



HAL
open science

Identification of potential biomarkers for predicting the early onset of diabetic cardiomyopathy in a mouse model

R. Johnson, X. Nxele, M. Cour, N. Sangweni, T. Jooste, N. Hadebe, E. Samodien, M. Benjeddou, M. Mazino, J. Louw, et al.

► To cite this version:

R. Johnson, X. Nxele, M. Cour, N. Sangweni, T. Jooste, et al.. Identification of potential biomarkers for predicting the early onset of diabetic cardiomyopathy in a mouse model. *Scientific Reports*, 2020, 10 (1), pp.12352. 10.1038/s41598-020-69254-x . inserm-03231675

HAL Id: inserm-03231675

<https://inserm.hal.science/inserm-03231675>

Submitted on 21 May 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



OPEN

Identification of potential biomarkers for predicting the early onset of diabetic cardiomyopathy in a mouse model

Rabia Johnson^{1,2}✉, Xolisa Nxele^{1,3}, Martin Cour^{4,5}, Nonhlakanipho Sangweni^{1,2}, Tracey Jooste^{1,2}, Nkanyiso Hadebe^{5,6}, Ebrahim Samodien¹, Mongi Benjeddu³, Mikateko Mazino⁷, Johan Louw^{1,8} & Sandrine Lecour⁵

Type 2 diabetes (T2D) is characterized by metabolic derangements that cause a shift in substrate preference, inducing cardiac interstitial fibrosis. Interstitial fibrosis plays a key role in aggravating left ventricular diastolic dysfunction (LVDD), which has previously been associated with the asymptomatic onset of heart failure. The latter is responsible for 80% of deaths among diabetic patients and has been termed diabetic cardiomyopathy (DCM). Through in silico prediction and subsequent detection in a leptin receptor-deficient db/db mice model (db/db), we confirmed the presence of previously identified potential biomarkers to detect the early onset of DCM. Differential expression of Lysyl Oxidase Like 2 (*LOXL2*) and Electron Transfer Flavoprotein Beta Subunit (*ETFβ*), in both serum and heart tissue of 6–16-week-old db/db mice, correlated with a reduced left-ventricular diastolic dysfunction as assessed by high-resolution Doppler echocardiography. Principal component analysis of the combined biomarkers, *LOXL2* and *ETFβ*, further displayed a significant difference between wild type and db/db mice from as early as 9 weeks of age. Knockdown in H9c2 cells, utilising siRNA of either *LOXL2* or *ETFβ*, revealed a decrease in the expression of Collagen Type I Alpha1 (*COL1A1*), a marker known to contribute to enhanced myocardial fibrosis. Additionally, receiver-operating curve (ROC) analysis of the proposed diagnostic profile showed that the combination of *LOXL2* and *ETFβ* resulted in an area under the curve (AUC) of 0.813, with a cut-off point of 0.824, thus suggesting the favorable positive predictive power of the model and further supporting the use of *LOXL2* and *ETFβ* as possible early predictive DCM biomarkers.

Diabetes affects 463 million people worldwide and this number is said to increase to 700 million by 2045¹. Diabetes and its complications are inextricably linked to cardiovascular dysfunction, which is currently the leading cause of mortality worldwide, affecting 17.9 million individuals². This association was first reported in the Framingham Heart study and since then numerous reports have come to the forefront, confirming a 2–4 times increased susceptibility of diabetic individuals to heart failure (HF)^{3,4}. Coronary artery disease (CAD) is the major type of CVD responsible for HF in diabetic individuals, however diabetic cardiomyopathy (DCM) is an established complication of diabetes mellitus (DM) existing in the absence of CAD or hypertension⁵. Furthermore, DCM is referred to as the silent killer, due to the manifestation of a long subclinical period in which the disease exists with no overt clinical symptoms^{6–8}.

¹Biomedical Research and Innovation Platform (BRIP), South African Medical Research Council (SAMRC), Tygerberg 7505, South Africa. ²Division of Medical Physiology, Faculty of Health Sciences, Stellenbosch University, Tygerberg, South Africa. ³Department of Biotechnology, University of Western Cape, Cape Town, South Africa. ⁴Hospices Civils de Lyon, Hôpital Edouard Herriot, Service de Médecine Intensive-Réanimation, Lyon Cedex 03, France. ⁵Hatter Institute for Cardiovascular Research in Africa (HICRA), Faculty of Health Sciences, University of Cape Town, Observatory, South Africa. ⁶Department of Anaesthesia, Faculty of Health Sciences, University of Cape Town, Observatory, South Africa. ⁷Biostatistics Research Unit, South African Medical Research Council (SAMRC), Cape Town, South Africa. ⁸Department of Biochemistry and Microbiology, University of Zululand, KwaDlangezwa 3886, Richards Bay, South Africa. ✉email: rabia.johnson@mrc.ac.za

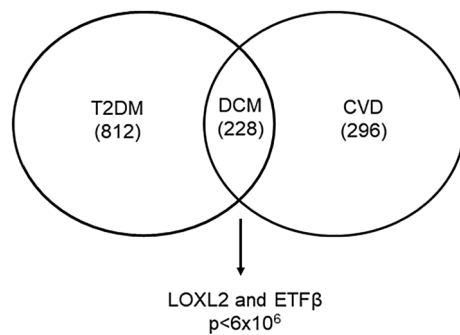


Figure 1. Venn diagram of differentially expressed genes (DEGs). Within the type 2 diabetes mellitus (T2DM) and cardiovascular disease (CVD) datasets, 812 and 296 candidate genes were identified respectively. Integration of the T2DM and CVD datasets, a diabetic cardiomyopathy (DCM) dataset was generated, in which 2 possible candidate genes were identified based on their Wilcoxon score (p value $< 6 \times 10^{-6}$).

DCM proceeds in three different stages: asymptomatic, diastolic and systolic stages, respectively. In its asymptomatic stage, the disease is characterized by abnormal left ventricular relaxation, referred to as left ventricular dysfunction (LVD). These functional abnormalities can be seen in 40–60% of asymptomatic diabetic cases using Doppler echocardiography, Tissue Doppler Imaging (TDI) echocardiography analysis or Speckle Track Imaging (STI) echocardiography⁹. Also, diabetic patients with subclinical diastolic dysfunction (DD) have a 5-year mortality rate of 30% compared to the 12.1% for non-diabetic patients with no diastolic dysfunction¹⁰. As DCM enters its later stage, it progresses from DD to overt heart failure with preserved ejection fraction, which has no proven effective treatment nor a biomarker available that can detect the early onset of DCM⁸. TDI- and STI echocardiograph analysis are the only imaging technologies available to detect DCM in its asymptomatic stage. However, these techniques are not always available, especially in developing countries. Current diagnostic markers include N-terminal pro b-type brain natriuretic peptide (NT-proBNP) and cardiac troponin T (cTn-T). However, none of these markers are able to detect DCM, either in its asymptomatic stage or early enough to have a therapeutic impact.

As such, there is a need for a prognostic marker that can improve early detection of DCM prior to the onset of irreversible complications. However, despite numerous clinical and pharmacological intervention studies, DCM remains elusive warranting new strategies to identify asymptomatic detection of the disease state. In this study, using an *in silico* predictive method the combination use of two possible biomarkers was identified, Lysyl Oxidase Like 2 (LOXL2) and Electron Transfer Flavoprotein Beta Subunit (ETFβ). The presence of the identified markers was then screened in the serum of db/db mice with confirmed LVDD.

Results

DCM biomarker prediction. Meta-analysis of publicly available microarray data sets using ArrayExpress resulted in approximately 15,485 and 16,450 common Ensemble IDs for T2DM and CVD, respectively. Upon statistical differential expression analysis, using the QD binning approach, 812 and 296 differential expressed genes (DEGS) were identified for T2DM and CVD, respectively. Thereafter, using a similar quantile discretization (QD) analysis, T2DM and CVD genes were integrated, and 228 common genes were identified. Due to limited serum availability in the animal study and to prevent bias, the top 2 genes were selected according to their Wilcoxon score (p value $< 6 \times 10^{-6}$), namely *LOXL2* and *ETFβ*, for further downstream analysis (Fig. 1).

Morphometric analysis of risk factors associated with DCM. To assess the presence of cardiac risk factors in our db/db model, fasting blood glucose levels (FBG) and serum lipids levels were investigated (Table 1). In this study, diabetic (db/db) mice presented with significantly increased body weight/tibia length at age 6, 9, 11 and 16 weeks when compared to the non-diabetic (db/+) control. There were no differences in heart weight/tibia length (data not shown), as well as the heart rate between db/db mice and aged-matched controls ($p > 0.05$). Fasting blood glucose levels, cholesterol, triglycerides and low-density lipoproteins (LDL) of db/db mice were significantly elevated from 9 weeks onwards when compared to their age-matched controls. The observed effects are characteristic of metabolic syndrome that continued throughout the study.

mRNA expression analysis of *LOXL2* and *ETFβ* in cardiac tissue. The expression of the two candidate biomarkers was verified in cardiac tissue of leptin-receptor-deficient mice. The results obtained showed a marked increase in *LOXL2* mRNA expression at 11-weeks (sevenfold, $p < 0.001$) when comparing db/db mice to the db/+ control. This expression difference was maintained at both 14-weeks (fourfold, $p < 0.1$) and 16-weeks (sixfold, $p < 0.01$) (Fig. 2A). *ETFβ* mRNA expression, on the other hand, tends to increase until 11-weeks (1.8-fold) when comparing db/db mice to db/+ control, after which a tendency to decrease in expression was observed at 14- (1.2-fold) and 16-weeks (1.3-fold), (Fig. 2B).

LOXL2, *ETFβ* and proBNP protein are expressed in the serum of obese mice. To explore the effect of *LOXL2* and *ETFβ* as a prognostic marker, we tested if the identified proteins are released from the car-

	Wild type (db/+)					Obese (db/db)				
	6 weeks	9 weeks	11 weeks	14 weeks	16 weeks	6 weeks	9 weeks	11 weeks	14 weeks	16 weeks
BW (g)/tibia length (mm)	1.21 ± 0.03	1.28 ± 0.06	1.34 ± 0.03	1.38 ± 0.02	1.33 ± 0.03	1.68 ± 0.06***	1.96 ± 0.03***	2.11 ± 0.07***	1.80 ± 0.14***	1.69 ± 0.11**
FBG (mmol/L)	8.33 ± 0.58	7.09 ± 0.29	8.91 ± 5.15	6.94 ± 0.71	10.46 ± 8.07	12.50 ± 0.52	22.74 ± 2.76***	28.30 ± 1.88***	29.50 ± 1.44***	27.33 ± 9.20***
Cholesterol (mol/L)	1.90 ± 0.07	2.37 ± 0.41	1.75 ± 0.03	1.85 ± 0.05	2.12 ± 0.17	3.90 ± 0.21***	3.45 ± 0.59*	3.79 ± 0.24**	3.90 ± 0.34**	3.77 ± 0.20***
LDL (mmol/L)	0.30 ± 0.00	0.30 ± 0.00	n/a	0.30 ± 0.00	0.30 ± 0.00	0.55 ± 0.05**	0.37 ± 0.08	n/a	0.50 ± 0.15	0.53 ± 0.10**
Triglycerides (mmol/L)	0.84 ± 0.14	0.77 ± 0.08	n/a	1.10 ± 0.13	0.78 ± 0.17	2.43 ± 0.22**	2.10 ± 0.56	n/a	3.11 ± 0.36	1.86 ± 0.65**

Table 1. Risk markers of cardiovascular dysfunction in db/db mice compared to their wild type control (db/+). BW body weight, LDL low-density lipoprotein, FBG fasting blood glucose levels. * $p \leq 0.015$, ** $p < 0.01$ *** $p \leq 0.001$ versus wild type control (db/+) at the same time point.

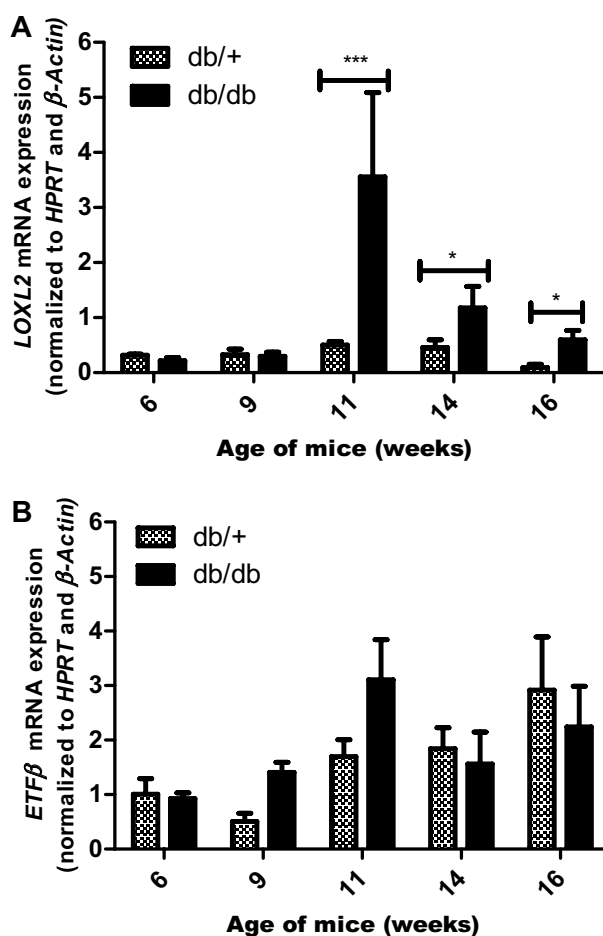


Figure 2. mRNA expression of LOXL2 and ETF β in hearts tissue of 6–16-week-old db/db mice. Differential mRNA expression of (A) Lysyl Oxidase Like 2 (LOXL2) and (B) Electron transfer flavoprotein subunit beta (ETF β) in heart tissue of leptin receptor-deficient mice (db/db) compared to aged matched lean control (db/+). Results are expressed as mean \pm SEM of $n = 4$ –8 animals per group. * $p < 0.05$ and *** $p \leq 0.001$ compared to wild type control (db/+).

diac interstitial space into the circulation. Results obtained showed that LOXL2 levels were significantly elevated in the serum of obese animals at 9- ($p < 0.01$), and 11-weeks ($p < 0.01$) when compared to age-matched controls, whereas serum levels at 6-, 14- and 16-weeks showed an increasing trend ($p > 0.05$) (Fig. 3A). Next serum protein levels of ETF β were investigated and data obtained showed that protein levels of db/db mice showed no change at 6 and 9 weeks ($p > 0.05$) and were significantly reduced at 11- ($p < 0.01$), 14- ($p < 0.001$) and 16-weeks ($p < 0.05$) when compared to the control (Fig. 3B). Interestingly, measurements of serum NT-pro-BNP (the known HF biomarker) was unchanged between 6 and 16 weeks. ($p > 0.05$) (Fig. 3C).

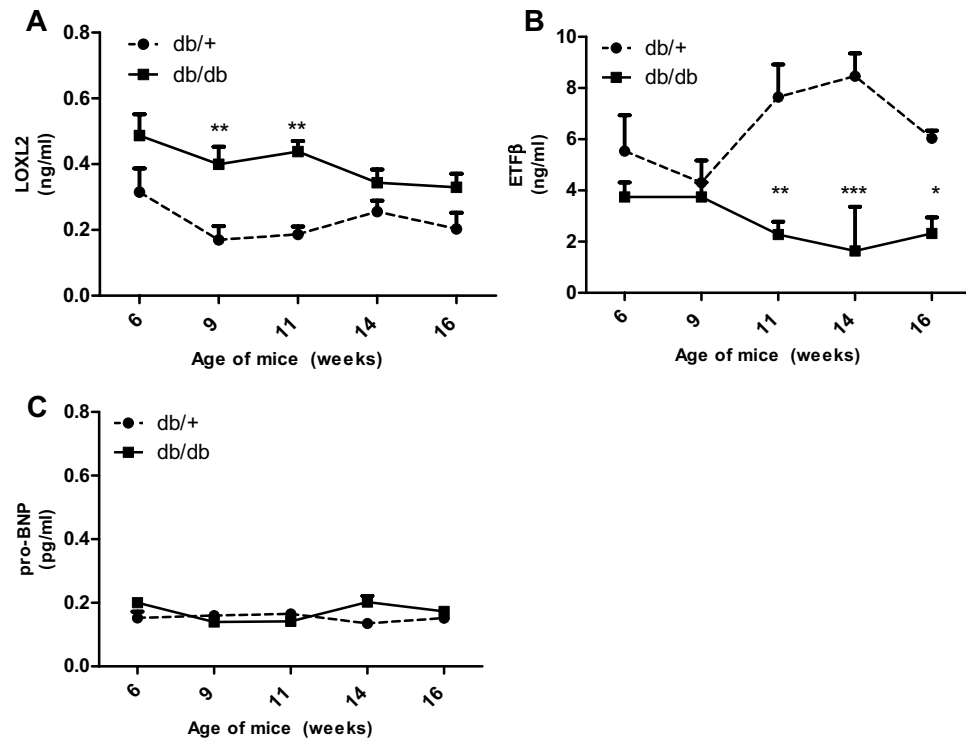


Figure 3. Expression of serum LOXL2 and ETFβ in 6–16-week-old Leptin receptor-deficient db/db mice and aged matched controls. (A) Lysyl Oxidase Like 2 (LOXL2), (B) Electron transfer flavoprotein subunit beta (ETFβ) and (C) N-terminal pro b-type natriuretic peptide protein expression in serum of leptin receptor-deficient mice (db/db) compared to aged matched lean control (db/+). Results are expressed as mean ± SEM of n = 8 animals per group. * $p < 0.05$, ** $p < 0.01$ and *** $p < 0.001$ compared to wild type control (db/+) at the same point.

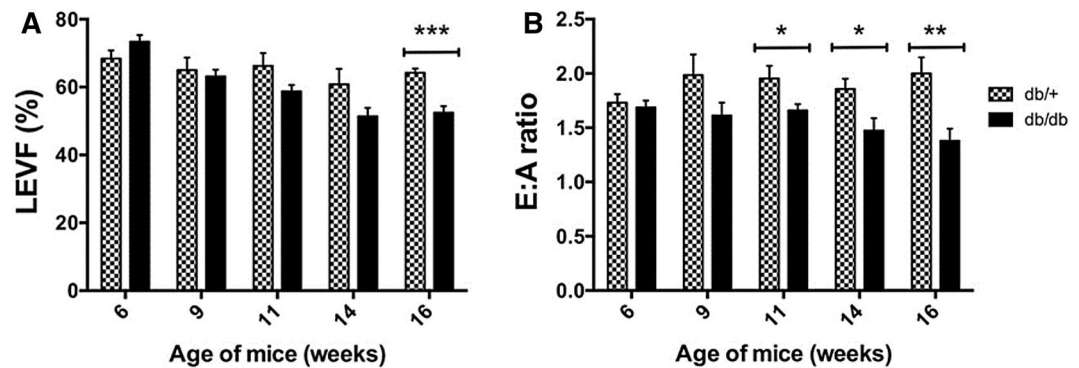


Figure 4. High-resolution Doppler echocardiography analysis in 6–16-week-old Leptin receptor-deficient db/db mice and their age matched db/+ heterozygous non-diabetic control group. (A) Measurement of left ventricular ejection fraction (LVEF) as a marker of systolic function; (B) Measurement of mitral E:A ratio as a marker of left ventricular diastolic function. Results are expressed as mean ± SEM of 6–8 animals per group. Statistical significance was evaluated using Mann–Whitney test. * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$ compared to aged matched wild type control.

High resolution Doppler echocardiography analysis as the gold standard to detect diabetic cardiomyopathy.

High-resolution Doppler echocardiography analysis was used to record structural and functional measurements of left ventricular remodeling in db/db and control mice as summarized in Fig. 4 and Supplementary Table S1. No significant difference in left ventricular internal diameter, left ventricular posterior wall thickness-end diastole and left ventricular mass at 6, 9, 11, 14 and 16 weeks of age were observed in db/db compared to db/+ control ($p > 0.05$). As shown in Fig. 4, significant impairment in LV systolic dysfunction was observed at 16 weeks in db/db mice while diastolic dysfunction, as assessed by a decrease in mitral E:A ratio, was diagnosed as early as 11 weeks (Fig. 4). Diastolic dysfunction was also confirmed by a significant increase in E

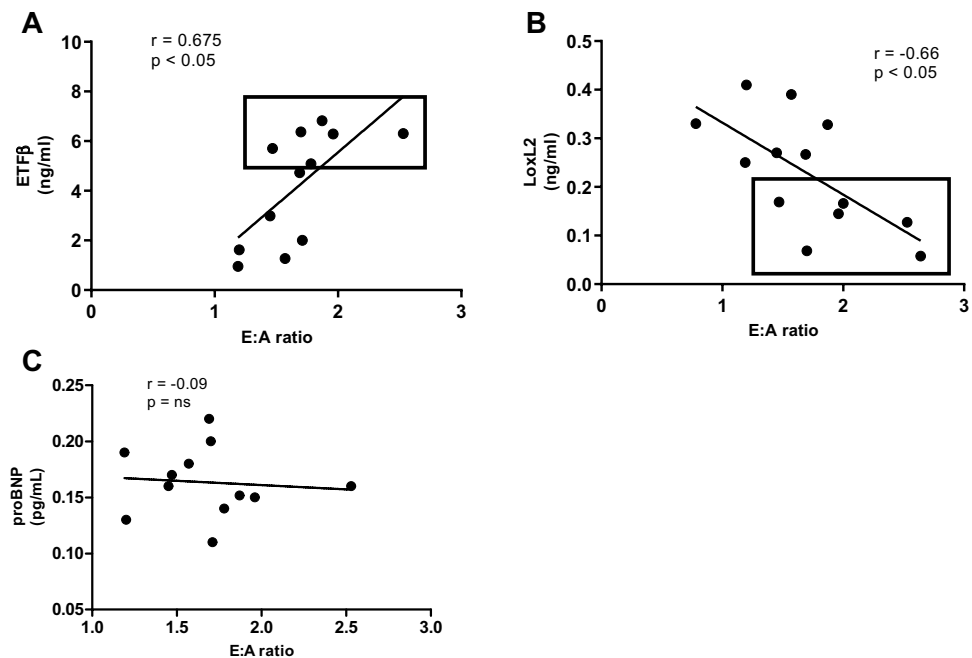


Figure 5. Correlation analysis of serum LOXL2, ETF β and proBNP with mitral E:A ratio in Leptin receptor-deficient db/db mice at 16 weeks. Mitral E:A ratio is a marker of left ventricular diastolic dysfunction, (A) Lysyl Oxidase Like 2 (LOXL2), was negatively and (B) Electron transfer flavoprotein subunit beta (ETF β) was positively correlated with mitral E:A ratio, while no correlation was observed with (C) N-terminal pro b-type natriuretic peptide (proBNP).

wave deceleration time (E_{DT}) and in isovolumic relaxation time (IVRT) in db/db compared to db/+ at both 14 and 16 weeks (Supplementary Table S1).

Serum LOXL2, ETF β and NT-proBNP correlation analysis with mitral E:A ratio. In clinical practice the mitral E:A ratio is currently used as an indication of the degree of HF, guiding the implementation of interventions. Therefore, we examined the correlation between serum levels of the two candidate biomarkers (LOXL2, ETF β) as well as NT-proBNP, using the Pearson's correlation test, and observed no linear relationship between serum levels of NT-proBNP and the mitral E:A ratio ($r = 0.05$) (Fig. 5C). However, a strong (downhill) negative linear relationship was observed between serum LOXL2 levels and the E:A ratio ($r = -0.65$, $p = 0.01$), while a positive correlation was observed between ETF β and the E:A ratio (Pearson correlation, $r = 0.65$, $p < 0.001$) (Fig. 5A,B). Based on the correlation analysis, we suggest that LOXL2 and ETF β could serve as potential biomarkers for the diagnosis of DCM.

siRNA analysis of LOXL2 and ETF β . In order to assess the functional role LOXL2 and ETF β may play in collagen crosslinking, H9c2 cells treated with siRNA were exposed to high glucose (HG) and palmitate (PAL). Results obtained showed that HG and PAL significantly increased scr_LOXL2, while decreasing scr ETF β when compared to the untreated control ($p < 0.001$) (Fig. 6A,C). Alternatively, cells treated with either si_LOXL2 or si ETF β showed significantly decreased mRNA expression when compared to the control ($p < 0.001$ and $p < 0.01$, respectively) (Fig. 6A,C). Subsequently, the role of LOXL2 and ETF β on the expression on COL1A was assessed. Results obtained showed that H9c2 cells transfected with the si_LOXL2, and post-treated with HG and PAL, resulted in a significant decrease in COL1A mRNA expression ($p < 0.01$) when compared to the scrambled (Fig. 6B). Similarly, H9c2 cells transfected with si ETF β showed significantly decreased COL1A mRNA expression (Fig. 6D).

Determination of sensitivity and specificity of LOXL2 and ETF β as a predictive biomarker. In order to evaluate the diagnostic ability of the two candidate biomarkers (LOXL2 and ETF β) and its possible use in a clinical setting, a generalised linear model was used to test for significant differences ($p < 0.001$) between db/+ and db/db mice at each week and compared to that of NT-proBNP. The results obtained showed no significant differences for NT-proBNP and LOXL2 at all the evaluated time points. Conversely, ETF β showed significant differences at week 11 and 14 ($p < 0.01$) (Table 2). Subsequently, the results for the three biomarkers (NT-proBNP, LOXL2 and ETF β) were combined to perform principal component analysis (PCA). Results showed that the first principal component accounted for almost 89% of the variation in the data, which was the linear combination of the three biomarkers (data not shown). To conduct ROC curve analysis the mice groups were either specified (db/+ or db/db) or unspecified. Receiver Operating Characteristic (ROC) curves, for NT-proBNP and LOXL2

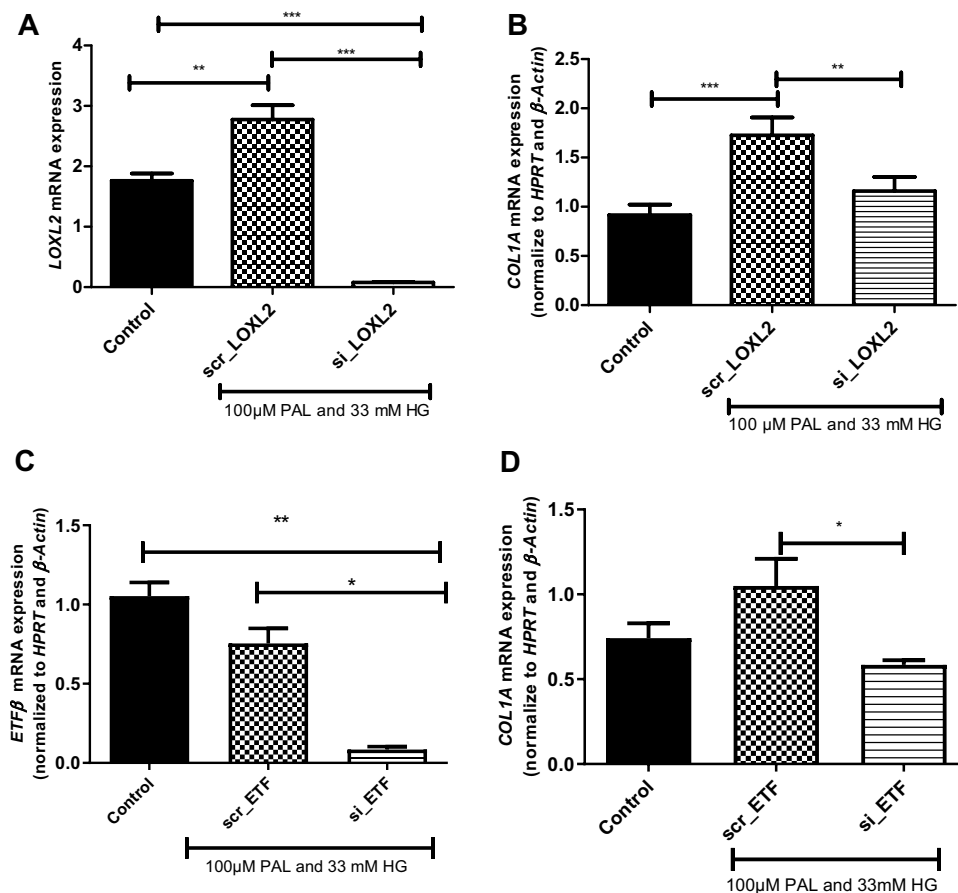


Figure 6. Effect of LOXL2 knockdown on *Col1A* and *ETFβ* expression. The degree of knockdown after H9c2 cardiomyocytes transfected with (A) *siLOXL2* and (C) *siETFβ* was exposed to 100 μ M Palmitate (PAL) and 33 mM glucose (HG). The relative mRNA expression of the fibrosis gene, *COL1A* in (B) *LOXL2* and (D) *ETFβ* after transfection of cells with either small interfering RNA (siRNA) or scrambled RNA (scrRNA). The expression levels were normalized relative to the control. Statistical significance was evaluated using the one-way ANOVA with a Tukey Post-hoc t-test. This graph represents the SEM of triplicate samples. $p < 0.05$, $p < 0.01$ and $p < 0.001$ when compared to control or scrambled control.

	Mice age (weeks)	t value	p value
NT-proBNP	6	-2.05	0.096
	9	1.33	0.23
	11	1.85	0.1
	14	-1.05	0.34
	16	-1.22	0.25
ETFβ	6	1.94	0.09
	9	0.59	0.57
	11	3.91	0.001*
	14	3.67	0.001*
	16	3.08	0.022
LOXL2	6	0.11	0.92
	9	-2.39	0.04
	11	-4.29	0.09
	14	0.23	0.82
	16	-1.98	0.08

Table 2. Statistical analysis for all biomarkers. *ETFβ, at 11 and 14 weeks have a significant difference ($p = 0.01$).

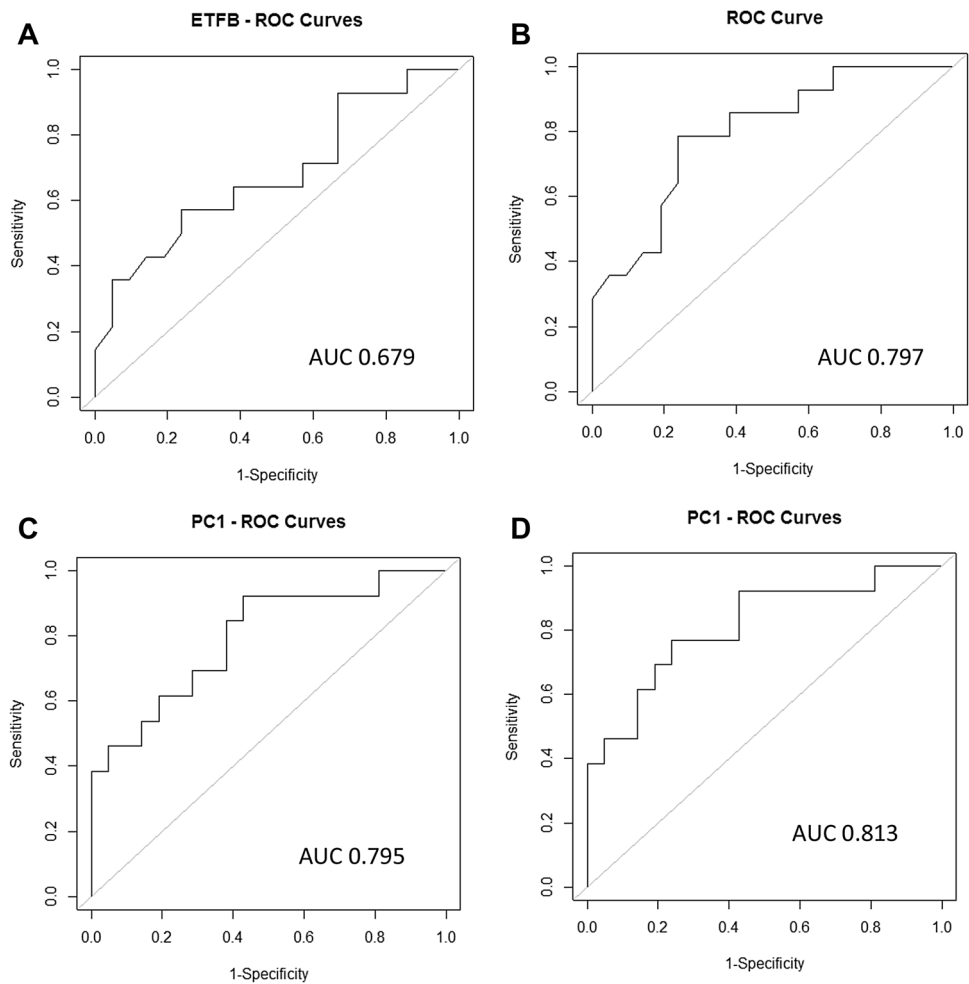


Figure 7. Receiver operating characteristics (ROC) of LOXL2 and ETF β between 9 and 14 weeks. ROC curve analysis of ETF β with (A) groups not specified and (B) groups specified. (C) Linear combination of LOXL2 and ETF β when groups are not specified and when (D) groups are specified a good predictive cut-off point of 0.824 was obtained. Logistic regression was used to perform ROC, $p < 0.05$.

generated area under the curve (AUC) values that remained non-significant in both specified and unspecified groups (data not shown). However, results obtained for ETF β demonstrated an AUC of 0.679 when the groups were not specified (Fig. 7A), which increased to 0.797 when they were specified (Fig. 7B). These results suggested that ETF β may be sensitive and specific enough to detect DCM.

Combination use of LOXL2 and ETF β . Since there was a significant difference in the expression of both biomarkers, LOXL2 and ETF β , between mice at 9 weeks to 14 weeks, a principal component analysis was performed by combining the two datasets to further improve their diagnostic ability. This linear combination was also used to generate ROC curves. Data presented for the linear combination of LOXL2 and ETF β showed an AUC of 0.795 with a cutoff point of 0.789 when the groups were not specified (Fig. 7C). However, the AUC was significantly increased to 0.813 when the groups were specified, with a cutoff value of 0.824 (Fig. 7D). The latter demonstrated a good predictive power of the model, thus implying that, compared with ETF β alone, the combination of the ETF β and LOXL2 as a biomarker is more sensitive and potentially has a higher diagnostic value.

Discussion

Currently, there is no single prognostic test that is sensitive enough to detect the pre-clinical onset of LVDD^{11,12}. Although brain natriuretic peptide (BNP) and NT-pro-BNP are well-established serum markers used to detect cardiac abnormalities, they are unable to predict the early onset of DCM^{12,13}. This was confirmed in the current study. Furthermore, TDI echocardiography, Phase-Magnetic Resonance Imaging and 2D/3D-Speckle Tracking echocardiography are all non-invasive diagnostic methods with the ability to diagnose and stratify LVDD in diabetic patients. However, these tests are expensive and not done routinely as they are specialized equipment that requires trained personal which is not available in all healthcare facilities. Recently, sodium-glucose linked transporter (SGLT2) inhibitors have been recognized as a new class of drugs that can protect the diabetic

myocardium. However, clinicians are currently faced with the dilemma of when to start treating diabetic individuals with SGLT2 inhibitors. Thus, a routine biomarker would greatly aid the confirmation of the presence of cardiac dysfunction and would allow an opportunity for the implementation of corrective therapy.

In the current study, we confirmed the occurrence of two potential serum biomarkers, LOXL2 and ETF β , to detect early onset of DCM. The presence of serum LOXL2 was first reported by Yang followed by Zhao who argued that augmented LOXL2 enhances collagen crosslinking that consequently increases cardiac fibrosis^{14,15}. Pertaining to ETF β , Lu reported that decreased ETF β expression was implicated in mitochondrial shift in substrate preference^{16,17}. Also, in a review conducted by Lorenzo and colleagues, the mentioned disease pathophysiology came under investigation and the authors concluded that both processes (augmented fibrosis and mitochondrial dysfunction) play a major role in the development of diabetes-induced HF^{11,18}. However, none of the mentioned studies aimed to investigate the combined use of the identified serum biomarkers. As such, our current study is the first to investigate the use of LOXL2 and ETF β as a combination biomarker for early detection of DCM.

The db/db mouse model is a well-established heart failure animal model in which mice develop diabetes by 8 weeks of age^{19,20} with increased LDL, cholesterol and triglycerides observed by week 9^{21,22}. We observed that by 6 weeks, diabetic mice weighed significantly more and presented with increased FBG, triglycerides, LDL and cholesterol levels. The latter are known features of DCM and were confirmed in a study conducted by Dludla, where the authors showed that increased FBG and lipid profiles are key risk predictors of cardiac fibrosis and subsequent HF²². Furthermore, it has been argued by Fang that myocardial functional changes do not progress in parallel with histological changes⁸. Instead, diabetic individuals have a long latent phase in which functional abnormalities, induced by the hyperglycaemic state, precede the onset of any structural changes. This was confirmed by Liu who argued that DCM has a long latent phase in which subtle structural modifications occur in the absence of symptomatic onset of the disease²³. The current study confirmed this with echocardiography changes confirming the presence of reduced ejection fraction whilst subtle changes in fibrosis was observed at 16 weeks (Supplementary Data, Figure S1). However, changes in LOXL2 were concomitant with echocardiography data. This was confirmed by Alex who further argued that the db/db mouse model in the C57BL6J background is a pathophysiological relevant model to study diabetes-induced cardiac dysfunction as it displays a marked increase in cardiac risk predictors, however, cardiac fibrosis using this specific model are only detected at 6 months only (24 weeks)²⁴. Taken together, these results confirm that this model displays a T2DM phenotype, whilst high cholesterol and LDL levels are important risk parameters for CVD development, thus, making it a clinically relevant model to test the identified biomarkers.

During disease progression, cardiac hypertrophy is usually characterized by an increase in interstitial fibrosis with augmented triglycerides and LDL affecting oxidative phosphorylation, myocardial structure as well as gene and protein expression of essential genes within this process²². In the present study, we found an increase in mRNA expression of *LOXL2*, after 11 weeks of age, which was concomitant with an impairment in LV diastolic function. This was the first study to show such an increase in diabetic cardiomyopathy. The observed increase at 11 weeks of *LOXL2* expression was confirmed by various other HF studies other than diabetes, where the authors showed an increase in mRNA expression of *LOXL2* was positively correlated with increased tissue fibrosis^{14,25,26}. Similarly, in a study performed by Zhao, it was argued that serum LOXL2 levels were positively correlated to the degree of left atrial fibrosis in Atrial fibrillation (AF) patients¹⁵. Furthermore, findings from Lu and later studies by Karthik argued that ETF β may be modulated during the development of insulin resistance in pancreatic β cells whereas the upregulation reduced cardiovascular dysfunction^{17,27}. Conversely, Boudina showed that after exposing db/db mice to increased palmitate, protein expression levels of ETF β were significantly decreased, however no coordinate increase in oxidative phosphorylation was observed²⁸. Nonetheless, more recently in a study done by Ruiz-Pinto, it was found that susceptible variants in the ETF β is a major contributor to anthracycline-mediated oxidative stress and myocardial injury. Inferring that decreased ETF β expression is linked to myocardial dysfunction²⁹. Our study made use of a db/db mice model who confirmed previous findings from Yang¹⁴ and Lu¹⁷, using different models showing that differential expression of *LOXL2* and *ETF β* was concomitant to increased ROS. Inferring that LOXL2 and ETF β might be implicated in the disease pathophysiology of DCM.

During disease progression, increased gluco- and lipotoxicity affect mitochondrial substrate preference ensuing in the enlargement of the heart muscle and hindering the ability of the myocardium to pump blood through the body effectively. As cardiomyopathy progresses, the heart becomes weaker due to reduced ejection fraction and is less able to pump blood through the body effectively. Although the precise age of onset in heart muscle deterioration in DCM is still incompletely understood, the development of LVD can directly be associated with increased fibrosis and development of LV wall thickness and decreased mitral E:A ratio. Mitral E:A ratio and ejection fraction are currently used as measurements of the hearts' ability to function optimally. In our study, we found that our identified serum markers correlated with cardiac dysfunction when the disease was still in its asymptomatic stage. This implies that the change in the circulatory markers (LOXL2 and ETF β) are as a result of the diabetic state that induces functional changes prior to the onset of any pathological changes. As such, the observed differential expression of the biomarkers is complicit to the maladaptive structural modifications that follow. According to Fuentes-Antras the maladaptive structural changes underlies the observed changes as shown by echocardiography, confirming that the hyperglycaemic state is the cause of LVD³⁰.

Furthermore, this study showed that knockdown of either LOXL2 or ETF β was able to reduce the expression of *COL1A*, a gene associated with increased fibrosis. Increased interstitial fibrosis, in the stressed heart, triggers the formation of enhanced collagen deposition that stiffens the heart muscle, while impairing diastolic relaxation and filling. Increased collagen deposits are not only concomitant to increased *COL1A* expression but also reduced ejection fraction and LV remodeling. Our data presented, therefore, infers that collagen formation might be modulated by LOXL2 and ETF β . Our findings are supported by two independent research groups reporting on the role of LOXL2 and ETF β and their association with increased fibrosis^{14,31,32}. In addition, ROC curve

analysis strongly supports that the identified markers were sensitive enough to detect the early onset of DCM, as confirmed by the AUC of 0.813 which is an indication of a good predictive test.

Study limitations

A limitation to this study was the inability to show the role ETF β plays in mitochondrial bioenergetics and the effect a loss in ETF β enzyme activity has on the respiratory chain, and subsequent ATP production. According to Boudina²⁸, ETF β is an electron acceptor that transfers electrons between Acyl-CoA substrates to the main respiratory chain, ubiquinone, which links the oxidation of fatty acids to the mitochondrial respiratory system. As such, a loss of function or decreased ETF β is known to alter cardiac mitochondrial flux, which consequently is the hallmark of DCM. To address this future studies should consider assessing the link between ETF β and respiratory chain function and mitochondrial ultrastructural abnormalities within the disease state.

Furthermore, it would have been of interest to follow disease progression from asymptomatic until a late stage of DCM, using LOXL2 and ETF β as serum biomarkers. Also, scarring as a consequence of hyperglycaemia is a progressive disease that worsens over time, with no cure and attaining treatment early might slow progression whilst maintaining the quality of life of the affected individual. Based on findings presented including studies performed by Yang¹⁴ and Lu¹⁷ it could be suggested that those diabetic patients (in this study reference is made to mice) with early dysregulation of serum LOXL2 and ETF β levels may be treated prophylactically in order to slow the progress of CVD. The proposed implementation strategy might be cost-intensive, however, in time patients might benefit from such treatment through increased life expectancy. As such, a follow-up human study should be initiated to prove such a hypothesis as the scope of this study did not encompass this important aspect. In the case of whether the identified markers can be used as either a preventative or even therapeutic intervention, it requires further investigation in order to support the clinical implication of the attained results.

Conclusion

It is well known that that increased hyperglycemia and augmented lipid accumulation are the main drivers of myocardial dysfunction and subsequent heart failure. In our study, a change in previously identified possible cardiac markers, LOXL2 and ETF β , was confirmed in the serum of db/db mice and correlated with increased LVDD, as observed with high-resolution Doppler echocardiography^{14,17,27,28}. We observed a significant increase in FBG and serum lipids levels from 9 weeks onwards. This was concomitant with increased LOXL2 and decreased ETF β serum levels at 9 and 11 weeks, respectively. Confirming a diabetic phenotype that was sustained until termination. As such, it would have been of interest to investigate if the identified changes occur only in the diabetic phenotype, regardless of whether they develop CVD and/or whether only the diabetic animals that eventually develop CVD had differentially regulated levels of the identified markers. Further investigation would need to be conducted to address this issue.

Furthermore, this study brings further insights to suggest that LOXL2 and ETF β might be implicated in the pathophysiology of DCM. Mechanistically, we propose that both identified biomarkers were able to regulate the expression of COLA1, a known cardiac fibrotic marker¹⁴. Most importantly, our data strongly suggest that the combined use of LOXL2 and ETF β displayed a good predictive power to detect the early onset of DCM and as such, support that the disease progression can be slowed through early intervention.

Materials and methods

In silico prediction of DCM biomarkers. *Data acquisition and processing.* In order to identify candidate genes differentially expressed in diabetic cardiomyopathy (DCM), the ArrayExpress database (<https://www.ebi.ac.uk/arrayexpress>) was screened for studies with publicly available microarray datasets belonging to Homo sapiens over 35 years of age. These studies included healthy controls and patients suffering from either type 2 diabetes mellitus (T2DM) or cardiovascular disease (CVD). To provide specificity for the disease state of interest (DCM), studies involving ischemia, CAD and hypertension were excluded; while studies inclusive of LVD and left ventricular ejection fraction (EF) were included with no restriction on ethnic group, gender, tissue or cell type.

For studies, utilising the Affymetrix and Agilent platform, raw datasets were extracted from Gene Expression Omnibus (GEO) using R scripting and the R package, *GEOquery*. Samples within these datasets fulfilling the specified criteria were then Robust-Multi array Average (RMA) normalised using the R package, *Simpleaffy*. Additionally, studies using a custom spotted oligonucleotide array was also included. These raw datasets were imported into R using the *ArrayExpress* package, while the *Limma* package was used for normalisation. Subsequently, microarray probe identifiers (IDs) in all the processed datasets were converted to Ensemble IDs or gene names, using a custom Python script. Probe sets, mapping multiple Ensembl IDs and technical replicates, were averaged to resolve probe redundancy using R and Ruby scripting.

Dataset integration and candidate gene identification. Differential expression analysis was performed in three stages: 1) to obtain differentially expressed genes (DEGs) for T2DM only, 2) to obtain DEGs for CVD only and 3) to obtain DEGs for integrated datasets from both T2DM and CVD, representing the DCM group. To this end, common Ensemble IDs were combined using a quantile discretization (QD) data binning approach, which integrates different microarray data at the gene expression level. In this study, a QD range of 128 was used to combine datasets in the T2DM and CVD subgroups only for statistical differential expression analysis, while a QD range of 1,028 was used for the integration (i.e. DCM) subgroup. Genes with a *p*-value of < 0.05 were regarded as significantly different using the Wilcoxon Rank Sum Test with Bonferroni and Benjamin-Hochberg correction for multiple testing. To prevent bias and due to limited samples, only the top 2 genes based on their Wilcoxon score were used for subsequent biomarker analysis.

In vivo validation. *Animals sample size and randomization.* Male C57BLKS/J leptin-receptor-deficient mice were obtained from Jackson's Laboratories (Sacramento, USA) and housed at the Primate Unit and Delft Animal Centre (PUDAC) of the South African Medical Research Council (SAMRC) in a controlled environment with a 12 h light/dark cycle in a temperature range of 23–25°C (relative humidity: ~50%). The mice received standard laboratory chow pellets (Afresh Vention, Cape Town, SA) ad libitum and had free access to water. At 6 weeks, based on their genetic background, mice were divided into 2 groups: (1) homozygous leptin receptor-deficient diabetic (db/db, n=40) mice and (2) heterozygous leptin-receptor-deficient non-diabetic wild type controls (db/+, n=40). Each group was then further divided into a total of 5 subgroups (week 6, 9, 11, 14, 16), with n=6–8 animals per subgroup. After each week, (6, 8, 9, 11, 14 and 16) high-resolution Doppler echocardiography was performed, and animals were terminated to collect blood and heart tissue for subsequent serum biomarker analysis. Ethical clearance for the use of animals was obtained from South African Medical Research Council (SAMRC) (ECRA 07/13) and all experiments were conducted in accordance with the National Institute of Health Guide for the Care and use of laboratory animals.

Echocardiography analysis. The mice were placed in a holding chamber and gassed with 1.0–2.5% isoflurane, whereafter they were removed and positioned in the supine position on a warming pad. Closed chest high-resolution Doppler echocardiography was performed with a VEVO 2,100 ultrasound system (Fujifilm, Visualsonics, Ontario, Canada) and a 30 MHz linear array transducer at the respective time intervals. Left ventricular internal diameter, interventricular septum thickness, posterior wall thickness were measured in systole and diastole (Figure S2). EF was measured as a marker of systolic function. Mitral E and A wave flow velocity, E:A ratio, deceleration time of mitral E wave velocity and isovolumic relaxation time (IVRT) were assessed as measurements of LVD using Doppler echocardiography (Figure S2). All measurements were made off-line on the mean of at least three consecutive cardiac cycles with the software resident on the ultrasound system.

Body weights and fasting blood glucose levels. Body weights were measured weekly, while 4 h FBG levels were monitored by tail prick using a OneTouch Select glucometer (Lifespan, Milpitas, CA, USA).

Cardiovascular disease risk predictors. On the day of termination, mice were anaesthetised with halothane before blood was collected from the abdominal vena cava for subsequent lipid profile and serum biomarker analysis. Serum was isolated from peripheral blood by centrifugation at 4,000 g for 15 min. Thereafter, the serum was removed and divided into two parts; one part was sent to PathCare Medical Diagnostic Laboratories (Cape Town, RSA) for total cholesterol, triglycerides and LDL analysis; while the other half was stored at -80°C for subsequent serum biomarker analysis.

Detection of LOXL2, ETF β and proBNP in serum. LOXL2, ETF β and NT-proBNP concentrations were measured with enzyme-linked immunosorbent assay (ELISA) (MYbiosource, Berlin, Germany), according to the manufacturer's instructions.

Functional analysis using an in vitro model. *Cell culture.* H9c2 cardiomyoblasts were cultured in supplemented Dulbecco's Modified Eagle's Medium (10% fetal bovine serum, 100 μ g/mL penicillin and 100 μ g/mL streptomycin) under standard tissue culture conditions (37 °C in humidified air and 5% CO₂). Cells were seeded in a 6-well plate at a seeding density of 2×10^4 cells/well and left for 2–3 days until confluent. On day 4, cells were utilised for transfection experiments using siRNA.

Knockdown of LOXL2 and ETF β using small Interfering RNA (siRNA). Knockdown experiments for the two selected candidate biomarkers were done using a Lipofectamine RNAimax reagent (ThermoFisher Scientific, Inc, Waltham, MA, USA), according to the manufacturer's instructions. Briefly, H9c2 cells at approximately 70% confluence were transfected with either siLOXL2, siETF β or scrRNA for LOXL2 or ETF β for 24 h, respectively. Thereafter, transfected cells were exposed to both high glucose (HG; 33 mM) and palmitate (PAL; 100 μ M) for 6 h. The degree of knockdown after 24 h of transfection was confirmed by RT-PCR. Non-transfected cells, exposed to either 5.5 mM glucose or 33 mM glucose, served as controls, respectively.

Gene expression analysis using quantitative real-time polymerase chain reaction (qRT-PCR). Total RNA was extracted using the TRIzol method (ThermoFisher Scientific Inc, Waltham, MA, USA), and subsequently purified with an RNeasy mini kit according to the manufacturer's instructions. The extracted RNA was treated with DNaseI (Applied Biosystems, Foster City, CA, USA) to eliminate genomic DNA contamination and reversed transcribed to generate cDNA with a Superscript first-strand cDNA synthesis kit (Applied Biosystems, Foster City, CA, USA) as per previously described protocol³³. Thereafter, qRT-PCR analysis was performed using 5 μ L Taqman Universal PCR master mix, 0.5 μ L predesigned and optimized TaqMan gene expression assays (LOXL2: Mm00804740_m1; ETF β : Mm005033401_m1) and 1 μ L cDNA on an ABI 7,500 Instrument (Applied Biosystems, Life Technologies, USA). The PCR reaction was as follows: 50 °C for 1 min and 95 °C for 10 min; followed by 95 °C for 15 s and 60 °C for 30 s (40 cycles). Gene expression data was normalized to that of Actin- β (ACT- β) and hypoxanthine-guanine phosphoribosyltransferase (HPRT) and relative gene expression was calculated using the standard curve method.

Statistical analysis. All data obtained throughout this study are expressed as the mean \pm standard error of the mean (SEM) where appropriate. Results for in vitro experiments were expressed as the mean of three inde-

pendent biological experiments, with each experiment containing at least three technical replicates, unless otherwise stated. For in vivo experiments, each treatment group included six mice. Differences between the means of groups were first tested for normality with Shapiro–Wilk tests. In the instances where data sets were normally distributed, they were assessed for statistical significance using one-way or two-way multivariate ANOVA and Student t-tests, while non-parametric Mann–Whitney U tests were used to analyse data sets deviating from¹ a normal distribution. To evaluate the diagnostic potential of each biomarker, principal component (PCI) analysis was performed by combining the data for the identified biomarkers and receiver operating characteristic (ROC) curve analysis, where an area under the curve (AUC) greater than 0.5 indicated diagnostic value. Subsequently, the ROC curve analysis was used to calculate corresponding sensitivity and specificity. To measure the strength and nature of the relationship between the expression levels of the candidate biomarkers and mitral E:A ratio, correlation analysis was performed using a Pearson and Spearman coefficient test. Statistical analyses were performed using GraphPad Prism software version 5.00 (GraphPad Software, Inc., La Jolla, CA, USA). In all instances, statistical significance was inferred for a *p*-value less than, or equal to 0.05.

Received: 5 December 2019; Accepted: 23 June 2020

Published online: 23 July 2020

References

1. IDF. International Diabetes Federation (IDF) *Diabetes Atlas* (9th Edition) Global fact sheet. 1–2 (2019).
2. WHO. World Health Organization. *Hypertensive Fact Sheet* (2019).
3. Rubler, S. *et al.* New type of cardiomyopathy associated with diabetic glomerulosclerosis. *Am. J. Cardiol.* **30**, 595–602 (1972).
4. Bertolucci, M. C. & Rocha, V. Z. Cardiovascular risk assessment in patients with diabetes. *Diabetol. Metab. Syndr.* **9**, 25. <https://doi.org/10.1186/s13098-017-0225-1> (2017).
5. Miki, T., Yuda, S., Kouzu, H. & Miura, T. Diabetic cardiomyopathy: pathophysiology and clinical features. *Heart Fail. Rev.* **18**, 149–166. <https://doi.org/10.1007/s10741-012-9313-3> (2013).
6. Boudina, S. & Abel, E. D. Diabetic cardiomyopathy revisited. *Circulation* **115**, 3213–3223. <https://doi.org/10.1161/circulationaha.106.679597> (2007).
7. Guo, R. & Nair, S. Role of microRNA in diabetic cardiomyopathy: from mechanism to intervention. *Biochem. Biophys. Acta.* **2070–2077**, 2017. <https://doi.org/10.1016/j.bbadis.2017.03.013> (1863).
8. Fang, Z. Y., Prins, J. B. & Marwick, T. H. Diabetic cardiomyopathy: evidence, mechanisms, and therapeutic implications. *Endocr. Rev.* **25**, 543–567. <https://doi.org/10.1210/er.2003-0012> (2004).
9. Lee, W. S. & Kim, J. Diabetic cardiomyopathy: where we are and where we are going. *Korean J. Int. Med.* **32**, 404–421. <https://doi.org/10.3904/kjim.2016.208> (2017).
10. Bui, A. L., Horwich, T. B. & Fonarow, G. C. Epidemiology and risk profile of heart failure. *Nat. Rev. Cardiol.* **8**, 30–41. <https://doi.org/10.1038/nrcardio.2010.165> (2011).
11. Lorenzo-Almoros, A. *et al.* Diagnostic approaches for diabetic cardiomyopathy. *Cardiovasc. Diabetol.* **16**, 28 (2017).
12. Leon, L. E., Rani, S., Fernandez, M., Larico, M. & Calligaris, S. D. Subclinical detection of diabetic cardiomyopathy with microRNAs: challenges and perspectives. *J. Diabetes Res.* **2016**, 6143129. <https://doi.org/10.1155/2016/6143129> (2016).
13. Romano, S. *et al.* Early diagnosis of left ventricular diastolic dysfunction in diabetic patients: a possible role for natriuretic peptides. *Cardiovasc. Diabetol.* **9**, 89. <https://doi.org/10.1186/1475-2840-9-89> (2010).
14. Yang, J. *et al.* Targeting LOXL2 for cardiac interstitial fibrosis and heart failure treatment. *Nat. Commun.* **7**, 13710. <https://doi.org/10.1038/ncomms13710> (2016).
15. Zhao, Y. *et al.* Increased serum lysyl oxidase-like 2 levels correlate with the degree of left atrial fibrosis in patients with atrial fibrillation. *Biosci. Rep.* **37**. <https://doi.org/10.1042/bsr20171332> (2017).
16. Jiang, J. *et al.* Increased mitochondrial ROS formation by acetaminophen in human hepatic cells is associated with gene expression changes suggesting disruption of the mitochondrial electron transport chain. *Toxicol. Lett.* **234**, 139–150. <https://doi.org/10.1016/j.toxlet.2015.02.012> (2015).
17. Lu, H., Yang, Y., Allister, E. M., Wijesekara, N. & Wheeler, M. B. The identification of potential factors associated with the development of type 2 diabetes: a quantitative proteomics approach. *Mol. Cell. Proteomics MCP* **7**, 1434–1451. <https://doi.org/10.1074/mcp.M700478-MCP200> (2008).
18. Lorenzo, O., Ramirez, E., Picatoste, B., Egado, J. & Tunon, J. Alteration of energy substrates and ROS production in diabetic cardiomyopathy. *Mediat. Inflamm.* **2013**, 461967. <https://doi.org/10.1155/2013/461967> (2013).
19. Belke, D. D. & Dillmann, W. H. Altered cardiac calcium handling in diabetes. *Curr. Hypertens. Rep.* **6**, 424–429 (2004).
20. Kobayashi, K. *et al.* The db/db mouse, a model for diabetic dyslipidemia: molecular characterization and effects of Western diet feeding. *Metab. Clin. Exp.* **49**, 22–31 (2000).
21. Inoue, T. *et al.* Downregulation of adipose triglyceride lipase in the heart aggravates diabetic cardiomyopathy in db/db mice. *Biochem. Biophys. Res. Commun.* **438**, 224–229. <https://doi.org/10.1016/j.bbrc.2013.07.063> (2013).
22. Dludla, P. V. *et al.* Age-dependent development of left ventricular wall thickness in type 2 diabetic (db/db) mice is associated with elevated low-density lipoprotein and triglyceride serum levels. *Heart Vessels* **32**, 1025–1031. <https://doi.org/10.1007/s00380-017-0978-3> (2017).
23. Liu, Q., Wang, S. & Cai, L. Diabetic cardiomyopathy and its mechanisms: Role of oxidative stress and damage. *J. Diabetes Invest.* **5**, 623–634. <https://doi.org/10.1111/jdi.12250> (2014).
24. Alex, L., Russo, I., Holoborodko, V. & Frangogiannis, N. G. Characterization of a mouse model of obesity-related fibrotic cardiomyopathy that recapitulates features of human heart failure with preserved ejection fraction. *Am. J. Physiol. Heart Circul. Physiol.* **315**, H934–H949. <https://doi.org/10.1152/ajpheart.00238.2018> (2018).
25. Gonzalez-Santamaria, J. *et al.* Matrix cross-linking lysyl oxidases are induced in response to myocardial infarction and promote cardiac dysfunction. *Cardiovasc. Res.* **109**, 67–78. <https://doi.org/10.1093/cvr/cvv214> (2016).
26. Chen, L., Li, S. & Li, W. LOX/LOXL in pulmonary fibrosis: potential therapeutic targets. *J. Drug Targeting* <https://doi.org/10.1080/1061186x.2018.1550649> (2018).
27. Karthik, D., Vijayakumar, R., Pazhanichamy, K. & Ravikumar, S. A proteomics approach to identify the differential protein level in cardiac muscle of diabetic rat. *Acta Biochim. Pol.* **61**, 285–293 (2014).
28. Boudina, S. *et al.* Mitochondrial energetics in the heart in obesity-related diabetes: direct evidence for increased uncoupled respiration and activation of uncoupling proteins. *Diabetes* **56**, 2457–2466. <https://doi.org/10.2337/db07-0481> (2007).
29. Ruiz-Pinto, S. *et al.* Exome array analysis identifies ETFB as a novel susceptibility gene for anthracycline-induced cardiotoxicity in cancer patients. *Breast Cancer Res. Treat.* **167**, 249–256. <https://doi.org/10.1007/s10549-017-4497-9> (2018).

30. Fuentes-Antras, J. *et al.* Targeting metabolic disturbance in the diabetic heart. *Cardiovasc. Diabetol.* **14**, 17. <https://doi.org/10.1186/s12933-015-0173-8> (2015).
31. Hirokawa, S., Shimanuki, T., Kitajima, H., Nishimori, Y. & Shimosaka, M. Identification of ETFB as a candidate protein that participates in the mechanoregulation of fibroblast cell number in collagen gel culture. *J. Dermatol. Sci.* **64**, 119–126. <https://doi.org/10.1016/j.jdermsci.2011.08.003> (2011).
32. Hirokawa, S., Shimanuki, T., Kitajima, H., Nishimori, Y. & Shimosaka, M. Knockdown of electron transfer flavoprotein beta subunit reduced TGF-beta-induced alpha-SMA mRNA expression but not COL1A1 in fibroblast-populated three-dimensional collagen gel cultures. *J. Dermatol. Sci.* **68**, 179–186. <https://doi.org/10.1016/j.jdermsci.2012.09.012> (2012).
33. Johnson, R. *et al.* Aspalathin, a dihydrochalcone C-glucoside, protects H9c2 cardiomyocytes against high glucose induced shifts in substrate preference and apoptosis. *Mol. Nutr. Food Res.* **60**, 922–934. <https://doi.org/10.1002/mnfr.201500656> (2016).

Acknowledgements

The work reported herein was made possible through funding by the South Africa Medical Research Council's Biomedical Research and Innovation Platform baseline funding as well as Division of Research Capacity Development Internship Scholarship and Intra-mural Post-doctoral Program, South African Rooibos Council and the National Research Foundation (NRF) Thuthuka Programme Grant 107261. NRF echocardiograph large equipment Grant at the University of Cape Town. We would like to thank Charna Chapman, Desmond Linden, Samira Ghoor, Ruzayda van Aarde and Joritha van Heerden for technical support.

Author contributions

RJ conceptualized, designed, wrote and edited the manuscript. X.N. conducted the experiments and collected and analyzed the data. T.J. performed the mRNA expression analysis and edited the manuscript. N.S. performed siRNA expression analysis. E.S. contributed to the siRNA design, analysis and edited the manuscript. M.M. performed the receiver operating characteristic (ROC) analysis. M.C. performed TDI echocardiography, analyzed the data and edited the manuscript. N.H. performed TDI echocardiography and edited the manuscript. S.L. performed TDI echocardiography and edited the manuscript. M.B. reviewed and edited the manuscript. J.L. acquired funding.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information is available for this paper at <https://doi.org/10.1038/s41598-020-69254-x>.

Correspondence and requests for materials should be addressed to R.J.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2020