the B7 family that is highly expressed in several solid and hematological cancers, was recently identified as a ligand for KIR3DL3 [97]. HLA class I molecules also play a predominant role in NK cell education or licensing. Four models of NK cell education, which allow for the functional development and tolerance to self in NK cells, have been described [98]. Among them, the “arming” or “licensing” model, which was initially described in mice, involves the engagement of an inhibitory NK receptor with a self-MHC class I molecule, and this engagement educates NK cells to become functionally competent. Conversely, the absence of engagement of the inhibitory NK receptor with its ligand induces so-called “unlicensed” NK cells [99]. The education of KIR⁺ NK cells allows these cells to distinguish healthy cells with normal expression levels of HLA class I molecules from those with no or decreased expression of HLA class I molecules, thus referring to the “missing self” theory [7]. Different studies demonstrated the importance of KIR2DL1/2 [7] and 3DL1 [100] in NK cell education. KIR–HLA interactions could also occur in *cis* on NK cells [100]. Overall, NK cell education models are based on inhibitory KIRs, but activating KIRs could also play a role. Indeed, KIR2DS1⁺ NK cells are alloreactive toward C2 targets, but only in C2-individuals [85]. In C2 individuals, KIR2DS1⁺ NK cells become anergic due to the strong and prolonged exposure of 2DS1 to its autologous C2 ligand, which limits the autoreactive immune response [101].

In addition to the phenotype, allelic polymorphisms in KIRs influence the functionality of KIR⁺ NK cells. Thus, the lowest recognition of C2 ligands by KIR2DL1⁺ NK cells was observed for the KIR2DL1*004 allotype [34,74,75]. This decreased recognition was reported to be due to the mutation of Arg²⁴⁵ to Cys²⁴⁵. Indeed, this mutation plays a role in the transduction of inhibitory signals, as KIR2DL1 allotypes carrying Arg²⁴⁵ recruit SHP-2 and β -arrestin 2 molecules, both of which are important signaling molecules mediated by 2DL1 [74]. Recently, we reported that the KIR2DL1*002 allotype (Arg²⁴⁵) interacts with all C2 targets as observed with the 2DL1*004 allotype (Cys²⁴⁵), suggesting that polymorphic positions other than that of the intracytoplasmic 245 residue may play a crucial role [34]. To a lesser extent, the KIR2DL2/3 allotypes impact the inhibition of KIR2DL2/3⁺ NK cell degranulation, showing a better inhibition with the 2DL2 allotype than with the 2DL3 allotype [102]. The allelic polymorphism of HLA-C could also affect the binding affinity of KIR2DL/HLA-C and the consequent NK cell functionality. Similarly to KIR2D, allelic polymorphisms in KIRs modulate the functionality of KIR3D⁺ NK cells, as was found through extensive studies of KIR3DL1 [30,103,104].

HLA-Cw molecules are more specifically dedicated to sensing the level of alteration and/or decreases in HLA class I expression at the surface of tumor cells than to the presentation of peptides to TcR [105]. As described previously, all HLA-Cw molecules are ligands for KIRs in an allele-specific manner [48,106]. In healthy individuals, the expression level of HLA-Cw molecules on fresh PBMCs is lower than that of HLA-A and -B molecules [105]. The differential expression of HLA-Cw molecules according to tissue specificity must be further investigated. The level of HLA-Cw expression varies depending on the HLA-C alleles, HLA haplotypes and polymorphisms encountered in the coding and regulatory regions of the HLA-C gene [107–112]. Genome-wide association studies (GWAS) performed in US blood donors showed a biallelic polymorphism of some SNPs located in the 5'UTR region –800 bp [109] and –35 kb [107] upstream of the initiation codon of the HLA-C gene or in the 3'UTR region [110] that had an effect on the HLA-C expression level. Usually, the expression level of HLA-Cw4 or -Cw6 is higher than that of HLA-Cw2 or -Cw8, but a low expression level was reported for the HLA-Cw7 and -Cw3 molecules. While heterogeneity in the HLA-Cw expression level is observed according to the polymorphisms in the coding and regulatory regions, some discordances remain depending on the population studied (mostly US populations), the methodologies used (e.g., SNPs, evaluation of the HLA-Cw expression level using the HLA-C/HLA-E-specific DT9 mAb, or quantification of mRNA), and the allelic specificities of HLA-C under study.

At present, it is difficult to correlate all HLA-C specificities, SNP polymorphisms, levels of HLA-C transcripts, and the corresponding HLA-Cw expression levels in both healthy individuals and people with diseases.

Overall, the variability in KIR gene content, KIR allele polymorphisms, the HLA class I environment, and the clonal expression of KIRs result in the inter- and intraindividual diversity of the KIR⁺ NK cell repertoire (Figure 2).

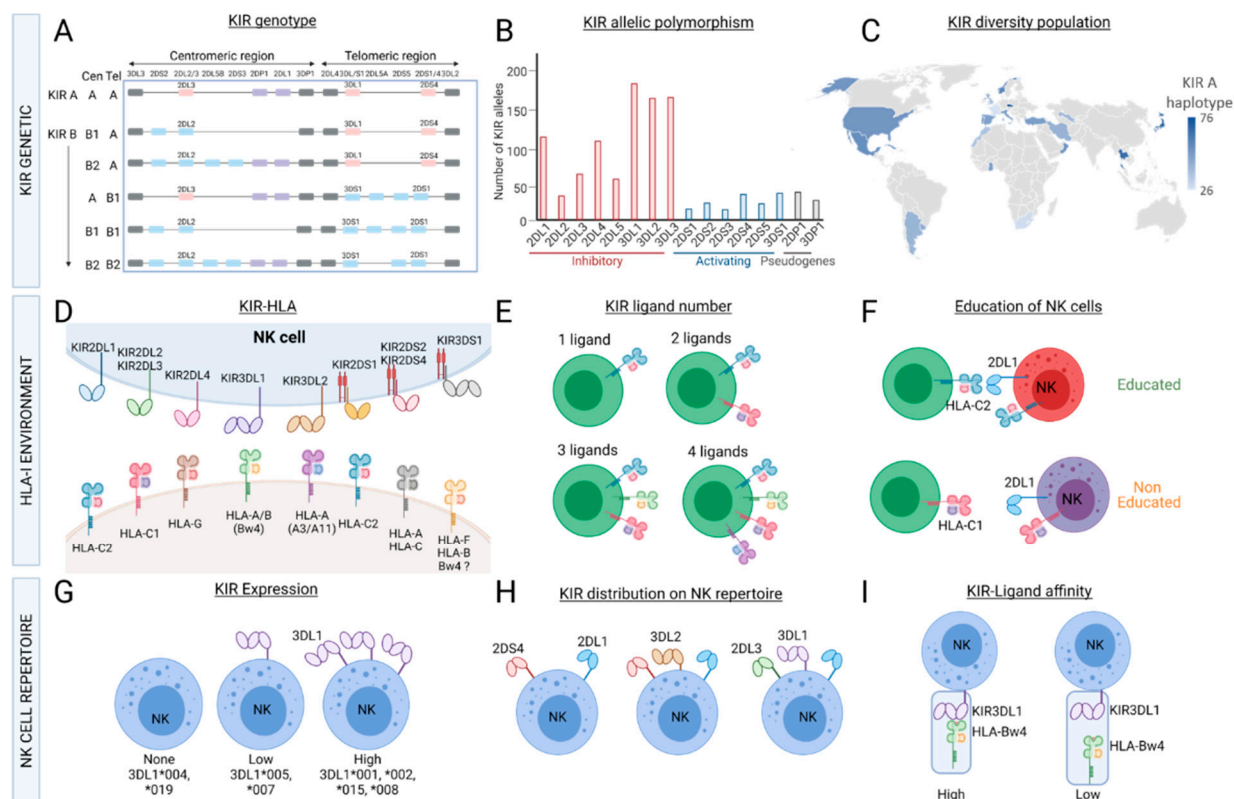


Figure 2. Variability of the KIR⁺ NK cell repertoire. KIR genes (7–14) are located in the leucocyte region complex (LRC) on chromosome 19. Depending on the KIR gene content, different KIR haplotypes (A and B) and centromeric/telomeric motifs are defined (A). KIR genes exhibit an allelic polymorphism (<https://www.ebi.ac.uk/ipd/kir/>) (accessed date: 26 July 2021). (B), and the diversity of KIR haplotypes and/or alleles can be observed worldwide (source provided by Dr J. Hollenbach from the 15th IHWG KIR component) (C). HLA class I molecules are ligands of KIRs (D). Depending on the typing of HLA-A, -B and -C, individuals can present from one to four KIR ligands (E). The HLA class I environment also play a major role in NK cell licensing (F). The allelic polymorphism of KIRs has an impact on the NK cell repertoire, as illustrated for the expression of different KIR3DL1 allotypes (G). KIRs are clonally distributed on NK cells; thus, the heterogeneous repertoire of NK cell subsets is based on the expression of different combinations of KIRs with different frequencies (H). The functions (cytotoxicity, cytokine production) of KIR⁺ NK cells and KIR/KIR ligand affinities (I) could be affected by both KIR and HLA class I polymorphisms.

3. Impacts of the KIR and HLA Class I Genes on the Antileukemic Effects of NK Cells in Patients with Acute Leukemia

3.1. Expression of HLA Class I Molecules on Leukemic Cells

Impaired recognition of tumor cells is associated with the loss of or decreased HLA expression [15]. This mechanism, which has been described in various solid tumors, could be related to genomic alterations of HLA genes, phenotypic alterations of HLA molecules, or changes in other elements involved in the antigenic presentation machinery. Toffalori et al. elegantly reported an immunological signature of relapse associated with a loss of HLA expression in AML patients [113]. In leukemic patients, the decreased expression of HLA class I molecules may depend on the loci, alleles, tissues, or cells, and it impacts

different HLA molecules, leading to a loss of HLA heterozygosity or a complete HLA haplotype loss [114–116]. The nature of HLA class I downregulation also varies depending on the origin of the leukemia [117]. The decrease in HLA expression occurs most frequently at the time of relapse, thus constituting a potential immunological signature [118]. In 2009, after haplo-identical HLA HSCT, Vago et al. reported a loss of the mismatched HLA haplotype in one-third of the patients (mostly with AML) who relapsed [119]. In a collaborative study, at the time of diagnosis of AML patients, we reported a loss of heterozygosity in the complete HLA region in half of the patients, as well as the loss of the HLA-A, -B, and -C genes—or only the HLA-A gene—in the remaining patients [120].

The impact of the HLA-Cw expression level on clinical outcomes after mismatched HLA-C HSCT remains relatively undocumented. HLA-C mismatches are usually reported to be associated with a higher incidence of acute graft-versus-host disease (GvHD) and higher mortality [121]. However, some frequently encountered HLA-C mismatches, such as HLA-C*03:03/*03:04 or HLA-C*07:01/*07:02, which correspond to a low HLA-C expression level in recipients and are evaluated using the expected MFI of the corresponding HLA-C specificities, are called permissive in the absence of deleterious effects after unrelated mismatched HLA-C HSCT [122–124]. In contrast, no effects caused by HLA-C level expression, which was evaluated by using the polymorphism of a miRNA in the 3'UTR, were reported after an unrelated HSCT [125]. These discordances could rely on HSCT cohorts or methods to correlate the levels of HLA-C expression according to the HLA-C polymorphism. The level HLA-Cw expression on leukemic cells or on reconstituting immune cells after HSCT is poorly documented due to the lack of specific mAbs available. One could expect that the level of HLA-Cw expression could modulate NK cell functions and their development through their *trans*- and *cis*-interactions with KIRs, respectively.

3.2. Antileukemic NK Cell Function in Patients with Acute Leukemia

In the context of acute leukemia, NK-cell-mediated immunosurveillance requires the establishment of an immunological synapse between NK cells and the target; this synapse involves the interactions between inhibitory and activating NK receptors and their ligands expressed on leukemic cells. NK cells express a wide range of activating receptors that recognize induced or upregulated molecules on tumor cells [19]. Moreover, cognate ligands are heterogeneously expressed on leukemic cells. In addition to the absence of expression or downregulation of HLA molecules, it was reported that the presence of activating ligands on the surface of leukemic cells was associated with the reduction of relapse incidence and improved overall survival of patients with acute leukemia [126]. We recently reported that ALL targets were better recognized by NK cells than AML targets. However, there is a broad interindividual disparity in NK cell responses against the same leukemic target, which highlights bad and good NK responders. Most effective NK cell subsets against different ALL targets express NKG2A, and these represent the most frequent subset in the NK cell repertoire. In contrast, minority CD57⁺ and/or KIR⁺ NK cell subsets are more efficient against AML targets [127]. Overall, our data may help to optimize the selection of HSC donors on the basis of immunogenetic KIR/HLA markers and to identify the best NK cell candidates in immunotherapies for AML patients.

4. Selection of HSC Donors Based on the HLA and KIR Genes

4.1. Selection of HSC Donors Based on HLA Matching

The matching of HLA class I (HLA-A, -B, -C) and class II (HLA-DRB1, -DQB1) genes between the donor and the recipient remains the gold standard [128,129] in order to prevent acute GvHD occurrence, to improve overall survival (OS), and to provide a beneficial GvL effect that is driven by alloreactive T and/or NK cells. The inheritance of HLA genes from the parental haplotypes in blocks results in 25% likelihood of finding a 10/10 HLA-identical donor among a patient's siblings. In the absence of related HLA-identical donors, an unrelated HLA-matched donor is sought in the registries. In an unrelated HSCT, an important feature to consider is that the HLA alleles, haplotype frequencies, and LD

between HLA genes may differ across populations [63]. Although there are more than 30 million potential unrelated HSC donors currently registered worldwide, finding suitably matched donors can be difficult for patients that harbor rare HLA alleles and/or because of the quality of available genotyping information. In the absence of a related or unrelated 10/10 HLA-matched donor, a 9/10 HLA-matched donor with selective mismatches could be selected. From 2010 to 2015, umbilical cord blood donors represented another option. For adult patients, two UCB donors were selected, and the level of HLA matching between the donor and recipient in double umbilical cord blood transplantation (dUCBT) was based on the HLA-A, -B, and -DRB1 genes (4/6 to 6/6 HLA matched). The impact of HLA-C matching remains to be clarified. Over the last 5 years, unmanipulated T-cell-repleted haplo-identical HSCT using cyclophosphamide post-transplantation (haplo-PTCy) as prophylaxis against GvHD has been recommended in the absence of a 10/10 HLA-identical donor. Haplo-PTCy is increasing in use and has shown similar outcomes to those observed after unrelated or even related HLA-matched HSCT [130,131]. Here, donors and recipients share only one HLA haplotype, resulting in a 50% likelihood of finding a haplo-identical donor among one's siblings and, therefore, increasing the number of potential HSC donors. The levels of HLA matching between a HSC donor and a recipient depending on the stem-cell sources are summarized in Figure 3.

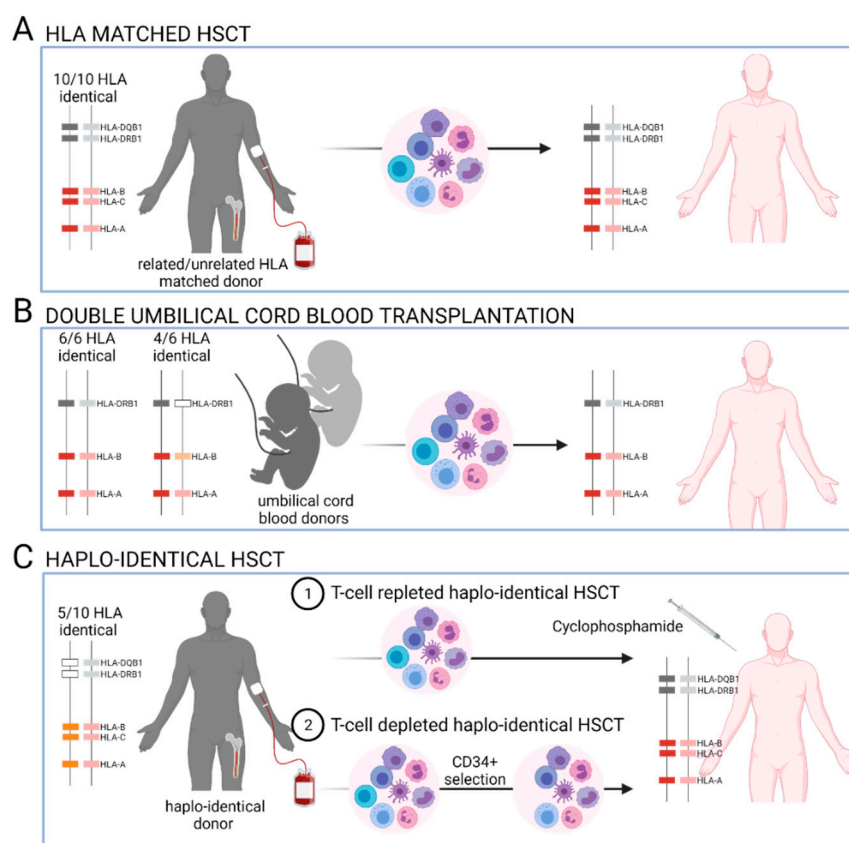


Figure 3. Donor/recipient HLA matching prior to allogeneic hematopoietic stem-cell transplantations (HSCT). The matching of the HLA-A, -B, -C, -DRB1, and -DQB1 genes between donors and recipients in both haplotypes is usually sought prior to related or unrelated HSCT (A). In the absence of related/unrelated 10/10 HLA-identical donors, double-umbilical cord blood transplantation (dUCBT) could be proposed so that only the matching of the HLA-A, -B, and -DR genes would be considered. In this context, the two UCB units and the recipient usually have 4/6 to 6/6 HLA matches (B). More recently, a donor with a 5/10 HLA-haplo-identical match is sought. The use of cyclophosphamide post-grafting as a prophylaxis against GvHD permits the deletion of the reconstitution of alloreactive T cells from T-repleted grafts (C).

4.2. KIR Models Used to Evaluate NK Cell Alloreactivity and Clinical Outcomes after Allogeneic HSCT

To evaluate the potential alloreactivity of KIR⁺ NK cells, the impacts of KIR ligands, KIR genes/genotypes, and KIR/KIR ligand (KIR-L) mismatches have been deeply investigated since 2000 in terms of HSCT outcomes; due to the heterogeneity of the HSCT cohorts and/or the predictive KIR models used, the results were discordant (Figure 4) [132,133].

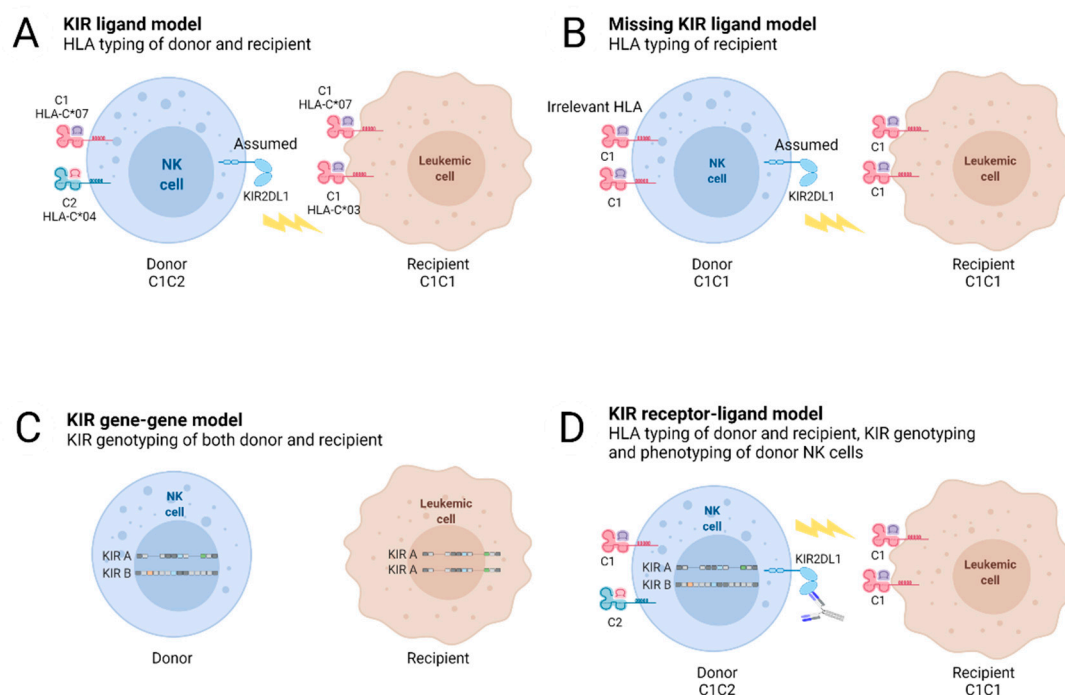


Figure 4. Genetic KIR/KIR ligand models defined to predict potential KIR⁺ NK cell alloreactivity (adapted from Symons et al. [134]). Genotyping of HLA class I and/or KIR genes permits the establishment of different KIR/KIR ligand models for use in HSCT. Knowledge of the HLA class I typing of donor/recipient pairs permits the identification of KIR ligand mismatches in the graft-versus-host (GvH) direction in HLA-mismatched HSCT, as defined by the KIR ligand model (A) or by the KIR ligands in recipients that show missing KIR ligands (B). In these contexts, the expression of KIR is only predictive. KIR genotyping of donors and recipients permits the assignment of donor and/or recipient KIR genes that impact the HSCT outcome, as defined by the KIR gene/gene model (C). Genotyping of HLA class I genes in donor/recipient pairs and of KIR genes in donors permits the identification of both educated KIR⁺ NK cells and missing KIR ligands in recipients (D). In this KIR receptor–ligand model, the expression of KIRs could be assessed using appropriated anti-KIR monoclonal antibodies (mAbs) or could be deduced from the donor’s KIR alleles (D).

The beneficial effect of NK cell alloreactivity based on HLA class I mismatches in terms of KIR ligands in the graft-versus-host direction (GvH) was initially underlined in the context of T-cell-depleted haplo-identical HSCTs, specifically for AML patients without relapse and aGvHD [135,136]. In parallel, the impacts of KIR ligands, donor KIR genes, and KIR-L mismatches on clinical outcomes were investigated after related or unrelated HSCTs, including HLA-matched and HLA-mismatched pairs. Notably, as HLA class I and KIR genes are located on different chromosomes, HLA-matched donor/recipient pairs could present relevant KIR gene mismatches. In related HLA-matched HSCT, the absence of KIR ligands in the recipient (missing KIR ligand, i.e., the absence of Bw4, C1 or C2) for the donor’s inhibitory KIRs was found to either be a beneficial prognostic factor by increasing OS and disease-free survival (DFS) and/or by decreasing relapse incidence for AML patients [137,138] or have no effect [101]. In addition to genetic studies of KIR, different profiles of NK cell reconstitution were reported with the establishment of a donor-like KIR repertoire, showing a gradual reconstitution from 6 to 9 months or longer to 1 year after related HLA-matched HSCT [137]. NKG2A⁺ KIR[−] NK cells were shown to

dominate early after HSCT, and NK cells that lacked inhibitory KIRs for self-HLA class I ligands remained tolerant at different points in T-cell-repleted and -depleted HLA-matched sibling HSCTs [101].

In unrelated HSCT, having a missing KIR ligand was found to be a predictive factor for relapse in adult patients with AML in HLA-mismatched pairs [139]. Interestingly, in a large series of unrelated HSCTs, including HLA-matched and single-mismatched pairs, Venstrom et al. reported a reduction in relapses in AML patients that were grafted with KIR2DS1+/C1+ donors [140]. In addition to KIR2DS1, we reported a beneficial impact of KIR3DL1/S1+ grafts in Bw4+ recipients with myeloid diseases on relapse incidence after unrelated HSCT [141]. Venstrom et al. also confirmed the beneficial impact of KIR3DS1+ donors by decreasing treatment-related mortality (TRM) and aGvHD, but found that there was no impact on relapses [142]. More broadly, a specific donor KIR B gene content score that is ≥ 2 [143], donor KIR B genotypes [144], or donor KIR CenB02 [145] should be favored for AML patients in order to decrease relapse incidence, which suggests that there is a need to detect the activating KIR genes. Notably, one prospective and multicentric study underlined the feasibility of donor KIR genotyping without delaying grafting [146]. In contrast, the assignment of donor KIR alleles is more time-consuming, and their effect on HSCT outcomes is primarily focused on limited KIR genes with phenotypic and functional characteristics of KIR⁺ NK cells, as was reported for KIR2DL1 [147]. Interestingly, a beneficial effect of specific donor KIR3DL1-HLA-B subtypes associated with weak or a lack of inhibition of NK cells was reported in a large series of AML patients that included unrelated 10/10 and 9/10 HLA-matched pairs [148]. Conversely, the genetic combination of donor KIR3DL1+ and patient HLA-A*24+ is associated with a higher risk of AML relapse following allogeneic HSCT [149]. Schetelig et al. reported that there were no correlations between KIR3DL1/HLA-B combinations and relapse or OS in unrelated HSCTs, which also included 10/10 and 9/10 HLA-matched pairs [150]. Recently, a retrospective study performed on a large cohort of AML patients reported that there was no impact of KIR3DL1/2DS1 or a haplotype-motif-based donor selection algorithm on OS and relapse after unrelated HLA matched HSCTs [151]. In line with HSCT protocols, the impact of KIR-L mismatches was investigated in adult patients after dUCBTs [152–158]. Once again, conflicting data were reported, which were linked either to the competing KIR models used or the heterogeneity of the patients and treatment modalities.

For the last 5 years, haplo-PTCy has offered a new means of identifying the KIR and/or HLA class I markers that are beneficial in improving the GvL effect driven by NK cells [159,160]. In this context, donors and recipients share only one HLA haplotype; thus, HLA class I mismatches could represent KIR-L mismatches. Donors and recipients could also have different KIR genes. In terms of inhibitory KIR gene mismatches, Symons et al. [134] reported a beneficial effect on the improvement of OS and the decrease in relapse and non-relapse mortality (NRM) after haplo-PTCy in 86 patients with lymphoid and myeloid diseases. Bastos-Oreiro et al. [161] reported the beneficial effect of at least one inhibitory KIR gene mismatch on the improvement of event-free survival (EFS) and the decrease in relapses after haplo-PTCy in 33 patients with various diseases. However, the specificity of the inhibitory KIR gene mismatches was not mentioned. Wanquet et al. [162] reported decreased relapses in patients with active diseases with KIR-L mismatches, but there were no beneficial effects of these mismatches for patients in complete remission after haplo-PTCy. Solomon et al. [163] reported a beneficial effect of KIR-L mismatches on OS and relapses after haplo-PTCy in 208 patients that received either a myeloablative conditioning (MAC) or a reduced-intensity conditioning (RIC) regimen. In 51 patients, which were all treated with an RIC regimen and infused with PBSCs, we reported a beneficial effect of KIR2D/HLA-C mismatches in the GvH direction, by decreasing the relapse incidence while increasing OS, aGvHD, and DFS after haplo-PTCy [164]. In contrast, in AML patients grafted with PBSCs, Shimoni et al. [165] reported a deleterious effect of KIR-L mismatches with increased relapses and decreased OS, after haplo-PTCy. In terms of KIR genotypes, Symons et al. [134] showed in 86 patients that KIR genotype AA recipients

from B⁺ KIR donors were associated with fewer relapses. The beneficial effect of donor B⁺ KIR was reported in both lymphoid and myeloid diseases and with RIC regimens [134]. Solomon et al. [163] reported a beneficial effect of the donor B⁺ KIR genotype on relapse incidence, particularly the presence of the KIR2DS2 gene in 208 patients with various diseases. Mixed bone marrow/PBSCs and both MAC and RIC regimens were used in these series. Lastly, Bastos-Oreiro et al. [161] reported that there was no protective effect of the donor B⁺ KIR genotype on relapse incidence after haplo-PTCy in a limited number of patients with mixed conditioning regimens. In terms of KIR gene motifs, we recently reported the impact of the donor KIR cenAA motif on relapse incidence after haplo-PTCy in 39 patients with myeloid diseases, which suggested that the GvL effect could be driven by specific KIR2DL1/3 allotypes [34]. The impact of KIR allele polymorphisms on haplo-PTCy outcomes has not been investigated so far.

Overall, these studies, which were mainly retrospective, demonstrated that the impacts of genetic combinations of KIR ligands, KIR genes, and/or KIR-L differ depending on diseases, immunosuppressive treatments, conditioning regimens, the stem-cell sources, the graft T-cell content, the GvHD prophylaxis, and the degree of HLA matching. However, HLA typing alone is not sufficient for providing the specificities of the donor KIR genes involved. Although the gene frequencies of KIR2DL1/3/3DL1 are close to 100%, allowing speculation about their presence depending on donor KIR ligands, the distinguishing between KIR2DL2 and 2DL3 is essential because their phenotypes and functions in NK cells differ [81]. KIR3DL1 allele typing also permits one to check for the absence of 3DL1*004, which is frequent in Caucasian people [46]. The reported and divergent effects of KIR/KIR ligand genetic combinations on HSCT outcomes (Table 1) suggest that donor KIR genotyping, including the detection of activating KIR genes, discrimination among KIR gene motifs, KIR allele polymorphisms, and the HLA class I environment, should be considered in order to predict the potential KIR⁺ NK cell alloreactivity after HSCT. Given the current literature, it is proposed that an HSC donor with B⁺ KIR gene content is selected for HLA-identical HSCTs, and an HSC donor with a KIR cenAA motif is selected for haplo-PTCy in order to limit the relapse incidence in adult patients with AML.

Table 1. Impacts of the genetic combinations of KIR/KIR ligands on clinical outcomes for adult AML patients, including selected studies that focused on both HLA-matched and HLA-mismatched HSCTs.

Authors	N	AML ¹	HSCT	HLA	Source ²	Depletion	Conditioning ³	Outcome ⁴
Hsu et al. [138]	178	32%	Related	Matched	BM	T-cell-depleted	MAC	AML patients with missing KIR ligands (HLA-C or HLA-B ligands): ↗ OS, ↗ DFS, ↘ relapse
Björklund et al. [101]	105	77%	Related	Matched	PBSC (67%) and BM (33%)	T-replete	MAC (72%) and RIC (28%)	No benefit of missing KIR ligand on DFS, relapse, and aGvHD
Hsu et al. [139]	1770	13%	Unrelated	Matched (48%) and mismatched (52%)	unknown	T-replete	MAC	AML patients with missing KIR ligand: ↘ relapse in mismatched HSCT but no impact on matched HSCT

Table 1. Cont.

Authors	N	AML ¹	HSCT	HLA	Source ²	Depletion	Conditioning ³	Outcome ⁴
Venstrom et al. [140]	1277	100%	Unrelated	Matched (52%) and mis-matched (48%)	BM (54%) and PBSC (46%)	T-replete (72%) and T-cell-depleted (28%)	MAC (85%) and RIC (15%)	Donor 2DS1+/C1+: ↘ TRM, ↘ relapse
Gagne et al. [141]	264	52% of acute leukemia	Unrelated	Matched (62%) and mis-matched (38%)	BM	T-replete	TBI based (70%)	Donor KIR3DL1/S1+ and recipient Bw4+: ↗ OS, ↘ relapse for HLA-matched and HLA-mismatched HSCs for malignant diseases
Venstrom et al. [142]	1087	28%	Unrelated	Matched (62%) and mis-matched (38%)	BM (97%)	T-replete (80%) and T-cell-depleted (20%)	Unknown	Donor KIR3DS1+: ↘ TRM, ↘ aGvHD, no impact on relapse
Cooley et al. [143]	1409	77%	Unrelated	Matched (50%) and mis-matched (50%)	BM (90%) and PBSC (10%)	T-replete	MAC	Donor CenBB or ≥ 2 KIR B gene content: ↗ DFS and ↘ relapse in matched and mismatched HSCs. Donor TelBB: ↘ relapse
Weisdorf et al. [144]	2662	100%	Unrelated	Matched (89%)	BM (15%) and PBSC (85%)	Unknown	MAC (59%) and RIC (41%)	Donor KIR B haplotype: ↘ relapse only in RIC patients. Donor KIR B genes/ recipient C1+: ↘ relapse
Boudreau et al. [148]	1328	100%	Unrelated	Matched (53%) and mis-matched (47%)	BM (54%) and PBSC (46%)	T-replete (92%)	MAC (84%) and RIC (16%)	Donor KIR3DL1/HLA-B subtype combinations linked to weak or non-inhibition: ↘ TRM, ↘ relapse. Distinct beneficial effect of donor KIR2DS1+/C1+
Ploeg et al. [149]	604	100%	Unrelated	Matched (64%) and mis-matched (36%)	BM (48%) and PBSC (52%)	T-replete (67%) and T-cell-depleted (33%)	MAC (85%) and RIC (15%)	Donor KIR3DL1+ and recipient HLA-A*24+: ↗ relapse

Table 1. Cont.

Authors	N	AML ¹	HSCT	HLA	Source ²	Depletion	Conditioning ³	Outcome ⁴
Ruggeri et al. [135]	92	62%	Related	Haplo-identical	unknown	T-cell depleted	Unknown	Donor/recipient KIR ligand mismatches in the GvH direction: no aGvHD and no relapse in AML patients
Symons et al. [134]	86	30%	Related	Haplo-identical	BM	T-replete	RIC, PTCy	Donor/recipient inh.KIR gene MM: \nearrow OS, \searrow relapse, donor KIR B/recipient KIR AA \nearrow OS, \searrow NRM
Bastos et al. [161]	33	30%	Related	Haplo-identical	unknown	T-replete	RIC (66%) and MAC (34%), PTCy	At least one donor/recipient inh KIR gene MM: \nearrow EFS, \searrow relapse. No protective effect of donor B genotype
Wanquet et al. [162]	144	35% of myeloid diseases	Related	Haplo-identical	PBSC (63%) and BM (37%)	T-replete	RIC (87%), MAC (13%), PTCy	Donor/recipient KIR ligand MM in the GvH direction: \searrow relapse, \nearrow PFS for patients with active disease but no beneficial effect for patients in complete remission
Solomon et al. [163]	208	34%	Related	Haplo-identical	PBSC and BM	T-replete	MAC (41%), RIC (59%), PTCy	Donor/recipient KIR ligand MM in the GvH direction or donor KIR2DS2+: \nearrow OS, \nearrow DFS, \searrow relapse. No impact on aGvHD
Shimoni et al. [165]	444	74%	Related	Haplo-identical	BM (53%) and PBSC (47%)	T-replete	MAC (55%) and RIC (45%), PTCy	Donor/recipient KIR ligand MM in the GvH direction: no impact on aGvHD, cGvHD and NRM. In AML patients grafted with PBSCs, \nearrow relapse and \searrow OS

Table 1. Cont.

Authors	N	AML ¹	HSCT	HLA	Source ²	Depletion	Conditioning ³	Outcome ⁴
Willem et al. [164]	51	35%	Related	Haplo-identical	PBSC	T-replete	RIC, PTCy	Donor inh.KIR2D/recipient HLA-C MM: \nearrow aGvHD, \nearrow OS, \nearrow DFS, \searrow relapse
Dubreuil et al. [34]	81	63% myeloid diseases	Related	Haplo-identical	PBSC	T-replete	RIC, PTCy	Donor CenAA: \searrow relapse only in patients with myeloid diseases

¹ AML, acute myeloid leukemia; ² BM, bone marrow; PBSCs, peripheral blood stem cells; ³ MAC, myeloablative conditioning; PTCy, post-transplant cyclophosphamide; RIC, reduced-intensity regimen; ⁴ aGvHD, acute graft-versus-host disease; CenAA, centromeric A KIR motif; CenBB, centromeric BB KIR motif; DFS, disease-free survival; HSCT, hematopoietic stem-cell transplantation; inh. KIR, inhibitory KIR; MM, mismatch; NRM, non-relapse mortality; OS, overall survival; PFS, progression-free survival; TRM, treatment-related mortality.

4.3. Reconstitution of NK Cells after T-Replete Haploidentical HSCT

After haplo-PTCy, NK cells rapidly reconstitute the recipient's immune system, as they are the first lymphocytes to mediate a GvL effect, even before T cells [164,166–168]. The prophylaxis against GvHD plays a major role in the modulation of the reconstitution of immune cells and possibly in their function after haplo-HSCT. Recovery of NK cells after haplo-PTCy was reported by Vago's group in 2018 [168]. A similar NK cell recovery was observed in patients who received different PTCy protocols. PTCy first eliminates proliferating donor NK cells; then, an IL15 peak correlated with the kinetics of NK cell recovery appears, and a second wave of reconstituted NK cells with an immature phenotype occurs 2 weeks after HSCT. The development of mature NK cells can take place up to 1 year after HSCT. PTCy decreases NK cell alloreactivity, and the GvL effect is correlated with residual mature NK cells. Overall, there are predictive factors that correspond to the absolute counts of NK cells and the proportion of mature NK cells at day +30. A high expression of KIRs on NK cells at day +30 was correlated with a decreased relapse incidence [168]. Consistently with the data from Russo et al., we evaluated NK cell reconstitution in a cohort of 51 adult patients after haplo-PTCy [164]. KIR2DL2/3⁺ and 3DL1⁺ NK cell recovery at day +30 was inversely impacted by KIR-L mismatches. The loss of KIR2DL2/3⁺ NK cells at day 30 in pairs with inhibitory KIR2D/HLA-C mismatches suggested a deletion of alloreactive KIR⁺ NK cells by PTCy [164]. After haplo-PTCy, the reconstitution of T and NK cells revealed increased NK and decreased T-cell frequencies in patients treated with PTCy + ATG compared to PTCy alone [167]. In patients without relapse, the frequency of NKG2A⁺ KIR2DL2/3/S2[−] NK cells was increased at day +60 and these cells were more numerous in patients with PTCy + ATG compared to PTCy alone. The use of the NKG2A, CD57, NKG2C, and KIR2DL3 markers expressed on NK cells at day +30 made it possible to define an immature NK cell repertoire (i.e., KIR2DL3[−]NKG2A⁺CD57[−]) associated with fewer relapses [167]. In contrast, the predominant KIR2DL3[−]NKG2A[−]CD57[−] NK cell subset was associated with a high incidence of relapse [167]. Overall, deciphering the heterogeneity of NK cells is crucial for identifying the NK cell subsets that are efficient against hematological diseases in order to optimize HSC donor selection and, more broadly, NK-cell-based immunotherapies, as discussed below.

5. NK-Cell-Based Immunotherapies Apart from HSCT

In leukemic patients, the functions of NK cells are usually impaired, which is particularly linked to the alteration of their repertoire. Thus, NK-cell-based immunotherapies are devoted to increasing their antileukemic potential in patients by using the addition of specific cytokines, mAbs, and/or the adoptive transfer of unmodified or engineered NK cells [18,19,169–174].

5.1. Immune Molecules for Boosting NK Cell Functions in Leukemic Patients

Among the various cytokines, IL-2 and IL-15 are of interest for promoting the survival, proliferation, differentiation, and activation of NK cells. In particular, ALT-803, a super-agonist of IL-15 that was administrated to 33 patients who relapsed after HSCT, showed a clinical benefit in 19 patients in a phase I trial [175]. In clinical studies that combined ALT-803 with adoptive transfer of NK cells, ALT-803 was expected to enhance the activation, survival, and expansion of donor-derived NK cells. Moreover, the cocktail of IL-12, IL-15, and IL-18 used in vitro to promote NK cell proliferation and activation unleashes tumor killing of NK cells via KIR downregulation [176]. Apart from cytokines, mAbs that are specific for tumor antigens could be used to enhance the ADCC function of NK cells through the binding of the IgG Fc fragment with the CD16 receptor. In particular, an Fc-engineered CD133 mAb showed an improved affinity to NK cells and potent NK cell degranulation against AML [177]. Among mAbs, bi- or tri-specific killer engagers (BiKEs, TriKEs) are used to modulate NK cell activity. They are designed to contain variable single-chain fragments that function against both tumor-associated antigens and NK-activating receptors in order to create an immunological synapse between NK cells and tumor cells [178]. In particular, it has been reported that the CD16x33 BiKE targets the CD33 myeloid antigen and triggers the lytic activity of NK cells and the release of cytotoxic granules against AML cells [179] in vitro. Moreover, the 161533 TriKE, which consists of an anti-CD16 scFv, a modified IL-15 linker, and an anti-CD33 scFv, could trigger ADCC while increasing both NK cell survival and proliferation against AML targets in vivo [180]. NK-cell-based immunotherapies using mAbs could also target immune checkpoints to raise the inhibition driven by NK cells. Interestingly, lirulimab and monalizumab target the pan-KIR and NKG2A inhibitory NK cell receptors, respectively [181]. PD-1 could also represent an interesting target [171]. KIR/HLA interactions could also have an impact on the ADCC function of NK cells, as we recently reported in lymphoma patients who received a rituximab-based anticancer immunotherapy [182].

5.2. Adoptive NK Cell Immunotherapies

Infusions of allogeneic NK cells that are activated ex vivo or CAR-modified NK cells represent promising immunotherapies against hematologic malignancies. In contrast to T lymphocytes, allogeneic NK cells are not HLA-restricted and can be more safely used. Another advantage is the wide range of NK cell sources such as PB, UCB, embryonic stem cells, induced pluripotent stem cells, and NK cell lines [183,184]. In several clinical trials of NK-cell-based immunotherapies, patients received chemotherapy prior to NK cell infusion. Interestingly, the drugs used could stimulate the expression of activating ligands or either downregulate the expression of inhibitory ligands on tumor cell [185] or promote the recruitment of NK cells to the tumor sites by increasing the secretion of selected chemokines. In addition to the adoptive transfer of unmodified NK cells, CAR NK cells could be used. Similarly to CAR T cells, CAR NK cells are composed of an extracellular signaling domain that recognizes specific antigens on tumors, a transmembrane region, and an intracellular domain that activates pathways to enhance the lysis of target cells. CAR NK cells eliminate tumors not only through the ability of CAR to specifically recognize antigen-expressing tumors, but also through NK cell receptors themselves [20]. Preclinical trials of NK cells demonstrated their clinical benefits in AML patients after HSCT. However, the clinical activity of NK cells remains modest at best, which limits their current therapeutic use. CAR-modified NK92 and primary NK cells are currently used against hematological cancers. The tumor antigen that is targeted is often CD19 or CD20, which makes it possible to target B-cell malignancies, such as B-ALL or CD33, in order to target AML [19]. However, some issues related to the survival rate and cytotoxicity of NK cells after thawing need to be solved in order to expand the use of CAR NK cells in clinical applications. New strategies, including the integration of suicide genes into CAR constructs and bispecific CAR molecules, are being developed to increase both the safety and the efficacy of CAR

NK cell therapy [20]. The NK-cell-based immunotherapies used for the treatment of acute leukemia are summarized in Figure 5.

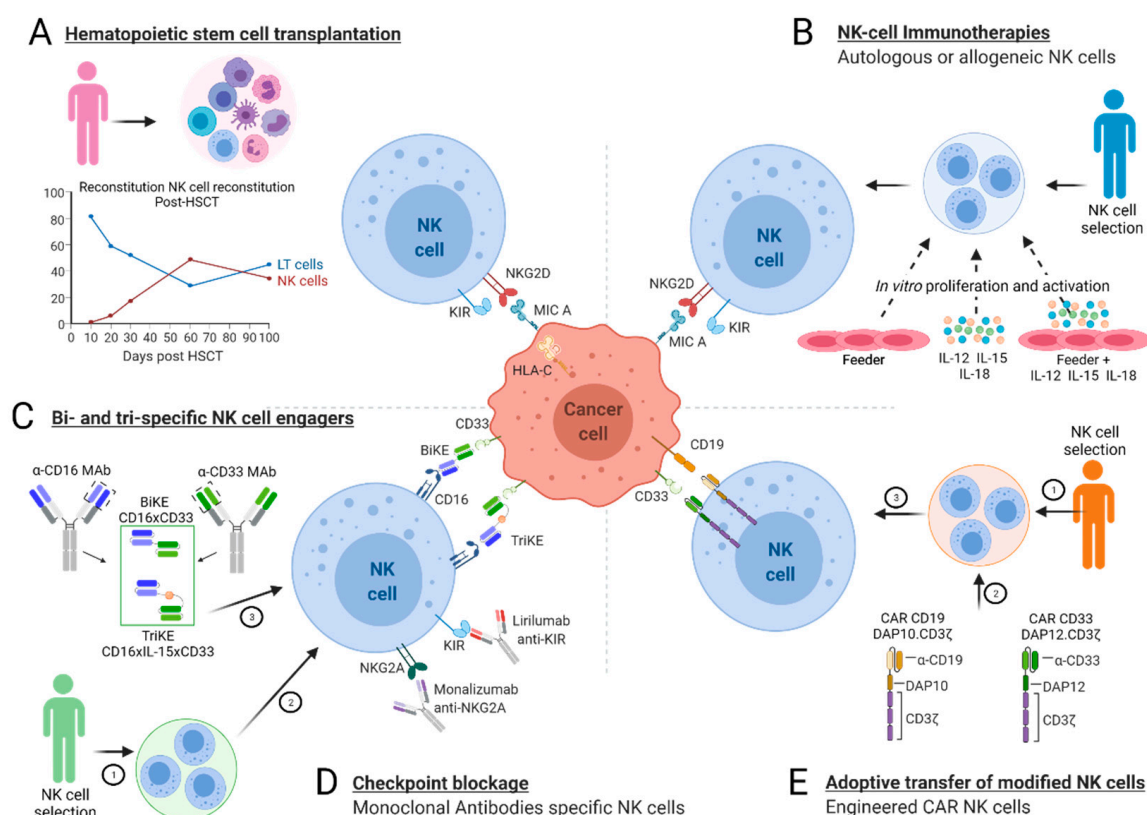


Figure 5. NK-cell-based immunotherapies for acute leukemia patients. Because of their antileukemic (GvL) effects, NK cells play a major role, as they are the first cells to reconstitute the recipient's immune system after allogeneic hematopoietic stem-cell transplantation (HSCT) (A). Apart from HSCT, the efficiency of NK-cell based immunotherapies could be enhanced with the infusion of autologous or allogeneic NK cells that are activated in vitro using a feeder and/or specific cytokines (B). The use of bi- or tri-specific killer engagers (BiKEs, TriKEs) (C) or specific anti-NK cell receptor monoclonal antibodies (mAbs) (D) could also enhance the GvL function of NK cells. Adoptive transfer of engineered NK cells armed with chimeric antigen receptors (CARs) (E) is another emerging alternative.

Overall, NK-cell-based immunotherapies could be combined to boost the efficacy of NK cells against leukemia, depending on the patients. However, given the heterogeneity of NK cells, the source of NK cells is still a key element, and allogeneic donors should be carefully selected by using specific KIR/HLA markers.

6. Conclusions

Polymorphic KIR and HLA class I genes play a major role in the structuration of the functional NK cell repertoire. In the context of HSCT, they are of special interest for the promotion of a beneficial GvL effect in leukemic patients. Haplo-PTCy makes it possible to choose the best HSC donor among different ones. Until now, few studies have focused on the relevance of polymorphic KIR/KIR ligands to the outcomes of haplo-PTCy. One could expect that the consideration of the KIR gene content or specific KIR alleles could help to refine the scores used for HSC donor selection, as well as to evaluate their influence on haplo-PTCy outcomes in patients with high-risk and acute leukemia. The ultimate goal is to define the best KIR and/or HLA markers during HSC donor selection with respect to a beneficial GvL effect and a reduced GvHD incidence after haplo-PTCy. More broadly, the inclusion of KIR/HLA markers should improve the efficiency of allogeneic NK-cell-based immunotherapies, such as treatment with CAR NK cells.

Author Contributions: Writing—original draft preparation, L.D. and K.G.; review and editing, L.D., P.C., C.R. and K.G. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by EFS Centre-Pays de la Loire and by grants from the International Research Group on unrelated Hematopoietic Stem-Cell Transplantation (IRGHET), Etablissement Français du Sang (EFS), Leucémie Espoir Atlantique Famille (LEAF), and La Ligue contre le Cancer and Agence de la Biomédecine (ABM). L.D. is a PhD student supported by la Région Pays de la Loire/EFS Centre Pays de la Loire (N°2018-09766).

Acknowledgments: The authors acknowledge MDPI for English language editing. The figures were created using the BioRender software (www.BioRender.com).

Conflicts of Interest: The authors declare no conflict of interest.

References

- Cooper, M.; Fehniger, T.; Caligiuri, M.A. The biology of human natural killer-cell subsets. *Trends Immunol.* **2001**, *22*, 633–640. [[CrossRef](#)]
- Vivier, E.; Artis, D.; Colonna, M.; Dieffenbach, A.; Di Santo, J.P.; Eberl, G.; Koyasu, S.; Locksley, R.M.; McKenzie, A.N.; Mebius, R.E.; et al. Innate Lymphoid Cells: 10 Years On. *Cell* **2018**, *174*, 1054–1066. [[CrossRef](#)]
- Schwane, V.; Huynh-Tran, V.H.; Vollmers, S.; Yakup, V.M.; Sauter, J.; Schmidt, A.H.; Peine, S.; Altfeld, M.; Richert, L.; Körner, C. Distinct Signatures in the Receptor Repertoire Discriminate CD56bright and CD56dim Natural Killer Cells. *Front. Immunol.* **2020**, *11*, 568927. [[CrossRef](#)]
- Kärre, K. Natural killer cell recognition of missing self. *Nat. Immunol.* **2008**, *9*, 477–480. [[CrossRef](#)] [[PubMed](#)]
- Ljunggren, H.-G.; Kärre, K. In search of the ‘missing self’: MHC molecules and NK cell recognition. *Immunol. Today* **1990**, *11*, 237–244. [[CrossRef](#)]
- Djaoud, Z.; Parham, P. HLAs, TCRs, and KIRs, a Triumvirate of Human Cell-Mediated Immunity. *Annu. Rev. Biochem.* **2020**, *89*, 717–739. [[CrossRef](#)]
- Anfossi, N.; André, P.; Guia, S.; Falk, C.S.; Roetynck, S.; Stewart, C.A.; Bresó, V.; Frassati, C.; Revirón, D.; Middleton, D.; et al. Human NK Cell Education by Inhibitory Receptors for MHC Class I. *Immunity* **2006**, *25*, 331–342. [[CrossRef](#)]
- Valiante, N.M.; Uhrberg, M.; Shilling, H.G.; Lienert-Weidenbach, K.; Arnett, K.; D’Andrea, A.; Phillips, J.H.; Lanier, L.L.; Parham, P. Functionally and Structurally Distinct NK Cell Receptor Repertoires in the Peripheral Blood of Two Human Donors. *Immunity* **1997**, *7*, 739–751. [[CrossRef](#)]
- Horowitz, A.; Strauss-Albee, D.; Leipold, M.; Kubo, J.; Nemat-Gorgani, N.; Dogan, O.C.; Dekker, C.L.; Mackey, S.; Maecker, H.; Swan, G.E.; et al. Genetic and Environmental Determinants of Human NK Cell Diversity Revealed by Mass Cytometry. *Sci. Transl. Med.* **2013**, *5*, 208ra145. [[CrossRef](#)] [[PubMed](#)]
- Parham, P. Co-evolution of lymphocyte receptors with MHC class I. *Immunol. Rev.* **2015**, *267*, 1–5. [[CrossRef](#)]
- Björkström, N.; Riese, P.; Heuts, F.; Andersson, S.; Fauriat, C.; Ivarsson, M.A.; Björklund, A.T.; Tullberg, M.F.; Michaëlsson, J.; Rottenberg, M.; et al. Expression patterns of NKG2A, KIR, and CD57 define a process of CD56dim NK-cell differentiation uncoupled from NK-cell education. *Blood* **2010**, *116*, 3853–3864. [[CrossRef](#)]
- Djaoud, Z.; David, G.; Bressollette, C.; Willem, C.; Rettman, P.; Gagne, K.; Legrand, N.; Mehlal, S.; Cesbron, A.; Imbert-Marcille, B.-M.; et al. Amplified NKG2C+NK Cells in Cytomegalovirus (CMV) Infection Preferentially Express Killer Cell Ig-like Receptor 2DL: Functional Impact in Controlling CMV-Infected Dendritic Cells. *J. Immunol.* **2013**, *191*, 2708–2716. [[CrossRef](#)] [[PubMed](#)]
- Malmberg, K.-J.; Béziat, V.; Ljunggren, H.-G. Spotlight on NKG2C and the human NK-cell response to CMV infection. *Eur. J. Immunol.* **2012**, *42*, 3141–3145. [[CrossRef](#)] [[PubMed](#)]
- Ljunggren, H.-G.; Malmberg, K.-J. Prospects for the use of NK cells in immunotherapy of human cancer. *Nat. Rev. Immunol.* **2007**, *7*, 329–339. [[CrossRef](#)] [[PubMed](#)]
- Zeiser, R.; Vago, L. Mechanisms of immune escape after allogeneic hematopoietic cell transplantation. *Blood* **2019**, *133*, 1290–1297. [[CrossRef](#)]
- Vacca, P.; Montaldo, E.; Croxatto, D.; Moretta, F.; Bertina, A.; Vitale, C.; Locatelli, F.; Mingari, M.C.; Moretta, L. NK Cells and Other Innate Lymphoid Cells in Hematopoietic Stem Cell Transplantation. *Front. Immunol.* **2016**, *7*, 188. [[CrossRef](#)]
- Triplett, B.M.; Horwitz, E.M.; Iyengar, R.; Turner, V.; Holladay, M.S.; Gan, K.; Behm, F.G.; Leung, W. Effects of activating NK cell receptor expression and NK cell reconstitution on the outcomes of unrelated donor hematopoietic cell transplantation for hematologic malignancies. *Leukemia* **2009**, *23*, 1278–1287. [[CrossRef](#)] [[PubMed](#)]
- Hu, W.; Wang, G.; Huang, D.; Sui, M.; Xu, Y. Cancer Immunotherapy Based on Natural Killer Cells: Current Progress and New Opportunities. *Front. Immunol.* **2019**, *10*, 1205. [[CrossRef](#)]
- Shimasaki, N.; Jain, A.; Campana, D. NK cells for cancer immunotherapy. *Nat. Rev. Drug Discov.* **2020**, *19*, 200–218. [[CrossRef](#)]
- Wang, W.; Jiang, J.; Wu, C. CAR-NK for tumor immunotherapy: Clinical transformation and future prospects. *Cancer Lett.* **2020**, *472*, 175–180. [[CrossRef](#)]
- Chen, S. Releasing the brake in CAR natural killer cells. *Blood* **2021**, *137*, 579–581. [[CrossRef](#)]

22. Borrego, F.; Kabat, J.; Kim, D.-K.; Lieto, L.; Maasho, K.; Peña, J.; Solana, R.; Coligan, J.E. Structure and function of major histocompatibility complex (MHC) class I specific receptors expressed on human natural killer (NK) cells. *Mol. Immunol.* **2002**, *38*, 637–660. [\[CrossRef\]](#)
23. Trowsdale, J.; Barten, R.; Haude, A.; Stewart, C.A.; Beck, S.; Wilson, M.J. The genomic context of natural killer receptor extended gene families. *Immunol. Rev.* **2001**, *181*, 20–38. [\[CrossRef\]](#)
24. Hsu, K.C.; Chida, S.; Geraghty, D.E.; Dupont, B. The killer cell immunoglobulin-like receptor (KIR) genomic region: Gene-order, haplotypes and allelic polymorphism. *Immunol. Rev.* **2002**, *190*, 40–52. [\[CrossRef\]](#) [\[PubMed\]](#)
25. Abraham, J.P.; Barker, D.J.; Robinson, J.; Maccari, G.; Marsh, S.G.E. The IPD Databases: Cataloguing and Understanding Allele Variants. *Methods Mol. Biol.* **2018**, *1802*, 31–48. [\[CrossRef\]](#)
26. O'Connor, G.M.; McVicar, D.W. The Yin-Yang of KIR3DL1/S1: Molecular Mechanisms and Cellular Function. *Crit. Rev. Immunol.* **2013**, *33*, 203–218. [\[CrossRef\]](#)
27. Vilches, C.; Pando, M.J.; Parham, P. Genes encoding human killer-cell Ig-like receptors with D1 and D2 extracellular domains all contain untranslated pseudoexons encoding a third Ig-like domain. *Immunogenetics* **2000**, *51*, 639–646. [\[CrossRef\]](#) [\[PubMed\]](#)
28. Wilson, M.J.; Torkar, M.; Trowsdale, J. Genomic organization of a human killer cell inhibitory receptor gene. *Tissue Antigens* **1997**, *49*, 574–579. [\[CrossRef\]](#)
29. Uhrberg, M.; Valiante, N.M.; Shum, B.P.; Shilling, H.G.; Lienert-Weidenbach, K.; Corliss, B.; Tyan, D.; Lanier, L.L.; Parham, P. Human Diversity in Killer Cell Inhibitory Receptor Genes. *Immunity* **1997**, *7*, 753–763. [\[CrossRef\]](#)
30. Yawata, M.; Yawata, N.; Draghi, M.; Little, A.-M.; Partheniou, F.; Parham, P. Roles for HLA and KIR polymorphisms in natural killer cell repertoire selection and modulation of effector function. *J. Exp. Med.* **2006**, *203*, 633–645. [\[CrossRef\]](#)
31. Augusto, D.; Norman, P.J.; Dandekar, R.; Hollenbach, J.A. Fluctuating and Geographically Specific Selection Characterize Rapid Evolution of the Human KIR Region. *Front. Immunol.* **2019**, *10*, 989. [\[CrossRef\]](#)
32. Hollenbach, J.A.; Augusto, D.; Alaez, C.; Bubnova, L.N.; Fae, I.; Fischer, G.; Gonzalez-Galarza, F.F.; Gorodezky, C.; Karabon, L.; Kuśnierz, P.; et al. 16th IHIW: Population Global Distribution of Killer Immunoglobulin-like Receptor (KIR) and Ligands. *Int. J. Immunogenet.* **2012**, *40*, 39–45. [\[CrossRef\]](#) [\[PubMed\]](#)
33. Jiang, W.; Johnson, C.; Jayaraman, J.; Simecek, N.; Noble, J.; Moffatt, M.F.; Cookson, W.O.; Trowsdale, J.; Traherne, J.A. Copy number variation leads to considerable diversity for B but not A haplotypes of the human KIR genes encoding NK cell receptors. *Genome Res.* **2012**, *22*, 1845–1854. [\[CrossRef\]](#)
34. Dubreuil, L.; Maniangu, B.; Chevallier, P.; Quémener, A.; Legrand, N.; Béné, M.C.; Willem, C.; David, G.; Alizadeh, M.; Makanga, D.R.; et al. Centromeric KIR AA Individuals Harbor Particular KIR Alleles Conferring Beneficial NK Cell Features with Implications in Haplo-Identical Hematopoietic Stem Cell Transplantation. *Cancers* **2020**, *12*, 3595. [\[CrossRef\]](#) [\[PubMed\]](#)
35. Middleton, D.; Meenagh, A.; Gourraud, P.-A. KIR haplotype content at the allele level in 77 Northern Irish families. *Immunogenetics* **2007**, *59*, 145–158. [\[CrossRef\]](#)
36. Pyo, C.-W.; Guethlein, L.A.; Vu, Q.; Wang, R.; Abi-Rached, L.; Norman, P.; Marsh, S.G.E.; Miller, J.S.; Parham, P.; Geraghty, D.E. Different Patterns of Evolution in the Centromeric and Telomeric Regions of Group A and B Haplotypes of the Human Killer Cell Ig-Like Receptor Locus. *PLoS ONE* **2010**, *5*, e15115. [\[CrossRef\]](#) [\[PubMed\]](#)
37. Vendelbosch, S.; De Boer, M.; Gouw, R.A.T.W.; Ho, C.K.Y.; Geissler, J.; Swelsen, W.T.N.; Moorhouse, M.J.; Lardy, N.M.; Roos, D.; Berg, T.K.V.D.; et al. Extensive Variation in Gene Copy Number at the Killer Immunoglobulin-Like Receptor Locus in Humans. *PLoS ONE* **2013**, *8*, e67619. [\[CrossRef\]](#)
38. Traherne, J.A.; Martin, M.; Ward, R.; Ohashi, M.; Pellett, F.; Gladman, D.; Middleton, D.; Carrington, M.; Trowsdale, J. Mechanisms of copy number variation and hybrid gene formation in the KIR immune gene complex. *Hum. Mol. Genet.* **2009**, *19*, 737–751. [\[CrossRef\]](#)
39. Robinson, J.; Halliwell, J.A.; McWilliam, H.; López, R.; Marsh, S.G.E. IPD—The Immuno Polymorphism Database. *Nucleic Acids Res.* **2012**, *41*, D1234–D1240. [\[CrossRef\]](#)
40. Schellekens, J.; Gagne, K.; Marsh, S.G.E. Natural Killer Cells and Killer-Cell Immunoglobulin-Like Receptor Polymorphisms: Their Role in Hematopoietic Stem Cell Transplantation. *Methods Mol. Biol.* **2013**, *1109*, 139–158. [\[CrossRef\]](#)
41. Maniangu, B.; Legrand, N.; Alizadeh, M.; Guyet, U.; Willem, C.; David, G.; Charpentier, E.; Walencik, A.; Retière, C.; Gagne, K.; et al. Killer Immunoglobulin-Like Receptor Allele Determination Using Next-Generation Sequencing Technology. *Front. Immunol.* **2017**, *8*, 547. [\[CrossRef\]](#)
42. Wagner, I.; Schefzyk, D.; Pruschke, J.; Schöfl, G.; Schöne, B.; Gruber, N.; Lang, K.; Hofmann, J.; Gnahn, C.; Heyn, B.; et al. Allele-Level KIR Genotyping of More Than a Million Samples: Workflow, Algorithm, and Observations. *Front. Immunol.* **2018**, *9*, 2843. [\[CrossRef\]](#)
43. Norman, P.J.; Hollenbach, J.A.; Nemat-Gorgani, N.; Marin, W.M.; Norberg, S.J.; Ashouri, E.; Jayaraman, J.; Wroblewski, E.E.; Trowsdale, J.; Rajalingam, R.; et al. Defining KIR and HLA Class I Genotypes at Highest Resolution via High-Throughput Sequencing. *Am. J. Hum. Genet.* **2016**, *99*, 375–391. [\[CrossRef\]](#)
44. Meenagh, A.; Gonzalez, A.; Sleator, C.; McQuaid, S.; Middleton, D. Investigation of killer cell immunoglobulin-like receptor gene diversity, KIR2DL1 and KIR2DS1. *Tissue Antigens* **2008**, *72*, 383–391. [\[CrossRef\]](#)
45. Pando, M.J.; Gardiner, C.; Gleimer, M.; McQueen, K.L.; Parham, P. The Protein Made from a Common Allele of KIR3DL1 (3DL1*004) Is Poorly Expressed at Cell Surfaces due to Substitution at Positions 86 in Ig Domain 0 and 182 in Ig Domain 1. *J. Immunol.* **2003**, *171*, 6640–6649. [\[CrossRef\]](#)

46. Gagne, K.; Willem, C.; Legrand, N.; Djaoud, Z.; David, G.; Rettman, P.; Bressollette-Bodin, C.; Senitzer, D.; Esbelin, J.; Cesbron-Gautier, A.; et al. Both the nature of KIR3DL1 alleles and the KIR3DL1/S1 allele combination affect the KIR3DL1 NK-cell repertoire in the French population. *Eur. J. Immunol.* **2013**, *43*, 1085–1098. [[CrossRef](#)] [[PubMed](#)]
47. Shilling, H.G.; Guethlein, L.A.; Cheng, N.W.; Gardiner, C.M.; Rodriguez, R.; Tyan, D.; Parham, P. Allelic Polymorphism Synergizes with Variable Gene Content to Individualize Human KIR Genotype. *J. Immunol.* **2002**, *168*, 2307–2315. [[CrossRef](#)] [[PubMed](#)]
48. Hilton, H.; Guethlein, L.A.; Goyos, A.; Nemat-Gorgani, N.; Bushnell, D.A.; Norman, P.; Parham, P. Polymorphic HLA-C Receptors Balance the Functional Characteristics of KIR Haplotypes. *J. Immunol.* **2015**, *195*, 3160–3170. [[CrossRef](#)] [[PubMed](#)]
49. Huhn, O.; Chazara, O.; Ivarsson, M.A.; Retière, C.; Venkatesan, T.C.; Norman, P.J.; Hilton, H.G.; Jayaraman, J.; Traherne, J.A.; Trowsdale, J.; et al. High-Resolution Genetic and Phenotypic Analysis of KIR2DL1 Alleles and Their Association with Pre-Eclampsia. *J. Immunol.* **2018**, *201*, 2593–2601. [[CrossRef](#)] [[PubMed](#)]
50. Vargas, L.D.B.; Dourado, R.M.; Amorim, L.M.; Ho, B.; Calonga-Solís, V.; Issler, H.C.; Marin, W.M.; Beltrame, M.H.; Petzl-Erler, M.L.; Hollenbach, J.A.; et al. Single Nucleotide Polymorphism in KIR2DL1 Is Associated With HLA-C Expression in Global Populations. *Front. Immunol.* **2020**, *11*, 1881. [[CrossRef](#)]
51. Bono, M.; Pende, D.; Bertaina, A.; Moretta, A.; Della Chiesa, M.; Sivori, S.; Zecca, M.; Locatelli, F.; Moretta, L.; Bottino, C.; et al. Analysis of KIR3DP1 Polymorphism Provides Relevant Information on Centromeric KIR Gene Content. *J. Immunol.* **2018**, *201*, 1460–1467. [[CrossRef](#)]
52. Spierings, E.; Fleischhauer, K. Histocompatibility. In *The EBMT Handbook: Hematopoietic Stem Cell Transplantation and Cellular Therapies*; Carreras, E., Dufour, C., Mohty, M., Kröger, N., Eds.; Springer: Cham, Switzerland, 2019; ISBN 978-3-030-02277-8.
53. Robinson, J.; Barker, D.J.; Georgiou, X.; Cooper, M.A.; Flicek, P.; Marsh, S.G.E. IPD-IMGT/HLA Database. *Nucleic Acids Res.* **2019**, *48*, D948–D955. [[CrossRef](#)] [[PubMed](#)]
54. Trowsdale, J.; Knight, J.C. Major Histocompatibility Complex Genomics and Human Disease. *Annu. Rev. Genom. Hum. Genet.* **2013**, *14*, 301–323. [[CrossRef](#)]
55. Robinson, J.; Guethlein, L.A.; Cereb, N.; Yang, S.Y.; Norman, P.J.; Marsh, S.G.E.; Parham, P. Distinguishing functional polymorphism from random variation in the sequences of >10,000 HLA-A, -B and -C alleles. *PLoS Genet.* **2017**, *13*, e1006862. [[CrossRef](#)]
56. Dos Santos, E.J.M.; McCabe, A.; Gonzalez-Galarza, F.F.; Jones, A.; Middleton, D. Allele Frequencies Net Database: Improvements for storage of individual genotypes and analysis of existing data. *Hum. Immunol.* **2016**, *77*, 238–248. [[CrossRef](#)] [[PubMed](#)]
57. Creary, L.E.; Gangavarapu, S.; Mallempati, K.C.; Montero-Martín, G.; Caillier, S.J.; Santaniello, A.; Hollenbach, J.A.; Oksenberg, J.R.; Fernández-Viña, M.A. Next-generation sequencing reveals new information about HLA allele and haplotype diversity in a large European American population. *Hum. Immunol.* **2019**, *80*, 807–822. [[CrossRef](#)]
58. Osoegawa, K.; Mallempati, K.C.; Gangavarapu, S.; Oki, A.; Gendzekhadze, K.; Marino, S.R.; Brown, N.K.; Bettinotti, M.P.; Weimer, E.; Montero-Martín, G.; et al. HLA alleles and haplotypes observed in 263 US families. *Hum. Immunol.* **2019**, *80*, 644–660. [[CrossRef](#)] [[PubMed](#)]
59. Li, H.; Ivarsson, M.A.; Walker-Sperling, V.E.; Subleski, J.; Johnson, J.; Wright, P.W.; Carrington, M.; Björkström, N.K.; McVicar, D.W.; Anderson, S.K. Identification of an elaborate NK-specific system regulating HLA-C expression. *PLoS Genet.* **2018**, *14*, e1007163. [[CrossRef](#)]
60. Mayor, N.P.; Robinson, J.; McWhinnie, A.J.M.; Ranade, S.; Eng, K.; Midwinter, W.; Bultitude, W.; Chin, C.-S.; Bowman, B.; Marks, P.; et al. HLA Typing for the Next Generation. *PLoS ONE* **2015**, *10*, e0127153. [[CrossRef](#)]
61. Schöfl, G.; Lang, K.; Quenzel, P.; Böhme, I.; Sauter, J.; Hofmann, J.A.; Pingel, J.; Schmidt, A.H.; Lange, V. 2.7 million samples genotyped for HLA by next generation sequencing: Lessons learned. *BMC Genom.* **2017**, *18*, 161. [[CrossRef](#)]
62. Carapito, R.; Radosavljevic, M.; Bahram, S. Next-Generation Sequencing of the HLA locus: Methods and impacts on HLA typing, population genetics and disease association studies. *Hum. Immunol.* **2016**, *77*, 1016–1023. [[CrossRef](#)]
63. Sanchez-Mazas, A.; Nunes, J.M.; Middleton, D.; Sauter, J.; Buhler, S.; McCabe, A.; Hofmann, J.; Baier, D.M.; Schmidt, A.H.; Nicoloso, G.; et al. Common and well-documented HLA alleles over all of Europe and within European sub-regions: A catalogue from the European Federation for Immunogenetics. *HLA* **2017**, *89*, 104–113. [[CrossRef](#)]
64. Carapito, R.; Bahram, S. Genetics, genomics, and evolutionary biology of NKG2D ligands. *Immunol. Rev.* **2015**, *267*, 88–116. [[CrossRef](#)] [[PubMed](#)]
65. Braud, V.M.; Allan, D.S.; McMichael, A.J. Functions of nonclassical MHC and non-MHC-encoded class I molecules. *Curr. Opin. Immunol.* **1999**, *11*, 100–108. [[CrossRef](#)]
66. Manser, A.R.; Weinhold, S.; Uhrberg, M. Human KIR repertoires: Shaped by genetic diversity and evolution. *Immunol. Rev.* **2015**, *267*, 178–196. [[CrossRef](#)]
67. Uhrberg, M. Shaping the human NK cell repertoire: An epigenetic glance at KIR gene regulation. *Mol. Immunol.* **2005**, *42*, 471–475. [[CrossRef](#)] [[PubMed](#)]
68. David, G.; Morvan, M.; Gagne, K.; Kerdudou, N.; Willem, C.; Devys, A.; Bonneville, M.; Folléa, G.; Bignon, J.-D.; Retière, C. Discrimination between the main activating and inhibitory killer cell immunoglobulin-like receptor positive natural killer cell subsets using newly characterized monoclonal antibodies. *Immunology* **2009**, *128*, 172–184. [[CrossRef](#)]
69. Schönberg, K.; Sribar, M.; Enczmann, J.; Fischer, J.C.; Uhrberg, M. Analyses of HLA-C-specific KIR repertoires in donors with group A and B haplotypes suggest a ligand-instructed model of NK cell receptor acquisition. *Blood* **2011**, *117*, 98–107. [[CrossRef](#)] [[PubMed](#)]

70. Gardiner, C.M.; Guethlein, L.A.; Shilling, H.G.; Pando, M.; Carr, W.H.; Rajalingam, R.; Vilches, C.; Parham, P. Different NK Cell Surface Phenotypes Defined by the DX9 Antibody Are Due to KIR3DL1 Gene Polymorphism. *J. Immunol.* **2001**, *166*, 2992–3001. [\[CrossRef\]](#) [\[PubMed\]](#)
71. Parham, P.; Norman, P.J.; Abi-Rached, L.; Guethlein, L.A. Variable NK Cell Receptors Exemplified by Human KIR3DL1/S1. *J. Immunol.* **2011**, *187*, 11–19. [\[CrossRef\]](#)
72. Li, H.; Pascal, V.; Martin, M.P.; Carrington, M.; Anderson, S.K. Genetic Control of Variegated KIR Gene Expression: Polymorphisms of the Bi-Directional KIR3DL1 Promoter Are Associated with Distinct Frequencies of Gene Expression. *PLoS Genet.* **2008**, *4*, e1000254. [\[CrossRef\]](#) [\[PubMed\]](#)
73. Li, H.; Wright, P.W.; McCullen, M.; Anderson, S.K. Characterization of KIR intermediate promoters reveals four promoter types associated with distinct expression patterns of KIR subtypes. *Genes Immun.* **2015**, *17*, 66–74. [\[CrossRef\]](#) [\[PubMed\]](#)
74. Bari, R.; Bell, T.; Leung, W.-H.; Vong, Q.P.; Chan, W.K.; Das Gupta, N.; Holladay, M.; Rooney, B.; Leung, W. Significant functional heterogeneity among KIR2DL1 alleles and a pivotal role of arginine245. *Blood* **2009**, *114*, 5182–5190. [\[CrossRef\]](#) [\[PubMed\]](#)
75. Le Luduec, J.-B.; Boudreau, J.E.; Freiberg, J.C.; Hsu, K.C. Novel Approach to Cell Surface Discrimination Between KIR2DL1 Subtypes and KIR2DS1 Identifies Hierarchies in NK Repertoire, Education, and Tolerance. *Front. Immunol.* **2019**, *10*, 734. [\[CrossRef\]](#) [\[PubMed\]](#)
76. Fan, Q.R.; Long, E.O.; Wiley, D.C. Crystal structure of the human natural killer cell inhibitory receptor KIR2DL1–HLA-Cw4 complex. *Nat. Immunol.* **2001**, *2*, 452–460. [\[CrossRef\]](#) [\[PubMed\]](#)
77. Rajagopalan, S.; Long, E.O. The Direct Binding of a p58 Killer Cell Inhibitory Receptor to Human Histocompatibility Leukocyte Antigen (HLA)-Cw4 Exhibits Peptide Selectivity. *J. Exp. Med.* **1997**, *185*, 1523–1528. [\[CrossRef\]](#)
78. Saunders, P.M.; Vivian, J.; O'Connor, G.; Sullivan, L.; Pymm, P.G.; Rossjohn, J.; Brooks, A.G. A bird's eye view of NK cell receptor interactions with their MHC class I ligands. *Immunol. Rev.* **2015**, *267*, 148–166. [\[CrossRef\]](#)
79. Boyington, J.C.; Motyka, S.A.; Schuck, P.; Brooks, A.; Sun, P.D. Crystal structure of an NK cell immunoglobulin-like receptor in complex with its class I MHC ligand. *Nature* **2000**, *405*, 537–543. [\[CrossRef\]](#)
80. Moesta, A.; Norman, P.; Yawata, M.; Yawata, N.; Gleimer, M.; Parham, P. Synergistic Polymorphism at Two Positions Distal to the Ligand-Binding Site Makes KIR2DL2 a Stronger Receptor for HLA-C Than KIR2DL3. *J. Immunol.* **2008**, *180*, 3969–3979. [\[CrossRef\]](#)
81. David, G.; Djaoud, Z.; Willem, C.; Legrand, N.; Rettman, P.; Gagne, K.; Cesbron, A.; Retière, C. Large Spectrum of HLA-C Recognition by Killer Ig-like Receptor (KIR)2DL2 and KIR2DL3 and Restricted C1 Specificity of KIR2DS2: Dominant Impact of KIR2DL2/KIR2DS2 on KIR2D NK Cell Repertoire Formation. *J. Immunol.* **2013**, *191*, 4778–4788. [\[CrossRef\]](#)
82. Frazier, W.R.; Steiner, N.; Hou, L.; Dakshanamurthy, S.; Hurley, C.K. Allelic Variation in KIR2DL3 Generates a KIR2DL2-like Receptor with Increased Binding to its HLA-C Ligand. *J. Immunol.* **2013**, *190*, 6198–6208. [\[CrossRef\]](#) [\[PubMed\]](#)
83. Lunemann, S.; Martrus, G.; Hölzemer, A.; Chapel, A.; Ziegler, M.; Körner, C.; Beltran, W.G.; Carrington, M.; Wedemeyer, H.; Altfeld, M. Sequence variations in HCV core-derived epitopes alter binding of KIR2DL3 to HLA-C*03:04 and modulate NK cell function. *J. Hepatol.* **2016**, *65*, 252–258. [\[CrossRef\]](#) [\[PubMed\]](#)
84. Eragopalan, S.; Long, E.O. KIR2DL4 (CD158d): An activation receptor for HLA-G. *Front. Immunol.* **2012**, *3*, 258. [\[CrossRef\]](#)
85. Morvan, M.; David, G.; Sébille, V.; Perrin, A.; Gagne, K.; Willem, C.; Kerdudou, N.; Denis, L.; Clemenceau, B.; Folléa, G.; et al. Autologous and allogeneic HLA KIR ligand environments and activating KIR control KIR NK-cell functions. *Eur. J. Immunol.* **2008**, *38*, 3474–3486. [\[CrossRef\]](#)
86. Moretta, A.; Sivori, S.; Vitale, M.; Pende, D.; Morelli, L.; Augugliaro, R.; Bottino, C.; Moretta, L. Existence of both inhibitory (p58) and activatory (p50) receptors for HLA-C molecules in human natural killer cells. *J. Exp. Med.* **1995**, *182*, 875–884. [\[CrossRef\]](#) [\[PubMed\]](#)
87. Liu, J.; Xiao, Z.; Ko, H.L.; Shen, M.; Ren, E.C. Activating killer cell immunoglobulin-like receptor 2DS2 binds to HLA-A*11. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 2662–2667. [\[CrossRef\]](#)
88. Naiyer, M.M.; Cassidy, S.A.; Magri, A.; Cowton, V.; Chen, K.; Mansour, S.; Kranidioti, H.; Mbirbindi, B.; Rettman, P.; Harris, S.; et al. KIR2DS2 recognizes conserved peptides derived from viral helicases in the context of HLA-C. *Sci. Immunol.* **2017**, *2*, eaal5296. [\[CrossRef\]](#)
89. Rettman, P.; Blunt, M.D.; Fulton, R.J.; Vallejo, A.F.; Bastidas-Legarda, L.Y.; España-Serrano, L.; Polak, M.E.; Al-Shamkhani, A.; Retiere, C.; Khakoo, S.I. Peptide: MHC-based DNA vaccination strategy to activate natural killer cells by targeting killer cell immunoglobulin-like receptors. *J. Immunother. Cancer* **2021**, *9*, e001912. [\[CrossRef\]](#)
90. Graef, T.; Moesta, A.; Norman, P.; Abi-Rached, L.; Vago, L.; Aguilar, A.M.O.; Gleimer, M.; Hammond, J.; Guethlein, L.A.; Bushnell, D.A.; et al. KIR2DS4 is a product of gene conversion with KIR3DL2 that introduced specificity for HLA-A*11 while diminishing avidity for HLA-C. *J. Exp. Med.* **2009**, *206*, 2557–2572. [\[CrossRef\]](#)
91. Vitale, M.; Sivori, S.; Pende, D.; Augugliaro, R.; Di Donato, C.; Amoroso, A.; Malnati, M.; Bottino, C.; Moretta, L. Physical and functional independency of p70 and p58 natural killer (NK) cell receptors for HLA class I: Their role in the definition of different groups of alloreactive NK cell clones. *Proc. Natl. Acad. Sci. USA* **1996**, *93*, 1453–1457. [\[CrossRef\]](#)
92. Stern, M.; Ruggeri, L.; Capanni, M.; Mancusi, A.; Velardi, A. Human leukocyte antigens A23, A24, and A32 but not A25 are ligands for KIR3DL1. *Blood* **2008**, *112*, 708–710. [\[CrossRef\]](#) [\[PubMed\]](#)
93. Burian, A.; Wang, K.L.; Finton, K.A.K.; Lee, N.; Ishitani, A.; Strong, R.K.; Geraghty, D.E. HLA-F and MHC-I Open Conformers Bind Natural Killer Cell Ig-Like Receptor KIR3DS1. *PLoS ONE* **2016**, *11*, e0163297. [\[CrossRef\]](#) [\[PubMed\]](#)

94. Martin, M.P.; Gao, X.; Lee, J.-H.; Nelson, G.W.; Detels, R.; Goedert, J.J.; Buchbinder, S.; Hoots, K.; Vlahov, D.; Trowsdale, J.; et al. Epistatic interaction between KIR3DS1 and HLA-B delays the progression to AIDS. *Nat. Genet.* **2002**, *31*, 429–434. [[CrossRef](#)] [[PubMed](#)]
95. Döhning, C.; Scheidegger, D.; Samaridis, J.; Cella, M.; Colonna, M. A human killer inhibitory receptor specific for HLA-A1,2. *J. Immunol.* **1996**, *156*, 3098–3101. [[PubMed](#)]
96. Hansasuta, P.; Dong, T.; Thananchai, H.; Weekes, M.; Willberg, C.; Aldemir, H.; Rowland-Jones, S.; Braud, V.M. Recognition of HLA-A3 and HLA-A11 by KIR3DL2 is peptide-specific. *Eur. J. Immunol.* **2004**, *34*, 1673–1679. [[CrossRef](#)] [[PubMed](#)]
97. Bhatt, R.S.; Berjis, A.; Konge, J.C.; Mahoney, K.M.; Klee, A.N.; Freeman, S.S.; Chen, C.-H.; Jegede, O.A.; Catalano, P.J.; Pignon, J.-C.; et al. KIR3DL3 Is an Inhibitory Receptor for HHLA2 that Mediates an Alternative Immunoinhibitory Pathway to PD1. *Cancer Immunol. Res.* **2021**, *9*, 156–169. [[CrossRef](#)] [[PubMed](#)]
98. Höglund, P.; Brodin, P. Current perspectives of natural killer cell education by MHC class I molecules. *Nat. Rev. Immunol.* **2010**, *10*, 724–734. [[CrossRef](#)] [[PubMed](#)]
99. Yokoyama, W.M.; Kim, S. Licensing of natural killer cells by self-major histocompatibility complex class I. *Immunol. Rev.* **2006**, *214*, 143–154. [[CrossRef](#)]
100. Boudreau, J.; Mulrooney, T.J.; LE Ludec, J.-B.; Barker, E.; Hsu, K.C. KIR3DL1 and HLA-B Density and Binding Calibrate NK Education and Response to HIV. *J. Immunol.* **2016**, *196*, 3398–3410. [[CrossRef](#)]
101. Björklund, A.T.; Schaffer, M.; Fauriat, C.; Ringdén, O.; Remberger, M.; Hammarstedt, C.; Barrett, A.J.; Ljungman, P.; Ljunggren, H.-G.; Malmberg, K.-J. NK cells expressing inhibitory KIR for non-self-ligands remain tolerant in HLA-matched sibling stem cell transplantation. *Blood* **2010**, *115*, 2686–2694. [[CrossRef](#)]
102. Winter, C.C.; Gumperz, J.E.; Parham, P.; Long, E.O.; Wagtmann, N. Direct binding and functional transfer of NK cell inhibitory receptors reveal novel patterns of HLA-C allotype recognition. *J. Immunol.* **1998**, *161*, 571–577. [[PubMed](#)]
103. Carr, W.H.; Pando, M.J.; Parham, P. KIR3DL1 Polymorphisms That Affect NK Cell Inhibition by HLA-Bw4 Ligand. *J. Immunol.* **2005**, *175*, 5222–5229. [[CrossRef](#)]
104. O'Connor, G.M.; Guinan, K.; Cunningham, R.T.; Middleton, D.; Parham, P.; Gardiner, C.M. Functional polymorphism of the KIR3DL1/S1 receptor on human NK cells. *J. Immunol.* **2006**, *178*, 235–241. [[CrossRef](#)]
105. Anderson, S.K. Molecular evolution of elements controlling HLA-C expression: Adaptation to a role as a killer-cell immunoglobulin-like receptor ligand regulating natural killer cell function. *HLA* **2018**, *92*, 271–278. [[CrossRef](#)] [[PubMed](#)]
106. Hilton, H.G.; Parham, P. Missing or altered self: Human NK cell receptors that recognize HLA-C. *Immunogenetics* **2017**, *69*, 567–579. [[CrossRef](#)]
107. Apps, R.; Qi, Y.; Carlson, J.M.; Chen, H.; Gao, X.; Thomas, R.; Yuki, Y.; Del Prete, G.Q.; Goulder, P.; Brumme, Z.L.; et al. Influence of HLA-C Expression Level on HIV Control. *Science* **2013**, *340*, 87–91. [[CrossRef](#)] [[PubMed](#)]
108. Eisaie, M.; Roelen, D.L.; Ecarrington, M.; Petersdorf, E.W.; Claas, F.H.J.; Haasnoot, G.W.; Eoudshoorn, M. Association between CTL Precursor Frequency to HLA-C Mismatches and HLA-C Antigen Cell Surface Expression. *Front. Immunol.* **2014**, *5*, 547. [[CrossRef](#)]
109. Vince, N.; Li, H.; Ramsuran, V.; Naranbhai, V.; Duh, F.-M.; Fairfax, B.P.; Saleh, B.; Knight, J.; Anderson, S.; Carrington, M. HLA-C Level Is Regulated by a Polymorphic Oct1 Binding Site in the HLA-C Promoter Region. *Am. J. Hum. Genet.* **2016**, *99*, 1353–1358. [[CrossRef](#)]
110. Kulkarni, S.; Qi, Y.; O'Huigin, C.; Pereyra, F.; Ramsuran, V.; McLaren, P.; Fellay, J.; Nelson, G.; Chen, H.; Liao, W.; et al. Genetic interplay between HLA-C and MIR148A in HIV control and Crohn disease. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 20705–20710. [[CrossRef](#)]
111. Kaur, G.; Gras, S.; Mobbs, J.; Vivian, J.; Cortes, A.; Barber, T.; Kuttikkatte, S.B.; Jensen, L.T.; Attfield, K.; Dendrou, C.; et al. Structural and regulatory diversity shape HLA-C protein expression levels. *Nat. Commun.* **2017**, *8*, 15924. [[CrossRef](#)]
112. René, C.; Lozano, C.; Eliaou, J.-F. Expression of classical HLA class I molecules: Regulation and clinical impacts: Julia Bodmer Award Review 2015. *HLA* **2016**, *87*, 338–349. [[CrossRef](#)]
113. Toffalori, C.; Zito, L.; Gambacorta, V.; Riba, M.; Oliveira, G.; Bucci, G.; Barcella, M.; Spinelli, O.; Greco, R.; Crucitti, L.; et al. Immune signature drives leukemia escape and relapse after hematopoietic cell transplantation. *Nat. Med.* **2019**, *25*, 603–611. [[CrossRef](#)]
114. Verheyden, S.; Demanet, C. NK cell receptors and their ligands in leukemia. *Leukemia* **2007**, *22*, 249–257. [[CrossRef](#)]
115. Verheyden, S.; Ferrone, S.; Mulder, A.; Claas, F.H.; Schots, R.; De Moerloose, B.; Benoit, Y.; Demanet, C. Role of the inhibitory KIR ligand HLA-Bw4 and HLA-C expression levels in the recognition of leukemic cells by Natural Killer cells. *Cancer Immunol. Immunother.* **2008**, *58*, 855–865. [[CrossRef](#)] [[PubMed](#)]
116. Varbanova, V.; Naumova, E.; Mihaylova, A. Killer-cell immunoglobulin-like receptor genes and ligands and their role in hematologic malignancies. *Cancer Immunol. Immunother.* **2016**, *65*, 427–440. [[CrossRef](#)]
117. Masuda, K.; Hiraki, A.; Fujii, N.; Watanabe, T.; Tanaka, M.; Matsue, K.; Ogama, Y.; Ouchida, M.; Shimizu, K.; Ikeda, K.; et al. Loss or down-regulation of HLA class I expression at the allelic level in freshly isolated leukemic blasts. *Cancer Sci.* **2007**, *98*, 102–108. [[CrossRef](#)] [[PubMed](#)]
118. Ahci, M.; Toffalori, C.; Bouwmans, E.; Crivello, P.; Brambati, C.; Pultrone, C.; Stempelmann, K.; Bost, D.; Mazzi, B.; Beelen, D.W.; et al. A new tool for rapid and reliable diagnosis of HLA loss relapses after HSCT. *Blood* **2017**, *130*, 1270–1273. [[CrossRef](#)]

119. Vago, L.; Perna, S.K.; Zanussi, M.; Mazzi, B.; Barlassina, C.; Lupo-Stanghellini, M.T.; Perrelli, N.F.; Cosentino, C.; Torri, F.; Angius, A.; et al. Loss of Mismatched HLA in Leukemia after Stem-Cell Transplantation. *N. Engl. J. Med.* **2009**, *361*, 478–488. [\[CrossRef\]](#)
120. Dubois, V.; Sloan-Béna, F.; Cesbron, A.; Hepkema, B.G.; Gagne, K.; Gimelli, S.; Heim, D.; Tichelli, A.; Delaunay, J.; Drouet, M.; et al. Pretransplant HLA mistyping in diagnostic samples of acute myeloid leukemia patients due to acquired uniparental disomy. *Leukemia* **2012**, *26*, 2079–2085. [\[CrossRef\]](#) [\[PubMed\]](#)
121. Etiercy, J.-M. HLA-C Incompatibilities in Allogeneic Unrelated Hematopoietic Stem Cell Transplantation. *Front. Immunol.* **2014**, *5*, 216. [\[CrossRef\]](#)
122. Petersdorf, E.W.; Gooley, T.A.; Malkki, M.; Bacigalupo, A.P.; Cesbron, A.; Du Toit, E.; Ehninger, G.; Egeland, T.; Fischer, G.F.; Gervais, T.; et al. HLA-C expression levels define permissible mismatches in hematopoietic cell transplantation. *Blood* **2014**, *124*, 3996–4003. [\[CrossRef\]](#)
123. Fernandez-Viña, M.A.; Wang, T.; Lee, S.J.; Haagenson, M.; Aljurf, M.; Askar, M.; Battiwalla, M.; Baxter-Lowe, L.-A.; Gajewski, J.; Jakubowski, A.A.; et al. Identification of a permissible HLA mismatch in hematopoietic stem cell transplantation. *Blood* **2014**, *123*, 1270–1278. [\[CrossRef\]](#)
124. Fernández-Viña, M.A. HLA allotype expressivity in transplantation. *Blood* **2014**, *124*, 3839–3840. [\[CrossRef\]](#)
125. Hoff, G.A.; Fischer, J.C.; Hsu, K.; Cooley, S.; Miller, J.S.; Wang, T.; Haagenson, M.; Spellman, S.; Lee, S.J.; Uhrberg, M.; et al. Recipient HLA-C Haplotypes and microRNA 148a/b Binding Sites Have No Impact on Allogeneic Hematopoietic Cell Transplantation Outcomes. *Biol. Blood Marrow Transplant.* **2017**, *23*, 153–160. [\[CrossRef\]](#)
126. Mastaglio, S.; Wong, E.; Perera, T.; Ripley, J.; Blombery, P.; Smyth, M.J.; Koldej, R.; Ritchie, D. Natural killer receptor ligand expression on acute myeloid leukemia impacts survival and relapse after chemotherapy. *Blood Adv.* **2018**, *2*, 335–346. [\[CrossRef\]](#) [\[PubMed\]](#)
127. Makanga, D.R.; Da Rin de Lorenzo, F.; David, G.; Willem, C.; Dubreuil, L.; Legrand, N.; Guillaume, T.; Peterlin, P.; LeBourgeois, A.; Béné, M.C.; et al. Genetic and Molecular Basis of Heterogeneous NK Cell Responses against Acute Leukemia. *Cancers* **2020**, *12*, 1927. [\[CrossRef\]](#)
128. Lee, S.J.; Klein, J.; Haagenson, M.; Baxter-Lowe, L.A.; Confer, D.L.; Eapen, M.; Fernandez-Vina, M.; Flomenberg, N.; Horowitz, M.; Hurley, C.K.; et al. High-resolution donor-recipient HLA matching contributes to the success of unrelated donor marrow transplantation. *Blood* **2007**, *110*, 4576–4583. [\[CrossRef\]](#) [\[PubMed\]](#)
129. Petersdorf, E.W. Optimal HLA matching in hematopoietic cell transplantation. *Curr. Opin. Immunol.* **2008**, *20*, 588–593. [\[CrossRef\]](#)
130. Nagler, A.; Ruggeri, A. Haploidentical stem cell transplantation (HaploSCT) for patients with acute leukemia—an update on behalf of the ALWP of the EBMT. *Bone Marrow Transpl.* **2019**, *54*, 713–718. [\[CrossRef\]](#) [\[PubMed\]](#)
131. Mohty, M. Haploidentical transplantation: Finally, some light. *Blood* **2021**, *137*, 296–297. [\[CrossRef\]](#)
132. Bignon, J.-D.; Gagne, K. KIR matching in hematopoietic stem cell transplantation. *Curr. Opin. Immunol.* **2005**, *17*, 553–559. [\[CrossRef\]](#)
133. Leung, W. Use of NK cell activity in cure by transplant. *Br. J. Haematol.* **2011**, *155*, 14–29. [\[CrossRef\]](#)
134. Symons, H.J.; Leffell, M.S.; Rossiter, N.D.; Zahurak, M.; Jones, R.J.; Fuchs, E.J. Improved Survival with Inhibitory Killer Immunoglobulin Receptor (KIR) Gene Mismatches and KIR Haplotype B Donors after Nonmyeloablative, HLA-Haploidentical Bone Marrow Transplantation. *Biol. Blood Marrow Transpl. J. Am. Soc. Blood Marrow Transpl.* **2010**, *16*, 533–542. [\[CrossRef\]](#)
135. Ruggeri, L.; Capanni, M.; Urbani, E.; Perruccio, K.; Shlomchik, W.D.; Tosti, A.; Posati, S.; Rogaia, D.; Frassoni, F.; Aversa, F.; et al. Effectiveness of Donor Natural Killer Cell Alloreactivity in Mismatched Hematopoietic Transplants. *Science* **2002**, *295*, 2097–2100. [\[CrossRef\]](#)
136. Locatelli, F.; Pende, D.; Falco, M.; Della Chiesa, M.; Moretta, A.; Moretta, L. NK Cells Mediate a Crucial Graft-versus-Leukemia Effect in Haploidentical-HSCT to Cure High-Risk Acute Leukemia. *Trends Immunol.* **2018**, *39*, 577–590. [\[CrossRef\]](#)
137. Shilling, H.G.; McQueen, K.L.; Cheng, N.W.; Shizuru, J.A.; Negrin, R.S.; Parham, P. Reconstitution of NK cell receptor repertoire following HLA-matched hematopoietic cell transplantation. *Blood* **2003**, *101*, 3730–3740. [\[CrossRef\]](#)
138. Hsu, K.C.; Keever-Taylor, C.; Wilton, A.; Pinto, C.; Heller, G.; Arkun, K.; O'Reilly, R.J.; Horowitz, M.M.; Dupont, B. Improved outcome in HLA-identical sibling hematopoietic stem-cell transplantation for acute myelogenous leukemia predicted by KIR and HLA genotypes. *Blood* **2005**, *105*, 4878–4884. [\[CrossRef\]](#)
139. Hsu, K.C.; Gooley, T.; Malkki, M.; Pinto-Agnello, C.; Dupont, B.; Bignon, J.-D.; Bornhäuser, M.; Christiansen, F.; Gratwohl, A.; Morishima, Y.; et al. KIR Ligands and Prediction of Relapse after Unrelated Donor Hematopoietic Cell Transplantation for Hematologic Malignancy. *Biol. Blood Marrow Transpl.* **2006**, *12*, 828–836. [\[CrossRef\]](#) [\[PubMed\]](#)
140. Venstrom, J.M.; Pittari, G.; Gooley, T.A.; Cheurning, J.H.; Spellman, S.; Haagenson, M.; Gallagher, M.M.; Malkki, M.; Petersdorf, E.; Dupont, B.; et al. HLA-C-Dependent Prevention of Leukemia Relapse by Donor Activating KIR2DS1. *N. Engl. J. Med.* **2012**, *367*, 805–816. [\[CrossRef\]](#) [\[PubMed\]](#)
141. Gagne, K.; Busson, M.; Bignon, J.-D.; Balère-Appert, M.-L.; Loiseau, P.; Dormoy, A.; Dubois, V.; Perrier, P.; Jollet, I.; Bois, M.; et al. Donor KIR3DL1/3DS1 Gene and Recipient Bw4 KIR Ligand as Prognostic Markers for Outcome in Unrelated Hematopoietic Stem Cell Transplantation. *Biol. Blood Marrow Transpl.* **2009**, *15*, 1366–1375. [\[CrossRef\]](#) [\[PubMed\]](#)
142. Venstrom, J.M.; Gooley, T.A.; Spellman, S.; Pring, J.; Malkki, M.; Dupont, B.; Petersdorf, E.; Hsu, K.C. Donor activating KIR3DS1 is associated with decreased acute GVHD in unrelated allogeneic hematopoietic stem cell transplantation. *Blood* **2010**, *115*, 3162–3165. [\[CrossRef\]](#) [\[PubMed\]](#)

143. Cooley, S.; Weisdorf, D.J.; Guethlein, L.A.; Klein, J.P.; Wang, T.; Le, C.T.; Marsh, S.G.E.; Geraghty, D.; Spellman, S.; Haagenson, M.D.; et al. Donor selection for natural killer cell receptor genes leads to superior survival after unrelated transplantation for acute myelogenous leukemia. *Blood* **2010**, *116*, 2411–2419. [\[CrossRef\]](#) [\[PubMed\]](#)
144. Weisdorf, D.; Cooley, S.; Wang, T.; Trachtenberg, E.; Vierra-Green, C.; Spellman, S.; Sees, J.A.; Spahn, A.; Vogel, J.; Fehniger, T.A.; et al. KIR B donors improve the outcome for AML patients given reduced intensity conditioning and unrelated donor transplantation. *Blood Adv.* **2020**, *4*, 740–754. [\[CrossRef\]](#) [\[PubMed\]](#)
145. Guethlein, L.A.; Beyzaie, N.; Nemat-Gorgani, N.; Wang, T.; Ramesh, V.; Marin, W.M.; Hollenbach, J.A.; Schetelig, J.; Spellman, S.R.; Marsh, S.G.E.; et al. Following Transplantation for Acute Myelogenous Leukemia, Donor KIR Cen B02 Better Protects against Relapse than KIR Cen B01. *J. Immunol.* **2021**, *206*, ji2100119. [\[CrossRef\]](#)
146. Weisdorf, D.; Cooley, S.; Wang, T.; Trachtenberg, E.; Haagenson, M.D.; Vierra-Green, C.; Spellman, S.; Spahn, A.; Vogel, J.; Kobusingye, H.; et al. KIR Donor Selection: Feasibility in Identifying better Donors. *Biol. Blood Marrow Transpl.* **2019**, *25*, e28–e32. [\[CrossRef\]](#)
147. Bari, R.; Rujkijyanont, P.; Sullivan, E.; Kang, G.; Turner, V.; Gan, K.; Leung, W. Effect of Donor KIR2DL1 Allelic Polymorphism on the Outcome of Pediatric Allogeneic Hematopoietic Stem-Cell Transplantation. *J. Clin. Oncol.* **2013**, *31*, 3782–3790. [\[CrossRef\]](#) [\[PubMed\]](#)
148. Boudreau, J.E.; Giglio, F.; Gooley, T.A.; Stevenson, P.A.; LE Ludeuc, J.-B.; Shaffer, B.C.; Rajalingam, R.; Hou, L.; Hurley, C.K.; Noreen, H.; et al. KIR3DL1/HLA-B Subtypes Govern Acute Myelogenous Leukemia Relapse After Hematopoietic Cell Transplantation. *J. Clin. Oncol.* **2017**, *35*, 2268–2278. [\[CrossRef\]](#)
149. Van Der Ploeg, K.; Le Ludeuc, J.-B.; Stevenson, P.A.; Park, S.; Gooley, T.A.; Petersdorf, E.W.; Shaffer, B.C.; Hsu, K.C. HLA-A alleles influencing NK cell function impact AML relapse following allogeneic hematopoietic cell transplantation. *Blood Adv.* **2020**, *4*, 4955–4964. [\[CrossRef\]](#)
150. Schetelig, J.; Baldauf, H.; Heidenreich, F.; Massalski, C.; Frank, S.; Sauter, J.; Stelljes, M.; Ayuk, F.A.; Bethge, W.A.; Bug, G.; et al. External validation of models for KIR2DS1/KIR3DL1-informed selection of hematopoietic cell donors fails. *Blood* **2020**, *135*, 1386–1395. [\[CrossRef\]](#) [\[PubMed\]](#)
151. Schetelig, J.; Baldauf, H.; Koster, L.; Kuxhausen, M.; Heidenreich, F.; de Wreede, L.C.; Spellman, S.; van Gelder, M.; Bruno, B.; Onida, F.; et al. Haplotype Motif-Based Models for KIR-Genotype Informed Selection of Hematopoietic Cell Donors Fail to Predict Outcome of Patients With Myelodysplastic Syndromes or Secondary Acute Myeloid Leukemia. *Front. Immunol.* **2021**, *11*, 584520. [\[CrossRef\]](#)
152. Willemze, R.; Ebmt, P.; Rodrigues, C.A.; Labopin, M.; Sanz, G.; Michel, G.; Socié, G.; Rio, B.; Sirvent, A.; Renaud, M.; et al. KIR-ligand incompatibility in the graft-versus-host direction improves outcomes after umbilical cord blood transplantation for acute leukemia. *Leukemia* **2009**, *23*, 492–500. [\[CrossRef\]](#)
153. Brunstein, C.G.; Wagner, J.E.; Weisdorf, D.J.; Cooley, S.; Noreen, H.; Barker, J.N.; DeFor, T.; Verneris, M.R.; Blazar, B.R.; Miller, J.S. Negative effect of KIR alloreactivity in recipients of umbilical cord blood transplant depends on transplantation conditioning intensity. *Blood* **2009**, *113*, 5628–5634. [\[CrossRef\]](#)
154. Garfall, A.; Kim, H.T.; Sun, L.; Ho, V.T.; Armand, P.; Koreth, J.; Alyea, E.P.; Soiffer, R.J.; Antin, J.H.; Ritz, J.; et al. KIR ligand incompatibility is not associated with relapse reduction after double umbilical cord blood transplantation. *Bone Marrow Transpl.* **2013**, *48*, 1000–1002. [\[CrossRef\]](#)
155. Guillaume, N.; Loiseau, P.; Gagne, K.; Teissierenc, H.M.; Cayuela, J.-M.; Henry, G.; Robin, M.; De Latour, R.P.; Gluckman, E.; Socié, G.; et al. Natural killer cell licensing after double cord blood transplantation is driven by the self-HLA class I molecules from the dominant cord blood. *Haematologica* **2016**, *101*, e209–e212. [\[CrossRef\]](#)
156. Rettman, P.; Legrand, N.; Willem, C.; Lode, L.; Chevallier, P.; Cesbron, A.; Senitzer, D.; Retière, C.; Gagne, K. Use of killer cell immunoglobulin-like receptor genes as early markers of hematopoietic chimerism after double-umbilical cord blood transplantation. *Haematologica* **2015**, *100*, e475–e479. [\[CrossRef\]](#)
157. Rettman, P.; Willem, C.; Volteau, C.; Legrand, N.; Chevallier, P.; Lodé, L.; Esbelin, J.; Cesbron, A.; Bonneville, M.; Moreau, P.; et al. Impact of Graft-Versus-Graft Natural Killer Cell Alloreactivity on Single Unit Dominance After Double Umbilical Cord Blood Transplantation. *Transplantation* **2017**, *101*, 2092–2101. [\[CrossRef\]](#) [\[PubMed\]](#)
158. Rettman, P.; Malard, F.; Legrand, N.; Avinens, O.; Eliaou, J.-F.; Picard, C.; Dormoy, A.; Lafarge, X.; De Matteis, M.; Kennel, A.; et al. Impact of KIR/HLA genetic combinations on double umbilical cord blood transplantation outcomes. Results of a French multicentric retrospective study on behalf of the Société Francophone de Greffe de Moelle et de Thérapie Cellulaire (SFGM-TC) and the Société Francophone d’Histocompatibilité et d’Immunogénétique (SFHI). *Bone Marrow Transpl.* **2016**, *51*, 1499–1503. [\[CrossRef\]](#)
159. Baumeister, S.H.C.; Rambaldi, B.; Shapiro, R.M.; Romee, R. Key Aspects of the Immunobiology of Haploidentical Hematopoietic Cell Transplantation. *Front. Immunol.* **2020**, *11*, 191. [\[CrossRef\]](#)
160. Dubois, V.; Amokrane, K.; Beguin, Y.; Bruno, B.; Chevallier, P.; Delbos, F.; Devillier, R.; Giannoli, C.; Guidicelli, G.; Harif, M.; et al. Haploidentical hematopoietic stem cell transplant: How to choose the best donor? Guidelines from the Francophone Society of Bone Marrow Transplantation and Cellular Therapy (SFGM-TC). *Bull. Cancer* **2020**, *107*, S72–S84. [\[CrossRef\]](#) [\[PubMed\]](#)
161. Bastos-Oreiro, M.; Anguita, J.; Martínez-Laperche, C.; Fernández, L.; Buces, E.; Navarro-Bailón, A.; Pascual, C.; Pérez-Corral, A.M.; Balsalobre, P.; Muñoz, C.; et al. Inhibitory killer cell immunoglobulin-like receptor (iKIR) mismatches improve survival after T-cell-repleted haploidentical transplantation. *Eur. J. Haematol.* **2016**, *96*, 483–491. [\[CrossRef\]](#)

162. Wanquet, A.; Bramanti, S.; Harbi, S.; Fürst, S.; Legrand, F.; Faucher, C.; Granata, A.; Calmels, B.; Lemarie, C.; Picard, C.; et al. Killer Cell Immunoglobulin-Like Receptor–Ligand Mismatch in Donor versus Recipient Direction Provides Better Graft-versus-Tumor Effect in Patients with Hematologic Malignancies Undergoing Allogeneic T Cell–Replete Haploidentical Transplantation Followed by Post-Transplant Cyclophosphamide. *Biol. Blood Marrow Transplant. J. Am. Soc. Blood Marrow Transpl.* **2018**, *24*, 549–554. [\[CrossRef\]](#)
163. Solomon, S.R.; Aubrey, M.T.; Zhang, X.; Piluso, A.; Freed, B.M.; Brown, S.; Jackson, K.C.; Morris, L.E.; Holland, H.K.; Solh, M.M.; et al. Selecting the Best Donor for Haploidentical Transplant: Impact of HLA, Killer Cell Immunoglobulin-Like Receptor Genotyping, and Other Clinical Variables. *Biol. Blood Marrow Transplant. J. Am. Soc. Blood Marrow Transpl.* **2018**, *24*, 789–798. [\[CrossRef\]](#) [\[PubMed\]](#)
164. Willem, C.; Makanga, D.R.; Guillaume, T.; Maniangu, B.; Legrand, N.; Gagne, K.; Peterlin, P.; Garnier, A.; Béné, M.C.; Cesbron, A.; et al. Impact of KIR/HLA Incompatibilities on NK Cell Reconstitution and Clinical Outcome after T Cell–Replete Haploidentical Hematopoietic Stem Cell Transplantation with Posttransplant Cyclophosphamide. *J. Immunol.* **2019**, *202*, 2141–2152. [\[CrossRef\]](#)
165. Shimoni, A.; Labopin, M.; Finke, J.; Ciceri, F.; Deconinck, E.; Kröger, N.; Gramatzki, M.; Stelljes, M.; Blaise, D.; Stoezel, F.; et al. Donor selection for a second allogeneic stem cell transplantation in AML patients relapsing after a first transplant: A study of the Acute Leukemia Working Party of EBMT. *Blood Cancer J.* **2019**, *9*, 88. [\[CrossRef\]](#)
166. Retière, C.; Willem, C.; Guillaume, T.; Vié, H.; Gautreau-Rolland, L.; Scotet, E.; Saulquin, X.; Gagne, K.; Béné, M.C.; Imbert, B.-M.; et al. Impact on early outcomes and immune reconstitution of high-dose post-transplant cyclophosphamide vs anti-thymocyte globulin after reduced intensity conditioning peripheral blood stem cell allogeneic transplantation. *Oncotarget* **2018**, *9*, 11451–11464. [\[CrossRef\]](#) [\[PubMed\]](#)
167. Makanga, D.R.; Guillaume, T.; Willem, C.; Legrand, N.; Gagne, K.; Cesbron, A.; Gendzekhadze, K.; Peterlin, P.; Garnier, A.; Le Bourgeois, A.; et al. Posttransplant Cyclophosphamide and Antithymocyte Globulin versus Posttransplant Cyclophosphamide as Graft-versus-Host Disease Prophylaxis for Peripheral Blood Stem Cell Haploidentical Transplants: Comparison of T Cell and NK Effector Reconstitution. *J. Immunol.* **2020**, *205*, 1441–1448. [\[CrossRef\]](#) [\[PubMed\]](#)
168. Russo, A.; Oliveira, G.; Berglund, S.; Greco, R.; Gambacorta, V.; Cieri, N.; Toffalori, C.; Zito, L.; Lorentino, F.; Piemontese, S.; et al. NK cell recovery after haploidentical HSCT with posttransplant cyclophosphamide: Dynamics and clinical implications. *Blood* **2018**, *131*, 247–262. [\[CrossRef\]](#)
169. Davis, Z.B.; Vallera, D.A.; Miller, J.S.; Felices, M. Natural killer cells unleashed: Checkpoint receptor blockade and BiKE/TriKE utilization in NK-mediated anti-tumor immunotherapy. *Semin. Immunol.* **2017**, *31*, 64–75. [\[CrossRef\]](#) [\[PubMed\]](#)
170. Daher, M.; Rezvani, K. Next generation natural killer cells for cancer immunotherapy: The promise of genetic engineering. *Curr. Opin. Immunol.* **2018**, *51*, 146–153. [\[CrossRef\]](#)
171. Hodgins, J.J.; Khan, S.T.; Park, M.M.; Auer, R.C.; Ardolino, M. Killers 2.0: NK cell therapies at the forefront of cancer control. *J. Clin. Investig.* **2019**, *129*, 3499–3510. [\[CrossRef\]](#)
172. Rafei, H.; Daher, M.; Rezvani, K. Chimeric antigen receptor (CAR) natural killer (NK)-cell therapy: Leveraging the power of innate immunity. *Br. J. Haematol.* **2021**, *193*, 216–230. [\[CrossRef\]](#)
173. Lupo, K.B.; Matosevic, S. Natural Killer Cells as Allogeneic Effectors in Adoptive Cancer Immunotherapy. *Cancers* **2019**, *11*, 769. [\[CrossRef\]](#)
174. Gurney, M.; O'Dwyer, M. Realizing Innate Potential: CAR-NK Cell Therapies for Acute Myeloid Leukemia. *Cancers* **2021**, *13*, 1568. [\[CrossRef\]](#)
175. Romee, R.; Cooley, S.; Berrien-Elliott, M.M.; Westervelt, P.; Verneris, M.R.; Wagner, J.E.; Weisdorf, D.J.; Blazar, B.R.; Ustun, C.; DeFor, T.E.; et al. First-in-human phase 1 clinical study of the IL-15 superagonist complex ALT-803 to treat relapse after transplantation. *Blood* **2018**, *131*, 2515–2527. [\[CrossRef\]](#)
176. Ewen, E.-M.; Pahl, J.H.; Miller, M.; Watzl, C.; Cerwenka, A. KIR downregulation by IL-12/15/18 unleashes human NK cells from KIR/HLA-I inhibition and enhances killing of tumor cells. *Eur. J. Immunol.* **2018**, *48*, 355–365. [\[CrossRef\]](#) [\[PubMed\]](#)
177. Koerner, S.P.; André, M.C.; Leibold, J.S.; Kousis, P.C.; Kübler, A.; Pal, M.; Haen, S.; Bühring, H.-J.; Grosse-Hovest, L.; Jung, G.; et al. An Fc-optimized CD133 antibody for induction of NK cell reactivity against myeloid leukemia. *Leukemia* **2017**, *31*, 459–469. [\[CrossRef\]](#)
178. Smits, N.C.; Coupet, T.A.; Godbersen, C.; Sentman, C.L. Designing multivalent proteins based on natural killer cell receptors and their ligands as immunotherapy for cancer. *Expert Opin. Biol. Ther.* **2016**, *16*, 1105–1112. [\[CrossRef\]](#)
179. Wiernik, A.; Foley, B.; Zhang, B.; Verneris, M.R.; Warlick, E.D.; Gleason, M.K.; Ross, J.A.; Luo, X.; Weisdorf, D.J.; Walcheck, B.; et al. Targeting Natural Killer Cells to Acute Myeloid Leukemia In Vitro with a CD16 × 33 Bispecific Killer Cell Engager and ADAM17 Inhibition. *Clin. Cancer Res.* **2013**, *19*, 3844–3855. [\[CrossRef\]](#) [\[PubMed\]](#)
180. Vallera, D.A.; Felices, M.; McElmurry, R.; McCullar, V.; Zhou, X.; Schmohl, J.U.; Zhang, B.; Lenvik, A.J.L.; Panoskaltsis-Mortari, A.; Verneris, M.R.; et al. IL15 Trispecific Killer Engagers (TriKE) Make Natural Killer Cells Specific to CD33+ Targets While Also Inducing Persistence, In Vivo Expansion, and Enhanced Function. *Clin. Cancer Res.* **2016**, *22*, 3440–3450. [\[CrossRef\]](#) [\[PubMed\]](#)
181. André, P.; Denis, C.; Soulas, C.; Bourbon-Caillet, C.; Lopez, J.; Arnoux, T.; Bléry, M.; Bonnafous, C.; Gauthier, L.; Morel, A.; et al. Anti-NKG2A mAb Is a Checkpoint Inhibitor that Promotes Anti-tumor Immunity by Unleashing Both T and NK Cells. *Cell* **2018**, *175*, 1731–1743.e13. [\[CrossRef\]](#)

182. Makanga, D.R.; Jullien, M.; David, G.; Legrand, N.; Willem, C.; Dubreuil, L.; Walencik, A.; Touzeau, C.; Gastinne, T.; Tessoulin, B.; et al. Low Number of KIR Ligands in Lymphoma Patients Favors a Good Rituximab-Dependent NK Cell Response. *Oncoimmunology* **2021**, *10*, 1936392. [[CrossRef](#)] [[PubMed](#)]
183. Becker, P.S.A.; Suck, G.; Nowakowska, P.; Ullrich, E.; Seifried, E.; Bader, P.; Tonn, T.; Seidl, C. Selection and expansion of natural killer cells for NK cell-based immunotherapy. *Cancer Immunol. Immunother.* **2016**, *65*, 477–484. [[CrossRef](#)] [[PubMed](#)]
184. Guillerey, C.; Huntington, N.D.; Smyth, M. Targeting natural killer cells in cancer immunotherapy. *Nat. Immunol.* **2016**, *17*, 1025–1036. [[CrossRef](#)] [[PubMed](#)]
185. Shi, J.; Tricot, G.J.; Garg, T.K.; Malaviarachchi, P.A.; Szmania, S.M.; Kellum, R.E.; Storrie, B.; Mulder, A.; Shaughnessy, J.J.D.; Barlogie, B.; et al. Bortezomib down-regulates the cell-surface expression of HLA class I and enhances natural killer cell-mediated lysis of myeloma. *Blood* **2008**, *111*, 1309–1317. [[CrossRef](#)] [[PubMed](#)]