



## Bisphosphonates in common pediatric and adult bone sarcomas

Marie-Francoise Heymann, Frederic Lezot, Dominique Heymann

### ► To cite this version:

Marie-Francoise Heymann, Frederic Lezot, Dominique Heymann. Bisphosphonates in common pediatric and adult bone sarcomas. BONE, 2020, 139, pp.115523. 10.1016/j.bone.2020.115523 . inserm-03298321

HAL Id: inserm-03298321

<https://inserm.hal.science/inserm-03298321v1>

Submitted on 23 Jul 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

## **Bisphosphonates in common pediatric and adult bone sarcomas**

Marie-Francoise Heymann<sup>1,2</sup>, Frederic Lezot<sup>3</sup>, Dominique Heymann<sup>1,2,4</sup>

1. Institut de Cancérologie de l'Ouest, Saint-Herblain, France

2. Université de Nantes, Institut de Cancérologie de l'Ouest, Saint-Herblain, France

3. Université de Nantes, Inserm, U1238, Faculty of Medicine, Nantes, France

4. University of Sheffield, Dept of Oncology and Metabolism, School of Medicine, Sheffield, UK

### **Corresponding author:**

Prof. D. Heymann

Institut de Cancérologie de l'Ouest

Blvd Jacques Monod

44805 Saint-Herblain, France

Email: [dominique.heymann@univ-nantes.fr](mailto:dominique.heymann@univ-nantes.fr)

## **Abstract**

The therapeutic strategies proposed currently for bone sarcomas are based on neo-adjuvant chemotherapy, delayed en-bloc wide resection, and adjuvant chemotherapy. Unfortunately, bone sarcomas are characterized by high rates of poor drug response, with a high risk of drug resistance, local recurrence and/or a high propensity for induced metastases. The pathogenesis of bone sarcomas is strongly associated with dysregulation of local bone remodeling and increased osteolysis that plays a part in tumor development. In this context, bisphosphonates (BPs) have been proposed as a single agent or in combination with conventional drugs to block bone resorption and the vicious cycle established between bone and sarcoma cells. Pre-clinical *in vitro* studies revealed the potential “anti-tumor” activities of nitrogen-bisphosphonates (N-BPs). In pre-clinical models, N-BPs reduced significantly primary tumor growth in osteosarcoma and Ewing sarcoma, and the installation of lung metastases. In chondrosarcoma, N-BPs reduced the recurrence of local tumors after intralesional curettage, and increased overall survival. In pediatric and adult osteosarcoma patients, N-BPs have been assessed in combination with conventional chemotherapy and surgery in randomized phase 3 studies with no improvement in clinical outcome. The lack of benefit may potentially be explained by the biological impact of N-BPs on macrophage differentiation/recruitment which may alter CD8<sup>+</sup>-T lymphocyte infiltration. Thanks to their considerable affinity for the mineralized extracellular matrix, BPs are an excellent platform for drug delivery in malignant bone sites with reduced systemic toxicity, which opens up new opportunities for their future use.

## 1. Introduction

Bone sarcomas are malignant primary tumors in bone, represented by heterogeneous neoplastic entities of mesenchymal origin [1]. Based on their low frequency, with less than 2% of malignant tumors registered in the EUROCARE database, bone sarcomas are considered to be orphan tumors. Osteosarcoma, Ewing sarcoma and chondrosarcoma are the main members of this tumor family. Osteosarcoma and Ewing sarcomas are associated with a high ability to establish metastatic nodules in the lungs and bone, and chondrosarcomas are characterized by a high risk of local recurrence and metastatic disease for high-grade histological subtypes [1].

Initial development of bone sarcomas is explained by the conjunction of oncogenic events (e.g. mutation of *p53*, *Rb*, *EXT*; *fusion gene: EWS-FLI*) and dysregulation of the microenvironment which markedly contributes to the pathogenesis of the disease [3-7]. As bone is the natural initial environment for bone sarcomas, bone cells cooperate with tumor cells and facilitate their proliferation and migration. Osteoblasts and mesenchymal stem cells communicate directly with bone sarcoma cells using GAP junctions [8,9], but also indirectly through the release of cytokines/growth factors and the production of extracellular vesicles [10]. All bone sarcoma entities are characterized by a major alteration to local bone remodeling that can lead to bone fractures. This bone degradation is linked to the recruitment of osteoclast precursors and/or the activation of mature osteoclasts by soluble factors secreted by cancer cells. M-CSF and RANKL are the two main protagonists related to this process [3]. In turn, activated osteoclasts degrade bone and induce the release of growth factors initially trapped in the bone extracellular matrix. TGF- $\beta$  is one of the factors that stimulates cancer cell proliferation and cell migration [11,12]. A vicious cycle between resident cells and tumor cells is then established and plays a part in bone sarcoma growth and the associated metastatic process (Figure 1).

The activation of osteoclastogenesis in bone sarcomas has stimulated therapeutic interest in osteoclast targeting agents as a mean of blocking the functional dependency of tumor growth related to bone resorption [13,14]. Bisphosphonates (BPs) have been used successfully for skeletal diseases associated with exacerbated activation of osteoclasts. In this context, BPs were initially used in Paget's disease and malignant hypercalcemia such as bone metastases, and their clinical indications have progressively been extended to osteoporosis [15]. Osteoclasts/osteoclast precursors were the first targets of BPs, but decades of studies have

clearly demonstrated that BPs have numerous cellular targets including macrophages, endothelial cells and immune cells, and could also modulate the biology of cancer cells, at least *in vitro*. BP use has been studied extensively in prostate and breast cancer patients with bone metastases, and has been also assessed in bone sarcomas. The aim of the present review is to give a brief overview of the mechanisms of action of BPs in bone sarcomas, and to discuss their clinical relevance in these tumor entities.

## **1. BPs and osteosarcoma**

Osteosarcoma is the main bone sarcoma and represents around 50% of malignant primary bone tumors [1]. Osteosarcomas are characterized by a proliferation of mesenchymal cells responsible for the formation of an osteoid matrix. Even though a small number of osteosarcomas can be directly related to predisposition factors such as a mutation of *p53* (Li Fraumeny syndrome), fibrous dysplasia, Paget's disease and irradiation, most osteosarcomas are of unknown cause. The former entities are named secondary osteosarcomas and are more frequently observed around the age of 60 years, and the latter are called primary or idiopathic osteosarcomas and are characterized by a peak of incidence around the age of 18 years. They can be classified according their localization (e.g. medullary, intracortical, and juxtacortical osteosarcomas), as well as according to their histological subtypes (e.g. fibroblastic, chondroblastic, telangiectatic). Initial osteosarcoma growth is associated with a phase of osteoclastic activation/recruitment, which has led to the use of BPs [16]. Osteosarcoma can be then considered as a bone-forming tumor functionally related to osteoclasts. The local microenvironment is also characterized by a marked immune infiltrate composed of a high level of macrophages and lymphocytes [17].

### **1. BPs modulate the local osteosarcoma microenvironment**

#### **1.1. BP targeting of osteoclasts in osteosarcoma**

Osteoclasts and their precursors are the main cellular targets of BPs in bone, and inhibit bone resorption by inducing osteoclast cell death [18]. Interestingly, it has been observed that administration of nitrogen-containing BPs (N-BPs) such as zoledronate may interfere with the mechanical properties of bone, by altering bone remodeling and inhibiting the biological functions of both osteoclasts and osteoblasts [19]. For example, N-BP-treated mice showed reduced bone growth [19]. Low doses of N-BPs (0.02-0.025 mg/kg) were associated with a frequency of hypocalcemia that was significantly reduced by decreasing the dose of N-BPs used (0.0125 mg/kg). However, these doses were lower than those used in an oncological

context [20]. Higher doses of N-BPs (50 µg/kg every 2 days × 10) were then assessed in growing mice to determine their potential harmful impact on bone [21]. N-BPs had a transient inhibitory effect on bone length with bone-growth arrest during treatment. These effects were associated to a strong inhibition of resorption leading to an increase of the metaphyseal bone density [22, 23]. Osteoblast and osteoclast biomarkers decreased in serum. The bone growth arrest was stopped at the end of N-BP treatment. A young patient with an osteosarcoma treated with conventional chemotherapy combined with N-BP (50 µg/kg every 2 days × 10 iv injections) exhibited an increase in bone mineral density at the growth plate related to inhibition of bone resorption that was transient, as shown by X-ray radiography [21]. N-BPs inhibit osteoclast function transiently and could be used in young patients. Zoledronate can be safely combined with conventional chemotherapy with maximal tolerated doses mg/m<sup>2</sup> (max 4 mg) for patients with metastatic osteosarcoma [24]. However, more recently, Lezot *et al.* showed that high doses of zoledronate irreversibly disturbed teeth eruption and elongation, and delayed skull bone formation in a preclinical mouse model [25].

BPs were then assessed in immunocompetent and immunodeficient murine models of osteosarcoma. In an immunocompetent rat model (OSRGa cells), zoledronate reduced the growth of primary tumors and inhibited the osteolytic lesions associated with tumor growth [26]. When combined with conventional chemotherapy (Ifosfamide), it was more effective than single agents for preventing tumor recurrence and improving tissue repair. In immunodeficient mice (inoculation of human Saos2 osteosarcoma cells), zoledronate (s.c. twice weekly at 120 microg/kg) inhibited tumor growth in the orthotopic site [27]. On the contrary, Labrinidis *et al.* did not observe any effect of zoledronate on tumor growth when given as either a weekly or a single dose of 100 microg/kg (inoculation of human K-Hos osteosarcoma cells) [28]. However, they did observe a beneficial effect of zoledronate in that it prevented osteolytic lesions and significantly limited the amount of malignant ectopic bone formation [28]. In a xenogenic model (athymic rats) of canine osteosarcoma, zoledronate improved bone architecture and its mechanical properties when combined with PTH [29]. Similarly, Wolfe *et al.* observed that zoledronate had a significant bone protector effect in a murine model of canine osteosarcoma (OSCA40 cells inoculated intratibially in nude mice followed by limb amputation), with no effective inhibition of lung metastases [30]. In two different models (immunocompetent: murine POS-1 cells inoculated in C3H/He mice [31]; immunodeficient: murine LM8 cells injected into nude mice [32]), high doses of zoledronate significantly reduced the establishment of lung metastases and improved overall survival.

These results were controversial, and zoledronate did not show any therapeutic benefit in immunodeficient mice (human K-HOS cells) [28] nor in an immunocompetent rat osteosarcoma model (MSK- 8G osteosarcoma cells) [33]. The literature nevertheless described sporadic but encouraging progression-free survival in patients with high-grade osteosarcoma and progressive metastatic disease after polychemotherapy and treated with zoledronate alone [34]. However, two independent phase III clinical trials did not demonstrate any therapeutic benefit of combining zoledronate with conventional chemotherapy in pediatric and adult metastatic osteosarcoma patients [35,36]. These discrepancies may be explained by the differential origin of the cell lines used, the cancer cell inoculation protocol (e.g. intratibial, paraosseous sites), or the strain of mice/rat models (C3H/HE, Balbc nu/nu, NMRI Nude, Sprague Dawley, Fisher 344). Furthermore, there are major conflicts in the literature regarding the involvement of osteoclasts in metastasis. There is some evidence that osteoclasts are directly associated with poor outcomes in osteosarcoma [37] that may be related to the pro-angiogenic role of osteoclasts [38]. In contrast, other studies show that osteoclasts localized at the site of primary tumors are capable of preventing the metastatic process [37]. This apparent dual activity of osteoclasts may be related to the stage of the disease and should be reconsidered in light of recent knowledge acquired about osteoclasts [39]. Osteoclasts belong to a very heterogeneous family composed of multinucleated cells with pro-inflammatory and immune-tolerant activities that may explain the differential therapeutic benefit of BPs depending on the subsets of osteoclast.

## **1.2. Targeting of immune cells by BPs**

Tumor-associated macrophages (TAMs) are recruited by cancer cells and infiltrate the tumor mass. Their polarization appears to be controlled by the local microenvironment [40]. In 2015, Junankar *et al.* definitively established that macrophages are the main targets of BPs and consequently that the antitumor activity observed in bone and outside the skeleton is mediated at least partly via TAMs [41]. Osteosarcoma tissues are markedly infiltrated by TAMS [17] and their polarization is crucial in tumor development and the initiation of the metastatic process [42,43]. Zoledronate prevents M2-macrophage polarization by targeting the mevalonate pathway and RhoA geranyl-geranylation [44]. In addition, zoledronate promotes M1-macrophage polarization though the TL4 pathway [45]. The dysregulation of M1/M2 macrophage polarization induced by N-BPs may alter CD8+-T lymphocyte infiltration [46]. This functional impact on macrophage polarization may explain the lack of therapeutic benefit of N-BPs in osteosarcoma patients [44,45].

N-BPs induce intracellular accumulation of isopentenyl diphosphate (IPP) and dimethylallyl diphosphate (DMAPP) by inhibiting farnesyl pyrophosphate synthase [47-49]. Treatment of cancer cells *in vitro* by zoledronate results in the accumulation of IPP/DMAPP, cell death and then the release of the phosphoantigen that can trigger activation of V $\gamma$ 9V $\delta$ T lymphocytes [47-48]. Similarly, the peripheral blood monocytes targeted by N-BPs are responsible for V $\gamma$ 9V $\delta$ T lymphocyte activation, induced through accumulation of phosphoantigens and in a cell contact-dependent manner [49]. V $\gamma$ 9V $\delta$ T lymphocyte activation by N-BPs was a promising therapeutic line in osteosarcoma [50-54]. However, Fleming *et al.* recently revealed that  $\gamma$ T cells may promote tumor progression by mimicking Treg/Th2-like cells and inducing local immunosuppression (e.g. modulation of dendritic cell activity and inhibition of PD1-PDL1 axis) [55].

### **1.3. Direct *in vitro* activity of BPs in osteosarcoma**

Numerous publications have demonstrated the direct effects of BPs on osteosarcoma cells *in vitro* [56-68]. N-BPs inhibited the proliferation of osteosarcoma cells and induced cell death by blocking cell cycle progression in the S, G2/M phases independently of their p53 and Rb status [66,67]. This cell cycle arrest was due to activation of the intra-S DNA damage checkpoint, and more particularly to an increase in ATR, P-chk1, Wee1, and P-cdc2 phosphorylation levels and a decrease in cdc25c [66]. Zoledronate induced Rb phosphorylation and stimulated the progression in the G1/S phase of Rb wild-type osteosarcoma cells, leading to their accumulation in the S phase, where they became more sensitive to DNA damage. Failing to repair their DNA damage, the osteosarcoma cells initiated a feedback process by inhibiting Rb phosphorylation through cyclin/cdk complex regulation [66]. In addition, zoledronate induced atypical apoptosis [66,68]. Zoledronate-induced cell death was independent of caspase activation and involved the mitochondria pathway. N-BP-associated cell death was characterized by nuclear alterations and dysregulation of the Bax/Bcl2 balance. It also implicated up-regulation of mitochondrial permeability, with translocation of apoptosis-inducing factor (AIF) and endonuclease-G (EndoG) [66,68]. At the end of the molecular process, N-BPs markedly altered cytoskeletal organization and cell junctions, leading to inhibited cell migration and disturbance of focal adhesion kinases. More recently, AKT/GSK-3 $\beta$ / $\beta$ -Catenin signaling was also associated with N-BP-induced cell death [56]. Both N-BPs and non-N-BPs also directly modulated osteosarcoma cell migration and

invasion through the inhibition of metalloproteinase expression [69-71]. Finally, N-BPs suppressed human osteosarcoma cell metastasis by induction of EMT [72].

The development of a drug resistant phenotype can be observed after long-term treatment of osteosarcoma cells with N-BPs [73,74]. This phenotype was specific to N-BPs, and N-BP-resistant cells remained sensitive to non-N-BPs (e.g. clodronate or pamidronate) as well as being associated with a differential expression of farnesyl diphosphate synthase (FPPS), the main target of N-BPs [73]. In addition to this metabolic resistance, N-BPs increased stem cell markers such as NANOG, cMYC, OCT 4, and SOX-2 directly related to the drug resistance [74,75]. Zoledronate-induced cell resistance can be bypassed by inhibiting HSP-27 expression [76]. BPs have been also assessed in combination with other drugs or therapeutic approaches (Table 1) and showed beneficial effects. Despite the *in vitro* effect of BPs on osteosarcoma cells, no direct effects of BPs on bone sarcoma cells have been demonstrated *in vivo*.

## **2. Use of BPs in Ewing sarcoma**

Ewing sarcoma is the second most represented bone sarcoma in terms of frequency. It affects children and adolescents, with a peak of incidence at the age of 15 years. Like osteosarcoma, it also has a marked propensity for inducing lung metastases, which are responsible for dramatically low overall survival of around 20% at 5 years [1]. Histologically, Ewing sarcoma cells are CD99 positive undifferentiated round cells. They are characterized at the molecular level by the presence of EWS/FLi gene fusion. Like osteosarcoma, the development of Ewing sarcoma in bone sites is associated with the recruitment of osteoclastic precursors and the activation of osteoclasts, resulting in local bone destruction. In this context, BPs have been assessed in pre-clinical *in vivo* models and revealed a significant therapeutic benefit [89-91]. N-BPs inhibited the development of primary tumors only in bone sites and not in soft tissue. They also reduced dissemination via inhibition of matrix metalloproteinases [89,90]. Zhou *et al.* compared the effect of single treatments (zoledronate, paclitaxel) with combined therapy and showed that the most effective treatment was zoledronate associated with paclitaxel [89]. Similarly, minodronate induced synergistic inhibitory activity on tumor growth when combined with doxorubicin. In combination with ifosfamide, zoledronate showed synergistic effects on *in vivo* tumor growth [90]. The aim of Ewing 2008 (Clinical Trail.gov Identifier: NCT00987636), a randomized, phase 3 clinical trial, was to study the therapeutic benefit of associating N-BPs and conventional chemotherapy. More than 900 patients with localized and disseminated Ewing sarcoma were enrolled. The final analyses are

in progress and the results will be announced soon. *In vitro*, N-BPs exhibited marked activities on cancer cells and inhibited cell proliferation with a cell cycle arrest in the S, G2/M phase [90,92,93] and additive effects when combined with conventional chemotherapeutic drugs (e.g. doxorubicin, etoposide and vincristine) [94]. N-BPs may also alter the immune response in Ewing sarcoma. Mueller *et al.* analyzed the *in vitro* antitumor activities of NK cells, and revealed that zoledronate impeded *in vitro* NK cell expansion and cytolytic function against Ewing sarcoma cells. Consequently the combination of N-BPs with adoptive transfer does not appear recommended.

### **3. BPs in the treatment of chondrosarcoma**

With a peak of incidence at the age of 45 years, chondrosarcoma is a cartilaginous tumor characterized by malignant chondrocytes trapped in a poorly-vascularized hyaline-like extracellular matrix [95]. Depending on the histological subtype, chondrosarcomas are associated with somatic mutations in *IDH-1* or *IDH-2*, *EXT*, *p53*, *Rb*, or *COL2A1*. Chondrosarcomas are considered to be radio- and chemoresistant tumors, and consequently the surgical approach is the first therapeutic line for low-grade entities combined with chemotherapy or radiotherapy for the highest grade tumors. Chondrosarcoma is associated with high local recurrence and a metastatic potential for high-grade tumors. Like osteosarcoma and Ewing sarcoma, osteoclasts play a key role in the pathogenesis of chondrosarcoma and the inhibition of bone resorption impaired tumor growth [96,97], and slowed down recurrent progression after intralesional curettage [96]. *In vitro* investigations demonstrated that N-BPs induced chondrosarcoma cell death in a caspase-independent manner [96,98,99] and inhibited cell invasion through decreased secretion of MMP2 [100]. Treatment of chondrosarcoma cells by N-BPs induced the production of phosphoantigen which may stimulate the expansion of V $\gamma$ 9V $\delta$ 2 T cells as well as increase their cytotoxic activity [101]. A phase 1b clinical trial (Clinical Trail.gov Identifier: NCT03173976) is currently in progress to assess the safety and efficacy of neoadjuvant zoledronate in patients with resectable chondrosarcoma of any grade. Fifteen patients will be enrolled. Its completion date is estimated as July 2022.

### **4. Perspectives for BPs in bone sarcoma**

In addition to the three main entities of pediatric and adult sarcomas described above, BPs have been assessed in other malignant and benign primary bone tumors and tumor-like lesions (Table 3). A patient suffering from an advanced sacrum chordoma with zoledronate showed a

significant pain reduction which was refractory to analgesic, opioids and antiepileptic compounds. The tumor appeared unchanged during the treatment [102]. No large clinical trial has been set up yet. Because giant cell tumors of bone are characterized by the presence of numerous and large osteoclasts, BP-based therapeutic approaches were assessed in patients [103,104]. In both studies, no serious general adverse effects were observed but adjuvant treatment with zoledronate did not prevent the local recurrence of the tumors. Aneurysmal bone cysts are aggressive and expansile bone lesions associated with hyper active osteolytic process. Pamidronate and zoledronate showed therapeutic benefits with a reduction of the bone oedema and partial or complete ossification of their lesions [105,106]. Fibrous dysplasia is a benign fibro-osseous bone disease in which the bone is progressively replaced by a fibrous tissue and increased bone resorption associated with bone pain and fragility. In this context, BPs have been also assessed in fibrous dysplasia [107-113]. BP treatments did not arrest the expanding feature of these bone lesions but showed a significant improvement in bone pain, prevented fractures. Malignant transformations have been described for all of these bone diseases. Indeed, in addition to show morphological similarities between low-grade central osteosarcoma and fibrous dysplasia [114], osteosarcoma can arise in this bone disease [115]. Similarly, osteosarcomas arising in aneurysmal bone cyst [116] or in Paget disease [117,118] have been described. Such osteosarcomas could identify a subgroup of osteosarcoma for which BPs may represent an interesting option. In a similar manner, telangiectatic osteosarcoma may represent a specific subset of osteosarcoma associating giant cells enriched microenvironment [119]. BP therapies resulted in a decrease of osteoclast number and the development of a fibroblastic stroma [119]. It could be interesting to revisit the therapeutic approach of these rare osteosarcomas in the light of such observations.

As described above, from the *in vitro* and *in vivo* results of pre-clinical investigations, BPs have shown promising clinical advantages in the treatment of bone sarcomas. Unfortunately, the clinical trials have been disappointing in osteosarcoma with no improvement in clinical outcome. Thanks to their strong attraction for the bone mineral matrix, BPs have been used extensively in bone imaging activities [120]. This property has now been revisited in order to obtain functionalized BPs capable of delivering drugs to bone sites (Table 2). Various conjugation strategies are now available for functionalizing imaging agents, anti-tumor agents and nanocarriers with BPs. The readers will find more details in the excellent manuscript proposed by Xing *et al.* in the special issue on Bisphosphonates [132]. Their clinical advantages of functionalized BPs are not only to deliver high concentrations of drugs to the

tumor development site, but also to reduce the systemic effects of anti-cancer compounds, and consequently their toxicity. Some have already obtained orphan drug status and should be assessed soon in clinical trials.

## References

- [1] H.K. Brown, K. Schiavone, F. Gouin, M.F. Heymann, D. Heymann, Biology of bone sarcomas and new therapeutic developments, *Calcif Tissue Int.* 102 (2018) 174-195.
- [2] C.A. Stiller, A.W. Craft, I. Corazziari, Survival of children with bone sarcoma in Europe since 1978: results from the EUROCARE study. *Eur. J. Cancer* 37 (2001) 760–766.
- [3] Y. Wittrant, S. Théoleyre, C. Chipoy, M. Padrines, F. Blanchard, D. Heymann, F. Rédin, RANKL/RANK/OPG: new therapeutic targets in bone tumours and associated osteolysis. *Biochim Biophys Acta.* 1704 (2004) 49-57.
- [4] F. Lamoureux, P. Richard, Y. Wittrant, S. Battaglia, P. Pilet, V. Trichet, F. Blanchard, F. Gouin, B. Pitard, D. Heymann, F. Redini, Therapeutic relevance of osteoprotegerin gene therapy in osteosarcoma: blockade of the vicious cycle between tumor cell proliferation and bone resorption. *Cancer Res.* 67 (2007) 7308-7318.
- [5] V. Crenn, K. Biteau, J. Amiaud, C. Dumars, R. Guiho, L. Vidal, L.L. Nail, D. Heymann, A. Moreau, F. Gouin, F. Redini, Bone microenvironment has an influence on the histological response of osteosarcoma to chemotherapy: retrospective analysis and preclinical modeling. *Am. J. Cancer Res.* 7 (2017) 2333-2349.
- [6] F. Redini, D. Heymann, Bone tumor environment as a potential therapeutic target in ewing sarcoma. *Front. Oncol.* 33 (2015) 279.
- [7] E. David, F. Blanchard, M.F. Heymann, G. De Pinieux, F. Gouin, F. Rédi, D. Heymann, The bone niche of chondrosarcoma: a sanctuary for drug resistance, tumor growth and also a source of new therapeutic targets. *Sarcoma* 2011 (2011) 932451.
- [8] M. Tellez-Gabriel, C. Charrier, B. Brounais-Le Roye, M. Mullar, H.K. Brown, F. Verrecchia, D. Heymann, Analysis of gap junctional intercellular communications using a dielectrophoresis-based microchip. *Eur J Cell Biol.* 96 (2017) 110-118.
- [9] J. Talbot, R. Brion, G. Picarda, J. Amiaud, J. Chesneau, G. Bougras, V. Stresing, F. Tirode, D. Heymann, F. Redini, F. Verrecchia, Loss of connexin43 expression in Ewing's sarcoma cells favors the development of the primary tumor and the associated bone osteolysis. *Biochim. Biophys. Acta.* 1832 (2013) 553-564.
- [10] F. Perut, L. Roncuzzi, N. Baldini, The emerging roles of extracellular vesicles in osteosarcoma. *Front. Oncol.* 9 (2019) 1342.
- [11] A. Cappariello, N. Rucci, Tumour-derived extracellular vesicles (EVs): a dangerous "message in a bottle" for bone. *Int. J. Mol. Sci.* 20 (2019)

[12] F. Verrecchia, F. Redini, Transforming growth factor- $\beta$  signaling plays a pivotal role in the interplay between osteosarcoma cells and their microenvironment. *Front. Oncol.* 8 (2018) 133.

[13] D. Heymann, Y. Fortun, F. Rédini, M. Padrines, Osteolytic bone diseases: physiological analogues of bone resorption effectors as alternative therapeutic tools. *Drug Discov. Today* 10 5 (2005) 242-247.

[14] D. Heymann, B. Ory, F. Gouin, J.R. Green, F. Rédini, Bisphosphonates : new therapeutic agents for the treatment of bone tumors. *Mol. Med.* 10 (2004) 337-343.

[15] S.H. Ralston. **Bisphosphonates in the management of Paget's disease.** *Bone*, (<https://doi.org/10.1016/j.bone.2020.115465>)

[16] F.C. Kelleher, H. O'Sullivan. Monocytes, macrophages, and osteoclasts in osteosarcoma. *J. Adolesc. Young Adult Oncol.* 6 (2017) 396-405.

[17] M.F. Heymann, F. Lézot, D. Heymann, The contribution of immune infiltrates and the local microenvironment in the pathogenesis of osteosarcoma. *Cell. Immunol.* 343 (2019) 103711.

[18] M.J. Rogers, J.C. Crockett, F.P. Coxon, J. Mönkkönen, Biochemical and molecular mechanisms of action of bisphosphonates. *Bone* 49 (2011) 34-41.

[19] S. Pozzi, S. Valet, S. Mukherjee, D. Cirstea, N. Vaghela, L. Santo, E. Rosen, H. Ikeda, Y. Okawa, T. Kiziltepe, J. Schoonmaker, W. Xie, T. Hideshima, E. Weller, M.L. Bouxsein, N.C. Munshi, K.C. Anderson, N. Raje, High-dose zoledronic acid impacts bone remodeling with effects on osteoblastic lineage and bone mechanical properties. *Clin. Cancer Res.* 15 (2009) 5829-5839.

[20] Munns CF, Mansoor HR, Hong J, Briody J, Höglér W, McQuade M, D.G. Little, C.T. Cowell, Acute phase response and mineral status following low dose intravenous zoledronic acid in children. *Bone* 41 (2007) 366-370.

[21] S. Battaglia, S. Dumoucel, J. Chesneau, M.F. Heymann, G. Picarda, F. Gouin, N. Corradini, D. Heymann, F. Redini, Impact of oncopediatric dosing regimen of zoledronic acid on bone growth: preclinical studies and case report of an osteosarcoma pediatric patient. *J. Bone Miner. Res.* 26 (2011) 2439-2351.

[22] R. Schenk, P. Eggli, H. Fleisch, S. Rosini, Quantitative morphometric evaluation of the inhibitory activity of new aminobisphosphonates on bone resorption in the rat. *Calcif. Tissue Int.* 38(1986):342-9.

[23] R. Schenk, W.A. Merz, R. Mühlbauer, R.G.G. Russell, H. Fleisch, Effect of ethane-1-hydroxy-1,1-diphosphonate (EHDP) and dichloromethylene diphosphonate (Cl<sub>2</sub>MDP) on the calcification and resorption of cartilage and bone in the tibial epiphysis and metaphysis of rats. *Calcified Tissue Res.* 11(1973) 196-214.

- [24] R.E. Goldsby, T.M. Fan, D. Villaluna, L.M. Wagner, M.S. Isakoff, J. Meyer, R.L. Randall, S. Lee, G. Kim, M. Bernstein, R. Gorlick, M. Krailo, N. Marina, Feasibility and dose discovery analysis of zoledronic acid with concurrent chemotherapy in the treatment of newly diagnosed metastatic osteosarcoma: a report from the Children's Oncology Group. *Eur. J. Cancer* 49 (2013) 2384-2391.
- [25] F. Lézot, J. Chesneau, S. Battaglia, R. Brion, B. Castaneda, J.C. Farges, D. Heymann, F. Rémini, Preclinical evidence of potential craniofacial adverse effect of zoledronic acid in pediatric patients with bone malignancies. *Bone* 68 (2014) 146-152.
- [26] D. Heymann, B. Ory, F. Blanchard, M.F. Heymann, P. Coipeau, C. Charrier, S. Couillaud, J.P. Thiery, F. Gouin, F. Redini, Enhanced tumor regression and tissue repair when zoledronic acid is combined with ifosfamide in rat osteosarcoma. *Bone* 37 (2005) 74-86.
- [27] C.R. Dass, P.F. Choong, Zoledronic acid inhibits osteosarcoma growth in an orthotopic model. *Mol. Cancer Ther.* 6 (2007) 3263-3270.
- [28] A. Labrinidis, S. Hay, V. Liapis, V. Ponomarev, D.M. Findlay, A. Evdokiou, Zoledronic acid inhibits both the osteolytic and osteoblastic components of osteosarcoma lesions in a mouse model. *Clin. Cancer Res.* 15 (2009) 3451-3461.
- [29] Curtis RC, Custis JT, Ehrhart NP, Ehrhart EJ, Condon KW, Gookin SE, Donahue SW. Combination therapy with zoledronic acid and parathyroid hormone improves bone architecture and strength following a clinically-relevant dose of stereotactif radiation therapy for local treatment of canine osteosarcoma in athymic rat. *PLoS One* 11 (2016) e0158005
- [30] T.D. Wolfe, S.P. Pillai, B.E. Hildreth 3rd, L.G. Lanigan, C.K. Martin, J.L. Werberck, T.J. Rosol, Effect of zoledronic acid and amputation on bone invasion and lung metastasis of canine osteosarcoma in nude mice. *Clin. Exp. Metastasis* 28 (2011) 377-389.
- [31] B. Ory, M.F. Heymann, A. Kamijo, F. Gouin, D. Heymann, F. Redini F, Zoledronic acid suppresses lung metastases and prolongs overall survival of osteosarcoma-bearing mice. *Cancer* 104 (2005) 2522-2529.
- [32] K. Koto, N. Horie, S. Kimura, H. Murata, T. Sakabe, T. Matsui, M. Watanabe, S. Adachi, T. Maekawa , S. Fushiki, T. Kubo, Clinically relevant dose of zoledronic acid inhibits spontaneous lung metastasis in a murine osteosarcoma model. *Cancer Lett.* 274 (2009) 271-278.
- [33] A. Labrinidis, S. Hay, V. Liapis, D.M. Findlay, A. Evdokiou, Zoledronic acid protects against osteosarcoma-induced bone destruction but lacks efficacy against pulmonary metastases in a syngeneic rat model. *Int. J. Cancer* 127 (2010) 345-354.
- [34] R.M. Conry, M.G. Rodriguez, J.G. Pressey, Zoledronic acid in metastatic osteosarcoma: encouraging progression free survival in four consecutive patients. *Clin. Sarcoma Res.* 6 (2016) 6.
- [35]. S. Piperno-Neuman, M.C. Le Deley, F. Rémini, H. Pacquement, P. Marec-Bérard, P. Petit, H. Brisson, C. Lervat, J.C. Gentet, N. Entz-Werlé, A. Italiano, N. Corradini, E. Bompas, N. Penel, M.D. Tabone, A. Gomez-Brouchet, J.M. Guinebretière, E. Mascard, F. Gouin, A. Chevance, N. Bonnet, J.Y. Blay, L. Brugières; Sarcoma Group of UNICANCER; French

Society of Pediatric Oncology (SFCE); French Sarcoma Group (GSF-GETO), zoledronate in combination with chemotherapy and surgery to treat osteosarcoma (OS2006) : a randomized multicentre, open-label, phase 3 trial. Lancet Oncol. 17 (2016) 1070-1080.

[36].S. Li, P. Chen, Y. Pei, K. Zheng, W. Wang, E. Qiu, X. Zhang, Addition of zoledronate to chemotherapy in patients with osteosarcoma treated with limb-sparing surgery: a phase III clinical trial. Med. Sci. Monit. 25 (2019) 1429-1438.

[37] L. Endo-Munoz, A. Evdokiou A., N.A. Saunders, The role of osteoclasts and tumour-associated macrophages in osteosarcoma metastasis. Biochim. Biophys. Acta. 1826 (2012) 434-442.

[38] F.C. Cackowski, J.L. Anderson, K.D. Patrene, R.J. Choksi, S.D. Shapiro, J.J. Windle, H.C. Blair, G.D. Roodman, Osteoclasts are important for bone angiogenesis. Blood 115 (2010) 140-149.

[39] M.B. Made, L. Ibáñez, A. Wakkach, T.J. de Vries, A. Teti, F. Apparailly, C. Blin-Wakkach, Immune function and diversity of osteoclasts in normal and pathological conditions. Front. Immunol. 10 (2019) 1408.

[40] K. Mehla, P.K. Singh. Metabolic regulation of macrophage polarization in cancer. Trends Cancer. 5 (2019) 822-834.

[41] S. Junankar, G. Shay, J. Jurczyluk, N. Ali, J. Down, N. Pocock, A. Parker, A. Nguyen, S. Sun, B. Kashemirov, C.E. McKenna, P.I. Croucher, A. Swarbrick, K. Weilbaecher, T.G. Phan, M.J. Rogers, Real-time intravital imaging establishes tumor-associated macrophages as the extraskeletal target of bisphosphonate action in cancer. Cancer Discov. 5 (2015) 35-42.

[42] E.P. Buddingh, M.L. Kuijjer, R.A. Duim, H. Bürger, K. Agelopoulos K, O. Myklebost, M. Serra, F. Mertens, P.C. Hogendoorn, A.C. Lankester, A.M. Cleton-Jansen, Tumor-infiltrating macrophages are associated with metastasis suppression in high-grade osteosarcoma: a rationale for treatment with macrophage activating agents. Clin. Cancer Res. 17 (2011) 2110-2119.

[43] C. Dumars, J.M. Ngyuen, A. Gaultier, R. Lanel, N. Corradini, F. Gouin, D. Heymann, M.F. Heymann, Dysregulation of macrophage polarization is associated with the metastatic process in osteosarcoma. Oncotarget. 7 (2016) 78343-78354.

[44] G. Comito, C. Pons Segura, M.L. Taddei, M. Lanciotti, S. Serni, A. Morandi, P. Chiarugi, E. Giannoni, Zoledronic acid impairs stromal reactivity by inhibiting M2-macrophages polarization and prostate cancer-associated fibroblasts. Oncotarget. 2017 Jan 3;8(1):118-132.

[45] W. Zhu, R. Xu, J. Du, Y. Fu, S. Li, P. Zhang, L. Liu, H. Jiang, Zoledronic acid promotes TLR-4-mediated M1 macrophage polarization in bisphosphonate-related osteonecrosis of the jaw. FASEB J. 33 (2019) 5208-5219.

[46] A. Gomez-Bouchet, C. Illac, J. Gilhodes, C. Bouvier, S. Aubert, J.M. Guinebretiere, B. Marie, F. Larousserie, N. Entz-Werlé, G. de Pinieux, T. Filleron, V. Minard, V. Minville, E. Mascard, F. Gouin, M. Jimenez, M.C. Ledley, S. Piperno-Neumann, L. Brugieres, F. Rédi, 14

CD163-positive tumor-associated macrophages and CD8-positive cytotoxic lymphocytes are powerful diagnostic markers for the therapeutic stratification of osteosarcoma patients: An immunohistochemical analysis of the biopsies from the French OS2006 phase 3 trial. *Oncoimmunology* 6 (2017) e1331193.

[47] L.M. Mitrofan, J. Pelkonen, J. Mönkkönen, The level of ATP analog and isopentenyl pyrophosphate correlates with zoledronic acid-induced apoptosis in cancer cells in vitro. *Bone* 45 (2009) 1153-1160.

[48] J. Räikkönen, H. Mönkkönen, S. Auriola, J. Mönkkönen. Mevalonate pathway intermediates downregulate zoledronic acid-induced isopentenyl pyrophosphate and ATP analog formation in human breast cancer cells. *Biochem. Pharmacol.* 79 (2010) 777-783.

[49] A.J. Roelofs, M. Jauhainen, H. Mönkkönen, M.J. Rogers, J. Mönkkönen, K. Thompson, Peripheral blood monocytes are responsible for gammadelta T cell activation induced by zoledronic acid through accumulation of IPP/DMAPP. *Br. J. Haematol.* 144 (2009) 245-50.

[50] Z.X. Li, L.L. Sun, R.L. Cheng, Z.W. Sun, Z.M. Ye, Amplification of  $\gamma\delta$  T cells in PBMCs of healthy donors and osteosarcoma patients stimulated by zoledronate. *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi* 28 (2012) 822-824

[51] M. Liu, L.L. Sun, Y.J. Li, H.Y. Li, J. Zhang, B.H. Li, Z.M. Ye, Trastuzumab enhanced the cytotoxicity of V $\gamma$ 9V $\delta$ 2 T cells against zoledronate-sensitized osteosarcoma cells. *Int. Immunopharmacol.* 28 (2015) 160-167.

[52] H. Jiang, Q. Xu, C. Yang, Z.G. Cao, Z.X. Li, Z.M. Ye ZM. Gammadelta T cells stimulated by zoledronate kill osteosarcoma cells. *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi* 26 (2010) 1195-1197.

[53] Z. Li, H. Peng, Q. Xu, Z. Ye, Sensitization of human osteosarcoma cells to V $\gamma$ 9V $\delta$ 2 T-cells-mediated cytotoxicity by zoledronate. *J. Orthop. Res.* 30 (2012) 824-830.

[54] S. Wang, H. Li, C. Ye, P. Lin, B. Li, W. Zhang, L. Sun, Z. Wang, D. Xue, W. Teng, X. Zhou, N. Lin, Z. Ye, Vaproic acid combined with zoledronic acid enhanced gd T cell-mediated cytotoxicity against osteosarcoma cells via the accumulation of mevalonated pathway intermediate. *Front. Immunol.* 9 (2018) 9:377.

[55] C. Fleming, S. Morrissey, Y. Cai, J. Yan,  $\gamma\delta$  T Cells: unexpected regulators of cancer development and progression. *Trends Cancer* 3(2017) 561-570.

[56] S. Li, J.J. Li, Zoledronic acid modulates human osteosarcoma cells proliferation via GSK-3 $\beta$  activation. *Neoplasma* 66 (2019) 766-775.

[57] P.S. Mackie, J.L. Fisher, H. Zhou, P.F. Choong, Bisphosphonates regulate cell growth and gene expression in the UMR 106-01 clonal rat osteosarcoma cell line. *Br. J. Cancer.* 84 (2001) 951-958.

[58] J. Sonnemann, V. Eckervogt, B. Truckenbrod, J. Boos, W. Winkelmann, F. van Valen, The bisphosphonate pamidronate is a potent inhibitor of human osteosarcoma cell growth in vitro. *Anticancer Drugs* 12 (2001) 459-465.

- [59] J.P. Farese, J. Ashton, R. Milner, L.L. Ambrose, J. Van Gilder, The effect of the bisphosphonate alendronate on viability of canine osteosarcoma cells in vitro. *In Vitro Cell. Dev. Biol. Anim.* 40 (2004) 113-117.
- [60] N. Horie, H. Murata, Y. Nishigaki, T. Matsui, H. Segawa, M. Nogawa, T. Yuasa, S. Kimura, T. Maekawa, S. Fushiki, T. Kubo, The third-generation bisphosphonates inhibit proliferation of murine osteosarcoma cells with induction of apoptosis. *Cancer Lett.* 238 (2006) 111-118.
- [61] T. Kubo, S. Shimose, T. Matsuo, K. Tanaka, Y. Yasunaga, A. Sakai, M. Ochi, Inhibitory effects of a new bisphosphonate, minodronate, on proliferation and invasion of a variety of malignant bone tumor cells. *J. Orthop. Res.* 24 (2006) 1138-1144.
- [62] R. Inoue, N.A. Matsuki, G. Jing, T. Kanematsu, K. Abe, M. Hirata, The inhibitory effect of alendronate, a nitrogen-containing bisphosphonate on the PI3K-Akt-NF $\kappa$ B pathway in osteosarcoma cells. *Br. J. Pharmacol.* 146 (2005) 633-641.
- [63] M. Muraro, O.M. Mereuta, F. Carraro, E. Madon, F. Fagioli, Osteosarcoma cell line growth inhibition by zoledronate-stimulated effector cells. *Cell Immunol.* 249 (2007) 63-72.
- [64] B. Kubista, K. Trieb, F. Sevelda, C. Toma, F. Arrich, P. Heffeter, L. Elbling, H. Sutterlüty, K. Scotlandi, R. Kotz, M. Micksche, W. Berger, Anticancer effects of zoledronic acid against human osteosarcoma cells. *J. Orthop. Res.* 24 (2006) 1145-52.
- [65] R. Tenta, N. Pitulis, D. Tiblalexi, C. Consoulas, H. Katopodis, E. Konstantinidou, M. Manoussakis, D. Kletsas D, M.N. Alexis, A. Poyatzi, M. Koutsilieris M, Mechanisms of the action of zoledronic acid on human MG-63 osteosarcoma cells. *Horm. Metab. Res.* 40 (2008) 737-745.
- [66] B. Ory, F. Blanchard, S. Battaglia, F. Gouin, F. Rédini, D. Heymann, Zoledronic acid activates the DNA S-phase checkpoint and induces osteosarcoma cell death characterized by apoptosis-inducing factor and endonuclease-G translocation independently of p53 and retinoblastoma status. *Mol. Pharmacol.* 71 (2007) 333-343.
- [67] T. Iguchi, Y. Miyakawa, K. Saito, C. Nakabayashi, M. Nakanishi, H. Saya, Y. Ikeda, M. Kizaki, Zoledronate-induced S phase arrest and apoptosis accompanied by DNA damage and activation of the ATM/Chk1/cdc25 pathway in human osteosarcoma cells. *Int. J. Oncol.* 31 (2007) 285-291.
- [68] A. Evdokiou, A. Labrinidis, S. Bouralexis, S. Hay, D.M. Findlay, Induction of cell death of human osteogenic sarcoma cells by zoledronic acid resembles anoikis. *Bone* 33 (2003) 216-228
- [69] Z.F. Xin, Y.K. Kim, S.T. Jung, Risedronate inhibits human osteosarcoma cell invasion. *J. Exp. Clin. Cancer Res.* 28 (2009) 105.
- [70] Y.Y. Cheng, L. Huang, K.M. Lee, K. Li, S.M. Kumta, Alendronate regulates cell invasion and MMP-2 secretion in human osteosarcoma cell lines. *Pediatr. Blood Cancer* 42 (2004) 410-415.

- [71] P. Heikkilä, O. Teronen, M.Y. Hirn, T. Sorsa, T. Tervahartiala, T. Salo, Y.T. Konttinen, T. Halttunen, M. Moilanen, R. Hanemaaiger, M. Laitinen, Inhibition of matrix metalloproteinase-14 in osteosarcoma cells by clodronate. *J. Surg. Res.* 111 (2003) 45-52.
- [72] H.L. Cheng, C.W. Lin, J.S. Yang, M.J. Hsieh, S.F. Yang, K.H. Lu, Zoledronate blocks geranylgeranylation not farnesylation to suppress human osteosarcoma U2OS cells metastasis by EMT via Rho A activation and FAK-inhibited JNK and p38 pathways. *Oncotarget* 7 (2016) 9742-9758.
- [73] B. Ory, G. Moriceau, V. Trichet, F. Blanchard, M. Berreur, F. Rédini, M. Rogers, D. Heymann, Farnesyl diphosphate synthase is involved in the resistance to zoledronic acid of osteosarcoma cells. *J. Cell. Mol. Med.* 12 (2008) 928-941.
- [74] A Yoshiyama, T. Morii, K. Ohtsuka, H. Ohnishi, T. Tajima, T. Aoyagi, K. Mochizuki, K. Satomi, S. Ichimura, Development of stemness in cancer cell lines resistant to the anticancer effects of zoledronic acid. *Anticancer Res.* 36(2016):625-31.
- [75] H.K. Brown, M. Tellez-Gabriel, D. Heymann, Cancer stem cells in osteosarcoma. *Cancer Lett.* 386 (2017) 189-195.
- [76] T. Morii, K. Ohtsuka, H. Ohnishi, K. Mochizuki, K. Satomi, Inhibition of heat-shock protein 27 expression eliminates drug resistance of osteosarcoma to zoledronic acid. *Anticancer Res.* 30 (2010) 3565-3571.
- [77] N. Horie, H. Murata, S. Kimura, H. Takeshita, T. Sakabe, T. Matsui, T. Maekawa, T. Kubo, S. Fushiki, Combined effects of a third-generation bisphosphonate, zoledronic acid with other anticancer agents against murine osteosarcoma. *Br. J. Cancer* 96 (2007) 255-261.
- [78] M.S. Benassi, A. Chiechi, F. Ponticelli, L. Pazzaglia, G. Gamberi, L. Zanella, M.C. Manara, P. Perego, S. Ferrari, P. Picci, Growth inhibition and sensitization to cisplatin by zoledronic acid in osteosarcoma cells. *Cancer Lett.* 250 (2007) 194-205.
- [79] M.H. Moon, J.K. Jeong, J.S. Seo, J.W. Seol, Y.J. Lee, M. Xue, C.J. Jackson, S.Y. Parks Bisphosphonate enhances TRAIL sensitivity to human osteosarcoma cells via death receptor 5 upregulation. *Exp. Mol. Med.* 43 (2011) 138-145.
- [80] B. Ory, M. Baud'huin, F. Verrecchia, B.B. Royer, T. Quillard, J. Amiaud, S. Battaglia, D. Heymann, F. Redini, F. Lamoureux, Blocking HSP90 addiction inhibits tumor cell proliferation, metastasis development, and synergistically acts with zoledronic acid. *Clin. Cancer Res.* 22 (2016) 2520-2533.
- [81] F. Lamoureux, M. Baud'huin, B. Ory, R. Guiho, A. Zoubeidi, M. Gleave, D. Heymann, F. Rédini, Clusterin inhibition using OGX-011 synergistically enhances zoledronic acid activity in osteosarcoma. *Oncotarget* 5 (2014) 7805-7819.
- [82] C.C. Wu, Y.F. Huang, C.P. Hsieh, P.J. Chueh, Y.L. Chen, Combined use of zoledronic acid augments ursolic acid-induced apoptosis in human osteosarcoma cells through enhanced oxidative stress and autophagy. *Molecules* 21 (2016) pii: E1640.

- [83] G. Moriceau, B. Ory, L. Mitrofan, C. Riganti, F. Blanchard, R. Brion, C. Charrier, S. Battaglia, P. Pilet, M.G. Denis, L.D. Shultz, J. Mönkkönen, F. Rédini, D. Heymann, Zoledronic acid potentiates mTOR inhibition and abolishes the resistance of osteosarcoma cells to RAD001 (Everolimus): pivotal role of the prenylation process. *Cancer Res.* 70 (2010) 10329-31039
- [84] T. Kubo, S. Shimose, T. Matsuo, A. Sakai, M. Ochi. Efficacy of a nitrogen-containing bisphosphonate, minodronate, in conjunction with a p38 mitogen activated protein kinase inhibitor or doxorubicin against malignant bone tumor cells. *Cancer Chemother. Pharmacol.* 62 (2008) 111-116.
- [85] K. Ryu, H. Murata, K. Koto, N. Horie, T. Matsui, Y. Nishigaki, T. Sakabe, H. Takeshita, M. Itoi, S. Kimura, E. Ashihara, T. Maekawa, S. Fushiki, T. Kubo, Combined effects of bisphosphonate and radiation on osteosarcoma cells. *Anticancer Res.* 30 (2010) 2713-2720
- [86] E.H. Kim, M.S. Kim, K.H. Lee, J.S. Koh, W.G. Jung, C.B. Kong, Zoledronic acid is an effective radiosensitizer in the treatment of osteosarcoma. *Oncotarget* 7 (2016) 70869-70880.
- [87] K. Hoddinott, M.L Oblak, G.A. Wood, S. Boston, A.J. Mutsaers, Evaluation of effects of radiation therapy combined with either pamidronate or zoledronate on canine osteosarcoma cells. *Can. J. Vet. Res.* 83 (2019) 3-10.
- [88] P.G. Heymann, T. Ziebart, P.W. Kämmerer, R. Mandic, A. Saydali, A. Braun, A. Neff, G.F. Draenert, The enhancing effect of a laser photochemotherapy with cisplatin or zoledronic acid in primary human osteoblasts and osteosarcoma cells in vitro. *J. Oral Pathol. Med.* 45 (2016) 803-809.
- [89] Z. Zhou, H. Guan, X. Duan, E.S. Kleinerman, Zoledronic acid inhibits primary bone tumor growth in Ewing sarcoma. *Cancer* 104 (2005 ) 1713-1720.
- [90] G.A. Odri, S. Dumoucel, G. Picarda, S. Battaglia, F. Lamoureux, N. Corradini, J. Rousseau, F. Tirode, K. Laud, O. Delattre, F. Gouin, D. Heymann, F. Redini, Zoledronic acid as a new adjuvant therapeutic strategy for Ewing's sarcoma patients. *Cancer Res.* 70 (2010) 7610-7609.
- [91] G. Odri, P.P. Kim, F. Lamoureux, C. Charrier, S. Battaglia, J. Amiaud, D. Heymann, F. Gouin, F. Redini, Zoledronic acid inhibits pulmonary metastasis dissemination in a preclinical model of Ewing's sarcoma via inhibition of cell migration. *BMC Cancer* 14 (2014) 169.
- [92] T. Kubo, S. Shimose, T. Matsuo, A. Sakai, M. Ochi, Efficacy of a nitrogen-containing bisphosphonate, minodronate, in conjunction with a p38 mitogen activated protein kinase inhibitor or doxorubicin against malignant bone tumor cells. *Cancer Chemother. Pharmacol.* 62 (2008) 111-116.
- [93] J. Sonnemann, V. Eckervogt, B. Truckenbrod, J. Boos, W. Winkelmann, F van Valen, The bisphosphonate pamidronate is a potent inhibitor of Ewing's sarcoma cell growth in vitro. *Anticancer Drugs* 14 (2003) 767-771.
- [94] M.P. Dos Santo, C.B. de Farias, R. Roesler, A.L. Brunetto, AL. Abujamra, In vitro antitumor effect of sodium butyrate and zoledronic acid combined with traditional

chemotherapeutic drugs: a paradigm of synergistic molecular targeting in the treatment of Ewing sarcoma. *Oncol. Rep.* 31 (2014) 955-968

[95] S.K. Mueller, B. Altvater, C. Chen, S. Kailayangiri, M. Ahlmann, U. Dirksen, H. Juergens, C. Rossig, Zoledronic acid negatively affects the expansion of in vitro activated human NK cells and their cytolytic interactions with Ewing sarcoma cells. *Oncol. Rep.* 29 (2013) 2348-2354.

[96] F. Gouin, B. Ory, F. Rédini, D. Heymann, Zoledronic acid slows down rat primary chondrosarcoma development, recurrent tumor progression after intralesional curettage and increases overall survival. *Int. J. Cancer* 119 (2006) 980-984.

[97] J.E. Otero, J.W. Stevens, A.E. Malandra, D.C. Fredericks, P.R. Odgren, J.A. Buckwalter, J. Morcuende J., Osteoclast inhibition impairs chondrosarcoma growth and bone destruction. *J. Orthop. Res.* 32 (2014) 1562-1571.

[98] A. Streitbuerger, M. Henrichs, H. Ahrens, C. Lanvers-Kaminzky, F. Gouin, G. Gosheger, J. Hardes, Cytotoxic effect of clodronate and zoledronate on the chondrosarcoma cell lines HTB-94 and CAL-78. *Int. Orthop.* 35 (2011) 1369-1373.

[99] M. Susi, T. Morii, H. Yabe, K. Horiuchi, Y. Toyama, L. Weissbach, F.J. Hornicek, H. Morioka, Alendronate inhibits growth of high-grade chondrosarcoma cells. *Anticancer Res.* 29 (2009) 1879-1888.

[100] T. J. Lai, S.F. Hsu, T.M. Li, H.C. Hsu, J.G. Lin, C.J. Hsu, M.C. Chou, M.C. Lee, S.F. Yang, Y.C. Fong, Alendronate inhibits cell invasion and MMP-2 secretion in human chondrosarcoma cell line. *Acta Pharmacol. Sin.* 28 (2007) 1231-1235

[101] L. Sun, Y. Li, Z. Jiang, J. Zhang, H. Li, B. Li, Z. Ye, V $\gamma$ 9V $\delta$ 2 T cells and zoledronate mediate antitumor activity in an orthotopic mouse model of human chondrosarcoma. *Tumour Biol.* 37 (2016) 7333-7344.

[102] L. Montella, R. Addeo, V. Faiola, G. Cennamo, R. Guerrasi, E. Capasso, R. Mamome, M. Caraglia, S. Del Prete, Zoledronic acid in metastatic chondrosarcoma and advanced sacrum chordoma: two case reports. *J. Clin. Cancer Res.* 28 (2009) 7.

[103] F. Gouin, A.R. Rochwerger, A. Di Marco, P. Rosset, P. Bonnevialle, F. Fiorenza, P. Anract, Adjuvant treatment with zoledronic acid after extensive curettage for giant cell tumours of bone. *Eur. J. Cancer* 50 (2014) 2425-2431.

[104] A. Lipplaa, J.R. Kroep, L. van der Heijden, P.C. Jutte, P.C.W. Hogendoorn, S. Dijkstra, H. Gelberblom, Adjuvant zoledronic acid in high-risk giant cell tumor of bone: a multicenter randomized phase II trial. *Oncologist* 24 (2019) 889

[105] D.C. Kieser, S. Mazas, D.T. Cawley, T. Fujishiro, C. Tavolaro, L. Boissiere, I. Obeid, V. Pointillart, J.M. Vital, O. Gille, Bisphosphonate therapy for spinal aneurysmal bone cysts. *Eur. J. Spine* 27 (2018) 851-858.

[106] P.J. Simm, A. Biggin, M.R. Zacharin, C.P. Rodda, E. Tham, A. Siafarikas, C. Jefferies, P.L. Hofman, D.E. Jensen, H. Woodhead, J. Brown, B.J. Wheeler, D. Brookes, A. Lafferty,

C.F. Munns, on behalf of the APEG Bone Mineral Working Group, Consensus guidelines on the use of bisphosphonate therapy in children and adolescents. *J. Paediatrics Child Health* 54 (2018) 223-233.

[107] D. Liens, P.D. Delmas, P.J. Meunier, Long-term effects of intravenous pamidronate in fibrous dysplasia of bone. *Lancet* 343 (1994) 953-954.

[108] R.D. Chapurlat, P.D. Delmas, D. Liens, P.J. Meunier, Long-term effects of intravenous pamidronate in fibrous dysplasia of bone. *J. Bone Miner. Res.* 12 (1997) 1746-1752.

[109] R.D. Chapurlat, P. Hugueny, P.D. Delmas, P.J. Meunier, Treatment of fibrous dysplasia of bone with intravenous pamidronate: long-term effectiveness and evaluation of predictors of response to treatment. *Bone* 35 (2004) : 235-242.

[110] J.M. Lane, S.N. Khan, W.J. O'Connor, M. Nydick, J.P. Hommen, R. Schneider, E. Tomin, J. Brand, J. Curtin, Bisphosphonate therapy in fibrous dysplasia. *Clin. Orthop. Rel. Res.* 12 (1997) 1314-1315.

[111] M.S. Parisi, M.B. Oliveri, C.A. Mautalen, Bone mineral density response to long-term bisphosphonate therapy in fibrous dysplasia. *J. Clin. Densitom.* 4 (2001) 167-172.

[112] B. Chan, M. Zacharin, Pamidronate treatment of polyostotic fibrous dysplasia: failure to prevent expansion of dysplastic lesions during childhood. *J. Pediatr. Endocrinol. Metab.* 19 (2006) 75-80.

[113] Y. Wang, O. Wang, Y. Jiang, M. Li, W. Xia, X. Meng, X. Xing, Efficacy and safety of bisphosphonate therapy in McCune-Albright syndrome-related to polyostotic fibrous dysplasia : a single-center experience. *Endocr. Pract.* 25 (2019) 23-30.

[114] H.S. Vasiliadis, C. Arnoutoglou, S. Plakoutis, M. Doukas, A. Batistatou, T.A. Xenakis, Low-grade central osteosarcoma of distal femur, resembling fibrous dysplasia Word *J. Orthop.* 4 (2013) 327-332.

[115] Y. Sugiura, H. Kanda, N. Motoi, K. Nomura, K. Inamura, E. Okada, H. Mastumoto, T. Shimoji, S. Matsumoto, J. Nakayama, Y. Takazawa, Y., Ishikawa, R. Machinmai, Osteosarcoma arising in fibrous dysplasia, confirmed by mutational analysis of GNAS gene. *Pathol. Res. Pract.* 214 (2018) 318-324.

[116] A.P. Kansara, J.F. Wan, K.D. Devulapalli, A.E. Horvai, R.J. O'Donnell, T.M. Link, Malignant transformation of an aneurysmal bone cyst to fibroblastic osteosarcoma. *Am. J. Orthop.* 45 (2016) E367-E372

[117] L.A. Mansouri, J.M. Meadeb, J.A. Albert, H. Duval, G. Chalès, N. Hajjaj-Hassouni, P. Guggenbuhl, Bifocal sarcomatous transformation of Paget's disease: a case report. *Joint Bone Spine* 76 (2009) 574-575.

[118] A.T. Deyrup, A.G. Montag, C.Y. InwardsZ. Xu, R.G. Swee, K.K. Unni, Sarcomas arising in Paget disease of bone: a clinicopathologic analysis of 70 cases. *Arch. Pathol. Lab. Med.* 131 (2007) 942-946

- [119] L.T.C. Chow, Giant cell rich osteosarcoma revisited-diagnostic criteria and histopathologic patterns, Ki67, CDK4, and MDM2 expression, changes in response to bisphosphonate and denosumab treatment. *Virchows Arch.* 468 (2016) 741-755.
- [120] N.A. Nadar, N. Margiotta, M. Iafisco, J.J.J.P. van den Beucken, O.C. Boerman, S.C.G. Leeuwenburgh, bisphosphonate-functionalized imaging agents, anti-tumor agents and nanocarriers for treatment of bone cancer. *Adv. Healthc. Mater.* 6 (2017) 8.
- [121] E. David, S. Cagnol, J.Y. Goujon, M. Egorov, J. Taurelle, C. Benestau, L. Morandieu, C. Moal, M. Sicard, S. Pairel, D. Heymann, F. Redini, F. Gouin, R. Le Bot, 12b80 - hydroxybisphosphonate linked doxorubicin: bone targeted strategy for treatment of osteosarcoma. *Bioconjug. Chem.* 30 (2019) 1665-1676.
- [122] S. Rudnick-Glick, E. Corem-Salkmon, I. Grinberg, S. Margel, Targeted drug delivery of near IR fluorescent doxorubicin-conjugated poly(ethylene glycol) bisphosphonate nanoparticles for diagnosis and therapy of primary and metastatic bone cancer in a mouse model. *J. Nanobiotechnology* 14 (2016) 80.
- [123] Q. Yin, L. Tang, K. Cai, R. Tong, R. Sternberg, X. Yang, L.W. Dobrucki, L.B. Borst, D. Kamstock, Z. Song, W.G. Helferich, J. Cheng, T.M. Fan, Pamidronate functionalized nanoconjugates for targeted therapy of focal skeletal malignant osteolysis. *Proc. Natl. Acad. Sci. USA* 113 (2016) E4601-9.
- [124] V.M. Wu, J. Mickens, V. Uskoković, Bisphosphonate-functionalized hydroxyapatite nanoparticles for the delivery of the bromodomain inhibitor JQ1 in the treatment of osteosarcoma. *ACS Appl. Mater. Interfaces* 9 (2017) 25887-25904.
- [125] Y. Xi, T. Jiang, Y. Yu, J. Yu, M. Xue, N. Xu, J. Wen, W. Wan, H. He, Y. Shen, D. Chen, X. Ye, T.J. Webster, Dual targeting curcumin loaded alendronate-hyaluronan-octadecanoic acid micelles for improving osteosarcoma therapy. *Int. J. Nanomedicine* 14 (2019 ) 6425-6437.
- [126] Y. Sun, L. Chen, X. Wu, Q. Ding, Bifunctional bisphosphonate derivatives and platinum complexes with high affinity for bone hydroxyapatite. *Bioorg. Med. Chem. Lett.* 27 (2017) 1070-1075.
- [127] K. Farbod, K. Sariibrahimoglu, A. Curci, A. Hayrapetyan, J.N. Hakvoort, van den Beucken JJ, Iafisco M, Margiotta N, Leeuwenburgh SC. Controlled release of chemotherapeutic platinum-bisphosphonate complexes from injectable calcium phosphate cements. *Tissue Eng. Part A* 22 (2016) 788-800.
- [128] P. Liu, L. Sun, D.S. Zhou, P. Zhang, Y.H. Wang, D. Li, Q.H. Li, R.J. Feng, Development of alendronate-conjugated poly (lactic-co-glycolic acid)-dextrans nanoparticles for active targeting of cisplatin in osteosarcoma. *Sci Rep.* 2015 Dec 1;5:17387.
- [129] L. Yu, L. Cai, H. Hu, Y. Zhang Y, Experiments and synthesis of bone-targeting epirubicin with the water-soluble macromolecular drug delivery systems of oxidized-dextran. *J. Drug Target* 22 (2014) 343-351.

[130] X.N. Yang, J.C. Zeng, Y.C. Song, H. Zhan, F.X. Pei, Targeted antiosteosarcoma methotrexate-bisphosphonate conjugate induces apoptosis of osteosarcoma cells in vitro. Eur. Rev. Med. Pharmacol. Sci. 18 (2014) 2116-23.

[131] G.M. Mekhail, A.O. Kamel, G.A. Awad, N.D. Mortada, R.L. Rodrigo, P.A. Spagnuolo, S.D. Wettig, Synthesis and evaluation of alendronate-modified gelatin biopolymer as a novel osteotropic nanocarrier for gene therapy. Nanomedicine (Lond) 11 (2016) 2251-2273.

[132] L. Xing, F.H. Ebetino, R.K. Boeckman, V. Srinivasan, J. Tao, T.K. Sawyer, J. Li, Z. Yao, B.F. Boyce, Targeting anti-cancer agents to bone using bisphosphonates. Bone, in press (doi:10.1016/j.bone.2020.11549).

## **Figure Legends**

**Figure 1: The vicious cycle established between bone sarcoma cells and bone cells.** Cancer cells secrete pro-resorptive factors such as M-CSF and RANKL. These factors recruit osteoclast precursors and activate osteoclast differentiation and activation. In turn, activated osteoclasts initiate bone resorption and release the cytokines/growth factors trapped in the mineralized extracellular matrix (e.g. TGF- $\beta$ ) and which stimulate cancer cell proliferation, modify their differentiation and facilitate their migration to form distant metastatic foci. Osteoblasts and mesenchymal stem cells also play a part in tumor development by direct (e.g. GAP junctions) and indirect communications (e.g. extracellular vesicles or cytokine secretion). Immune cells (e.g. macrophages) play a part in establishing a permissive local microenvironment.

**Table 1. Drugs/therapies combined with bisphosphonates in osteosarcoma**

BPs	Drug/therapy combined with BPs	Effects	References
Zoledronate	Paclitaxel, Doxorubicin Gemcitabine	Increased sensitivity to paclitaxel, doxorubicine and gemcitabine	[77]
Zoledronate	Cisplatin	Sensitization to cisplatin	[78]
Zoledronate	TRAIL	Enhanced sensitivity to TRAIL	[79]
Zoledronate	PF4942847 (HSP90 inhibitor)	Synergistic inhibitory effect on tumor cell proliferation, metastasis development	[80]
Zoledronate	OGX-11 (clusterin inhibitor)	Synergistic activity	[81]
Zoledronate	Ursolic acid	Increased cell apoptosis	[82]
Zoledronate	RAD001 (Everolimus)	Potentiated mTOR inhibition and abolishes the resistance of osteosarcoma cells to RAD001	[83]
Minodronate	p38 inhibitor Doxorubicin	Increased efficacy of p38 inhibitor and Doxorubicin	[84]
Zoledronate Pamidronate	Radiation	Radiosensitization	[85-87]
Zoledronate	Photochemotherapy	Enhanced effect	[88]