



HAL
open science

Nascent RNA m6A modification at the heart of the gene-retrotransposon conflict

Victor Billon, Gael Cristofari

► **To cite this version:**

Victor Billon, Gael Cristofari. Nascent RNA m6A modification at the heart of the gene-retrotransposon conflict. *Cell Research*, 2021, Online ahead of print. 10.1038/s41422-021-00518-5 . inserm-03259604

HAL Id: inserm-03259604

<https://inserm.hal.science/inserm-03259604v1>

Submitted on 14 Jun 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Nascent RNA m6A methylation at the heart of the gene-retrotransposon conflict

Victor Billon¹ and Gael Cristofari^{1,*}

¹University Cote d'Azur, Inserm, CNRS, Nice, France

*correspondence: Gael.Cristofari@univ-cotedazur.fr

Retrotransposons are highly abundant mobile genetic elements in mammalian genomes and can interfere with many genomic processes, particularly the transcription of genes. A study by Xiong, Wang *et al.* reveals that segments of nascent RNAs containing retrotransposons are enriched in *N*⁶-methyladenosine (m6A) and act as transcriptional roadblocks for elongating RNA polymerase II, but that cellular proteins capable to bind to these RNAs, SAFB/SAFB2, can mitigate this effect.

Long interspersed element 1 (L1) retrotransposons are transposable elements capable of propagating within genomes by a copy-and-paste mechanism through an RNA intermediate and a reverse transcription step. They are present in many eukaryotic lineages, but have been, and still are, particularly active in mammals, acting as potent endogenous mutagens. They are strongly silenced by several layers of transcriptional and post-transcriptional mechanisms, both in the nucleus and in the cytoplasm, that restrict their expression and mobilization to the germ cells, the early embryo, and a very narrow set of adult somatic cells. Nevertheless, some of these elements manage to break free from these locks and insert into new genomic locations, often landing in introns, and occasionally leading to genetic disease¹.

Inserting a large piece of DNA into a gene is potentially more damaging than single nucleotide mutations. A full-length L1 copy is approximately 6kb in length, carries its own promoter sequence, as well as two open reading frames encoding the retrotransposition machinery, and ends with a polyadenylation signal¹. L1s also contain multiple cryptic splice sites, and binding sites for transcription factors and RNA binding proteins. Surprisingly, when insertion occurs into introns, transplanting so many cis-acting regulatory elements does not appear, in most cases, to prevent polymerase II from producing functional transcripts. This can be due in part to purifying selection that eliminates the most detrimental L1 insertions, particularly those in sense orientation relative to genes, or to specific protein RNA-binding complexes that can mask L1 sequences embedded in introns and prevent them to become functional².

In mammals, several DNA and RNA base modifications have been implicated in the regulation of transposable elements or in preventing damaging consequences of their activation. Well-known examples of such modifications include 5-methylcytosine in DNA, an epigenetic mark associated with transcriptional repression³, and adenosine-to-inosine (A-to-I) RNA editing that inhibits the activation of inflammatory pathways that could arise in response to double-stranded RNA structures formed by sense and antisense retrotransposon sequences⁴.

However, the most abundant mRNA modification, *N*⁶-adenosine methylation (m6A), has only recently been detected in RNA from retrotransposons⁵⁻⁷. First, a CRISPR screen performed in mouse embryonic stem cells and designed to identify regulators of retroviral-like retrotransposons, the IAP elements, discovered that the m6A writers METTL3 and METTL14 are key regulators of retrotransposon RNA stability. Notably, they exert opposing effects on different retrotransposon

families: while m6A destabilizes IAP RNA, it appears to stabilize L1 RNA⁵. Furthermore, m6A RNA modifications may regulate the formation or maintenance of retrotransposon-associated heterochromatin - at least at specific retrotransposon loci - possibly through chromatin-associated RNA^{6,8}.

In a new study published in *Cell Research*⁹, Wenbo Li and colleagues explore m6A RNA modifications in nascent transcripts of human cells using a newly developed method, termed MINT-seq, that provides much higher intronic coverage than previous studies. They observe that RNA segments derived from L1 elements and embedded in introns in the sense orientation relative to genes are particularly enriched in m6A modifications. Such m6A-rich domains, called MILs for m6A-rich intronic L1s, are characterized by decreased transcriptional elongation downstream of L1 (**Fig. 1a**). The levels of m6A enrichment in these intronic regions correlate with the extent of transcription block, and deletion or inversion of MILs by genome editing rescues efficient polymerase II progression. Although it is still unclear whether m6A is directly responsible for transcription attenuation, or if other unappreciated characteristics of MILs are involved, a first hint of the underlying mechanisms arose from the systematic analysis of publicly available RNA-protein cross-linking immunoprecipitation (CLIP) data. Several RNA-binding proteins are enriched at MILs, particularly SAFB and SAFB2. The authors demonstrate biochemically that these two poorly characterized proteins can specifically bind L1 RNA in the sense orientation, a property enhanced by m6A modifications. Upon depletion of SAFB/SAFB2, transcription blockade at MILs is exacerbated suggesting that these two factors limit MIL-induced transcriptional attenuation and thereby contribute to fine-tune transcription (**Fig. 1a**).

MINT-seq, as most short-read-based approaches, cannot discriminate transcripts produced from the internal L1 promoter and serving as intermediates of L1 replication, from genic transcripts that may contain L1 sequences in their introns¹⁰. However, looking at carefully chosen L1 loci, known to be autonomously transcribed, and using synthetic reporter constructs, the authors show that m6A is deposited both on passenger L1 sequences embedded in intronic nascent RNAs and on L1s transcribed from their own promoters. They could also address the effect of m6A methylation and its readers on L1 RNA stability and retrotransposition. m6A methylation stabilizes L1 RNA and increases L1 mobility, a process promoted by the m6A reader YTHDC1, and antagonized by SAFB/SAFB2 (**Fig. 1b**). In agreement with these results, another recent study concluded that m6A positively regulates L1 expression and retrotransposition, and further identified a cytoplasmic effector of this pathway, eIF3, which stimulates L1 RNA translation⁷.

Xiong, Wang *et al.* have uncovered the role of SAFB/SAFB2 proteins as peacekeepers that safeguard gene function against potent endogenous mutagens. Not only do these proteins limit the ability of L1 to propagate, but also once new L1 insertions occur, they alleviate L1-mediated transcription attenuation. Future work will decipher the full set of m6A writers, readers and erasers, and how they stand in the gene-retrotransposon conflict (**Fig. 1b inset**).

Acknowledgements

Work in the laboratory of GC is supported by the French government, through the Agence Nationale de la Recherche (Idex UCAJEDI, ANR-15-IDEX-01; Labex SIGNALIFE, ANR-11-LABX-0028-01; RetroMet, ANR-16-CE12-0020; ImpacTE, ANR-19-CE12-0032), the Fondation pour la Recherche Médicale (FRM, DEQ20180339170), Inserm (GOLD cross-cutting programme on genomic variability), CNRS (GDR 3546), and the University Hospital Federation (FHU) OncoAge.

Competing interest

GC is an unpaid associate editor of the journal Mobile DNA (Springer-Nature) and an unpaid review editor of the Genetics, Genomics and Epigenomics of Aging section within the Frontiers in Aging journal (Frontiers).

References

1. Kazazian, H. H. & Moran, J. V. Mobile DNA in Health and Disease. *N. Engl. J. Med.* **377**, 361–370 (2017).
2. Attig, J. *et al.* Heteromeric RNP Assembly at LINEs Controls Lineage-Specific RNA Processing. *Cell* **174**, 1067–1081.e17 (2018).
3. Deniz, Ö., Frost, J. M. & Branco, M. R. Regulation of transposable elements by DNA modifications. *Nat. Rev. Genet.* **20**, 417–431 (2019).
4. Eisenberg, E. & Levanon, E. Y. A-to-I RNA editing — immune protector and transcriptome diversifier. *Nat. Rev. Genet.* **19**, 473–490 (2018).
5. Chelmicki, T. *et al.* m6A RNA methylation regulates the fate of endogenous retroviruses. *Nature* **591**, 312–316 (2021).
6. Liu, J. *et al.* The RNA m6A reader YTHDC1 silences retrotransposons and guards ES cell identity. *Nature* **591**, 322–326 (2021).
7. Hwang, S.-Y. *et al.* L1 retrotransposons exploit RNA m6A modification as an evolutionary driving force. *Nat. Commun.* **12**, 880 (2021).
8. Xu, W. *et al.* METTL3 regulates heterochromatin in mouse embryonic stem cells. *Nature* **591**, 317–321 (2021).
9. Xiong, F. *et al.* RNA m6A Modification Orchestrates a LINE-1/Host Interaction That 2 Facilitates Retrotransposition and Contributes to Long Gene Vulnerability. *Cell Res.* **In Press**, (2021).
10. Lanciano, S. & Cristofari, G. Measuring and interpreting transposable element expression. *Nat. Rev. Genet.* **21**, 721–736 (2020).

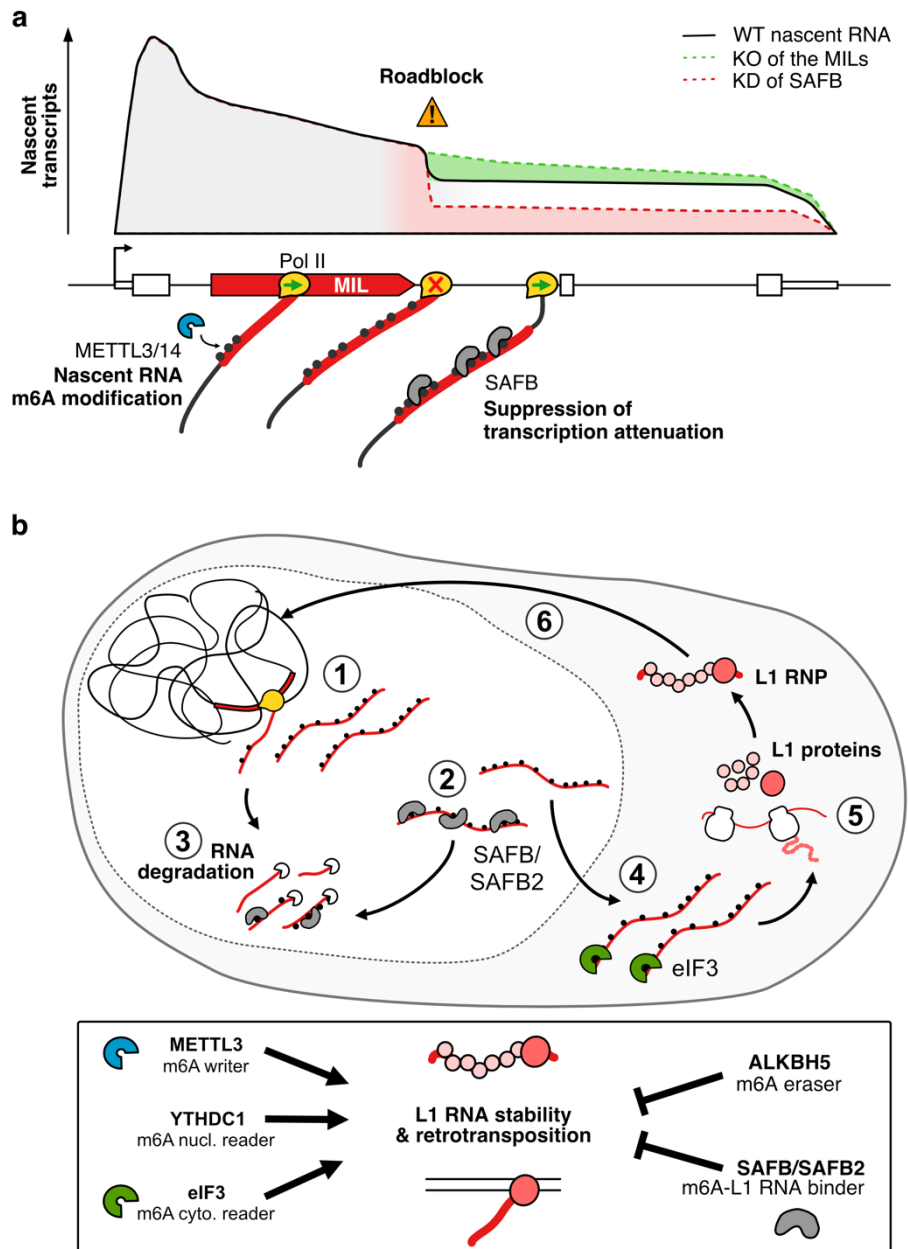


Fig. 1: A dual role for SAFB/SAFB2 in protecting the genome against L1 retrotransposons. a SAFB limits transcriptional attenuation caused by intronic L1s. RNA polymerase II (Pol II) transcribes a gene containing a sense-oriented intronic L1 (red). The nascent RNA segment containing the L1 sequence is m6A methylated (black circles) by METTL3/14 (blue) and bound by SAFB/SAFB2 (grey). The intronic L1 causes a transcription roadblock (dark line). Deletion (KO) or inversion of the L1 sequence prevents attenuation while depletion (KD) of SAFB increases it (dashed lines). **b** A model for the influence of m6A on L1 mobilization. When transcribed from its own promoter, L1 RNA can also be *N*⁶A-methylated by METTL3/14 (1). SAFB/SAFB2 proteins preferentially bind m6A-modified L1 RNA (2). Unmodified L1 RNAs and SAFB-bound L1 RNAs are destabilized (3). In the cytoplasm, m6A-modified L1 RNA can hijack the translation initiation factor eIF3 (green), enhancing L1 translation (4). L1 ribonucleoprotein particles (RNPs) formed by L1 proteins and RNA (5) drive reverse transcription and genomic integration (6). Inset, m6A pathway effectors regulating L1 mobilization.