

Plasma BCAA changes in Patients with NAFLD are Sex Dependent 1

Guillaume Grzych, Luisa Vonghia, Marie-Adélaïde Bout, Jonas Weyler, An Verrijken, Eveline Dirinck, Marie Joncquel, Luc van Gaal, Réjane Paumelle, Sven Francque, et al.

▶ To cite this version:

Guillaume Grzych, Luisa Vonghia, Marie-Adélaïde Bout, Jonas Weyler, An Verrijken, et al.. Plasma BCAA changes in Patients with NAFLD are Sex Dependent 1. Journal of Clinical Endocrinology and Metabolism, 2020, 105 (7), pp.dgaa175. 10.1210/clinem/dgaa175. inserm-02864390

HAL Id: inserm-02864390 https://inserm.hal.science/inserm-02864390

Submitted on 11 Jun2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 Plasma BCAA changes in Patients with NAFLD are Sex Dependent Guillaume Grzych^{1,2}, Luisa Vonghia^{3,4}, Marie-Adélaïde Bout², Jonas Weyler^{3,4}, An Verrijken^{4,5}, 2 Eveline Dirinck^{4,5}, Marie Joncguel², Luc Van Gaal^{4,5}, Réjane Paumelle¹, Sven Francgue^{3,4*}, 3 Anne Tailleux^{1*}, Joel T. Haas^{1*}, Bart Staels^{1,2*} 4 5 Affiliations: 6 7 ¹ Univ. Lille, Inserm, CHU Lille, Institut Pasteur de Lille, U1011- EGID, F-59000 Lille, France. ² CHU Lille, Service d'Hormonologie, Métabolisme, Nutrition, Oncologie, F-59000 Lille, 8 9 France 10 ³ Department of Gastroenterology and Hepatology, Antwerp University Hospital, 2650 11 Edegem/Antwerp, Belgium ⁴ Laboratory of Experimental Medicine and Pediatrics, Faculty of Medicine and Health 12 Sciences, University of Antwerp, 2610 Wilrijk/Antwerp, Belgium 13 ⁵ Department of Endocrinology, Diabetology and Metabolism, Antwerp University Hospital, 14 15 2650 Edegem/Antwerp, Belgium * These authors co-supervised the study 16 17 Corresponding Author: Joel T. Haas 18 19 Institut Pasteur de Lille, 1 rue du professeur Calmette, BP245, 59019 LILLE - France 20 Tel: +33320877171 - Fax: +33320877360 21 e-mail: joel.haas@pasteur-lille.fr 22 23 Disclosures Summary: BS and SF are consultants for Genfit S.A. SF and LV are consultants 24 for Inventiva. All other authors have nothing to declare. **Funding Sources:** 25 This work was supported by the French Agence Nationale pour la Recherche (ANR-10-LABX-26 27 46 and ANR-16-RHUS-0006) and grants from the European Commission: HEPADIP (Contract 28 LSHM-CT-2005-018734) and RESOLVE (Contract FP7-305707) projects. BS is a recipient of an

ERC Advanced Grant (694717). SF has a senior clinical research mandate from the Fund for
 Scientific Research (FWO) Flanders (1802154N).

31 Abstract

32 Context: Plasma branched chain amino acid (BCAA) concentrations correlate positively with 33 body mass index (BMI), measures of insulin resistance (IR) and severity of non-alcoholic 34 fatty liver disease (NAFLD). Moreover, plasma BCAA concentrations also differ between the 35 sexes, which display different susceptibilities to cardio-metabolic diseases.

36 **Objective:** Assess whether plasma BCAA concentrations associate with NAFLD severity

37 independently of BMI, IR and sex.

38 Patients: Patients visiting the obesity clinic of the Antwerp University Hospital were
 39 consecutively recruited from 2006 to 2014.

40 **Design and Setting:** A cross-sectional study cohort of 112 obese patients (59 women and 53

- 41 men) was divided into four groups according to NAFLD severity. Groups were matched for
- 42 sex, age, BMI, HOMA-IR and HbA₁c.

43 Main Outcome Measures: Fasting plasma BCAA concentrations were measured by tandem
 44 mass spectrometry using the aTRAQ[™] method.

45 **Results:** In the study cohort, a modest positive correlation was observed between plasma

46 BCAA concentrations and NAFLD severity, as well as a strong effect of sex on plasma BCAA

47 levels. Subgroup analysis by sex revealed that while plasma BCAA concentrations increased

48 with severity of NAFLD in women, they tended to decrease in men. Additionally, only

49 women displayed significantly increased plasma BCAAs with increasing fibrosis.

50 **Conclusion:** Plasma BCAA concentrations display sex-dimorphic changes with increasing 51 severity of NAFLD, independently of BMI, IR and age. Additionally, plasma BCAA are 52 associated with significant fibrosis in women, but not in men. These results highlight the 53 importance of a careful consideration of sex as a major confounding factor in cross-sectional 54 studies of NAFLD.

55 Introduction

Non-Alcoholic Fatty Liver Disease (NAFLD) is a major global public health problem. NAFLD covers a spectrum of conditions from simple hepatic triglyceride accumulation, termed non-alcoholic fatty liver (NAFL), to more severe non-alcoholic steatohepatitis (NASH) which combines NAFL with hepatic necroinflammation (1). Patients with NASH are at high risk for progressive liver fibrosis, hepatocellular carcinoma and cardiovascular disease (2). Recent studies indicate that NAFLD progression and incidence are significantly influenced by sex, suggesting NAFLD may be a sexually dimorphic disease (3,4).

Development of non-invasive diagnostic markers for NAFLD remains a major challenge. Current guidelines for accurate NAFLD diagnosis require liver biopsy (5,6), which is invasive and susceptible to inter-observer discrepancies (7,8). Non-invasive imaging, individual biochemical markers and combined clinical and biochemical score-based methods are being developed, but have not yet been accepted as a valid substitute for histological diagnosis. Thus, there is an urgent need to identify accurate markers of NAFLD stages, especially NASH and fibrosis, to simplify disease staging for diagnosis and monitoring treatment response.

71 Moreover, as biological plausibility is an important feature of these non-invasive markers,

72 their identification often yields insight into disease mechanisms.

73 The branched chain amino acids (BCAA) leucine, valine and isoleucine are essential amino 74 acids whose plasma concentrations are increased in metabolic diseases such as obesity and 75 type 2 diabetes (T2D) (9–11). Recently, plasma BCAA were also found to be increased in 76 NAFLD patients compared to patients without NAFLD (12–14). Interestingly, a recent large-77 scale population study demonstrated that plasma BCAA and their degradation products 78 were elevated in men compared to women (15). Given the known discrepancies between 79 men and women in metabolic disease susceptibility and the interaction between obesity, IR 80 and NAFLD (9,11,16–20), it is currently unclear whether changes in plasma BCAA are 81 specifically related to NAFLD progression or rather to accompanying metabolic alterations. 82 Consequently, the aim of this study was to determine whether changes in plasma BCAA 83 concentrations are specifically associated with severity of NAFLD in humans. We measured 84 plasma BCAA concentrations in a well-characterized patient cohort with histologically 85 assessed NAFLD. Our results show that plasma BCAA levels vary with increasing severity 86 from NAFL to NASH in a sex-dependent manner, independently of alterations in glucose 87 metabolism.

88

89 Methods

90 **Description of the patients**

91 Patients were consecutively recruited at the metabolic unit of the obesity clinic of the 92 Antwerp University Hospital (a tertiary referral facility) from October 2006 to May 2014. The 93 study protocol, part of the HEPADIP protocol (Belgian registration number B30020071389), 94 was approved by the Ethical Committee of the Antwerp University Hospital (file 6/25/125) 95 and required written informed consent of the patient. Inclusion criteria were previously 96 Briefly, patients underwent extensive metabolic characterization and described (21). 97 systematic non-invasive screening for the presence of NAFLD (e.g. plasma alanine 98 transaminase (ALT), aspartate transaminase (AST) and gamma glutamyltransferase (GGT) 99 measurements, and ultrasound features) and proposed for liver biopsy upon suspicion of 100 NAFLD. Liver biopsy was performed percutaneously (16G Menghini), transjugularly (16G 101 transjugular liver biopsy needle) or perioperatively (16G Trucut needle) for patients who 102 subsequently underwent bariatric surgery. The herein described assessment, including liver 103 biopsy, was performed before any treatment initiation or consideration.

104

105 Clinical assessment and biological assay

Fasting plasma glucose and insulin were measured as previously described (22). Insulin resistance (IR) was estimated using both the homeostatic model assessment of insulin resistance (HOMA-IR) calculated by [insulin (mU/L) × glucose (mmol/L)]/22.5 and using the updated homeostasis model assessment HOMA2 from Levy *et al.* (23) that estimates insulin resistance (HOMA2-IR), steady state beta cell function (HOMA2-B) and insulin sensitivity (HOMA2-S), as percentages of a normal reference population.

112

113 Liver histology

114 Hematoxylin and eosin, Sirius red, reticulin and Perls iron stains were routinely performed 115 on all liver biopsies and analyzed in batch by two experienced pathologists blinded to clinical 116 data. The histological features of NAFLD (steatosis, ballooning, lobular inflammation and 117 fibrosis) were assessed using the NASH Clinical Research Network Scoring System (5). 118 According to biopsy evaluation, patients were divided in 4 groups: Healthy, NAFL, NASH and 119 NASH with significant fibrosis. The healthy group was defined as steatosis=0, ballooning=0, lobular inflammation=0 and fibrosis=0. The NAFL group included patients with steatosis > 0 120 121 and allowing some presence of either ballooning or lobular inflammation, but not meeting 122 the criteria for NASH. NASH diagnosis required the combined presence of steatosis, 123 ballooning and lobular inflammation, according to current guidelines (24,25). Patients with 124 NASH were subsequently divided according to fibrosis stage: NASH without significant 125 fibrosis (fibrosis ≤ 1) and NASH with significant fibrosis (NASH-F, fibrosis ≥ 2).

126

127 Plasma amino acid measurements

128 Blood samples were collected in EDTA-coated tubes after an overnight fast. Isolated plasma 129 was aliquoted, immediately frozen and stored at -80°C until analysis. Targeted 130 metabolomics was performed to quantify plasma amino acids using the aTRAQ[™] Kit (Sciex, 131 Framingham, MA, USA) as described previously (26). Briefly, plasma amino acids were 132 amine-modified using the aTRAQ[™] labeling reagent, which provides a specific mass tag for 133 tandem mass spectrometry (MS). The tag is identified by MS/MS fragmentation of samples 134 and standards while in multiple reaction monitoring (MRM) mode. An internal standard (IS) 135 set of aTRAQ[™] labeled amino acids was used for detection and quantification by high 136 performance liquid chromatography (LC) (Shimadzu C18 column, Kyoto, Japan) associated 137 with MS/MS (Sciex 3200 Qtrap, Framingham, MA). Quantification was performed dividing

the peak area for each analyte by the peak area of the corresponding IS and multiplying bythe IS concentration.

140

141 Rationale of study group composition

142 As outlined above, BCAA metabolism and concentrations are influenced by BMI, diabetes 143 and sex. These factors are poorly accounted for when studying the relationship between BCAA concentrations and presence and severity of NAFLD. In order to unequivocally study 144 145 the relation between plasma BCAA concentrations and NAFLD severity, the following 146 approach was used. Patients were first classified into clearly distinct groups ("NAFLD 147 stages") based on their liver histological phenotype, as follows: healthy liver, NAFL, NASH 148 and NASH-F (see histology section above) or unclassified (i.e. patients not clearly falling into 149 one of the previously defined categories). Patients were subsequently selected from the 4 150 well defined NAFLD stages to compose groups of equal size that were further matched for 151 the following parameters: BMI, fasting plasma glucose, HOMA-IR, HbA1c and age. Finally, an 152 equal number of men and women was included in each study group when possible.

153

154 Statistical analysis

Patient selection was performed using regression-based matching in R (*matchit package* Rproject) considering clinical parameters as described above. Values are expressed as mean ± standard deviation (mean ± SD) or median ± Inter Quartile Range (IQR) as indicated in the figure legends. Statistical differences in clinical and biological parameters were assessed by Mann-Whitney or ANOVA test for quantitative values and Chi-squared test for categorical values. The Spearman rank test was used to determine correlation between variables. All

univariate correlations were then verified in a linear model adjusting for BMI, HOMA-IR and
 sex. P-values less than 0.05 were considered statistically significant.

163 **Results**

164 **Patient selection and matching**

165 Among all consecutively recruited patients with available clinical information (n=408) and 166 fasting plasma samples, four study groups corresponding to different stages of NAFLD were 167 established (study cohort) as described in the methods section. This process resulted in 4 168 clearly distinct liver phenotypical groups of about 30 patients, matched for BMI, fasting 169 plasma glucose (FPG), HOMA-IR, HbA₁c and age. For the group of healthy liver, a lower 170 number of patients who met both the histological phenotype and all matching criteria was 171 available, n = 23. As outlined in the methods section, an equal number of men and women 172 was included in each study group when possible. The clinical, biological and liver histological 173 characteristics of the patients are shown in Table 1. Overall, 112 patients were divided over 174 the 4 groups as follows: 23 healthy liver (15 women and 8 men), 30 with NAFL (15 women 175 and 15 men), 30 with NASH (15 women and 15 men) and 29 with NASH and significant 176 fibrosis (NASH-F; 14 women and 15 men). As per the matching protocol, the four groups did 177 not show significant differences in sex, age, BMI, FPG, HOMA-IR and HbA₁c (Table 1).

178

179 Plasma BCAA concentrations highly associate with sex, but not with NAFLD stages

Plasma concentrations of the three BCAA in the study cohort were measured and correlations between plasma BCAA concentrations and NAFLD stage were assessed. In agreement with previous studies (12–14), plasma leucine and, to a greater extent, plasma isoleucine correlated with NAFLD stage (Spearman coefficient: rho=0.17 and rho=0.25,

p=0.06 and p<0.01, respectively) (Figure 1A, 1C). However, plasma value concentrations did
 not significantly correlate with NAFLD stage (Figure 1B).

186 To assess which factor most contributed to plasma BCAA variation in the study cohort, we 187 calculated the correlations between BCAA and several known confounding factors (i.e. age, 188 BMI, HOMA-IR and sex). Interestingly, sex showed the strongest association with plasma 189 BCAA concentrations when compared to age, BMI, HOMA-IR and NAFLD stage (Figure 2). As 190 expected based on the patient stratification strategy, no or minor correlations were 191 observed between BCAA and age, BMI and HOMA-IR. Moreover, we calculated the 192 Spearman correlation coefficient between plasma BCAA and HOMA2 parameters (HOMA2-193 IR) and no significant correlations were observed (Figure 2).

194 To confirm this specific correlation of plasma BCAA with sex, independently of BMI, age and 195 glucose homeostasis parameters, a linear model including age, BMI, HOMA-IR, NAFLD stage 196 and sex was developed. This model confirmed an independent association between the 3 197 BCAA plasma concentrations and sex (p<0.001). We consequently analysed the BCAA 198 concentrations in the study cohort separating men and women, irrespective of NAFLD 199 status. The plasma concentrations of the 3 BCAA were higher in men compared to women 200 (p<0.001) (Figure 3A, 3B, 3C), indicating that sex is a major determinant of plasma BCAA 201 concentrations.

202

203 Sex differentially influences the correlation of plasma BCAA with NAFLD severity

Due to the strong effect of sex on plasma BCAA concentrations, men and women were subsequently analysed separately. Clinical, biological and liver histological characteristics of the patients separated according to sex (53 men and 59 women) and NAFLD stage are shown in **Table 2**. The NAFLD subgroups stratified by sex remained matched according to 208 age, BMI and glucose homeostasis parameters (Table 2), as was the case for the 4 initial 209 groups (Table 1). Moreover, average age, BMI, FPG, HOMA-IR, HbA1c and liver histological 210 characteristics were not different between men and women (Table 3). For women, 211 menopause status was collected, and the proportion of women post-menopause did not 212 differ with NAFLD severity (Table 2). Moreover, for women with NASH (n=29), we observed 213 no significant differences in the plasma BCAA concentrations between pre- (n=18) and post-214 menopausal (n=11) women (data not shown). The low numbers of post-menopausal 215 patients in the Healthy and NAFL groups did not permit the same comparison. Still, these 216 results suggest that menopause alone is unlikely to lead to a major difference in plasma 217 BCAA levels in the context of NAFLD.

Interestingly, plasma BCAA showed an inverse relation to NAFLD depending on patient sex.
In men, plasma leucine and valine negatively correlated with increasing NAFLD stage (Figure **4A, 4B**), whereas plasma isoleucine did not correlate with NAFLD stage (Figure 4C).
Conversely, in women, plasma leucine, valine and isoleucine concentrations all positively
correlated with NAFLD severity (plasma leucine: Spearman rho=0.43, p<0.01; plasma valine:
Spearman rho=0.39, p<0.01; plasma isoleucine: Spearman rho=0.43, p<0.01; Figure 4D, 4E, **4F**). These results indicate a sex-specific signature of plasma BCAA in NAFLD.

225

226 Sex-dependent association of plasma BCAA with NAFLD features

To better understand the specific disease features associated with plasma BCAA changes, plasma BCAA concentrations were correlated with each NAFLD histological characteristic separately in men and women. In men (n=53), with the exception of a modest inverse correlation of plasma valine with lobular inflammation (rho=-0.33, p=0.02), plasma BCAA concentrations did not significantly associate with any of the parameters tested (**Figure 5**). 232 Conversely, in women, the 3 plasma BCAA correlated positively with steatosis and fibrosis. 233 Additionally, plasma leucine and isoleucine concentrations significantly correlated with the 234 histological ballooning score (rho=0.36, p=0.004 for leucine and rho=0.33, p=0.009 for 235 isoleucine) (**Figure 5**).

236 Interestingly, all 3 plasma BCAA most strongly correlated with fibrosis in women only (Figure 237 5). To investigate whether BCAA could discriminate early from more advanced stages of 238 fibrosis in patients with NASH, women and men were subgrouped in 2 fibrosis stages: no or 239 low fibrosis (F0 to F1; F0-1) and significant fibrosis (F≥2). In men, no significant association 240 was found between plasma BCAA and fibrosis group (Figure 6A-C). Conversely, women 241 displayed a significant increase of all three plasma BCAAs with the presence of significant 242 fibrosis (leucine: 117±21 µmol/L vs 147±31 µmol/L; valine: 221±34 µmol/L vs 278±50 243 μ mol/L; isoleucine: 61±10 μ mol/L vs 81±16 μ mol/L; F0-1 vs F≥2, Figure 6D-F). Moreover, as 244 expected from the initial matching strategy, HOMA-IR, BMI, age and glucose homeostasis 245 parameters were not significantly different between the fibrosis subgroups.

246

247 **Discussion**

Here, we investigated the correlation of plasma BCAA levels with NAFLD severity and fibrosis in patient groups from a cross-sectional NAFLD cohort. To account for their potential confounding effects, groups were matched for age, BMI, glucose metabolism parameters and sex. Our results indicate that plasma BCAA correlate with severity of NAFLD independently of known confounders, but in a sex-dependent manner, increasing from NAFL to fibrotic NASH in women, while decreasing in NASH in men.

Our results extend recent findings from Gaggini *et al.* (14), who found a positive association of plasma BCAA with increasing NAFLD severity and fibrosis in a cohort of both obese and

256 non-obese NAFLD patients. However, in this study, sex ratio imbalances were present 257 between the disease groups (14). Indeed, there was a higher proportion of men in the obese 258 NAFLD group, whereas in the control group there was a higher proportion of women. Our 259 results suggest this may have exaggerated the association of elevated BCAA with NAFLD. 260 Lake et al. also found a positive association between plasma BCAA and NAFLD stages with 261 limited numbers of patients (12). However, non-standard NAFLD classification criteria were 262 used, and no data on sex distribution nor insulin-resistance was provided. Finally, our 263 findings in women are in agreement with a third study reporting a positive association of 264 plasma BCAA with increasing NAFLD severity in a cohort of obese, non-diabetic women (13).

265

266 There are numerous sources of seeming discordance in the literature among cohort studies 267 of metabolite changes in NAFLD. These can arise from the definition of experimental groups 268 (i.e. NAFLD vs Healthy or NASH versus NAFL, etc.) or differences in confounding parameters 269 between experimental and control groups. For example, we found that alterations in 270 circulating bile acids, previously reported to be elevated in NAFLD (27,28), may be due to 271 differences in insulin resistance rather than NASH per se in a cohort of non-diabetic NASH 272 patients (29). The present study, with clearly distinct groups based on their liver phenotype, 273 but matched for several potential confounders, allowed specific assessment of the 274 association of histological liver alterations with plasma metabolites, i.e. BCAA. Therefore, 275 studies designed to identify specific associations with NAFLD presence and severity should, 276 in our opinion, always be controlled for known confounding factors, especially sex, as shown 277 in the present study.

The effect of sex in cohort studies of NAFLD has recently been highlighted, as prevalence and severity of NAFLD is higher in men than in pre-menopausal women (3,30,31). Hence, a

280 specific metabolic signature dependent on patient sex may be expected in NAFLD. To our 281 knowledge, the present study is the first to reveal sex-dependent associations of plasma 282 metabolites with NAFLD severity. Indeed, plasma BCAA concentrations increased with 283 NAFLD severity in women whereas they decreased in men, independently of glucose 284 metabolism parameters. With respect to other clinical variables, we observed no significant 285 differences between men and women for plasma triglycerides, plasma total cholesterol, 286 ASAT or ALAT (not shown). As expected, plasma HDL-C was lower in men than in women, 287 and tended to decrease in both sexes with increasing NAFLD severity (not shown). Based on 288 the strong influence of patient sex in the determination of plasma metabolite composition 289 (15), we believe that untargeted metabolomic studies may reveal further examples of such 290 metabolic changes that may be key to explanations of divergent disease phenotypes 291 between men and women. The fact that NAFLD is associated with changes in sex hormone 292 concentrations such as sex hormone binding globulin or testosterone, may provide a 293 potential explanation for sex susceptibility differences in NAFLD (32,33). Our data suggest 294 that BCAA measurement could be useful to evaluate disease progression in women and thus 295 be integrated in future non-invasive scores for NAFLD screening. However, large-scale 296 prospective studies with paired biopsies, such as currently ongoing clinical trials, will be 297 necessary to fully test the applicability of such a strategy.

298

299 Whether BCAA are markers or actors in NAFLD progression remains unclear. BCAA 300 metabolism plays an important role in many cellular functions and signaling (34). Plasma 301 BCAA levels reflect net differences in BCAA intake from the diet and their catabolism. The 302 first two steps of BCAA catabolism, catalyzed by BCAT (branched-chain amino acid 303 transaminase, in peripheral tissues) and BCKDH (branched-chain α -ketoacid dehydrogenase,

304 mainly in liver), respectively, are reduced in insulin resistance and obesity (35). The 305 reductions in these enzymatic activities limit the availability of BCAA-derived succinyl-CoA 306 and acetyl-CoA, which would otherwise feed the TCA cycle. The combination of these 307 changes could lead to lower hepatic mitochondrial activity. Interestingly, BCAA (especially 308 leucine) are well-known activators of the mTOR pathway (36–38), whose activation may 309 further drive insulin resistance and abnormal lipid accumulation. In line, feeding obese mice 310 with BCAA was associated with chronic phosphorylation of mTOR and enhanced liver injury 311 in this model (39). However, the involvement of BCAA in liver injury in humans appears to 312 be inconsistent with these preclinical models. For example, in patients with cirrhosis, long-313 term oral BCAA supplementation reduced liver injury and related complications (40–42). 314 Consequently, elevated plasma BCAA in humans may be a protective mechanism to reduce 315 further hepatic injury in a chronic setting.

316

Although our findings do not prove causality of the observed association, they highlight that patient sex must be considered when developing metabolite-based diagnostic/evaluation scores for NAFLD.

320

321

322 Acknowledgements.

We thank Magali Defevre, Laurence George and Alain Kerckhove (Centre de Biologie et
 Pathologie Génétique, CHU Lille, France) for help with plasma amino acid measurements.
 325

- 326 **References**
- 327
- 328 Haas JT, Francque S, Staels B. Pathophysiology and Mechanisms of Nonalcoholic 1. 329 Fatty Liver Disease. Annu. Rev. Physiol. 2016;78(1):181-205. Kleiner DE, Brunt EM, Wilson LA, Behling C, Guy C, Contos M, Cummings O, 330 2. Yeh M, Gill R, Chalasani N, Neuschwander-Tetri BA, Diehl AM, Dasarathy S, Terrault 331 332 N, Kowdley K, Loomba R, Belt P, Tonascia J, Lavine JE, Sanyal AJ. Association of 333 Histologic Disease Activity With Progression of Nonalcoholic Fatty Liver Disease. JAMA 334 Netw. Open 2019;2(10). 335 Lonardo A, Nascimbeni F, Ballestri S, Fairweather D, Win S, Than TA, 3. 336 Abdelmalek MF, Suzuki A. Sex Differences in Nonalcoholic Fatty Liver Disease: State of 337 the Art and Identification of Research Gaps. Hepatology 2019;70(4):1457-1469. 338 4. Villanueva-Ortega E, Garcés-Hernández MJ, Herrera-Rosas A, López-Alvarenga JC, Laresgoiti-Servitje E, Escobedo G, Queipo G, Cuevas-Covarrubias S, 339 340 Garibay-Nieto GN. Gender-specific differences in clinical and metabolic variables 341 associated with NAFLD in a Mexican pediatric population. Ann. Hepatol. 2019;18(5):693-342 700. 343 Kleiner DE, Brunt EM, Van Natta M, Behling C, Contos MJ, Cummings OW, 5. Ferrell LD, Liu Y-C, Torbenson MS, Unalp-Arida A, Yeh M, McCullough AJ, Sanyal 344 345 AJ. Design and validation of a histological scoring system for nonalcoholic fatty liver 346 disease. Hepatology 2005;41(6):1313-1321. 347 Bedossa P. Current histological classification of NAFLD: strength and limitations. 6. 348 Hepatol. Int. 2013;7(2):765-770. 349 Rousselet M-C, Michalak S, Dupré F, Croué A, Bedossa P, Saint- André J-P, 7. 350 **Calès P.** Sources of variability in histological scoring of chronic viral hepatitis. *Hepatology* 351 2005;41(2):257-264. 352 Gawrieh S, Knoedler DM, Saeian K, Wallace JR, Komorowski RA. Effects of 8. interventions on intra- and interobserver agreement on interpretation of nonalcoholic fatty 353 354 liver disease histology. Ann. Diagn. Pathol. 2011;15(1):19-24. 355 Laferrère B, Reilly D, Arias S, Swerdlow N, Gorroochurn P, Bawa B, Bose M, 9. Teixeira J, Stevens RD, Wenner BR, Bain JR, Muehlbauer MJ, Haqq A, Lien L, Shah 356 SH, Svetkey LP, Newgard CB. Differential Metabolic Impact of Gastric Bypass Surgery 357 358 Versus Dietary Intervention in Obese Diabetic Subjects Despite Identical Weight Loss. Sci. 359 Transl. Med. 2011;3(80):80re2. 360 She P, Van Horn C, Reid T, Hutson SM, Cooney RN, Lynch CJ. Obesity-related 10. 361 elevations in plasma leucine are associated with alterations in enzymes involved in branched 362 chain amino acid (BCAA) metabolism. Am. J. Physiol. Endocrinol. Metab. 363 2007;293(6):E1552-E1563. 364 Newgard CB, An J, Bain JR, Muehlbauer MJ, Stevens RD, Lien LF, Haqq AM, 11. Shah SH, Arlotto M, Slentz CA, Rochon J, Gallup D, Ilkayeva O, Wenner BR, Yancy 365 WE, Eisenson H, Musante G, Surwit R, Millington DS, Butler MD, Svetkey LP. A 366 367 Branched-Chain Amino Acid-Related Metabolic Signature that Differentiates Obese and 368 Lean Humans and Contributes to Insulin Resistance. Cell Metab. 2009;9(4):311–326. Lake AD, Novak P, Shipkova P, Aranibar N, Robertson DG, Reily MD, Lehman-369 12. 370 McKeeman LD, Vaillancourt RR, Cherrington NJ. Branched chain amino acid 371 metabolism profiles in progressive human nonalcoholic fatty liver disease. Amino Acids 372 2015;47(3):603-615. 373 13. Hoyles L, Fernández-Real J-M, Federici M, Serino M, Abbott J, Charpentier J, 374 Heymes C, Luque JL, Anthony E, Barton RH, Chilloux J, Myridakis A, Martinez-Gili L, Moreno-Navarrete JM, Benhamed F, Azalbert V, Blasco-Baque V, Puig J, Xifra G, 375 376 Ricart W, Tomlinson C, Woodbridge M, Cardellini M, Davato F, Cardolini I, Porzio O,

377 Gentilieschi P, Lopez F, Foufelle F, Butcher SA, Holmes E, Nicholson JK, Postic C, Burcelin R, Dumas M-E. Molecular Phenomics and Metagenomics of Hepatic Steatosis in 378 379 Non-Diabetic Obese Women. Nat. Med. 2018;24(7):1070-1080. 380 Gaggini M, Carli F, Rosso C, Buzzigoli E, Marietti M, Latta VD, Ciociaro D, 14. Abate ML, Gambino R, Cassader M, Bugianesi E, Gastaldelli A. Altered amino acid 381 382 concentrations in NAFLD: Impact of obesity and insulin resistance. Hepatology 383 2018;67(1):145-158. 384 Krumsiek J, Mittelstrass K, Do KT, Stückler F, Ried J, Adamski J, Peters A, 15. Illig T, Kronenberg F, Friedrich N, Nauck M, Pietzner M, Mook-Kanamori DO, Suhre 385 K, Gieger C, Grallert H, Theis FJ, Kastenmüller G. Gender-specific pathway differences 386 387 in the human serum metabolome. *Metabolomics* 2015;11(6):1815–1833. 388 Ballestri S, Nascimbeni F, Baldelli E, Marrazzo A, Romagnoli D, Lonardo A. 16. 389 NAFLD as a Sexual Dimorphic Disease: Role of Gender and Reproductive Status in the 390 Development and Progression of Nonalcoholic Fatty Liver Disease and Inherent 391 Cardiovascular Risk. Adv. Ther. 2017;34(6):1291-1326. 392 Li L, Liu D-W, Yan H-Y, Wang Z-Y, Zhao S-H, Wang B. Obesity is an 17. 393 independent risk factor for non-alcoholic fatty liver disease: evidence from a meta-analysis of 394 21 cohort studies. Obes. Rev. 2016;17(6):510-519. Koves TR, Ussher JR, Noland RC, Slentz D, Mosedale M, Ilkayeva O, Bain J, 395 18. Stevens R, Dyck JRB, Newgard CB, Lopaschuk GD, Muoio DM. Mitochondrial Overload 396 397 and Incomplete Fatty Acid Oxidation Contribute to Skeletal Muscle Insulin Resistance. Cell 398 Metab. 2008;7(1):45-56. Zhou M, Jing S, Wu C-Y, Shu L, Dong W, Liu Y, Chen M, Wynn RM, Wang J, 399 19. 400 Wang J, Gui W-J, Qi X, Lusis AJ, Li Z, Wang W, Ning G, Yang X, Chuang DT, Wang Y. Sun H. Targeting BCAA Catabolism to Treat Obesity-Associated Insulin Resistance. 401 402 Diabetes 2019;68(9):1730-1746. 403 Neinast MD, Jang C, Hui S, Murashige DS, Chu Q, Morscher RJ, Li X, Zhan L, 20. 404 White E, Anthony TG, Rabinowitz JD, Arany Z. Quantitative Analysis of the Whole-405 Body Metabolic Fate of Branched-Chain Amino Acids. Cell Metab. 2019;29(2):417-429.e4. 406 Francque S, Verrijken A, Caron S, Prawitt J, Paumelle R, Derudas B, Lefebvre 21. 407 P, Taskinen M-R, Van Hul W, Mertens I, Hubens G, Van Marck E, Michielsen P, Van 408 **Gaal L, Staels B.** PPAR α gene expression correlates with severity and histological treatment 409 response in patients with non-alcoholic steatohepatitis. J. Hepatol. 2015;63(1):164-173. 410 Verrijken A, Beckers S, Francque S, Hilden H, Caron S, Zegers D, Ruppert M, 22. 411 Hubens G, Marck EV, Michielsen P, Staels B, Taskinen M-R, Hul WV, Gaal LV. A gene 412 variant of PNPLA3, but not of APOC3, is associated with histological parameters of NAFLD 413 in an obese population. *Obesity* 2013;21(10):2138–2145. 414 Levy JC, Matthews DR, Hermans MP. Correct Homeostasis Model Assessment 23. 415 (HOMA) Evaluation Uses the Computer Program. Diabetes Care 1998;21(12):2191-2192. 416 Chalasani N, Younossi Z, Lavine JE, Charlton M, Cusi K, Rinella M, Harrison 24. 417 SA, Brunt EM, Sanyal AJ. The diagnosis and management of nonalcoholic fatty liver 418 disease: Practice guidance from the American Association for the Study of Liver Diseases. 419 Hepatology 2018;67(1):328-357. 420 25. EASL-EASD-EASO Clinical Practice Guidelines for the management of non-421 alcoholic fatty liver disease. J. Hepatol. 2016;64(6):1388–1402. Filee R, Schoos R, Boemer F. Evaluation of Physiological Amino Acids Profiling by 422 26. 423 Tandem Mass Spectrometry. JIMD Rep. 2013;13:119–128. 424 27. Lake AD, Novak P, Shipkova P, Aranibar N, Robertson D, Reily MD, Lu Z, 425 Lehman-McKeeman LD, Cherrington NJ. Decreased Hepatotoxic Bile Acid Composition 426 and Altered Synthesis in Progressive Human Nonalcoholic Fatty Liver Disease. Toxicol. 427 Appl. Pharmacol. 2013;268(2):132-140.

428 28. Puri P, Daita K, Joyce A, Mirshahi F, Santhekadur PK, Cazanave S, Luketic 429 VA, Siddiqui MS, Boyett S, Min H-K, Kumar DP, Kohli R, Zhou H, Hylemon PB, Contos MJ, Idowu M, Sanyal AJ. The presence and severity of nonalcoholic steatohepatitis 430 431 is associated with specific changes in circulating bile acids. *Hepatology* 67(2):534–548. 432 Legry V, Francque S, Haas JT, Verrijken A, Caron S, Chávez-Talavera O, 29. 433 Vallez E, Vonghia L, Dirinck E, Verhaegen A, Kouach M, Lestavel S, Lefebvre P, Van 434 Gaal L, Tailleux A, Paumelle R, Staels B. Bile Acid Alterations Are Associated With 435 Insulin Resistance, but Not With NASH, in Obese Subjects. J. Clin. Endocrinol. Metab. 436 2017;102(10):3783-3794. 437 Tapper EB, Krajewski K, Lai M, Challies T, Kane R, Afdhal N, Lau D. Simple 30. 438 non-invasive biomarkers of advanced fibrosis in the evaluation of non-alcoholic fatty liver 439 disease. Gastroenterol. Rep. 2014;2(4):276-280. 440 31. Wang Z, Xu M, Hu Z, Hultström M, Lai E. Sex-specific prevalence of fatty liver 441 disease and associated metabolic factors in Wuhan, south central China. Eur. J. 442 Gastroenterol. Hepatol. 2014;26(9):1015-1021. Sarkar M, Wellons M, Cedars MI, VanWagner L, Gunderson EP, Ajmera V, 443 32. 444 Torchen L, Siscovick D, Carr JJ, Terry JG, Rinella M, Lewis CE, Terrault N. 445 Testosterone Levels in Pre-Menopausal Women are Associated With Nonalcoholic Fatty 446 Liver Disease in Midlife. Am. J. Gastroenterol. 2017;112(5):755-762. 447 Sarkar M, VanWagner LB, Terry JG, Carr JJ, Rinella M, Schreiner PJ, Lewis 33. 448 CE, Terrault N. Sex Hormone–Binding Globulin Levels in Young Men Are Associated 449 With Nonalcoholic Fatty Liver Disease in Midlife. Am. J. Gastroenterol. 2019;114(5):758-450 763. 451 34. Zhang S, Zeng X, Ren M, Mao X, Qiao S. Novel metabolic and physiological 452 functions of branched chain amino acids: a review. J. Anim. Sci. Biotechnol. 2017;8(1). 453 Lynch CJ, Adams SH. Branched-chain amino acids in metabolic signalling and 35. 454 insulin resistance. Nat. Rev. Endocrinol. 2014;10(12):723-736. 455 36. Um SH, D'Alessio D, Thomas G. Nutrient overload, insulin resistance, and 456 ribosomal protein S6 kinase 1, S6K1. Cell Metab. 2006;3(6):393-402. 457 Krebs M, Brunmair B, Brehm A, Artwohl M, Szendroedi J, Nowotny P, Roth E, 37. 458 Fürnsinn C, Promintzer M, Anderwald C, Bischof M, Roden M. The Mammalian Target 459 of Rapamycin Pathway Regulates Nutrient-Sensitive Glucose Uptake in Man. Diabetes 460 2007;56(6):1600-1607. 461 Wolfson RL, Chantranupong L, Saxton RA, Shen K, Scaria SM, Cantor JR, 38. 462 Sabatini DM. Sestrin2 is a leucine sensor for the mTORC1 pathway. Science 463 2016;351(6268):43-48. 464 Zhang F, Zhao S, Yan W, Xia Y, Chen X, Wang W, Zhang J, Gao C, Peng C, 39. 465 Yan F, Zhao H, Lian K, Lee Y, Zhang L, Lau WB, Ma X, Tao L. Branched Chain Amino Acids Cause Liver Injury in Obese/Diabetic Mice by Promoting Adipocyte Lipolysis and 466 467 Inhibiting Hepatic Autophagy. EBioMedicine 2016;13:157–167. Marchesini G, Bianchi G, Merli M, Amodio P, Panella C, Loguercio C, Rossi 468 40. 469 Fanelli F, Abbiati R. Nutritional supplementation with branched-chain amino acids in 470 advanced cirrhosis: a double-blind, randomized trial. Gastroenterology 2003;124(7):1792-471 1801. 472 41. Muto Y, Sato S, Watanabe A, Moriwaki H, Suzuki K, Kato A, Kato M, 473 Nakamura T, Higuchi K, Nishiguchi S, Kumada H. Effects of Oral Branched-Chain 474 Amino Acid Granules on Event-Free Survival in Patients With Liver Cirrhosis. Clin. 475 Gastroenterol. Hepatol. 2005;3(7):705-713. Fukui H, Saito H, Ueno Y, Uto H, Obara K, Sakaida I, Shibuya A, Seike M, 476 42. 477 Nagoshi S, Segawa M, Tsubouchi H, Moriwaki H, Kato A, Hashimoto E, Michitaka K, 478 Murawaki T, Sugano K, Watanabe M, Shimosegawa T. Evidence-based clinical practice

- 479 guidelines for liver cirrhosis 2015. J. Gastroenterol. 2016;51(7):629–650.

482 Figure 1. Plasma leucine and isoleucine but not valine are positively correlated to NAFLD stage.

483 Plasma BCAA concentrations according to NAFLD stage in Healthy (n=23), NAFL (n=30), NASH (n=30)

484 and NASH-F (n = 29) patients. Data are expressed as median \pm IQR. Rho coefficients and p-values

- 485 are obtained using the Spearman Rank test.
- 486

487 Figure 2. Plasma BCAA concentrations are strongly associated with sex.

Spearman correlations between plasma BCAA concentrations and NAFLD stage, sex, age, BMI and HOMA-IR and HOMA2-IR in the study cohort (n=112). Colors and area of circles reflect the Spearman rho values (red for positive, blue for inverse correlations). Only correlations with significant p-value (p<0.05) were represented.

492

493 Figure 3. Plasma BCAA are affected by sex.

494 BCAA concentrations in women (n=59) and men (n=53). Data are expressed as median \pm IQR. P-

495 values are obtained using the Mann-Whitney test.

496

497 Figure 4. Plasma BCAA variations related to NAFLD stage are sex dependent.

A to C: BCAA concentrations according to NAFLD stage for men in Healthy (n=8), NAFL (n=15), NASH (n=15) and NASH-F (n=15) patients. **D to F**: BCAA concentrations according to NAFLD stage for women in Healthy (n=15), NAFL (n=15), NASH (n=15) and NASH-F (n=14) patients. Data are expressed as median ± IQR. Rho coefficients and p-values are obtained using the Spearman Rank test.

503

504 Figure 5. Association between plasma BCAA concentrations and liver histological characteristics

505 are sex dependent.

506 Spearman correlations between plasma BCAA concentrations and liver histological characteristics in 507 the men (n=53) and the women (n=59) patients. Colors and area of circles reflect the

508	Spearman rho values (red for positive, blue for inverse correlations). P-values are obtained by
509	Spearman rank test, *: p-value<0.05, **, p-value < 0.01, ***P < 0.001, **** <p-value<0.0001.< td=""></p-value<0.0001.<>
510	
511	Figure 6. The association of plasma BCAA concentrations with fibrosis is sex dependent.
512	A to C: BCAA concentrations according to fibrosis stage (low fibrosis, F0-1, n=36 and high fibrosis
513	F≥2, n=17) in men group. D to F : BCAA concentrations according to fibrosis stage (low fibrosis, F0-1,
514	n=36 and high fibrosis F≥2, n=17) in women group. Data are expressed as median ± IQR. P-values are
515	obtained using the Mann-Whitney test.
516	
517	Table 1. Clinical, biological and liver histological characteristics of patients according to NAFLD
518	stage.
519	Data are expressed as mean ± standard deviation. P-values are obtained using the Chi-square
520	test for qualitative data and the ANOVA test for quantitative data. P-values <0.05 are considered
521	statistically significant (bold).
522	
523	Table 2. Clinical, biological and liver histological characteristics of the patients according to sex and
524	NAFLD stage.
525	Data are expressed as mean \pm standard deviation. P-values are obtained using the Chi-square test
526	for qualitative data and the ANOVA test for quantitative data. P-values <0.05 are considered
527	statistically significant (bold).
528	
529	Table 3. Clinical, biological and liver histological characteristics of the patients according to sex.
530	Data are expressed as mean \pm standard deviation. P-values are obtained using the Chi-square test
531	for qualitative data and the Mann-Whitney test for quantitative data. P-values <0.05 are considered
532	statistically significant (bold).
533	

Figure 1. Plasma leucine and isoleucine but not valine are positively correlated to NAFLD stage.



540 Figure 2. Plasma BCAA concentrations are strongly associated with sex.



- 546 Figure 3. Plasma BCAA are affected by sex.



- 551 Figure 4. Plasma BCAA variations related to NAFLD stage are sex dependent.





- 556 Figure 5. Association between plasma BCAA concentrations and liver histological characteristics
- 557 are sex dependent.



Men

Women

561 Figure 6. The association of plasma BCAA concentrations with fibrosis is sex dependent.



569	Table 1. Clinical, biological and liver histological characteristics of patients according to NAFLD
570	stage.

NAFLD Stage	All patients	Healthy	NAFL	NASH	NASH-F	p-value
n	112	23	30	30	29	
Sex (Women/Men)	59/53	15/8	15/15	15/15	14/15	0.61
Age (years)	42.6 ± 12.5	40 ± 12	43 ± 11	43 ± 13	43 ± 13	0.75
BMI (kg/m²)	40.5 ± 6.5	38 ± 4.9	42 ± 6.5	39 ± 4.5	41 ± 5.7	0.11
Fasting Plasma Glucose	86 ± 10	85 ± 9.9	86 ± 13.6	87 ± 11.9	89 ± 8.8	0.23
(mg/dL)						
HbA ₁ c (%)	5.6 ± 0.5	5.4 ± 0.3	5.6 ± 0.4	5.6 ± 0.4	5.5 ± 0.6	0.32
HOMA-IR	4.1 ± 2.8	4.2 ± 4.7	3.5 ± 1.8	4.0 ±2.4	4.3 ± 1.5	0.32
НОМА2-В (%)	181 ± 69	177 ± 65	186 ± 80	185 ± 73	174 ± 58	0.82
HOMA2-S (%)	57 ± 36	64 ± 46	48 ± 24	58 ± 28	60 ± 37	0.95
HOMA2-IR	2.3 ± 1.4	1.9 ± 0.9	2.8 ± 2	2.3 ± 1.2	2.1 ± 0.8	0.64
Steatosis Grade (0,1,2,3)	23/39/25/25	23/0/0/0	0/22/8/0	0/12/6/12	0/5/11/13	<0.0001
Ballooning (0,1,2)	43/46/23	23/0/0	20/7/3	0/18/12	0/21/8	<0.0001
Lobular Inflammation	41/44/21/6	23/0/0/0	18/10/2/0	0/20/8/2	0/14/11/4	<0.0001
(0,1,2,3)						
Fibrosis Stage (0,1,2,3,4)	69/12/23/8/0	23/0/0/0/0	26/2/2/0/0	20/10/0/0/0	0/0/21/8/0	<0.0001
572						
F7 0						

589	Table 2. Clinical, biological and liver histological characteristics of the patients according to sex and
590	NAFLD stage.

wen	Healthy	NAFL	NASH	NASH-F	p-value
n	8	15	15	15	
Age	38 ± 15	45 ± 11	41 ± 13	40 ± 11	0.48
BMI (kg/m²)	39 ± 4.6	41 ± 8.5	38 ± 4.6	39 ± 4.1	0.38
Fasting Plasma Glucose	89 ± 12	88 ± 14	85 ± 6	89 ± 9	0.99
(mg/dL)					
HbA1c (%)	5.5 ± 0.3	5.6 ± 0.5	5.6 ± 0.3	4.9 ± 0.3	0.99
HOMA-IR	5.8 ± 7.3	3.4 ± 1.2	3.6 ± 2.2	4 ± 1.3	0.95
НОМА2-В (%)	169 ± 33	162 ± 59	216 ± 79	171 ± 59	0.51
HOMA2-S (%)	56 ± 21	55 ± 26	51 ± 35	56 ± 23	0.94
HOMA2-IR	1.9 ± 0.6	2.3 ± 4.6	2.6 ± 1.2	2.1 ± 0.8	0.91
Steatosis Grade (0,1,2,3)	8/0/0/0	0/11/4/0	0/7/3/5	0/1/6/8	<0.0001
Ballooning (0,1,2)	8/0/0	7/6/2	0/10/5	0/11/4	<0.0001
Lobular Inflammation	8/0/0/0	11/3/1/0	0/10/4/1	0/8/5/2	<0.0001
(0,1,2,3)					
Fibrosis Stage (0,1,2,3,4)	8/0/0/0/0	12/1/2/0/0	10/5/0/0/0	0/0/11/4/0	<0.0001
Women	Healthy	NAFL	NASH	NASH-F	p-value
n	15	15	15	14	
n Age	15 41 ± 11	15 42 ± 13	15 46 ± 14	14 45 ± 15	0.28
n Age BMI (kg/m²)	15 41 ± 11 38 ± 5.2	15 42 ± 13 42 ± 4.8	$ 15 46 \pm 14 39 \pm 4.4 $	14 45 ± 15 42 ± 6.8	0.28 0.13
n Age BMI (kg/m²)	15 41 ± 11 38 ± 5.2	15 42 ± 13 42 ± 4.8	15 46 ± 14 39 ± 4.4	14 45 ± 15 42 ± 6.8	0.28 0.13
n Age BMI (kg/m ²) Fasting Plasma Glucose	15 41 ± 11 38 ± 5.2 83 ± 8	15 42 ± 13 42 ± 4.8 86 ± 13	$ 15 46 \pm 14 39 \pm 4.4 89 \pm 16 $	14 45 ± 15 42 ± 6.8 90 ± 9	0.28 0.13 0.16
n Age BMI (kg/m ²) Fasting Plasma Glucose (mg/dL)	15 41 ± 11 38 ± 5.2 83 ± 8	15 42 ± 13 42 ± 4.8 86 ± 13	$ 15 46 \pm 14 39 \pm 4.4 89 \pm 16 $	14 45 ± 15 42 ± 6.8 90 ± 9	0.28 0.13 0.16
n Age BMI (kg/m ²) Fasting Plasma Glucose (mg/dL) HbA ₁ c (%)	$ \begin{array}{r} 15 \\ 41 \pm 11 \\ 38 \pm 5.2 \\ \\ 83 \pm 8 \\ 5.4 \pm 0.4 \\ \end{array} $	$ 15 42 \pm 13 42 \pm 4.8 86 \pm 13 5.6 \pm 0.4 $	$ 15 46 \pm 14 39 \pm 4.4 89 \pm 16 5.6 \pm 0.6 $	$ \begin{array}{r} 14 \\ 45 \pm 15 \\ 42 \pm 6.8 \\ 90 \pm 9 \\ 5.6 \pm 0.8 \\ \end{array} $	0.28 0.13 0.16 0.25
n Age BMI (kg/m ²) Fasting Plasma Glucose (mg/dL) HbA ₁ c (%) HOMA-IR	$ \begin{array}{r} 15 \\ 41 \pm 11 \\ 38 \pm 5.2 \\ \\ 83 \pm 8 \\ 5.4 \pm 0.4 \\ 3.2 \pm 1.6 \\ \end{array} $	$ \begin{array}{r} 15 \\ 42 \pm 13 \\ 42 \pm 4.8 \\ \\ 86 \pm 13 \\ 5.6 \pm 0.4 \\ 3.6 \pm 2.1 \\ \end{array} $	$ 15 46 \pm 14 39 \pm 4.4 89 \pm 16 5.6 \pm 0.6 4.4 \pm 2.7 $	$ \begin{array}{r} 14 \\ 45 \pm 15 \\ 42 \pm 6.8 \\ \hline 90 \pm 9 \\ 5.6 \pm 0.8 \\ 4.7 \pm 1.8 \\ \end{array} $	0.28 0.13 0.16 0.25 0.06
n Age BMI (kg/m ²) Fasting Plasma Glucose (mg/dL) HbA ₁ c (%) HOMA-IR HOMA-2B (%)	$ \begin{array}{r} 15 \\ 41 \pm 11 \\ 38 \pm 5.2 \\ \\ 83 \pm 8 \\ 5.4 \pm 0.4 \\ 3.2 \pm 1.6 \\ 181 \pm 78 \\ \end{array} $	$ \begin{array}{r} 15 \\ 42 \pm 13 \\ 42 \pm 4.8 \\ \\ 86 \pm 13 \\ 5.6 \pm 0.4 \\ 3.6 \pm 2.1 \\ 211 \pm 93 \\ \end{array} $	$ \begin{array}{r} 15 \\ 46 \pm 14 \\ 39 \pm 4.4 \\ \\ 89 \pm 16 \\ 5.6 \pm 0.6 \\ 4.4 \pm 2.7 \\ 154 \pm 52 \\ \end{array} $	$ \begin{array}{r} 14 \\ 45 \pm 15 \\ 42 \pm 6.8 \\ \hline 90 \pm 9 \\ 5.6 \pm 0.8 \\ 4.7 \pm 1.8 \\ 177 \pm 59 \\ \end{array} $	0.28 0.13 0.16 0.25 0.06 0.42
n Age BMI (kg/m ²) Fasting Plasma Glucose (mg/dL) HbA ₁ c (%) HOMA-IR HOMA-2B (%) HOMA-2S (%)	$ \begin{array}{r} 15 \\ 41 \pm 11 \\ 38 \pm 5.2 \\ \\ 83 \pm 8 \\ 5.4 \pm 0.4 \\ 3.2 \pm 1.6 \\ 181 \pm 78 \\ 68 \pm 56 \\ \end{array} $	$ \begin{array}{r} 15 \\ 42 \pm 13 \\ 42 \pm 4.8 \\ \hline 5.6 \pm 0.4 \\ 3.6 \pm 2.1 \\ 211 \pm 93 \\ 41 \pm 21 \\ \end{array} $	$ \begin{array}{r} 15 \\ 46 \pm 14 \\ 39 \pm 4.4 \\ \\ 89 \pm 16 \\ 5.6 \pm 0.6 \\ 4.4 \pm 2.7 \\ 154 \pm 52 \\ 65 \pm 41 \\ \end{array} $	$ \begin{array}{r} 14 \\ 45 \pm 15 \\ 42 \pm 6.8 \\ \hline 90 \pm 9 \\ 5.6 \pm 0.8 \\ 4.7 \pm 1.8 \\ 177 \pm 59 \\ 64 \pm 48 \\ \end{array} $	0.28 0.13 0.16 0.25 0.06 0.42 0.85
n Age BMI (kg/m ²) Fasting Plasma Glucose (mg/dL) HbA ₁ c (%) HOMA-IR HOMA-2B (%) HOMA-2S (%) HOMA2-IR	$ \begin{array}{r} 15 \\ 41 \pm 11 \\ 38 \pm 5.2 \\ \\ 83 \pm 8 \\ 5.4 \pm 0.4 \\ 3.2 \pm 1.6 \\ 181 \pm 78 \\ 68 \pm 56 \\ 2.0 \pm 1.1 \\ \end{array} $	$ \begin{array}{r} 15 \\ 42 \pm 13 \\ 42 \pm 4.8 \\ \hline 86 \pm 13 \\ 5.6 \pm 0.4 \\ 3.6 \pm 2.1 \\ 211 \pm 93 \\ 41 \pm 21 \\ 3.3 \pm 2.4 \\ \end{array} $	$ \begin{array}{r} 15 \\ 46 \pm 14 \\ 39 \pm 4.4 \\ \overline{} \\ 89 \pm 16 \\ 5.6 \pm 0.6 \\ 4.4 \pm 2.7 \\ 154 \pm 52 \\ 65 \pm 41 \\ 2 \pm 1.1 \\ \end{array} $	$ \begin{array}{r} 14 \\ 45 \pm 15 \\ 42 \pm 6.8 \\ \hline 90 \pm 9 \\ 5.6 \pm 0.8 \\ 4.7 \pm 1.8 \\ 177 \pm 59 \\ 64 \pm 48 \\ 2.1 \pm 0.8 \\ \end{array} $	0.28 0.13 0.16 0.25 0.06 0.42 0.85 0.54
n Age BMI (kg/m ²) Fasting Plasma Glucose (mg/dL) HbA ₁ c (%) HOMA-IR HOMA-2B (%) HOMA-2S (%) HOMA2-IR	$ \begin{array}{r} 15 \\ 41 \pm 11 \\ 38 \pm 5.2 \\ \\ 83 \pm 8 \\ 5.4 \pm 0.4 \\ 3.2 \pm 1.6 \\ 181 \pm 78 \\ 68 \pm 56 \\ 2.0 \pm 1.1 \\ \end{array} $	$ \begin{array}{r} 15 \\ 42 \pm 13 \\ 42 \pm 4.8 \\ 86 \pm 13 \\ 5.6 \pm 0.4 \\ 3.6 \pm 2.1 \\ 211 \pm 93 \\ 41 \pm 21 \\ 3.3 \pm 2.4 \\ \end{array} $	$ \begin{array}{r} 15 \\ 46 \pm 14 \\ 39 \pm 4.4 \\ 89 \pm 16 \\ 5.6 \pm 0.6 \\ 4.4 \pm 2.7 \\ 154 \pm 52 \\ 65 \pm 41 \\ 2 \pm 1.1 \\ \end{array} $	$ \begin{array}{r} 14 \\ 45 \pm 15 \\ 42 \pm 6.8 \\ 90 \pm 9 \\ 5.6 \pm 0.8 \\ 4.7 \pm 1.8 \\ 177 \pm 59 \\ 64 \pm 48 \\ 2.1 \pm 0.8 \\ \end{array} $	0.28 0.13 0.16 0.25 0.06 0.42 0.85 0.54
n Age BMI (kg/m ²) Fasting Plasma Glucose (mg/dL) HbA ₁ c (%) HOMA-IR HOMA-2B (%) HOMA-2S (%) HOMA2-IR Menopause (no/yes)	$ \begin{array}{r} 15 \\ 41 \pm 11 \\ 38 \pm 5.2 \\ \\ 83 \pm 8 \\ 5.4 \pm 0.4 \\ 3.2 \pm 1.6 \\ 181 \pm 78 \\ 68 \pm 56 \\ 2.0 \pm 1.1 \\ \\ 13/2 \\ \end{array} $	$ \begin{array}{r} 15 \\ 42 \pm 13 \\ 42 \pm 4.8 \\ 86 \pm 13 \\ 5.6 \pm 0.4 \\ 3.6 \pm 2.1 \\ 211 \pm 93 \\ 41 \pm 21 \\ 3.3 \pm 2.4 \\ 12/3 \end{array} $	$ 15 46 \pm 14 39 \pm 4.4 89 \pm 16 5.6 \pm 0.6 4.4 \pm 2.7 154 \pm 52 65 \pm 41 2 \pm 1.1 9/6 $	$ \begin{array}{r} 14 \\ 45 \pm 15 \\ 42 \pm 6.8 \\ \hline 90 \pm 9 \\ 5.6 \pm 0.8 \\ 4.7 \pm 1.8 \\ 177 \pm 59 \\ 64 \pm 48 \\ 2.1 \pm 0.8 \\ \hline 9/5 \\ \end{array} $	0.28 0.13 0.16 0.25 0.06 0.42 0.85 0.54 0.31
n Age BMI (kg/m ²) Fasting Plasma Glucose (mg/dL) HbA ₁ c (%) HOMA-IR HOMA-2B (%) HOMA-2S (%) HOMA2-IR Menopause (no/yes)	$ \begin{array}{r} 15 \\ 41 \pm 11 \\ 38 \pm 5.2 \\ \\ 83 \pm 8 \\ 5.4 \pm 0.4 \\ 3.2 \pm 1.6 \\ 181 \pm 78 \\ 68 \pm 56 \\ 2.0 \pm 1.1 \\ \\ 13/2 \\ \end{array} $	$ \begin{array}{r} 15 \\ 42 \pm 13 \\ 42 \pm 4.8 \\ \hline 86 \pm 13 \\ 5.6 \pm 0.4 \\ 3.6 \pm 2.1 \\ 211 \pm 93 \\ 41 \pm 21 \\ 3.3 \pm 2.4 \\ \hline 12/3 \\ \end{array} $	$ \begin{array}{r} 15 \\ 46 \pm 14 \\ 39 \pm 4.4 \\ 89 \pm 16 \\ 5.6 \pm 0.6 \\ 4.4 \pm 2.7 \\ 154 \pm 52 \\ 65 \pm 41 \\ 2 \pm 1.1 \\ 9/6 \\ \end{array} $	$ \begin{array}{r} 14 \\ 45 \pm 15 \\ 42 \pm 6.8 \\ \hline 90 \pm 9 \\ 5.6 \pm 0.8 \\ 4.7 \pm 1.8 \\ 177 \pm 59 \\ 64 \pm 48 \\ 2.1 \pm 0.8 \\ \hline 9/5 \\ \end{array} $	0.28 0.13 0.16 0.25 0.06 0.42 0.85 0.54 0.31
n Age BMI (kg/m ²) Fasting Plasma Glucose (mg/dL) HbA ₁ c (%) HOMA-IR HOMA-2B (%) HOMA-2S (%) HOMA2-IR Menopause (no/yes) Steatosis Grade (0,1,2,3)	$ \begin{array}{r} 15 \\ 41 \pm 11 \\ 38 \pm 5.2 \\ \\ 83 \pm 8 \\ 5.4 \pm 0.4 \\ 3.2 \pm 1.6 \\ 181 \pm 78 \\ 68 \pm 56 \\ 2.0 \pm 1.1 \\ \\ 13/2 \\ 15/0/0/0 \\ \end{array} $	$ \begin{array}{r} 15 \\ 42 \pm 13 \\ 42 \pm 4.8 \\ 86 \pm 13 \\ 5.6 \pm 0.4 \\ 3.6 \pm 2.1 \\ 211 \pm 93 \\ 41 \pm 21 \\ 3.3 \pm 2.4 \\ 12/3 \\ 0/11/4/0 \\ \end{array} $	$ 15 46 \pm 14 39 \pm 4.4 89 \pm 16 5.6 \pm 0.6 4.4 \pm 2.7 154 \pm 52 65 \pm 41 2 \pm 1.1 9/6 0/5/3/7 $	$ \begin{array}{r} 14 \\ 45 \pm 15 \\ 42 \pm 6.8 \\ \hline 90 \pm 9 \\ 5.6 \pm 0.8 \\ 4.7 \pm 1.8 \\ 177 \pm 59 \\ 64 \pm 48 \\ 2.1 \pm 0.8 \\ \hline 9/5 \\ \hline 0/4/5/5 \\ \hline \end{array} $	0.28 0.13 0.16 0.25 0.06 0.42 0.85 0.54 0.31 <0.0001
n Age BMI (kg/m ²) Fasting Plasma Glucose (mg/dL) HbA ₁ c (%) HOMA-IR HOMA-2B (%) HOMA-2S (%) HOMA-2S (%) HOMA2-IR Menopause (no/yes) Steatosis Grade (0,1,2,3) Ballooning (0,1,2)	$ \begin{array}{r} 15 \\ 41 \pm 11 \\ 38 \pm 5.2 \\ \\ 83 \pm 8 \\ 5.4 \pm 0.4 \\ 3.2 \pm 1.6 \\ 181 \pm 78 \\ 68 \pm 56 \\ 2.0 \pm 1.1 \\ \\ 13/2 \\ \\ 15/0/0/0 \\ 15/0/0 \\ 15/0/0 \\ 15/0/0 \\ \end{array} $	$ \begin{array}{r} 15 \\ 42 \pm 13 \\ 42 \pm 4.8 \\ \\ 86 \pm 13 \\ 5.6 \pm 0.4 \\ 3.6 \pm 2.1 \\ 211 \pm 93 \\ 41 \pm 21 \\ 3.3 \pm 2.4 \\ \\ 12/3 \\ \\ 0/11/4/0 \\ 13/1/1 \\ \end{array} $	$ \begin{array}{r} 15 \\ 46 \pm 14 \\ 39 \pm 4.4 \\ \\ 89 \pm 16 \\ 5.6 \pm 0.6 \\ 4.4 \pm 2.7 \\ 154 \pm 52 \\ 65 \pm 41 \\ 2 \pm 1.1 \\ \\ 9/6 \\ \\ 0/5/3/7 \\ 0/8/7 \\ \end{array} $	$ \begin{array}{r} 14\\ 45 \pm 15\\ 42 \pm 6.8\\ \hline 90 \pm 9\\ \hline 5.6 \pm 0.8\\ 4.7 \pm 1.8\\ 177 \pm 59\\ 64 \pm 48\\ 2.1 \pm 0.8\\ \hline 9/5\\ \hline 0/4/5/5\\ 0/10/4\\ \end{array} $	0.28 0.13 0.16 0.25 0.06 0.42 0.85 0.54 0.54 (0.31) <0.0001
n Age BMI (kg/m ²) Fasting Plasma Glucose (mg/dL) HbA ₁ c (%) HOMA-IR HOMA-2B (%) HOMA-2B (%) HOMA-2S (%) HOMA2-IR Menopause (no/yes) Steatosis Grade (0,1,2,3) Ballooning (0,1,2) Lobular Inflammation	$ \begin{array}{r} 15 \\ 41 \pm 11 \\ 38 \pm 5.2 \\ \\ 83 \pm 8 \\ 5.4 \pm 0.4 \\ 3.2 \pm 1.6 \\ 181 \pm 78 \\ 68 \pm 56 \\ 2.0 \pm 1.1 \\ \\ 13/2 \\ 15/0/0/0 \\ 15/0/0 \\ 15/0/0 \\ 15/0/0 \\ 15/0/0 \\ 00 \\ 15/0/0 \\ 00 \\ 15/0/0 \\ 00 \\ 15/0/0 \\ 15/0 \\ 15/0$	$ \begin{array}{r} 15 \\ 42 \pm 13 \\ 42 \pm 4.8 \\ \\ 86 \pm 13 \\ 5.6 \pm 0.4 \\ 3.6 \pm 2.1 \\ 211 \pm 93 \\ 41 \pm 21 \\ 3.3 \pm 2.4 \\ \\ 12/3 \\ 0/11/4/0 \\ 13/1/1 \\ 7/7/1/0 \\ \end{array} $	$ \begin{array}{r} 15\\ 46 \pm 14\\ 39 \pm 4.4\\ \hline 89 \pm 16\\ \hline 5.6 \pm 0.6\\ 4.4 \pm 2.7\\ 154 \pm 52\\ 65 \pm 41\\ 2 \pm 1.1\\ \hline 9/6\\ \hline 0/5/3/7\\ 0/8/7\\ 0/10/4/1\\ \end{array} $	$ \begin{array}{r} 14\\ 45 \pm 15\\ 42 \pm 6.8\\ \hline 90 \pm 9\\ \hline 90 \pm 9\\ \hline 5.6 \pm 0.8\\ 4.7 \pm 1.8\\ 177 \pm 59\\ 64 \pm 48\\ 2.1 \pm 0.8\\ \hline 9/5\\ \hline 0/4/5/5\\ 0/10/4\\ 0/6/6/2\\ \end{array} $	0.28 0.13 0.16 0.25 0.06 0.42 0.85 0.54 0.54 0.31 <0.0001 <0.0001
n Age BMI (kg/m ²) Fasting Plasma Glucose (mg/dL) HbA ₁ c (%) HOMA-IR HOMA-2B (%) HOMA-2B (%) HOMA-2S (%) HOMA2-IR Menopause (no/yes) Steatosis Grade (0,1,2,3) Ballooning (0,1,2) Lobular Inflammation (0,1,2,3)	$ \begin{array}{r} 15 \\ 41 \pm 11 \\ 38 \pm 5.2 \\ \\ 83 \pm 8 \\ 5.4 \pm 0.4 \\ 3.2 \pm 1.6 \\ 181 \pm 78 \\ 68 \pm 56 \\ 2.0 \pm 1.1 \\ 13/2 \\ 15/0/0/0 \\ 15/0 \\ 15/0 \\ 15/0 \\ 1$	$ \begin{array}{r} 15 \\ 42 \pm 13 \\ 42 \pm 4.8 \\ 86 \pm 13 \\ 5.6 \pm 0.4 \\ 3.6 \pm 2.1 \\ 211 \pm 93 \\ 41 \pm 21 \\ 3.3 \pm 2.4 \\ 12/3 \\ 0/11/4/0 \\ 13/1/1 \\ 7/7/1/0 \\ 14 + 12 \\ 12/3 \\ $	$ \begin{array}{r} 15 \\ 46 \pm 14 \\ 39 \pm 4.4 \\ 89 \pm 16 \\ 5.6 \pm 0.6 \\ 4.4 \pm 2.7 \\ 154 \pm 52 \\ 65 \pm 41 \\ 2 \pm 1.1 \\ 9/6 \\ 0/5/3/7 \\ 0/8/7 \\ 0/10/4/1 \\ 10 + 10 + 10 + 10 + 10 + 10 + 10 $	$ \begin{array}{r} 14\\ 45 \pm 15\\ 42 \pm 6.8\\ \hline 90 \pm 9\\ \hline 5.6 \pm 0.8\\ 4.7 \pm 1.8\\ 177 \pm 59\\ 64 \pm 48\\ 2.1 \pm 0.8\\ \hline 9/5\\ \hline 0/4/5/5\\ 0/10/4\\ 0/6/6/2\\ \hline $	0.28 0.13 0.16 0.25 0.06 0.42 0.85 0.54 0.54 0.31 <0.0001 <0.0001 <0.0001

594 Table 3. Clinical, biological and liver histological characteristics of the patients according to sex

	Men	Women	p-value
n	53	59	
Age	41 ± 12	43 ± 12	0.89
BMI (kg/m²)	39 ± 5.5	40 ± 5.5	0.24
Facting Plasma Glucose (mg/dl)	87 + 9	86 + 10	0.29
HbA ₁ c (%)	5.5 ± 0.4	5.5 ± 0.5	0.58
HOMA-IR	4.2 ± 3.5	4.0 ± 2.1	0.77
НОМА2-В (%)	181 ± 65	181 ± 73	0.87
HOMA2-S (%)	54 ± 26	60 ± 44	0.93
HOMA2-IR	2.2 ± 1.2	2.3 ± 1.5	0.93
Steatosis Grade (0,1,2,3)	8/19/13/13	15/20/12/12	0.58
Ballooning (0,1,2)	15/27/11	28/19/12	0.08
Lobular Inflammation (0,1,2,3)	19/21/10/3	22/23/11/3	0.99
Fibrosis Stage (0,1,2,3,4)	30/6/13/4/0	39/6/10/4/0	0.74