

# Intestine-Liver Cross-talk in Type 2 Diabetes and Non-Alcoholic Fatty Liver Disease

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4	Intestine-Liver Cross-talk in Type 2 Diabetes and Non-Alcoholic
5	Fatty Liver Disease
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#### 14 Abstract:

15 Type 2 diabetes (T2D) and Non-Alcoholic Fatty Liver Disease (NAFLD) are pathologies whose prevalence continues to increase worldwide. Both diseases are precipitated by an excessive caloric 16 17 intake, which promotes insulin resistance and fatty liver. The role of the intestine and its crosstalk with 18 the liver in the development of these metabolic diseases is receiving increasing attention. Alterations in diet-intestinal microbiota interactions lead to the dysregulation of intestinal functions, resulting in 19 altered metabolite and energy substrate production and increased intestinal permeability. Connected 20 21 through the portal circulation, these changes in intestinal functions impact the liver and other metabolic 22 organs, such as visceral adipose tissue, hence participating in the development of insulin resistance, and worsening T2D and NAFLD. Thus, targeting the intestine may be an efficient therapeutic approach to 23 24 cure T2D and NAFLD.

In this review, we will first introduce the signaling pathways linking T2D and NAFLD. Next, we will address the role of the gut-liver crosstalk in the development of T2D and NAFLD, with a particular focus on the gut microbiota and the molecular pathways behind the increased intestinal permeability and inflammation. Finally, we will summarize the therapeutic strategies which target the gut and its functions and are currently used or under development to treat T2D and NAFLD.

#### 30 Abbreviations:

- 31 ALT: alanine aminotransferase
- 32 BA: bile acids
- 33 CYP7A1: cholesterol 7 alpha-hydroxylase
- 34 DDP-4: dipeptidyl peptidase-4
- 35 FGF: fibroblast growth factor
- 36 FMT: fecal microbiota transplantation
- 37 FXR: farnesoid X receptor
- 38 GLP-1: glucagon-like peptide-1
- 39 GLP-1R: glucagon-like peptide-1 receptor
- 40 HFD: high fat diet
- 41 HOMA-IR: homeostatic model assessment of insulin resistance
- 42 IgA: immunoglobulin A
- 43 IR: insulin resistance
- 44 JAM-A: junctional adhesion molecule A
- 45 LPS: lipopolysaccharide
- 46 MRI: magnetic resonance imaging
- 47 NAFL /NAFLD: non-alcoholic fatty liver/disease
- 48 NAS: non-alcoholic fatty liver disease activity score
- 49 NASH: non-alcoholic steatohepatitis
- 50 OCA: obeticholic acid
- 51 SCFA: short chain fatty acid
- 52 T2D: type 2 diabetes
- 53 TG: triglycerides
- 54 TGR5: takeda g-protein-coupled receptor 5
- 55 1. <u>T2D and NAFLD, two closely linked multi-organ pathologies</u>
- 56 Diabetes is the most common metabolic disease worldwide. With the western lifestyle pandemic,
- 57 this multifactorial disease has a worldwide prevalence estimated at 500 million patients [1]. The majority

of diabetic patients suffer from Type 2 Diabetes (T2D), which can be classified in different etiological
subgroups [2]. T2D is closely related to other metabolic diseases linked to excessive caloric intake and
sedentary life style, such as obesity [1]. The liver plays a central role in lipid and glucose metabolism, and
hepatic insulin resistance (IR) is a hallmark of both T2D and NAFLD [3,4]. Since >70% of T2D patients have
NAFLD [5–7], it has been recently proposed to rename NAFLD to "Metabolic Associated Fatty Liver
Disease" to highlight this association [6].

64 NAFLD is a progressive disease initiated by an increase of hepatic lipid content, termed steatosis, 65 the "Non-Alcoholic Fatty Liver" (NAFL) stage. When steatosis becomes chronic, a more complicated 66 histological pattern appears characterized by an increased number of inflammatory *foci* associated with 67 hepatocyte balloonisation, the "Non-Alcoholic SteatoHepatitis" (NASH) stage. NAFL and NASH are 68 reversible but, without life-style or medical intervention, liver fibrosis progressively appears developing 69 over decades to cirrhosis and hepatocarcinoma [8,9].

70 The development of diagnostic tools to detect the presence of NASH and its progression to 71 fibrosis is crucial given its increasing prevalence. Today, the gold standard of NASH diagnosis is based on 72 scoring of the histological features of NASH on hepatic biopsies [9]. However, needle biopsies are not 73 without risk and reader variability exists, precluding its application to large numbers of patients. Thus, non-invasive techniques based on biomarkers are being developed to facilitate diagnosis and stage 74 75 identification [10]. In addition, Magnetic Resonance Imaging (MRI) is used to estimate the hepatic fat 76 content and liver ultrasound elastography to determine the degree of fibrosis [11]. As NAFLD is linked to other metabolic diseases, these new diagnostic tools can be more easily applied to patients suffering 77 78 from common NAFLD co-morbidities, such as T2D and cardiovascular disease [8].

It is still unclear whether T2D and NAFLD develop independently or not, but several studies 79 80 indicate that the hepatic IR observed in T2D promotes NAFLD progression. Both metabolic diseases imply a suspended dialogue between the different metabolic organs (Figure 1). In the initial phase of glucose 81 82 intolerance, the chronic hyperinsulinemia leads to decreased hepatic insulin receptor signaling due to 83 reduced receptor synthesis and signal transduction defects, hence resulting in hepatic IR [12]. Moreover, the dialogue between adipose tissue and liver is impaired in obesity and NAFLD, with decreased 84 adiponectin secretion and increased release of free fatty acids by adipose tissue, which worsen hepatic 85 86 steatosis and fibrosis [13,14]. As a consequence to the loss of insulin sensitivity, hepatic steatosis 87 increases, associated with a dysregulation of liver glycogenesis, gluconeogenesis and lipid synthesis [3]. The sustained increase of bioactive lipids, such as ceramides and diacylglycerols, increase IR by altering 88 89 hepatic insulin receptor signaling. Indeed, diacylglycerols activate protein kinase C isoforms which 90 phosphorylate insulin receptor substrates [15,16]. Moreover, ceramides produced in hepatocytes or 91 released by the intestine [17,18] enhance protein kinase Cζ and phosphatase A2 which inhibit Akt2 [19], hence aggravating hepatic IR. Ceramides and diacylgycerols drive hepatocyte endoplasmic reticulum 92 93 stress, induce liver inflammation, via pathways including the Nucleotide-binding oligomerization domain-Like Receptor Pyrin domain-containing-3 (NLRP3) inflammasome, and apoptosis [20-22]. These 94 95 mechanisms promote the hepatic recruitment of immune cells, including macrophages, dendritic cells 96 and CD8<sup>+</sup> T lymphocytes [23–25], maintaining inflammation and activating hepatic stellate cells resulting 97 in fibrosis [26,27]. Muscle IR is linked to a decrease of muscular glycogen synthesis and an increase of hepatic de novo lipogenesis after carbohydrate ingestion [28]. In NAFLD patients, low skeletal muscle 98 mass is associated with liver fibrosis development [29] and an increase of muscle fat content is 99 100 associated with NASH [30]. Furthermore, sarcopenia independently associates with NAFLD in T2D,

suggesting that it may contribute to hepatic impairment in these patients [31]. Moreover, the hepatic damages in diabetic patients with advanced NAFLD may further promote organ dysfunctions, such as chronic kidney disease, cardiovascular complications or neurodegenerative diseases [32–36]. Thus, the dysregulation of cross-talk between liver and other metabolic organs plays a crucial role in the development of these metabolic diseases

106 In this review, we will discuss the role of the disrupted gut-liver crosstalk in T2D and NAFLD, 107 focusing on changes in gut microbiota, gut permeability, the entero-hepatic cycle, the endocrine 108 intestine and the immune system. Finally, we will discuss the intestine and its functions as potential 109 therapeutic targets for the treatment of T2D and NAFLD.

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#### 111 2. The role of gut-liver cross-talk disruption in T2D and NAFLD

The intestine is a complex tissue with many functions: it is not only a physical barrier controlling the absorption of nutrients, but also a site of hormone production and an immunological checkpoint. The gut *lumen* also hosts the microbiota, which are essential in the maintenance of a proper energy whole body balance. Through its direct anatomical connection with the liver via the portal vein, the intestine also directly regulates hepatic metabolism. Disruption of the dialogue between the intestine and the liver is likely a major contributor to the development of metabolic diseases such as T2D and NAFLD (Figure 2).

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121 **2.1** Dysregulation of intestinal lipid metabolism promotes dyslipidemia and hepatic steatosis

The main function of the intestine is the absorption of nutrients. For instance, dietary lipids like triglycerides (TG) are hydrolyzed by pancreatic lipase. Released fatty acids are then transported in enterocytes by specific intestinal fatty acid carriers. In the enterocyte, re-assembled TG are packaged into triglyceride-rich lipoproteins called chylomicrons, which are excreted at the basolateral side of enterocytes. Furthermore, the gut also plays a buffering role by storing lipids after high fat meals in enterocytes, which are then secreted at later stages depending on the metabolic conditions [37].

T2D patients display a postprandial dyslipidemia due to a combination of elevated chylomicron secretion and decreased intra-vascular clearance. Obese and T2D individuals exhibiting systemic IR display also lower insulin-responses in the intestine. The resulting decreased Akt phosphorylation in enterocytes results in an increased chylomicron production [38,39]. Moreover, obesity and IR lead to intestinal inflammation and oxidative stress, as demonstrated by higher intestinal levels of tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and cyclooxygenase-2 (COX-2) as well as increased basal and lipopolysaccharide (LPS)-induced interleukin 6 (IL-6) secretion [38].

135 NAFLD patients display also often a dyslipoproteinemia characterized by increased 136 apolipoprotein (Apo) B-containing lipoproteins [40]. An elevated TG absorption, associated with high 137 plasma free fatty acid levels, contributes to an increase of *de novo* hepatic ceramide synthesis, hepatic 138 steatosis and IR in NAFLD patients [41,42].

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## 2.2 Disruption of the intestinal barrier increases low grade inflammation and liver injury

The intestine is highly selective for the absorption of dietary nutrients and metabolites, while acting as a barrier to protect the organism against harmful molecules and bacterial translocation. The luminal mucus layer, which is mainly produced by mucin-producing goblet cells, prevents bacterial adhesion. The intestinal epithelial cells, which transport the luminal metabolites, also react to noxious

stimuli by secreting antimicrobial peptides [43]. The epithelial cell monolayer is glued together by inter-144 145 cell junctions, notably the tight junctions, which are composed of several proteins, such as zonula occludens 1 (ZO-1), junctional adhesion molecule A (JAM-A) and occludin. These cell junctions establish 146 intestinal integrity and allow specific permeability to control the passage from the lumen to the systemic 147 148 or lymphatic circulation. In the lamina propria, the intestinal immune system (see next section for details) produces chemokines, cytokines and immunoglobulins (Ig) in response to danger stimuli [44,45]. 149 150 Finally, beyond the lamina propria, the junctions between endothelial cells around the portal vein are 151 increasingly considered as the last layer of the intestinal barrier, called the gut vascular barrier [46].

In healthy conditions, most dietary nutrients cross the intestinal barrier through enterocytes, while fluids are mainly passively absorbed in a para-cellular manner [47]. In inflammatory states and metabolic diseases, tight junctions are disrupted leading to the uncontrolled passage of several microbiota-derived molecules, such as LPS, leading to low grade systemic inflammation and endotoxemia. Although there are only limited studies addressing whether intestinal permeability is dysregulated in T2D patients, a leaky gut has been shown to play a role in type 1 diabetes and NAFLD onset [48,49].

Miele *et al.* found an association between increased gut permeability and liver steatosis in biopsyproven NAFLD patients [50]. The urinary excretion of orally administrated Chromium-51 Ethylene Diamine Tetra-acetate, as an estimate of gut permeability, was higher in NAFLD patients and correlated positively with liver steatosis. Immunohistochemically assessed ZO-1 protein was also decreased in duodenal biopsies of the NAFLD patients [50]. JAM-A mRNA and protein expression levels are lower in colon biopsies from ultrasonography diagnosed NAFLD subjects [51]. In obese children with NAFLD, diagnosed using MRI and liver biopsy, the intestinal permeability increase positively correlated with liver

steatosis, but not with liver inflammation, ballooning nor NASH [52]. A large meta-analysis of adult and pediatric NAFLD cohorts did not observe a link between increased intestinal permeability and liver inflammation and fibrosis in NAFLD patients [49]. Thus, whereas steatosis and intestinal permeability appear connected, the role of intestinal permeability in inflammation and fibrosis is still under debate.

Feeding mice a high fat diet (HFD) increases hepatic steatosis and IR. These mice show body and liver weight increases, fasting hyperglycemia glucose intolerance and hyperinsulinemia [53]. Intestinal JAM-A deficient mice developed severe liver damage, illustrated by elevated plasma transaminase levels, when fed a NAFLD-inducing high fructose-high sucrose-high cholesterol diet. These mice also showed a rise of systemic LPS exposure, associated with exacerbated hepatic stellate cell activation, fibrosis and inflammation due to hepatic immune cell recruitment [51]. Moreover, HFD-feeding rapidly induced gut vascular barrier damage related to decreased Wnt/b-catenin signaling in endothelial cells [46].

Gut barrier integrity thus likely plays an important role in the development of metabolic diseases such as T2D and NAFLD. Although the chronology of events is still unclear, gut microbiota-derived endotoxemia may contribute to the altered gut liver cross-talk.

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# 2.3 Gut microbiota dysbiosis increases gut permeability and liver damage

The healthy human intestinal microbiota is composed of a large diversity of microorganisms, mainly anaerobic bacteria [54]. In a healthy state, the microbiota have symbiotic interactions with its host. It participates in nutrient absorption, assists in intestinal barrier function protection to prevent pathogen invasion, improves the function of the intestinal immune system and modulates metabolism [54]. The ability of the gut microbiota to process non-digestible components of the diet results in the production of key metabolites for the host such as short-chain fatty acids (SCFA; butyrate, propionate

and acetate) from dietary fibers. These SCFA induce intestinal L-cell release of Glucagon-Like Peptide-1 188 189 (GLP-1) [55] and butyrate improves the maintenance of the intestinal barrier and the intestinal immune system [56]. Gut bacteria also enzymatically modify host products, such as bile acids (BA), hence 190 191 impacting on host metabolism [57,58]. Simultaneously, certain gut microbiota can produce harmful 192 components, such as LPS by Gram-negative bacteria, or ethanol and acetaldehyde. In the healthy state, low production of these compounds may contribute to the prevention of intestinal toxicity [59]. 193 194 Quantitative or qualitative changes in the composition of the microbiota, called dysbiosis, may alter this 195 fine-tuned balance. Changes in the host's diet, administration of antibiotics, an opportunistic pathogen 196 or a pathology can disrupt this equilibrium [59].

Demonstration of the involvement of dysbiosis in the development of metabolic diseases has prompted studies aimed at understanding these variations and to develop new strategies to treat T2D and/or NAFLD through modulation of the gut microbiota. However, a "healthy or normal" microbiota is under the influence of the environment, genetics and geographical location of its host [60,61]. The gut microbiota of patients is therefore very heterogeneous, both at the phyla and family levels, which explains the complexity of studying the signature of a specific microbiome [62–64].

In humans, lower microbial diversity associates positively with dyslipidemia, IR, chronic inflammation, diabetes and obesity [60,61,65]. Furthermore, transplantation of fecal microbiota (FMT) from NAFLD patients into germ-free mice induces hepatic steatosis and inflammation, which are exacerbated by HFD-feeding [66]. The richness of only a few bacteria may vary specifically in metabolic diseases. For instance, *Akkermansia muciniphila* are decreased in obese and T2D patients [67,68] and high levels of *E. coli* correlate positively with hepatic fibrosis [69]. The accompanying increase in systemic LPS correlates with increased hepatic nuclear factor-kappa B (NF-kB) and Toll-Like Receptor 4 (TLR4) positive macrophages in biopsy-proven NASH patients. Circulating bacterial 16S rDNA levels correlate positively with liver damage in obese patients and measurement of its plasma concentrations has been suggested as a marker of liver fibrosis [70]. Although there is a strong assumption that these bacterial components originate from the gut, there is no clear physiological evidence that microbiota exist in blood and 16s rDNA levels may only reflect the increased intestinal permeability. Moreover, their exclusive intestinal origin is not unequivocally proven [70].

In rodents with metabolic disorders, a strong link between dysbiosis and increased intestinal permeability has been established. HFD-feeding of mice induces dysbiosis with gut vascular barrier damage and bacterial translocation to the liver. FMT from healthy mice to HFD mice restored intestinal permeability, as evidenced by the FITC-dextran assay, and metabolic disorders improved [46]. The same protection of the gut barrier is observed upon antibiotic treatment of HFD-fed mice [71]. Treatment with the TLR4 inhibitor improves the liver of mice on a NAFLD-inducing diet [72].

222 Other products of Gram-negative bacteria, such as endogenous ethanol, from carbohydrate fermentation, and acetaldehyde, an ethanol metabolite, stimulate the release of pro-inflammatory 223 224 cytokines from enterocytes and disrupt gut tight junctions [73,74]. In rodent models of NAFLD, 225 endogenous gut-produced ethanol increases hepatic TNF-α expression and inhibits the tricarboxylic acid 226 cycle, resulting in TG accumulation in the liver. Ethanol may increase the activity of cytochrome P450 227 2E1 which produces free radicals through the its oxidation in the liver [59,75,76]. This observation 228 challenged the name "NAFLD", as alcohol may contribute to the disease without exogenous intake [77]. In T2D and NAFLD, other metabolites, such as SCFA, are produced and/or modified by the 229 microbiota and have an impact on disease progression. An overall decrease in butyrate-producing 230 231 bacteria is found in NAFLD and T2D [78]. In humans, low levels of butyrate are associated with decreased

mucus production and a leaky gut [65]. In the colonic lamina propria of mice, butyrate has been shown 232 233 to induce the expression of Foxp3 in CD4<sup>+</sup> lymphocytes, inducing a T-regulatory profile [79]. In vitro butyrate treatment of dendritic cells reduced the expression of pro-inflammatory cytokines in response 234 to LPS [80]. Treatment of HFD-fed mice with butyrate improved villi morphology and gut tight junction 235 236 protein expression [81], which may be due to the regulation of IgA production by B cells, hence protecting the mucus layer [82]. Butyrate-treated HFD-fed mice have lower levels of hepatic TG and 237 238 cholesterol associated with decreased expression of fibrosis genes (TGF- $\beta$ , SMAD2, SMAD7, and  $\alpha$ -SMA) 239 and pro-inflammatory (MCP-1, TNF- $\alpha$ , IL-1, IL-2, IL-6, IFN-y). Thus, decreased systemic butyrate appears 240 to exacerbate NAFLD progression [81]. On the other hand, propionate and acetate increase liver TG 241 storage and hepatic gluconeogenesis in a NAFLD mouse model [59].

Another microbial product with effects on cardio-metabolic diseases is the choline-derivative 242 243 trimethylamine, which increases liver glucose intolerance in mice [83]. In the liver, TMA is oxidized to 244 Trimethylamine N-oxide (TMAO), which has been associated to increased atherosclerosis and cardiovascular risks [83]. Moreover, diabetic patients display increased blood levels of imidazole 245 propionate, a microbiota-derived metabolite of histidine, which induces liver IR [84]. Imidazole 246 propionate alters insulin signaling in the liver through activation of p62 and the mechanistic target of 247 rapamycin complex 1 (mTORC1) resulting in insulin receptor degradation, thus exacerbating IR in T2D 248 249 patients [85].

Overall, the gut microbiota, through modulation of the gut barrier and through the beneficial effects of its products, plays a key role in liver health and dysbiosis may promote the progression of T2D and NAFLD.

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#### 2.4 Alterations of the entero-hepatic cycle of bile acids in NAFLD

Bile acids (BA) are amphipathic molecules acting as detergents to facilitate lipid absorption in the intestine. BA are produced in the liver from cholesterol, secreted in bile and stored in the gallbladder to be released in the intestine after meal ingestion to participate to lipid absorption. In the intestine, the microbiota generates the secondary BA after deconjugation by bacterial bile salt hydrolase, dehydroxylation and epimerization. The majority of BA (95%) are re-absorbed in the distal intestine and recycled to the liver via the portal vein, the remainder being lost in feces [86].

Several studies reported changes in BA composition under different metabolic situations, with results often differing between studies possibly due to differences between the cohorts [87]. In T2D patients, both increased [88] or unchanged [89] plasma BA concentrations have been reported. Higher total plasma BA concentrations have been found in diabetic and non-diabetic NASH patients [87], but the NASH-related increases in plasma BA levels depend on the severity of insulin resistance [90].

Differences between conjugated and unconjugated BA found in NAFLD patients may be due, at least in part, to the decreased colonization of bile salt hydrolase expressing bacteria in the intestine [78]. Interestingly, plasma BA concentrations also increase with progression of hepatic fibrosis [91].

Besides their role in lipid absorption, BA signal via several receptors, amongst which the nuclear receptor Farnesoid X Receptor (FXR) and the Takeda G-protein-coupled Receptor 5 (TGR5) which are expressed in gut and liver. Both FXR and TGR5 regulate, in an opposite manner, GLP-1 secretion by enteroendocrine L cells. TGR5 activation hence improves glucose homeostasis through the release of pancreatic β-cell insulin [86]. Treatment of western diet-fed obese mice with the TGR5 agonist RDX8940 improved IR and hepatic steatosis [92]. TGR5 activation in rodents also improved hepatic inflammation by mechanisms involving reduced NF-κB signaling [93]. Furthermore, TGR5 is expressed in various

immune cells, including dendritic cells, Kupffer cells and macrophages, and its activation decreases
cytokine production and reduces the low grade inflammation observed in metabolic diseases [86].

Under physiological conditios, FXR in enterocytes induces the expression of Fibroblast Growth 278 Factor 19 (FGF19) (and its rodent ortholog FGF15). The peptide secreted in the portal circulation targets 279 280 FGFR4/BKlotho receptor on hepatocytes to repress cholesterol 7 alpha-hydroxylase (CYP7A1) expression and subsequently BA synthesis. Inhibition of BA synthesis reduces hepatic BA toxicity and protects 281 282 against inflammation [94]. In rodents, FGF15-deficiency increases hepatic steatosis and ER stress, which 283 is reversed by adeno-associated virus-mediated expression of FGF19 in the liver [94]. Moreover, FGF19 284 overexpression reduces plasma alanine aminotransferase (ALT) concentrations and improves NASH and 285 fibrosis in a diet-induced NASH model [95]. These mice also lose weight and display improved glucose tolerance [95]. Direct FXR activation in the liver also decreases BA synthesis due to the induction of the 286 287 Cyp7a1 inhibitor Small Heterodimer Partner (SHP) [86]. Finally, with the FGF19 analogue aldafermin 288 improves liver histology in NASH patients (see the next chapter).

Thus, increasing hepatic and intestinal FXR activation, which represses BA synthesis and hepatic 289 290 fibrosis, is a promising therapeutic strategy to treat NAFLD, which explains the development of numerous FXR agonists. In addition to its approved use in the treatment of primary biliary cholangitis 291 292 [96], the FXR agonist obeticholic acid (OCA) is the most advanced compound for fibrosis treatment in 293 NASH [97], whereas tropifexor and nidufexor are other FXR agonists under development [98,99]. 294 Although FXR activation may be a promising strategy to treat fibrosis, the role of FXR in T2D or NAFLD progression remains unclear. Prawitt and colleagues have shown that whole-body FXR knock-out mice 295 are protected against HFD-induced obesity [100]. The improved glycemic control observed in these mice 296 297 is due, at least in part, to a de-repression of GLP-1 secretion by gut L-cells [101]. To decipher the specific

role of FXR in the gut, several studies used intestine-specific FXR-deficient mice. Similarly to what is 298 299 observed in whole-body FXR knock-out mice, intestinal FXR knock-out mice are protected against HFD-300 induced obesity [102]. These mice show lower HFD-induced hepatic SREBP1C-dependent steatosis due to a decrease in intestinal ceramide synthesis [18,103]. However, whether these mice are also protected 301 302 from NASH and fibrosis is unknown at present. All these data highlight the complex role of intestinal FXR in NAFLD progression, also illustrated by the fact that whole-body FXR knock-out mice present gut barrier 303 304 defects when fed a normal chow diet [104] and hepatic steatosis when fed a cholesterol-supplemented 305 diet [105].

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#### 2.5 Gut immune system disruption increases liver inflammation

308 The intestinal immune system, which is an integral part of the gut barrier, contributes to shape 309 gut microbiota tolerance and to maintain gut integrity. Found in intestinal crypts, Paneth cells are 310 specialized cells that secrete antimicrobial peptides which modulate the gut microbiome and the local inflammatory response and contribute to the protection of the organism [106]. Moreover, secondary 311 312 lymphoid organs, the Peyer's patches, are structures localized in the small intestine that contribute to immune system education and tolerance towards food-derived antigens. Specialized M epithelial cells 313 allow antigen transcytosis from the intestinal lumen to the subepithelial dome of Peyer's patches, where 314 315 they induce, via dendritic cell-mediated antigen presentation, the production of immunoglobulin IgA and 316 IgM by plasmocytes [107,108]. IgAs are dimeric immunoglobulins secreted in the mucus layer to protect the mucosa against pathogens [109]. Lymphoid cells are also found outside of Peyer's patches in the 317 intestinal tissue. Indeed, intra-epithelial lymphocytes localized between the enterocytes are resident 318 319 cells contributing to the intestinal barrier [110], and regulatory lymphocytes promoting immune

tolerance [111] as well as resident self-maintaining macrophages which control gut homeostasis [112]
 are also found in the *lamina propria*. Finally, other immune cells such as innate lymphoid cells and
 mucosal-associated invariant T cells are found in the intestine and participate in the mucosal defense
 against pathogens and in the regulation of tissue inflammation [113,114].

324 The intestinal immune system equilibrium, however, is challenged in obesity and T2D leading to an increased low grade chronic inflammation, dysbiosis and gut permeability [115]. In rodents, HFD 325 feeding modifies the gut immune system through changes in intestinal Pattern Recognition Receptor 326 327 (PRR) expression, cytokine production and immune cell populations [116]. Moreover, intestinal IgA<sup>+</sup> B 328 cells are important for the maintenance of glucose tolerance: loss of IgA production results in an increased intestinal pro-inflammatory T cell response in HFD-fed mice [109]. HFD feeding also enhances 329 330 the pool of F4/80<sup>+</sup>CD11b<sup>+</sup>CD11c<sup>+</sup> pro-inflammatory macrophages in the colonic *lamina propria* [117]. 331 Experiments in macrophage CCR2<sup>-</sup> and epithelial CCL2-deficient mice showed that colonic macrophage 332 infiltration increases IR independently of body weight gain [117].

Obese patients display an increased infiltration of CD68<sup>+</sup> macrophages in the jejunal epithelium, 333 which correlates with decreased enterocyte insulin signaling [118]. Obese patients also show jejunal 334 mucosa alterations in CD8<sup>+</sup> T cell subtypes, with a decrease of CD8 $\alpha\alpha^+$  and an increase of CD8 $\alpha\beta^+$  T cells, 335 336 resulting in enhanced pro-inflammatory cytokine secretion [118]. In NAFLD patients, diagnosed using 337 ultrasonography and elevated transaminases, an increased immune cell infiltration in the colonic 338 mucosa has been observed [51]. The epithelial density of CD8<sup>+</sup> T cells also correlated positively with the NAS score, although NAS scores were low excluding the presence of overt NASH [118]. Furthermore, 339 340 bariatric surgery-induced weight loss and improved metabolism are accompanied by increased fecal IgA 341 levels [109].

Whether intestinal immune system dysregulation is a cause or a consequence of gut barrier disruption remains unclear [116]. As gut immune population homeostasis impacts on IR, targeting immune cell populations in the intestinal tissue may be a promising strategy for both T2D and NAFLD.

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#### 346 3 Targeting the intestine to treat T2D and NAFLD

As first-line treatment of T2D, weight-reducing strategies, essentially by lifestyle interventions, also efficiently cure NAFLD. A reduction of calorie intake together with increased physical activity improves NAFL, as assessed by MRI liver fat and plasma ALT level analysis [119], as well as later stage biopsy-confirmed NASH and fibrosis [120].

Here, we discuss therapeutic approaching targeting the intestine and the gut-liver axis, already used in the clinic or in a late stage clinical development (Table 1). It should be noted that these studies are difficult to compare due to the heterogeneity of patients and diagnostic methods used for NASH patient stratification.

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#### 356 **3.1 Bariatric surgery**

Bariatric surgery is a therapeutic option for obese patients with BMI>40, or >35 when comorbidities are present, such as T2D or NASH. These surgical interventions, beyond their effect on weight, cure not only T2D [121], but also improve NASH [122]. Indeed, bariatric surgery induces histological reversion of NASH in 85% of patients one year after the intervention [123], an effect which is sustained 5 years after surgery [124]. Similar results were obtained in patients with non-invasively diagnosed for NASH, combining elastography and MRI [11]. In addition, results from a retrospective study revealed that patients are less likely to progress to cirrhosis after bariatric surgery [125]. Nevertheless, the type of surgical procedure seems to have an impact, since sleeve gastrectomy is less efficient than Roux-en-Y gastric bypass in reducing the Fatty Liver Index and BARD non-invasive scores of NAFL and fibrosis [126].

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#### 368 **3.2 FGF19 analogs**

As discussed above, several preclinical studies identified beneficial effects of the intestinal FGF19 369 pathway on liver health [94,95]. Therefore, aldafermin (NGM282), a recombinant non-tumorigenic 370 371 modified FGF19 peptide, was developed. Twelve weeks of treatment with aldafermin reduced liver fat and ALT levels in biopsy-proven NASH patients, without improving IR [127]. Aldafermin treatment 372 373 reduced the NAS score, and plasma markers of collagen in a dose-escalating phase 2 study, suggesting an improvement of hepatic fibrosis [127,128]. In addition to the reduction of liver fat, 24 week treatment 374 375 with aldafermin (1 mg) resulted in histological improvement of NASH, i.e. lower steatosis, inflammation, 376 ballooning and fibrosis scores without weight loss or effects on IR [129]. Unfortunately, the ALPINE 2/3 study did not meet its primary endpoint, liver fibrosis improvement, and, even though the secondary 377 endpoint, ie NASH resolution without fibrosis worsening, was achieved, the clinical development of 378 379 aldafermin for NASH treatment was stopped [130].

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#### 382 3.3 FXR agonists

Rather than injecting exogenous recombinant FGF19, endogenous enterocyte FGF19 production is induced by FXR agonists, such as OCA, hence increasing peripheral blood FGF19 levels in T2D and NAFLD patients [131]. This synthetic BA is presently the most clinically advanced FXR-targeting therapy.

In NASH patients, long-term OCA treatment improves hepatic histological markers of NASH [97,132].
 However, glucose control and IR may deteriorate after OCA treatment and plasma LDL-cholesterol levels
 increased requiring statin treatment [97].

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Other therapies targeting FXR with (semi-)synthetic compounds show interesting results in preclinical models and, at present, in a few clinical studies. INT-767, a modified BA dual FXR/TGR5 agonist with intestinal tropism, reduces histological features of NASH and lowers fibrosis [133], while promoting brown adipogenesis in rodents [134]. Other synthetic FXR agonists are tropifexor, which reduces plasma TG and induces intestinal FGF15 production in rats [99], and nidufexor, which improves hepatic steatosis and NASH in murine models of T2D and NASH murine model. These compounds are now under evaluation in phase 2 clinical trials for NASH treatment [98,135].

Finally, the intestine-restricted FXR agonist fexaramine[136] improves insulin sensitivity in mice through a microbiota-dependent increase of plasma GLP-1 concentrations [137], but its potential effect on NASH has not yet been reported.

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# 3.4 Targeting the incretin pathway

Complementary to physical exercise and dietary intervention, activation of the GLP-1R either by subcutaneous injection of semi-synthetic recombinant GLP-1R agonists or *per os* treatment with a gliptin, which inhibits the GLP-1-degrading dipeptidyl peptidase-4 (DPP-4) hence increasing endogenous active GLP-1 concentrations, are largely used therapeutic options for the treatment of obesity and T2D. As these metabolic disorders are linked to NAFLD, they are also being tested for this indication in more recent clinical trials.

DPP-4 inhibitor monotherapy with sitagliptin did not improve hepatic steatosis in a two-year clinical trial [138]. In combination with metformin, sitagliptin was shown to decrease the hepatic fat fraction as assessed by MRI, without improving fibrosis in T2D NAFLD patients [139]. Therefore, the efficacy of DPP-4 inhibitors, if any, appears limited to steatosis, without effects on more advanced stages of NASH and fibrosis.

GLP-1R agonist treatment reduces body weight and improves glucose homeostasis. Therefore, 413 they are interesting candidates for the management of NAFLD. The LEAN trial was the first study to show 414 415 efficacy of liraglutide to lower NASH in biopsy-proven NASH patients [140]. More recent studies only 416 show results in patients with less advanced hepatic disease [139,141–143]. In monotherapy, liraglutide 417 was found as efficient as lifestyle interventions to improve NAFL, fasting glycemia and IR assessed by the 418 homeostatic model assessment of insulin resistance (HOMA-IR) [141]. In T2D and NAFLD patients, daily 419 liraglutide and weekly dulaglitide treatment also improved liver steatosis [139,142–144]. Moreover, 420 weekly injections of semaglutide for 72 weeks reversed NASH and reduced ALT levels in NASH patients [145]. Although it is unclear at present whether fibrosis improves upon GLP-1R agonist treatment, this 421 422 compound is of high interest given the fact that it can also be administered orally.

Finally, cotadutide, a dual GLP-1R/glucagon receptor agonist, which mimics the effects of oxyntomodulin, an intestinal peptide dual glucagon receptor and GLP-1R activator, reduced body weight and improved liver histology in mice [146].

426 **3.5 Targeting microbiota** 

427 As discussed above, the gut microbiota plays an important role in health and metabolic diseases, 428 as demonstrated by the fecal microbiota transplantation (FMT) in germ free mice [66]. Lean donor FMT 429 to patients with metabolic syndrome improved peripheral insulin sensitivity [147]. However, FMT with

a healthy donor graft did not improve the hepatic fat fraction nor IR in NAFLD patients [148].
Nevertheless, improved intestinal barrier function was observed, as assessed by the lactulose/mannitol
urinary test [148]. Except for the treatment of intestinal *C. difficile* infection, application of FMT is limited
due to problems of patient acceptance and lack of definition of an healthy graft [149].

434 Since germ-free mice were protected against diet-induced weight gain and hyperglycemia, the use of antibiotics to modulate the microbiota may also be considered. However, antibiotic treatment 435 increased TG levels [150] and penicillin treatment worsened IR and NAFLD in rodent models of HFD-436 437 induced obesity [151]. Such approach has only been tested in a limited manner in humans. Rifaximin, a 438 broad spectrum antibiotic, lowered ALT levels, endotoxemia and HOMA-IR in histologically-diagnosed obese NASH patients [152]. Treatment combining metrodinazole, an antiprotozoal antibiotic, and the 439 440 prebiotic inulin decreased transaminase levels in prediabetic patients, an effect not observed with inulin alone [153]. The lack of bacterial species-selective antibiotics and the apparition of antibiotic resistance 441 442 strongly limit the use of antibiotics for metabolic disorders, which should be restricted to infections.

Rather than replacing or destroying the commensal bacterial ecosystem, gut microbiota can be 443 more subtly modulated in different ways. Prebiotics are substrates selectively used by host micro-444 organisms which confer a health benefit by modulating bacterial metabolite production, eg from non-445 digestible carbohydrates that are transformed into SCFA [154]. Inulin treatment for 14 weeks decreased 446 447 HFD-induced hepatic injury in mice, as illustrated by decreased ALT levels, lower numbers of hepatic 448 macrophages, improved hepatic steatosis and improved glucose tolerance in HFD-fed mice [155]. Prebiotic treatment of non-diabetic patients lowered transaminases serum levels, serum TG and fasting 449 glycemia [156]. As microbial selection by prebiotics is difficult to control and depends on dietary habits 450 451 or geographic origin, exogenous intake of non-pathogenic bacteria can be an option. Probiotics are live

microorganisms which, when administered in proper doses, confer a health benefit to the host [157]. In 452 453 HFD-fed mice, Lactobacillus acidophilus NS1 supplementation improved hepatic steatosis and reduced fatty acid synthesis [158]. Administration of Akkermansia muciniphila modestly improved hepatic 454 histology and reduced hepatic IL-6 expression [68]. Rather than using a single bacteria species, some 455 456 studies evaluated the efficiency of probiotic mixtures on NAFLD. A mixture of 6 probiotic agents lowered hepatic fat content in 12 weeks-treated NAFL patients, without any effect on IR [159]. VSL#3, another 457 probiotic mixture, reduced hepatic steatosis and increased GLP-1 levels, without reducing ALT levels nor 458 459 IR in obese NAFLD children [160].

Finally, pre- and probiotics can be combined into symbiotics. Similar as in HFD-fed rodents [161], 460 an 8 week treatment with a symbiotic containing 7 probiotics and fructo-oligosaccharides improved 461 462 hepatic steatosis without decreasing ALT in NAFLD patients [162]. Twenty-eight weeks of treatment with the same symbiotic improved insulin sensitivity, reduced ALT levels and improved fibrosis in Fibroscan 463 464 diagnosed NAFL patients [163]. However, a recent study reported that combination of Bifidobacterium animalis subspecies lactis BB-12 and fructooligosaccharides for 10-14 months failed to reduce hepatic 465 466 fat content in NAFLD patients, whereas the degree of weight loss was the only variable correlated to steatosis [164]. 467

468 Overall, the benefits of microbiota-based approaches on NAFLD and T2D appear modest. The use 469 of these medications seems limited to the reduction of hepatic steatosis and cytolysis, with sometimes 470 benefits on IR. They thus appear indicated in the prevention and treatment of early, less severe forms of 471 NAFLD.

472

#### 473 **4** <u>Conclusion</u>

474	The metabolic consequences of an altered liver-intestine dialogue reveal its central role in
475	maintenance of energy homeostasis. As T2D and NAFLD impact both organs, targeting the intestine and
476	its functions seems to be a promising therapeutic strategy in the management of obesity, T2D and
477	NAFLD. Further research on the intestine-liver connection is therefore warranted.
478	
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493

# 494 Author contributions

Margaux Nawrot and Simon Peschard wrote the manuscript and made figures. Sophie Lestavel and Bart
Staels revised the article critically for intellectual content and gave final approval of the submitted
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# 498 5 Bibliography

- Evans M, Morgan AR, Patel D, Dhatariya K, Greenwood S, Newland-Jones P, et al. Risk Prediction of the
   Diabetes Missing Million: Identifying Individuals at High Risk of Diabetes and Related Complications.
   Diabetes Ther. 2021 Jan;12(1):87–105.
- Ahlqvist E, Storm P, Käräjämäki A, Martinell M, Dorkhan M, Carlsson A, et al. Novel subgroups of adultonset diabetes and their association with outcomes: a data-driven cluster analysis of six variables. Lancet Diabetes Endocrinol. 2018 May;6(5):361–9.
- 5053.Santoleri D, Titchenell PM. Resolving the Paradox of Hepatic Insulin Resistance. Cell Mol Gastroenterol506Hepatol. 2019;7(2):447–56.
- Haas JT, Francque S, Staels B. Pathophysiology and Mechanisms of Nonalcoholic Fatty Liver Disease. Annu
   Rev Physiol. 2016;78:181–205.
- Younossi ZM, Golabi P, de Avila L, Paik JM, Srishord M, Fukui N, et al. The global epidemiology of NAFLD
   and NASH in patients with type 2 diabetes: A systematic review and meta-analysis. Journal of Hepatology.
   2019 Oct 1;71(4):793–801.
- Eslam M, Newsome PN, Sarin SK, Anstee QM, Targher G, Romero-Gomez M, et al. A new definition for
   metabolic dysfunction-associated fatty liver disease: An international expert consensus statement. J
   Hepatol. 2020 Jul;73(1):202–9.
- Mantovani A, Petracca G, Beatrice G, Tilg H, Byrne CD, Targher G. Non-alcoholic fatty liver disease and
   risk of incident diabetes mellitus: an updated meta-analysis of 501 022 adult individuals. Gut. 2021 May
   1;70(5):962–9.
- Lonardo A, Nascimbeni F, Mantovani A, Targher G. Hypertension, diabetes, atherosclerosis and NASH:
   Cause or consequence? J Hepatol. 2018 Feb;68(2):335–52.
- Sheka AC, Adeyi O, Thompson J, Hameed B, Crawford PA, Ikramuddin S. Nonalcoholic Steatohepatitis: A
   Review. JAMA. 2020 24;323(12):1175–83.
- Harrison SA, Ratziu V, Boursier J, Francque S, Bedossa P, Majd Z, et al. A blood-based biomarker panel
   (NIS4) for non-invasive diagnosis of non-alcoholic steatohepatitis and liver fibrosis: a prospective
   derivation and global validation study. The Lancet Gastroenterology & Hepatology. 2020 Nov
   1;5(11):970–85.
- Allen AM, Shah VH, Therneau TM, Venkatesh SK, Mounajjed T, Larson JJ, et al. Multiparametric Magnetic
   Resonance Elastography Improves the Detection of NASH Regression Following Bariatric Surgery. Hepatol
   Commun. 2020 Feb;4(2):185–92.
- Petersen MC, Shulman GI. Mechanisms of Insulin Action and Insulin Resistance. Physiol Rev. 2018 Oct
   1;98(4):2133–223.
- 13. Khan RS, Bril F, Cusi K, Newsome PN. Modulation of Insulin Resistance in Nonalcoholic Fatty Liver Disease.
   Hepatology. 2019;70(2):711–24.

- 533 14. Donnelly KL, Smith CI, Schwarzenberg SJ, Jessurun J, Boldt MD, Parks EJ. Sources of fatty acids stored in
   534 liver and secreted via lipoproteins in patients with nonalcoholic fatty liver disease. J Clin Invest. 2005 May
   535 2;115(5):1343–51.
- Bezy O, Tran TT, Pihlajamäki J, Suzuki R, Emanuelli B, Winnay J, et al. PKCδ regulates hepatic insulin
  sensitivity and hepatosteatosis in mice and humans. J Clin Invest. 2011 Jun;121(6):2504–17.
- 53816.Perry RJ, Samuel VT, Petersen KF, Shulman GI. The role of hepatic lipids in hepatic insulin resistance and539type 2 diabetes. Nature. 2014 Jun 5;510(7503):84–91.
- Holland WL, Bikman BT, Wang L-P, Yuguang G, Sargent KM, Bulchand S, et al. Lipid-induced insulin
   resistance mediated by the proinflammatory receptor TLR4 requires saturated fatty acid-induced
   ceramide biosynthesis in mice. J Clin Invest. 2011 May;121(5):1858–70.
- 54318.Jiang C, Xie C, Li F, Zhang L, Nichols RG, Krausz KW, et al. Intestinal farnesoid X receptor signaling544promotes nonalcoholic fatty liver disease. J Clin Invest. 2015 Jan 2;125(1):386–402.
- 545 19. Bikman BT, Summers SA. Ceramides as modulators of cellular and whole-body metabolism. J Clin Invest.
  546 2011 Nov;121(11):4222–30.
- Pihlajamäki J, Kuulasmaa T, Kaminska D, Simonen M, Kärjä V, Grönlund S, et al. Serum interleukin 1
  receptor antagonist as an independent marker of non-alcoholic steatohepatitis in humans. J Hepatol.
  2012 Mar;56(3):663–70.
- Schwabe RF, Luedde T. Apoptosis and necroptosis in the liver: a matter of life and death. Nat Rev
   Gastroenterol Hepatol. 2018 Dec;15(12):738–52.
- Shulman GI. Ectopic fat in insulin resistance, dyslipidemia, and cardiometabolic disease. N Engl J Med.
   2014 Sep 18;371(12):1131–41.
- Haas JT, Vonghia L, Mogilenko DA, Verrijken A, Molendi-Coste O, Fleury S, et al. Transcriptional Network
   Analysis Implicates Altered Hepatic Immune Function in NASH development and resolution. Nat Metab.
   2019 Jun;1(6):604–14.
- Breuer DA, Pacheco MC, Washington MK, Montgomery SA, Hasty AH, Kennedy AJ. CD8+ T cells regulate
   liver injury in obesity-related nonalcoholic fatty liver disease. Am J Physiol Gastrointest Liver Physiol. 2020
   Feb 1;318(2):G211–24.
- 560 25. Dudek M, Pfister D, Donakonda S, Filpe P, Schneider A, Laschinger M, et al. Auto-aggressive CXCR6+ CD8 T
   561 cells cause liver immune pathology in NASH. Nature. 2021 Mar 24;
- Bhattacharjee J, Kirby M, Softic S, Miles L, Salazar-Gonzalez R-M, Shivakumar P, et al. Hepatic Natural
   Killer T-cell and CD8+ T-cell Signatures in Mice with Nonalcoholic Steatohepatitis. Hepatol Commun. 2017
   Jun;1(4):299–310.
- 56527.Arriazu E, Ge X, Leung T-M, Magdaleno F, Lopategi A, Lu Y, et al. Signalling via the osteopontin and high566mobility group box-1 axis drives the fibrogenic response to liver injury. Gut. 2017 Jun;66(6):1123–37.

- Petersen KF, Dufour S, Savage DB, Bilz S, Solomon G, Yonemitsu S, et al. The role of skeletal muscle insulin
   resistance in the pathogenesis of the metabolic syndrome. PNAS. 2007 Jul 31;104(31):12587–94.
- Hsieh Y-C, Joo SK, Koo BK, Lin H-C, Kim W. Muscle alterations are independently associated with
  significant fibrosis in patients with nonalcoholic fatty liver disease. Liver Int. 2021 Mar;41(3):494–504.
- 30. Nachit M, Kwanten WJ, Thissen J-P, Beeck BOD, Gaal LV, Vonghia L, et al. Muscle fat content is strongly
  associated with NASH: a longitudinal study in patients with morbid obesity. Journal of Hepatology
  [Internet]. 2021 Apr 14 [cited 2021 Apr 16];0(0). Available from: https://www.journal-ofhepatology.eu/article/S0168-8278(21)00234-8/abstract
- 575 31. Seo DH, Lee Y-H, Park SW, Choi YJ, Huh BW, Lee E, et al. Sarcopenia is associated with non-alcoholic fatty 576 liver disease in men with type 2 diabetes. Diabetes Metab. 2020 Oct;46(5):362–9.
- Targher G, Day CP, Bonora E. Risk of cardiovascular disease in patients with nonalcoholic fatty liver
   disease. N Engl J Med. 2010 Sep 30;363(14):1341–50.
- Targher G, Lonardo A, Byrne CD. Nonalcoholic fatty liver disease and chronic vascular complications of
   diabetes mellitus. Nat Rev Endocrinol. 2018 Feb;14(2):99–114.
- Mantovani A, Turino T, Lando MG, Gjini K, Byrne CD, Zusi C, et al. Screening for non-alcoholic fatty liver
  disease using liver stiffness measurement and its association with chronic kidney disease and
  cardiovascular complications in patients with type 2 diabetes. Diabetes Metab. 2020 Sep;46(4):296–303.
- Weinstein G, Zelber-Sagi S, Preis SR, Beiser AS, DeCarli C, Speliotes EK, et al. Association of Nonalcoholic
   Fatty Liver Disease With Lower Brain Volume in Healthy Middle-aged Adults in the Framingham Study.
   JAMA Neurol. 2018 Jan;75(1):97–104.
- Weinstein G, Davis-Plourde K, Himali JJ, Zelber-Sagi S, Beiser AS, Seshadri S. Non-alcoholic fatty liver
   disease, liver fibrosis score and cognitive function in middle-aged adults: The Framingham Study. Liver
   Int. 2019 Sep;39(9):1713–21.
- 59037.Stahel P, Xiao C, Nahmias A, Lewis GF. Role of the Gut in Diabetic Dyslipidemia. Front Endocrinol591(Lausanne). 2020;11:116.
- Veilleux A, Grenier E, Marceau P, Carpentier AC, Richard D, Levy E. Intestinal lipid handling: evidence and
   implication of insulin signaling abnormalities in human obese subjects. Arterioscler Thromb Vasc Biol.
   2014 Mar;34(3):644–53.
- S95 39. Nogueira J-P, Maraninchi M, Béliard S, Padilla N, Duvillard L, Mancini J, et al. Absence of acute inhibitory
  effect of insulin on chylomicron production in type 2 diabetes. Arterioscler Thromb Vasc Biol. 2012
  S97 Apr;32(4):1039–44.
- 40. Nass KJ, van den Berg EH, Faber KN, Schreuder TCMA, Blokzijl H, Dullaart RPF. High prevalence of
  apolipoprotein B dyslipoproteinemias in non-alcoholic fatty liver disease: The lifelines cohort study.
  Metabolism. 2017 Jul;72:37–46.
- 41. Tilg H, Moschen AR, Roden M. NAFLD and diabetes mellitus. Nat Rev Gastroenterol Hepatol. 2017
  Jan;14(1):32–42.

- 42. Luukkonen PK, Zhou Y, Sädevirta S, Leivonen M, Arola J, Orešič M, et al. Hepatic ceramides dissociate
  steatosis and insulin resistance in patients with non-alcoholic fatty liver disease. J Hepatol. 2016
  May;64(5):1167–75.
- 43. Pelaseyed T, Bergström JH, Gustafsson JK, Ermund A, Birchenough GMH, Schütte A, et al. The mucus and
  mucins of the goblet cells and enterocytes provide the first defense line of the gastrointestinal tract and
  interact with the immune system. Immunological Reviews. 2014;260(1):8–20.
- 44. Turner JR. Intestinal mucosal barrier function in health and disease. Nat Rev Immunol. 2009
  Nov;9(11):799–809.
- 611 45. Camilleri M. Leaky gut: mechanisms, measurement and clinical implications in humans. Gut. 2019
  612 Aug;68(8):1516–26.
- 46. Mouries J, Brescia P, Silvestri A, Spadoni I, Sorribas M, Wiest R, et al. Microbiota-driven gut vascular
  barrier disruption is a prerequisite for non-alcoholic steatohepatitis development. J Hepatol.
  2019;71(6):1216–28.
- 616 47. Schoultz I, Keita ÅV. The Intestinal Barrier and Current Techniques for the Assessment of Gut
  617 Permeability. Cells. 2020 Aug 17;9(8).
- 61848.Bosi E, Molteni L, Radaelli MG, Folini L, Fermo I, Bazzigaluppi E, et al. Increased intestinal permeability619precedes clinical onset of type 1 diabetes. Diabetologia. 2006 Dec;49(12):2824–7.
- 49. De Munck TJI, Xu P, Verwijs HJA, Masclee AAM, Jonkers D, Verbeek J, et al. Intestinal permeability in
  human nonalcoholic fatty liver disease: A systematic review and meta-analysis. Liver Int. 2020
  Dec;40(12):2906–16.
- 62350.Miele L, Valenza V, La Torre G, Montalto M, Cammarota G, Ricci R, et al. Increased intestinal permeability624and tight junction alterations in nonalcoholic fatty liver disease. Hepatology. 2009 Jun;49(6):1877–87.
- Rahman K, Desai C, Iyer SS, Thorn NE, Kumar P, Liu Y, et al. Loss of Junctional Adhesion Molecule A
  Promotes Severe Steatohepatitis in Mice on a Diet High in Saturated Fat, Fructose, and Cholesterol.
  Gastroenterology. 2016;151(4):733-746.e12.
- 52. Pacifico L, Bonci E, Marandola L, Romaggioli S, Bascetta S, Chiesa C. Increased circulating zonulin in
   children with biopsy-proven nonalcoholic fatty liver disease. World Journal of Gastroenterology. 2014
   Dec 7;20(45):17107–14.
- 631 53. Cani PD, Amar J, Iglesias MA, Poggi M, Knauf C, Bastelica D, et al. Metabolic endotoxemia initiates obesity
  632 and insulin resistance. Diabetes. 2007 Jul;56(7):1761–72.
- 54. Sonnenburg JL, Bäckhed F. Diet-microbiota interactions as moderators of human metabolism. Nature.
  2016 Jul 7;535(7610):56–64.
- 55. Ducastel S, Touche V, Trabelsi M-S, Boulinguiez A, Butruille L, Nawrot M, et al. The nuclear receptor FXR
  inhibits Glucagon-Like Peptide-1 secretion in response to microbiota-derived Short-Chain Fatty Acids. Sci
  Rep. 2020 13;10(1):174.

- 638 56. Mathewson ND, Jenq R, Mathew AV, Koenigsknecht M, Hanash A, Toubai T, et al. Gut microbiome639 derived metabolites modulate intestinal epithelial cell damage and mitigate graft-versus-host disease.
  640 Nat Immunol. 2016 May;17(5):505–13.
- 57. Bäckhed F, Ding H, Wang T, Hooper LV, Koh GY, Nagy A, et al. The gut microbiota as an environmental
  factor that regulates fat storage. PNAS. 2004 Nov 2;101(44):15718–23.
- 64358.Wahlström A, Sayin SI, Marschall H-U, Bäckhed F. Intestinal Crosstalk between Bile Acids and Microbiota644and Its Impact on Host Metabolism. Cell Metab. 2016 Jul 12;24(1):41–50.
- 645 59. Chu H, Duan Y, Yang L, Schnabl B. Small metabolites, possible big changes: a microbiota-centered view of
  646 non-alcoholic fatty liver disease. Gut. 2019 Feb;68(2):359–70.
- 647 60. Qin J, Li Y, Cai Z, Li S, Zhu J, Zhang F, et al. A metagenome-wide association study of gut microbiota in type 648 2 diabetes. Nature. 2012 Oct 4;490(7418):55–60.
- 649 61. Karlsson FH, Tremaroli V, Nookaew I, Bergström G, Behre CJ, Fagerberg B, et al. Gut metagenome in
  650 European women with normal, impaired and diabetic glucose control. Nature. 2013 Jun;498(7452):99–
  651 103.
- 62. Human Microbiome Project Consortium. Structure, function and diversity of the healthy human
  microbiome. Nature. 2012 Jun 13;486(7402):207–14.
- 63. Aron-Wisnewsky J, Vigliotti C, Witjes J, Le P, Holleboom AG, Verheij J, et al. Gut microbiota and human
  NAFLD: disentangling microbial signatures from metabolic disorders. Nat Rev Gastroenterol Hepatol.
  2020 May;17(5):279–97.
- 657 64. Fan Y, Pedersen O. Gut microbiota in human metabolic health and disease. Nat Rev Microbiol. 2021
  658 Jan;19(1):55–71.
- 65965.Le Chatelier E, Nielsen T, Qin J, Prifti E, Hildebrand F, Falony G, et al. Richness of human gut microbiome660correlates with metabolic markers. Nature. 2013 Aug 29;500(7464):541–6.
- 66. Chiu C-C, Ching Y-H, Li Y-P, Liu J-Y, Huang Y-T, Huang Y-W, et al. Nonalcoholic Fatty Liver Disease Is
  662 Exacerbated in High-Fat Diet-Fed Gnotobiotic Mice by Colonization with the Gut Microbiota from Patients
  663 with Nonalcoholic Steatohepatitis. Nutrients. 2017 Nov 6;9(11).
- 664 67. Cani PD, de Vos WM. Next-Generation Beneficial Microbes: The Case of Akkermansia muciniphila. Front
  665 Microbiol. 2017;8:1765.
- 666 68. Kim S, Lee Y, Kim Y, Seo Y, Lee H, Ha J, et al. Akkermansia muciniphila Prevents Fatty Liver Disease,
  667 Decreases Serum Triglycerides, and Maintains Gut Homeostasis. Appl Environ Microbiol [Internet]. 2020
  668 Mar 18 [cited 2021 Feb 22];86(7). Available from: https://aem.asm.org/content/86/7/e03004-19
- 669 69. Loomba R, Seguritan V, Li W, Long T, Klitgord N, Bhatt A, et al. Gut Microbiome-Based Metagenomic
  670 Signature for Non-invasive Detection of Advanced Fibrosis in Human Nonalcoholic Fatty Liver Disease.
  671 Cell Metab. 2017 May 2;25(5):1054-1062.e5.

- 672 70. Lelouvier B, Servant F, Païssé S, Brunet A-C, Benyahya S, Serino M, et al. Changes in blood microbiota
  673 profiles associated with liver fibrosis in obese patients: A pilot analysis. Hepatology. 2016
  674 Dec;64(6):2015–27.
- 675 71. Cani PD, Bibiloni R, Knauf C, Waget A, Neyrinck AM, Delzenne NM, et al. Changes in Gut Microbiota
  676 Control Metabolic Endotoxemia-Induced Inflammation in High-Fat Diet–Induced Obesity and Diabetes in
  677 Mice. Diabetes. 2008 Jun 1;57(6):1470–81.
- 678 72. Carpino G, Del Ben M, Pastori D, Carnevale R, Baratta F, Overi D, et al. Increased Liver Localization of
  679 Lipopolysaccharides in Human and Experimental NAFLD. Hepatology. 2020 Aug;72(2):470–85.
- Karal Strain Stra
- 682 74. Chaudhry KK, Shukla PK, Mir H, Manda B, Gangwar R, Yadav N, et al. Glutamine supplementation
  683 attenuates ethanol-induced disruption of apical junctional complexes in colonic epithelium and
  684 ameliorates gut barrier dysfunction and fatty liver in mice. The Journal of Nutritional Biochemistry. 2016
  685 Jan 1;27:16–26.
- 75. Zhu L, Baker SS, Gill C, Liu W, Alkhouri R, Baker RD, et al. Characterization of gut microbiomes in
  nonalcoholic steatohepatitis (NASH) patients: a connection between endogenous alcohol and NASH.
  Hepatology. 2013 Feb;57(2):601–9.
- Raucy JL, Lasker J, Ozaki K, Zoleta V. Regulation of CYP2E1 by Ethanol and Palmitic Acid and CYP4A11 by
   Clofibrate in Primary Cultures of Human Hepatocytes. Toxicological Sciences. 2004 Jun 1;79(2):233–41.
- 691 77. Eslam M, Sanyal AJ, George J, International Consensus Panel. MAFLD: A Consensus-Driven Proposed
   692 Nomenclature for Metabolic Associated Fatty Liver Disease. Gastroenterology. 2020 May;158(7):1999 693 2014.e1.
- 78. Yang G, Wei J, Liu P, Zhang Q, Tian Y, Hou G, et al. Role of the gut microbiota in type 2 diabetes and
  related diseases. Metabolism. 2021 Apr;117:154712.
- Furusawa Y, Obata Y, Fukuda S, Endo TA, Nakato G, Takahashi D, et al. Commensal microbe-derived
  butyrate induces the differentiation of colonic regulatory T cells. Nature. 2013 Dec 19;504(7480):446–50.
- Arpaia N, Campbell C, Fan X, Dikiy S, van der Veeken J, deRoos P, et al. Metabolites produced by
  commensal bacteria promote peripheral regulatory T-cell generation. Nature. 2013 Dec
  19;504(7480):451–5.
- 701 81. Zhou D, Pan Q, Xin F-Z, Zhang R-N, He C-X, Chen G-Y, et al. Sodium butyrate attenuates high-fat diet702 induced steatohepatitis in mice by improving gut microbiota and gastrointestinal barrier. World J
  703 Gastroenterol. 2017 Jan 7;23(1):60–75.
- 70482.Isobe J, Maeda S, Obata Y, Iizuka K, Nakamura Y, Fujimura Y, et al. Commensal-bacteria-derived butyrate705promotes the T-cell-independent IgA response in the colon. Int Immunol. 2020 Apr 12;32(4):243–58.
- 70683.Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, Dugar B, et al. Gut flora metabolism of707phosphatidylcholine promotes cardiovascular disease. Nature. 2011 Apr 7;472(7341):57–63.

- Koh A, De Vadder F, Kovatcheva-Datchary P, Bäckhed F. From Dietary Fiber to Host Physiology: ShortChain Fatty Acids as Key Bacterial Metabolites. Cell. 2016 Jun 2;165(6):1332–45.
- Koh A, Molinaro A, Ståhlman M, Khan MT, Schmidt C, Mannerås-Holm L, et al. Microbially Produced
  Imidazole Propionate Impairs Insulin Signaling through mTORC1. Cell. 2018 Nov 1;175(4):947-961.e17.
- 86. Chávez-Talavera O, Tailleux A, Lefebvre P, Staels B. Bile Acid Control of Metabolism and Inflammation in
  Obesity, Type 2 Diabetes, Dyslipidemia, and Nonalcoholic Fatty Liver Disease. Gastroenterology. 2017
  May;152(7):1679-1694.e3.
- 87. Chávez-Talavera O, Haas J, Grzych G, Tailleux A, Staels B. Bile acid alterations in nonalcoholic fatty liver
  716 disease, obesity, insulin resistance and type 2 diabetes: what do the human studies tell? Curr Opin
  717 Lipidol. 2019 Jun;30(3):244–54.
- 88. Sonne DP, van Nierop FS, Kulik W, Soeters MR, Vilsbøll T, Knop FK. Postprandial Plasma Concentrations of
  Individual Bile Acids and FGF-19 in Patients With Type 2 Diabetes. J Clin Endocrinol Metab. 2016
  Aug;101(8):3002–9.
- 89. Brufau G, Stellaard F, Prado K, Bloks VW, Jonkers E, Boverhof R, et al. Improved glycemic control with
  colesevelam treatment in patients with type 2 diabetes is not directly associated with changes in bile acid
  metabolism. Hepatology. 2010 Oct;52(4):1455–64.
- 90. Grzych G, Chávez-Talavera O, Descat A, Thuillier D, Verrijken A, Kouach M, et al. NASH-related increases
   in plasma bile acid levels depend on insulin resistance. JHEP Rep. 2021 Apr;3(2):100222.
- 91. Caussy C, Hsu C, Singh S, Bassirian S, Kolar J, Faulkner C, et al. Serum bile acid patterns are associated
  with the presence of NAFLD in twins, and dose-dependent changes with increase in fibrosis stage in
  patients with biopsy-proven NAFLD. Aliment Pharmacol Ther. 2019 Jan;49(2):183–93.
- Finn PD, Rodriguez D, Kohler J, Jiang Z, Wan S, Blanco E, et al. Intestinal TGR5 agonism improves hepatic
   steatosis and insulin sensitivity in Western diet-fed mice. Am J Physiol Gastrointest Liver Physiol. 2019
   Mar 1;316(3):G412–24.
- Wang Y-D, Chen W-D, Yu D, Forman BM, Huang W. The G-protein-coupled bile acid receptor, Gpbar1
  (TGR5), negatively regulates hepatic inflammatory response through antagonizing nuclear factor κ lightchain enhancer of activated B cells (NF-κB) in mice. Hepatology. 2011 Oct;54(4):1421–32.
- Alvarez-Sola G, Uriarte I, Latasa MU, Fernandez-Barrena MG, Urtasun R, Elizalde M, et al. Fibroblast
  growth factor 15/19 (FGF15/19) protects from diet-induced hepatic steatosis: development of an FGF19based chimeric molecule to promote fatty liver regeneration. Gut. 2017 Oct 1;66(10):1818–28.
- 738 95. Zhou M, Learned RM, Rossi SJ, DePaoli AM, Tian H, Ling L. Engineered FGF19 eliminates bile acid toxicity
  739 and lipotoxicity leading to resolution of steatohepatitis and fibrosis in mice. Hepatol Commun. 2017
  740 Dec;1(10):1024–42.
- 741 96. Markham A, Keam SJ. Obeticholic Acid: First Global Approval. Drugs. 2016 Aug;76(12):1221–6.

- 97. Neuschwander-Tetri BA, Loomba R, Sanyal AJ, Lavine JE, Van Natta ML, Abdelmalek MF, et al. Farnesoid X
  nuclear receptor ligand obeticholic acid for non-cirrhotic, non-alcoholic steatohepatitis (FLINT): a
  multicentre, randomised, placebo-controlled trial. Lancet. 2015 Mar 14;385(9972):956–65.
- P8. Chianelli D, Rucker PV, Roland J, Tully DC, Nelson J, Liu X, et al. Nidufexor (LMB763), a Novel FXR
  Modulator for the Treatment of Nonalcoholic Steatohepatitis. J Med Chem. 2020 Apr 23;63(8):3868–80.
- 747 99. Tully DC, Rucker PV, Chianelli D, Williams J, Vidal A, Alper PB, et al. Discovery of Tropifexor (LJN452), a
  748 Highly Potent Non-bile Acid FXR Agonist for the Treatment of Cholestatic Liver Diseases and Nonalcoholic
  749 Steatohepatitis (NASH). J Med Chem. 2017 Dec 28;60(24):9960–73.
- Prawitt J, Abdelkarim M, Stroeve JHM, Popescu I, Duez H, Velagapudi VR, et al. Farnesoid X Receptor
   Deficiency Improves Glucose Homeostasis in Mouse Models of Obesity. Diabetes. 2011 Jul 1;60(7):1861–
   71.
- 753101.Trabelsi M-S, Daoudi M, Prawitt J, Ducastel S, Touche V, Sayin SI, et al. Farnesoid X receptor inhibits754glucagon-like peptide-1 production by enteroendocrine L cells. Nat Commun. 2015 Jul 2;6(1):1–13.
- Li F, Jiang C, Krausz KW, Li Y, Albert I, Hao H, et al. Microbiome remodelling leads to inhibition of
   intestinal farnesoid X receptor signalling and decreased obesity. Nat Commun. 2013 Sep 24;4(1):2384.
- Jiang C, Xie C, Lv Y, Li J, Krausz KW, Shi J, et al. Intestine-selective farnesoid X receptor inhibition improves
   obesity-related metabolic dysfunction. Nat Commun. 2015 Dec 15;6(1):1–18.
- 104. Inagaki T, Moschetta A, Lee Y-K, Peng L, Zhao G, Downes M, et al. Regulation of antibacterial defense in
  the small intestine by the nuclear bile acid receptor. Proc Natl Acad Sci U S A. 2006 Mar 7;103(10):3920–
  5.
- Sinal CJ, Tohkin M, Miyata M, Ward JM, Lambert G, Gonzalez FJ. Targeted Disruption of the Nuclear
   Receptor FXR/BAR Impairs Bile Acid and Lipid Homeostasis. Cell. 2000 Sep 15;102(6):731–44.
- 106. Lueschow SR, McElroy SJ. The Paneth Cell: The Curator and Defender of the Immature Small Intestine.
   Front Immunol. 2020;11:587.
- Reboldi A, Cyster JG. Peyer's patches: organizing B-cell responses at the intestinal frontier. Immunol Rev.
   2016 May;271(1):230–45.
- Magri G, Comerma L, Pybus M, Sintes J, Lligé D, Segura-Garzón D, et al. Human Secretory IgM Emerges
   from Plasma Cells Clonally Related to Gut Memory B Cells and Targets Highly Diverse Commensals.
   Immunity. 2017 Jul 18;47(1):118-134.e8.
- 109. Luck H, Khan S, Kim JH, Copeland JK, Revelo XS, Tsai S, et al. Gut-associated IgA+ immune cells regulate
   obesity-related insulin resistance. Nat Commun. 2019 Aug 13;10(1):3650.
- 110. Olivares-Villagómez D, Van Kaer L. Intestinal Intraepithelial Lymphocytes: Sentinels of the Mucosal
   Barrier. Trends Immunol. 2018 Apr;39(4):264–75.

- Hadis U, Wahl B, Schulz O, Hardtke-Wolenski M, Schippers A, Wagner N, et al. Intestinal tolerance
   requires gut homing and expansion of FoxP3+ regulatory T cells in the lamina propria. Immunity. 2011
   Feb 25;34(2):237–46.
- 112. De Schepper S, Verheijden S, Aguilera-Lizarraga J, Viola MF, Boesmans W, Stakenborg N, et al. Self Maintaining Gut Macrophages Are Essential for Intestinal Homeostasis. Cell. 2018 Oct 4;175(2):400 415.e13.
- 113. Vivier E, Artis D, Colonna M, Diefenbach A, Di Santo JP, Eberl G, et al. Innate Lymphoid Cells: 10 Years On.
  Cell. 2018 Aug 23;174(5):1054–66.
- 783 114. Exley MA. What Makes MAITs Wait? Immunity. 2016 Jan 19;44(1):7–9.
- Scheithauer TPM, Rampanelli E, Nieuwdorp M, Vallance BA, Verchere CB, van Raalte DH, et al. Gut
   Microbiota as a Trigger for Metabolic Inflammation in Obesity and Type 2 Diabetes. Front Immunol.
   2020;11:571731.
- 116. Winer DA, Winer S, Dranse HJ, Lam TKT. Immunologic impact of the intestine in metabolic disease. J Clin
   Invest. 2017 03;127(1):33–42.
- 117. Kawano Y, Nakae J, Watanabe N, Kikuchi T, Tateya S, Tamori Y, et al. Colonic Pro-inflammatory
   Macrophages Cause Insulin Resistance in an Intestinal Ccl2/Ccr2-Dependent Manner. Cell Metab. 2016
   09;24(2):295–310.
- Monteiro-Sepulveda M, Touch S, Mendes-Sá C, André S, Poitou C, Allatif O, et al. Jejunal T Cell
   Inflammation in Human Obesity Correlates with Decreased Enterocyte Insulin Signaling. Cell Metab. 2015
   Jul 7;22(1):113–24.
- 119. Wong VW-S, Chan RS-M, Wong GL-H, Cheung BH-K, Chu WC-W, Yeung DK-W, et al. Community-based
   lifestyle modification programme for non-alcoholic fatty liver disease: a randomized controlled trial. J
   Hepatol. 2013 Sep;59(3):536–42.
- Romero-Gómez M, Zelber-Sagi S, Trenell M. Treatment of NAFLD with diet, physical activity and exercise.
   Journal of Hepatology. 2017 Oct 1;67(4):829–46.
- Buchwald H, Buchwald JN. Metabolic (Bariatric and Nonbariatric) Surgery for Type 2 Diabetes: A Personal
   Perspective Review. Diabetes Care. 2019 Feb;42(2):331–40.
- Souto KP, Meinhardt NG, Ramos MJ, Ulbrich-Kulkzynski JM, Stein AT, Damin DC. Nonalcoholic fatty liver
   disease in patients with different baseline glucose status undergoing bariatric surgery: analysis of
   intraoperative liver biopsies and literature review. Surg Obes Relat Dis. 2018 Jan;14(1):66–73.
- Lassailly G, Caiazzo R, Buob D, Pigeyre M, Verkindt H, Labreuche J, et al. Bariatric Surgery Reduces
   Features of Nonalcoholic Steatohepatitis in Morbidly Obese Patients. Gastroenterology. 2015
   Aug;149(2):379–88; quiz e15-16.
- Lassailly G, Caiazzo R, Ntandja-Wandji L-C, Gnemmi V, Baud G, Verkindt H, et al. Bariatric Surgery Provides
   Long-term Resolution of Nonalcoholic Steatohepatitis and Regression of Fibrosis. Gastroenterology. 2020
   Oct;159(4):1290-1301.e5.

- Wirth K, Sheka A, Kizy S, Irey R, Benner A, Sieger G, et al. Bariatric Surgery is Associated With Decreased
   Progression of Nonalcoholic Fatty Liver Disease to Cirrhosis: A Retrospective Cohort Analysis. Annals of
   Surgery. 2020 Jul;272(1):32–9.
- 814 126. Borges-Canha M, Neves JS, Mendonça F, Silva MM, Costa C, Cabral PM, et al. The Impact of Bariatric
   815 Surgery on Hepatic Function and Predictors of Liver Steatosis and Fibrosis. Obes Surg. 2020
   816 Aug;30(8):2935–41.
- Harrison SA, Rinella ME, Abdelmalek MF, Trotter JF, Paredes AH, Arnold HL, et al. NGM282 for treatment
  of non-alcoholic steatohepatitis: a multicentre, randomised, double-blind, placebo-controlled, phase 2
  trial. Lancet. 2018 24;391(10126):1174–85.
- Harrison SA, Rossi SJ, Paredes AH, Trotter JF, Bashir MR, Guy CD, et al. NGM282 Improves Liver Fibrosis
   and Histology in 12 Weeks in Patients With Nonalcoholic Steatohepatitis. Hepatology. 2020
   Apr;71(4):1198–212.
- Harrison SA, Neff G, Guy CD, Bashir MR, Paredes AH, Frias JP, et al. Efficacy and Safety of Aldafermin, an
   Engineered FGF19 Analog, in a Randomized, Double-Blind, Placebo-Controlled Trial of Patients With
   Nonalcoholic Steatohepatitis. Gastroenterology. 2021 Jan;160(1):219-231.e1.
- NGM Bio Reports Topline Results from 24-Week Phase 2b ALPINE 2/3 Study of Aldafermin in NASH |
   NGM Biopharmaceuticals, Inc. [Internet]. [cited 2021 Jun 23]. Available from:
   https://ir.ngmbio.com/news-releases/news-release-details/ngm-bio-reports-topline-results-24-week phase-2b-alpine-23-study/
- Mudaliar S, Henry RR, Sanyal AJ, Morrow L, Marschall H-U, Kipnes M, et al. Efficacy and safety of the
   farnesoid X receptor agonist obeticholic acid in patients with type 2 diabetes and nonalcoholic fatty liver
   disease. Gastroenterology. 2013 Sep;145(3):574-582.e1.
- Younossi ZM, Ratziu V, Loomba R, Rinella M, Anstee QM, Goodman Z, et al. Obeticholic acid for the
   treatment of non-alcoholic steatohepatitis: interim analysis from a multicentre, randomised, placebo controlled phase 3 trial. The Lancet. 2019 Dec 14;394(10215):2184–96.
- Roth JD, Feigh M, Veidal SS, Fensholdt LK, Rigbolt KT, Hansen HH, et al. INT-767 improves
   histopathological features in a diet-induced ob/ob mouse model of biopsy-confirmed non-alcoholic
   steatohepatitis. World J Gastroenterol. 2018 Jan 14;24(2):195–210.
- 134. Comeglio P, Cellai I, Mello T, Filippi S, Maneschi E, Corcetto F, et al. INT-767 prevents NASH and promotes
   visceral fat brown adipogenesis and mitochondrial function. J Endocrinol. 2018 Aug;238(2):107–27.
- Hernandez ED, Zheng L, Kim Y, Fang B, Liu B, Valdez RA, et al. Tropifexor-Mediated Abrogation of
   Steatohepatitis and Fibrosis Is Associated With the Antioxidative Gene Expression Profile in Rodents.
   Hepatology Communications. 2019;3(8):1085–97.
- Bownes M, Verdecia MA, Roecker AJ, Hughes R, Hogenesch JB, Kast-Woelbern HR, et al. A chemical,
   genetic, and structural analysis of the nuclear bile acid receptor FXR. Mol Cell. 2003 Apr;11(4):1079–92.

- Pathak P, Xie C, Nichols RG, Ferrell JM, Boehme S, Krausz KW, et al. Intestine farnesoid X receptor agonist
  and the gut microbiota activate G-protein bile acid receptor-1 signaling to improve metabolism.
  Hepatology. 2018;68(4):1574–88.
- 849138.Cui J, Philo L, Nguyen P, Hofflich H, Hernandez C, Bettencourt R, et al. Sitagliptin vs. placebo for non-850alcoholic fatty liver disease: A randomized controlled trial. J Hepatol. 2016 Aug;65(2):369–76.
- Yan J, Yao B, Kuang H, Yang X, Huang Q, Hong T, et al. Liraglutide, Sitagliptin, and Insulin Glargine Added
   to Metformin: The Effect on Body Weight and Intrahepatic Lipid in Patients With Type 2 Diabetes Mellitus
   and Nonalcoholic Fatty Liver Disease. Hepatology. 2019 Jun;69(6):2414–26.
- Armstrong MJ, Gaunt P, Aithal GP, Barton D, Hull D, Parker R, et al. Liraglutide safety and efficacy in
   patients with non-alcoholic steatohepatitis (LEAN): a multicentre, double-blind, randomised, placebo controlled phase 2 study. The Lancet. 2016 Feb 13;387(10019):679–90.
- 141. Khoo J, Hsiang JC, Taneja R, Koo S-H, Soon G-H, Kam CJ, et al. Randomized trial comparing effects of
  weight loss by liraglutide with lifestyle modification in non-alcoholic fatty liver disease. Liver Int.
  2019;39(5):941–9.
- Guo W, Tian W, Lin L, Xu X. Liraglutide or insulin glargine treatments improves hepatic fat in obese
   patients with type 2 diabetes and nonalcoholic fatty liver disease in twenty-six weeks: A randomized
   placebo-controlled trial. Diabetes Res Clin Pract. 2020 Dec;170:108487.
- 143. Feng W, Gao C, Bi Y, Wu M, Li P, Shen S, et al. Randomized trial comparing the effects of gliclazide,
  liraglutide, and metformin on diabetes with non-alcoholic fatty liver disease. Journal of Diabetes.
  2017;9(8):800–9.
- Kuchay MS, Krishan S, Mishra SK, Choudhary NS, Singh MK, Wasir JS, et al. Effect of dulaglutide on liver
   fat in patients with type 2 diabetes and NAFLD: randomised controlled trial (D-LIFT trial). Diabetologia.
   2020 Nov;63(11):2434–45.
- 145. Newsome PN, Buchholtz K, Cusi K, Linder M, Okanoue T, Ratziu V, et al. A Placebo-Controlled Trial of
   Subcutaneous Semaglutide in Nonalcoholic Steatohepatitis. N Engl J Med. 2020 Nov 13;
- 871 146. Boland ML, Laker RC, Mather K, Nawrocki A, Oldham S, Boland BB, et al. Resolution of NASH and hepatic
  872 fibrosis by the GLP-1R and GCGR dual-agonist cotadutide via modulating mitochondrial function and
  873 lipogenesis. Nature Metabolism. 2020 May;2(5):413–31.
- 147. Vrieze A, Van Nood E, Holleman F, Salojärvi J, Kootte RS, Bartelsman JFWM, et al. Transfer of intestinal
  microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome.
  Gastroenterology. 2012 Oct;143(4):913-916.e7.
- 148. Craven L, Rahman A, Nair Parvathy S, Beaton M, Silverman J, Qumosani K, et al. Allogenic Fecal
  Microbiota Transplantation in Patients With Nonalcoholic Fatty Liver Disease Improves Abnormal Small
  Intestinal Permeability: A Randomized Control Trial. Am J Gastroenterol. 2020 Jul;115(7):1055–65.
- van Nood E, Vrieze A, Nieuwdorp M, Fuentes S, Zoetendal EG, de Vos WM, et al. Duodenal infusion of
   donor feces for recurrent Clostridium difficile. N Engl J Med. 2013 Jan 31;368(5):407–15.

- Bäckhed F, Manchester JK, Semenkovich CF, Gordon JI. Mechanisms underlying the resistance to diet induced obesity in germ-free mice. PNAS. 2007 Jan 16;104(3):979–84.
- Mahana D, Trent CM, Kurtz ZD, Bokulich NA, Battaglia T, Chung J, et al. Antibiotic perturbation of the
   murine gut microbiome enhances the adiposity, insulin resistance, and liver disease associated with high fat diet. Genome Med. 2016 Apr 27;8(1):48.
- Abdel-Razik A, Mousa N, Shabana W, Refaey M, Elzehery R, Elhelaly R, et al. Rifaximin in nonalcoholic
   fatty liver disease: hit multiple targets with a single shot. Eur J Gastroenterol Hepatol. 2018
   Oct;30(10):1237–46.
- 153. Chong CYL, Orr D, Plank LD, Vatanen T, O'Sullivan JM, Murphy R. Randomised Double-Blind Placebo Controlled Trial of Inulin with Metronidazole in Non-Alcoholic Fatty Liver Disease (NAFLD). Nutrients.
   2020 Mar 27;12(4).
- 893 154. Gibson GR, Hutkins R, Sanders ME, Prescott SL, Reimer RA, Salminen SJ, et al. Expert consensus
  894 document: The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus
  895 statement on the definition and scope of prebiotics. Nat Rev Gastroenterol Hepatol. 2017 Aug;14(8):491–
  896 502.
- Bao T, He F, Zhang X, Zhu L, Wang Z, Lu H, et al. Inulin Exerts Beneficial Effects on Non-Alcoholic Fatty
   Liver Disease via Modulating gut Microbiome and Suppressing the Lipopolysaccharide-Toll-Like Receptor
   4-Mψ-Nuclear Factor-κB-Nod-Like Receptor Protein 3 Pathway via gut-Liver Axis in Mice. Front
   Pharmacol. 2020;11:558525.
- Behrouz V, Aryaeian N, Zahedi MJ, Jazayeri S. Effects of probiotic and prebiotic supplementation on
   metabolic parameters, liver aminotransferases, and systemic inflammation in nonalcoholic fatty liver
   disease: A randomized clinical trial. J Food Sci. 2020 Oct;85(10):3611–7.
- Hill C, Guarner F, Reid G, Gibson GR, Merenstein DJ, Pot B, et al. Expert consensus document. The
   International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and
   appropriate use of the term probiotic. Nat Rev Gastroenterol Hepatol. 2014 Aug;11(8):506–14.
- 907 158. Park S-S, Lee Y-J, Song S, Kim B, Kang H, Oh S, et al. Lactobacillus acidophilus NS1 attenuates diet-induced
  908 obesity and fatty liver. J Endocrinol. 2018 May;237(2):87–100.
- Ahn SB, Jun DW, Kang B-K, Lim JH, Lim S, Chung M-J. Randomized, Double-blind, Placebo-controlled Study
   of a Multispecies Probiotic Mixture in Nonalcoholic Fatty Liver Disease. Sci Rep. 2019 Apr 5;9(1):5688.
- 911 160. Alisi A, Bedogni G, Baviera G, Giorgio V, Porro E, Paris C, et al. Randomised clinical trial: The beneficial
  912 effects of VSL#3 in obese children with non-alcoholic steatohepatitis. Aliment Pharmacol Ther. 2014
  913 Jun;39(11):1276–85.
- 161. Lensu S, Pariyani R, Mäkinen E, Yang B, Saleem W, Munukka E, et al. Prebiotic Xylo-Oligosaccharides
   Ameliorate High-Fat-Diet-Induced Hepatic Steatosis in Rats. Nutrients. 2020 Oct 22;12(11).
- 916 162. Asgharian A, Askari G, Esmailzade A, Feizi A, Mohammadi V. The Effect of Symbiotic Supplementation on
   917 Liver Enzymes, C-reactive Protein and Ultrasound Findings in Patients with Non-alcoholic Fatty Liver

- 918Disease: A Clinical Trial. Int J Prev Med [Internet]. 2016 Mar 10 [cited 2021 Feb 22];7. Available from:919https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4809112/
- 163. Eslamparast T, Poustchi H, Zamani F, Sharafkhah M, Malekzadeh R, Hekmatdoost A. Synbiotic
  supplementation in nonalcoholic fatty liver disease: a randomized, double-blind, placebo-controlled pilot
  study. Am J Clin Nutr. 2014 Mar;99(3):535–42.
- 923 164. Scorletti E, Afolabi PR, Miles EA, Smith DE, Almehmadi A, Alshathry A, et al. Synbiotics Alter Fecal
   924 Microbiomes, But Not Liver Fat or Fibrosis, in a Randomized Trial of Patients With Nonalcoholic Fatty
   925 Liver Disease. Gastroenterology. 2020 May;158(6):1597-1610.e7.
- 165. Neuschwander-Tetri BA, Loomba R, Sanyal AJ, Lavine JE, Van Natta ML, Abdelmalek MF, et al. Farnesoid X
   nuclear receptor ligand obeticholic acid for non-cirrhotic, non-alcoholic steatohepatitis (FLINT): a
   multicentre, randomised, placebo-controlled trial. Lancet. 2015 Mar 14;385(9972):956–65.

Figure 1: The cross-talk between liver and metabolic organs is altered in T2D and NAFLD. Common
 dysregulations observed in T2D and NAFLD highlighting the central role of the liver and its dialogue with
 metabolic organs.

933 (Abbreviation: FFA: Free Fatty Acid)

934

**Figure 2: Gut dysfunctions alter liver health in T2D and/or NAFLD.** Left panel shows the multiple pathways of intestine-liver cross-talk in the healthy state. Enterocytes mediate the post-prandial absorption of nutrients, such as fats, regulating TG storage and chylomicron synthesis. The mucus layer and TJ complexes protect against bacterial translocation and endotoxemia. SCFA produced by the intestinal microbiota improve the gut barrier through mucin and IgA production, by goblet cells and LB respectively. Conjugated and deconjugated BA via binding to TGR5 receptor enhance GLP-1 release to maintain glucose homeostasis, and to FXR increase FGF15/19 synthesis.

942 Right panel shows the dysregulations of these pathways in T2D and/or NAFLD in relation to excess calorie intake and their impact on the liver. Increased FA absorption promotes gut and liver inflammation, and 943 944 liver steatosis. The gut barrier disruption is illustrated by a thinner mucus layer and a decrease of TJ, 945 which is exacerbated by microbiota alterations. Dysbiosis, associated with increased LPS and ethanol 946 levels, exacerbates gut inflammation, bacteria translocation in the portal circulation and endotoxemia 947 increasing liver steatosis, inflammation and fibrosis. Moreover, the change of SCFA producing bacteria 948 worsens gut permeability by decreasing mucin and IgA levels, promoting liver steatosis and IR. Simultaneously, alterations in intestinal BA metabolism, such as decreased BA deconjugation, in these 949 pathological contexts may lead to reduced levels of GLP-1 and FGF15/19, hence impacting on liver 950 951 injuries such as steatosis and fibrosis.

952	(Abbreviations: FA: Fatty Acid; LPS: lipopolysaccharide; EtOH: Ethanol; LT: T lymphocyte; LB: B
953	Lymphocyte; DC: dendritic cell; BA: Bile Acids; SCFA: short chain fatty acid; TGR5: takeda g-protein-
954	coupled receptor 5; FXR: farnesoid X receptor; GLP-1: Glucagon-like peptide-1; FGF15/19: Fibroblast
955	growth factor 15 or 19; TG: triglycerides; TJ: Tight Junctions; IgA: Immunoglobulin A; ZO-1: Zonula
956	Occludens 1; JAM-A: junctional adhesion molecule A; BSH: Bile Salt Hydrolase; ER stress: Endoplasmic
957	Reticulum Stress; IR: Insulin resistance; T2D: Type 2 Diabetes; NAFLD: Non-Alcoholic Fatty Liver diseases)
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959	Table 1: Clinical trials on NAFLD treatments targeting gut functions.
960	Abbreviations: ALT : alanine aminotransferase; AST : aspartate aminotransferase; BMI : body mass index,
961	GGT : gamma-glutamyl transferase; MRI : magnetic resonance imaging; NAFLD : non-alcoholic fatty liver
962	disease; NAS : NAFLD activity score; NASH : non-alcoholic steatohepatitis; OCA : obeticholic acid; OGTT :
963	oral glucose tolerance test; T2D : type 2 diabetes; TG : triglyceride
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Figure 2

Treatment	n NAFLD	NAFLD	Diagnostic	n T2D	Treatment effects on NAFLD	Treatment effects on T2D/IR	Study
	patients	type		patients			
Bariatric surgery		_					
	109	NASH	Liver biopsy	69	NASH reversion at 1 year post-surgery in 85% of patients	Reduction of fasting glycemia and insulin resistance from baseline	[123]
	64	NASH	Liver biopsy	?	NASH reversion at 5 year post surgery in 85% of patients	Reduction of fasting glycemia, HbA1c, Insulin resistance index from baseline	[124]
	2942	NAFL	?	987	Reduced risk to develop hepatic cirrhosis if patients underwent bariatric surgery		[125]
	1955	NAFL	FLI score : TG, BMI, GGT and waist circumference BARD score : BMI, AST/ALT, diabetes	522	Reduction of both FLI and BRAD score 1 year after surgery		[126]
FGF19 analogs							
Aldafermin (NGM282)							
	27 (3 mg) 28 (6 mg)	NASH	Liver biopsy MRI ALT	15 (3 mg) 17 (6 mg)	Decrease of hepatic fat content > 30% in 85% of patients, independently of dose Decrease of circulating fibrosis markers	No impact on fasting glycemia or insulin resistance	[127]
	24 (1 mg) 19 (3 mg)	NASH	Liver biopsy MRI ALT	9 (1 mg) 7 (3 mg)	Dose dependent improvement of NASH and fibrosis		[128]
	53 (1 mg)	NASH	Liver biopsy MRI ALT	32	Reduction of hepatic fat content, NAS score, fibrosis and ALT	No impact on fasting glycemia, HbA1c or HOMA-IR	[129]
FXR agonists							
Obeticholic Acid (OCA)							
	20 (25 mg) 21 (50 mg)	NAFL	Ultrasonography (17 + 18) ALT (3 + 5) Liver biopsy (0 + 2)	20 (25 mg) 21 (50 mg)	Increase plasma FGF19 Reduction of circulating fibrosis markers	Improve insulin sensitivity	[131]
	141	NASH	Liver biopsy	75	Liver histology improvement Decrease of weight loss, ALT, GGT	Increase of insulin resistance and fasting insulimemia	[165]
	312 (10 mg) 308 (25 mg)	NASH	Liver biopsy	171 (10 mg) 171 (25 mg)	Dose dependent fibrosis reduction Reduction of NAS score for the 25 mg dose	In T2D patients, early increase of glycemia and HbA1c, which return to placebo level by month 6	[132]

Targeting Incretin path	way						
Sitagliptin	_						
	25	NAFL	MRI + ALT	12	Not better than placebo to improve liver fat content	No effect on glycemia, insulinemia, HbA1c, IR	[138]
+ Metformin	27	NAFL	MRI	27	Reduction of hepatic fat fraction	Reduction of HbA1c	[139]
Liraglutide							
	23	NASH	Liver biopsy	9	NASH resolution Reduction of NAS score components	Reduction of HbA1c and fasting glycemia	[140]
	14	NAFL	MRI + ALT	?	Reduction of liver fat fraction from baseline Decrease of AST and ALT	Reduction of Insulin resistance, glycemia and insulinemia from baseline	[141]
	31	NAFL	Steatosis on liver biopsy or imaging technique	31	Reduction of intrahepatic content of lipids Decrease of AST and ALT from baseline	Reduction of insulin resistance	[142]
	29	NAFL	Ultrasonography	29	Reduction of hepatic fat Decrease of AST and ALT	Improvement in fasting glycemia, HbA1c and glucose tolerance after OGTT	[143]
+ Metformin	24	NAFL	MRI	27	Reduction of hepatic fat fraction	Reduction of HbA1c, fasting glycemia and post prandial plasma glucose	[139]
Semaglutide							
	80 (0.1 mg) 78 (0.2 mg) 82 (0.4 mg)	NASH	Liver biopsy	49 (0.1 mg) 51 (0.2 mg) 49 (0.4 mg)	NASH Resolution Reduction of ALT and AST		[145]
Targeting microbiota							
Allogenic FMT							
	15	NAFL	Liver biopsy + FibroScan + ultrasonography	?	No effect	No effect	[148]
Antibiotics							
Rifaximin	25	NASH	Liver biopsy + elevated ALT level	11	Reduction at 3 and 6 months of ALT, NAFLD-liver fat score, endotoxemia, IL6 Decrease at 6 months of AST Increase at 3 and 6 month of IL10	Reduction at 3 and 6 months of insulin resistance Reduction at 6 month of glycemia and insulinemia	[152]
Metrodinazole + inulin	20	?	Liver biopsy or BMI>27 + T2D or metabolic	7	Reduction of ALT	No effect	[153]

			1					
			elevated ALT					
Prebiotics								
Inulin	29	NAFL	Ultrasonography + ALT	0	Reduction of ALT, AST and TG	Reduction of fasting glycemia	[156]	
Probiotics								
Lactobacillus acidophilus, Lactobacillusrhamnosus, Lactobacillus paracasei, Pediococcus pentosaceus, Bifidobacterium lactis & Bifidobacterium breve	30	NAFL	MRI	?	Reduction of intrahepatic fat Reduction of TG	No changes in fasting glycemia, HOMA-IR or insulinemia	[159]	
VSL#3	22 children	NASH	Liver biopsy + ALT	?	No effect on ALT Improvement of fatty liver	No effect on insulin resistance or fasting glycemia Increase of GLP-1	[160]	
Symbiotics								
Lactobacillus acidophilus, Lactobacillus rhamnosus, Lactobacillus casei, Lactobacillus bulgaricus, Bifidobacterium Iongum, Bifidobacterium breve & Streptococcus thermophilus + fructooligosaccharides	38	NAFL	Ultrasonography	?	No effect on ALT Improvement of steatosis		[162]	
	26	NAFL	Ultrasonography + elevated ALT	?	Decrease of ALT Improvement of fibrosis score	Reduction of insulin resistance, fasting glycemia and insulinemia	[163]	
Bifidobacterium animalis sub lactis BB12 + fructooligosaccha <u>rides</u>	45	NAFL	MRI	17	No improvement in fatty liver Reduction of NAFLD fibrosis score vs baseline, but not vs placebo		[164]	

ALT : alanine aminotransferase, AST : aspartate aminotransferase, BMI : body mass index, GGT : gamma-glutamyl transferase, MRI : magnetic
 resonance imaging, NAFLD : non-alcoholic fatty liver disease, NAS : NAFLD activity score, NASH : non-alcoholic steatohepatitis, OCA : obeticholic
 acid, OGTT : oral glucose tolerance test, T2D : type 2 diabetes, TG : triglycerides

#### Table 1