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4 Running head: MICB, CMV and GVHD in HCT

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74 ABSTRACT

Graft-versus-host disease (GVHD) and cytomegalovirus (CMV)-related complications 75 leading causes of mortality after unrelated-donor hematopoietic cell are 76 transplantation (UD-HCT). The non-conventional MHC class I gene MICB, alike 77 MICA, encodes a stress-induced polymorphic NKG2D ligand. However, unlike MICA, 78 MICB interacts with the CMV-encoded UL16, which sequestrates MICB 79 intracellularly, leading to immune evasion. Here, we retrospectively analyzed the 80 impact of mismatches in MICB amino acid position 98 (MICB98), a key polymorphic 81 residue involved in UL16-binding, in 943 UD-HCT pairs who were allele-matched at 82 83 HLA-A, -B, -C, -DRB1, -DQB1 and MICA loci. HLA-DP typing was further available. MICB98 mismatches were significantly associated with an increased incidence of 84 acute (grade II-IV: HR, 1.20; 95% CI, 1.15 to 1.24; P < 0.001; grade III-IV: HR, 2.28; 85 95% CI, 1.56 to 3.34; P < 0.001) and chronic GVHD (HR, 1.21; 95% CI, 1.10 to 1.33; 86 P < 0.001). MICB98 matching significantly reduced the effect of CMV status on 87 overall mortality from a hazard ratio of 1.77 to 1.16. MICB98 mismatches showed a 88 GVHD-independent association with a higher incidence of CMV infection/reactivation 89 (HR, 1.84; 95% CI, 1.34 to 2.51; P < 0.001). Hence selecting a *MICB98*-matched 90 91 donor significantly reduces the GVHD incidence and lowers the impact of CMV status on overall survival. 92

93

94 **INTRODUCTION**

Unrelated-donor hematopoietic cell transplantation (HCT) is an established 95 treatment for a wide range of immunological and hematologic disorders, malignant or 96 otherwise ¹. Although more than 50,000 HCTs are performed annually worldwide ^{2, 3}, 97 adverse clinical outcomes occur frequently. One of the most common life-threatening 98 complications is graft-versus-host disease (GVHD), which greatly hampers the 99 successful outcome of this powerful and sometimes unique curative option. In GVHD, 100 the donor's immune cells attack the patient's organs and tissues, impairing their 101 ability to function and increasing the patient's susceptibility to infection. The 102 organs/tissues most frequently targeted are the skin, the gastrointestinal tract and the 103 liver. Despite the availability of effective immunosuppressive drugs, the incidence of 104 GVHD remains alarmingly high: up to 35% experience grade III-IV acute GVHD and 105 40% to 50% experience chronic GVHD ⁴⁻⁶. 106

Cytomegalovirus (CMV) infection/reactivation represents another leading 107 cause of morbidity and mortality in patients undergoing allogeneic HCT because it 108 frequently causes serious complications, e.g., pneumonia, hepatitis, gastroenteritis, 109 retinitis, and encephalitis ⁷⁻¹¹. Because of the immunosuppressive regimen, 110 allogeneic HCT patients are indeed at a higher risk for CMV infection and/or 111 reactivation. The incidence of CMV infection has been reported to vary between 40 112 and 80% in CMV seropositive allogeneic HCT patients not treated with anti-viral 113 prophylaxis drugs, which currently represents most of the allogeneic HCT recipients 114 ¹²⁻¹⁸. In seronegative patients receiving a transplant from a seropositive donor, the 115 rate of primo infection is approximately 30%¹². Despite the implementation of 116 prophylaxis, monitoring, and pre-emptive treatment of CMV reactivation/infection, 117 cases of CMV seropositivity of the donor and/or the recipient show decreased 118

survival rates compared to CMV-seronegative recipients who undergo allograft from 119 19 donors 16, New strategies **CMV-seronegative** for preventing CMV 120 reactivation/infection in transplant recipients therefore remain an important objective 121 for the improvement of allogeneic HCT. 122

Increasing the degree of human leukocyte antigen (HLA) matching is one of 123 the most important strategies to lower the risks of both GVHD and CMV infections. 124 The former is a direct consequence of better HLA-matching, whereas the latter is an 125 indirect effect due to the well-described association of CMV infection with GVHD 126 occurrence ^{20, 21}. However, even in genotypically HLA-matched donors and 127 128 recipients, the incidence of grade III-IV acute GVHD and CMV reactivation/infection can be as high as 30% and 80%, respectively ^{13, 22}. For CMV infection/reactivation, 129 other risk factors include age, source of stem cells, disease, and donor (D)/recipient 130 (R) CMV serological status ^{23, 24}. 131

The MHC-encoded non-conventional MHC class I chain-related (MIC) genes A 132 (MICA) and B (MICB) encode polymorphic cell surface proteins which bind to 133 NKG2D; an activating immune receptor expressed by cytotoxic T and NK cells ^{25, 26}. 134 This interaction is seminal in defense both against infections and malignancies. 135 Moreover, *MICB*^{27, 28} happens to be one of the most promising candidates to explain, 136 at least partially, GVHD and CMV complications that cannot be attributed to classical 137 HLA genes or the related *MICA* gene incompatibilities ²⁹⁻³¹. *MICB* is indeed highly 138 47 alleles 139 polymorphic, with reported to date (http://www.ebi.ac.uk/ipd/imgt/hla/stats.html). It encodes a cell-surface glycoprotein 140 up-regulated by cell stress ^{25, 32}. The gene is located 130 kb and 83 kb centromeric to 141 HLA-B and MICA, respectively, and was discovered by us over 20 years ago²⁵. 142 MICB is highly similar to MICA in terms of sequence (83% shared amino acid 143

sequence identity), linkage disequilibrium with HLA-B, protein structure (HLA-like 144 structure without association to ß₂-microglobulin) and constitutive expression pattern 145 (restricted to epithelial cells, fibroblasts, monocytes, dendritic cells and endothelial 146 cells) ^{26, 33, 34}. MICB is a ligand for the activating NKG2D receptor expressed on the 147 surface of cytotoxic CD8⁺ $\alpha\beta$ and $\gamma\delta$ T lymphocytes and natural killer cells ³⁵. 148 Interestingly, and in contrast to MICA, MICB binds the CMV protein UL16, which 149 sequestrates MICB intracellularly in an immune escape mechanism ³⁶. Different 150 MICB alleles are not equal with respect to binding to UL16. MICB*008 has been 151 shown to have a decreased binding capacity to UL16 compared to other alleles ³⁷. 152 *MICB*008* is characterized by a polymorphism at amino acid position 98, causing an 153 isoleucine (IIe) to methionine (Met) exchange in the α^2 domain of the MICB protein. 154 The variation IIe > Met is exclusively present in $MICB^*008$ and is the unique 155 156 polymorphic position that is in direct contact with UL16 through hydrophobic contacts (distance<4.0 Å) with leucine 161 of UL16 38 . 157

Several lines of evidence indicate that *MICB* could play a role in triggering 158 GVHD and/or modulating CMV infection/reactivation: (1) the localized expression in 159 epithelial cells of the gastrointestinal tract, whose damage during GVHD plays a 160 major pathophysiologic role in the amplification of systemic disease ³⁹; (2) the 161 common features with MICA that have repeatedly been shown to be involved in 162 GVHD ^{29, 30, 40-42}; and (3) the binding of MICB to the UL16 protein ³⁶. The present 163 study hence aims to show the effect of MICB matching at amino acid position 98, 164 representing about 6% of transplantations, on the outcome of unrelated donor HCT in 165 a cohort of 943 donor/recipient pairs matched for HLA-A, -B, -C, -DRB1, -DQB1, and 166 MICA. 167

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169 **PATIENTS AND METHODS**

170 STUDY DESIGN AND OVERSIGHT

This retrospective study was designed to test whether donor-recipient matching at 171 amino acid position 98 of the MICB protein (MICB98) improves the outcome of 172 unrelated HCT. Patients from six French and three Dutch centers and their donors 173 were included: the unrelated donors originated from national or international donor 174 registries. Genomic DNA samples and high-resolution HLA-A, -B, -C, -DRB1, -DQB1, 175 -DPB1 and MICA typing data were collected. Clinical information was made available 176 by the SFGM-TC and the HOVON Data Center from the EBMT (European group for 177 178 Blood and Marrow Transplantation) ProMISe patient database. All authors vouch for the accuracy and completeness of the results. This study, conducted under the 179 auspices of SFGM-TC and the Dutch-Belgian Cooperative Trial Group for 180 Hematology Oncology (HOVON), was approved by institutional review boards of the 181 participating centers and was performed according to the principles of the Declaration 182 of Helsinki. Written informed consent was obtained from all participants. 183

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185 PATIENTS AND DONORS

The study population consisted of 943 patients who underwent unrelated HCT for the treatment of blood disorders between 2005 and 2013. All patients received a first allogeneic transplant using bone marrow or peripheral blood stem cells, and donorrecipients were matched for 12 of the 12 possible alleles at *HLA-A*, *-B*, *-C*, *-DRB1*, *-DQB1*, and *MICA* loci (Table 1).

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192 MICB GENOTYPING AT AMINO ACID POSITION 98

The polymorphic nucleotide position 363 (C/G; rs3134900) causes an isoleucine (IIe) to methionine (Met) change at amino acid position 98 in the α 2 domain of the MICB protein. Both patients and unrelated donors were genotyped for this position by Sanger sequencing of *MICB's* exon 3, following previously described protocols ⁴³. The sequences were analyzed using Seqscape v2.6 (Life Technologies, USA) to assign genotypes.

199

200 **DEFINITIONS**

Grading of acute and chronic GVHD was performed according to the classification of 201 Glucksberg et al. ⁴⁴. For acute GVHD, severe corresponds to grades III and IV. CMV 202 positivity of the donor and/or the recipient was defined by the presence of anti-CMV 203 IgG in the serum of the donor and/or the recipient. CMV reactivation was defined as 204 the time from transplantation to the first CMV infection episode. In addition to clinical 205 206 examination, CMV infection/reactivation episodes were characterized at a molecular level by a viral load > 10^4 copies/ml as determined by quantitative PCR on whole 207 blood. Overall survival (OS) was defined as the time from transplantation to death by 208 any cause. Relapse-free survival (RFS) was defined as the time to relapse of primary 209 disease or death by any cause, whichever came first. Non-relapse mortality (NRM) 210 corresponds to mortality within the first complete remission of disease. Causes of 211 death unrelated to transplantation included deaths related to relapse, progression of 212 the original disease, secondary malignancy, and cell therapy (non-HCT). OS, RFS, 213 NRM, GVHD and CMV reactivation were censored at the time of the last follow-up. 214 Incidences of clinical outcomes were defined as the cumulative probability of the 215 outcomes at any given point. 216

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218 STATISTICAL ANALYSIS

219 After validating that the data meet requested assumptions, the distribution of each covariate between the MICB98 matched and mismatched groups was assessed by 220 Pearson's Chi square test or Fisher's exact test for small sample sizes. The 221 variances between the two groups were similar for the different variables assessed in 222 our models and statistical tests (average variances in the matched and mismatched 223 groups were 1.36 and 1.40, respectively). Multivariable competing risk regression 224 analyses were performed for acute GVHD II-IV, acute GVHD III-IV, chronic GVHD, 225 relapse, NRM and CMV reactivation, using an extended Fine and Grav model ⁴⁵⁻⁴⁷. 226 For OS and RFS, Cox proportional regression models were used ⁴⁸. Competing 227 events were defined as death without GVHD and relapse for GVHD endpoints (acute 228 and chronic GVHD); death from any cause other than transplantation for NRM; 229 230 relapse and death for CMV reactivation; and non-relapse mortality for relapse. All statistical models were adjusted for center effect and covariates defining the 231 European Society for Blood and Marrow Transplantation risk score: patient age, 232 disease stage at transplantation, time to transplantation, and donor-recipient sex 233 combination. In addition to these, the following relevant variables were included: 234 HLA-DPB1 matching (T-cell epitope matching level as defined by Fleischhauer et 235 al.⁴⁹), patient-donor serological status for cytomegalovirus, year of transplantation, 236 source of stem cells, conditioning regimen, GVHD prophylaxis, treatment with 237 antithymocyte globulin or Alemtuzumab, and disease category. Interactions between 238 patient-donor serological status for cytomegalovirus and matching at amino acid 239 position 98 of *MICB* were also assessed in the multivariable analyses. ^{50, 51}All models 240 were checked for interactions and proportional hazards assumptions. All statistical 241 analyses were conducted using the computing environment R⁵². 242

244 **RESULTS**

The demographics of the study population are shown in Table 1. The median 245 246 post-transplant follow-up was 36 months (mean: 37 months; range: 1 to 105 months), and the median patient age was 53 years (mean: 48 years; range: 1 to 73 years). 247 The patients suffered from both malignant and non-malignant diseases. Most 248 transplants were performed with non-myeloablative/reduced intensity conditioning 249 250 regimens (67%); in vivo T-cell depletion was performed in the majority of cases (73%), and peripheral blood was the main source for stem cells (79%). All 251 donor/patient pairs were fully typed at high resolution (2nd field) for HLA-A, -B, -C, -252 DRB1, -DQB1, -DPB1 and MICA²⁹ and were matched for 12 out of 12 alleles at HLA-253 A, -B, -C, -DRB1, -DQB1 and MICA loci. Among the 943 transplantations, 394 254 (41.8%) had non-permissive HLA-DPB1 mismatches. Fifty-six (5.9%) transplants 255 were MICB98 mismatched. The mismatch vectors of these 56 transplants were graft-256 versus-host (n=22), host-versus-graft (n=33) and bidirectional (n=1). Except for the 257 patient-donor CMV status, all relevant covariates for the analyzed clinical outcomes 258 were equally distributed in the MICB98 matched and -mismatched patients (Table 1). 259 Organ-specific sub-analyses showed that the MICB98 matching effect was more 260 important in the gut and the skin than in the liver (supplemental Figure 1). MICB98 261 mismatches were significantly associated with an increased incidence of acute 262 GVHD (hazard ratio (HR) for grades II-IV: 1.20; 95% CI, 1.15 to 1.24; P < 0.001; for 263 grades III-IV: 2.28; 95% CI, 1.56 to 3.34; P < 0.001) (Table 2). At day 100 post-HCT, 264 the cumulative incidences of severe (grades III-IV) acute GVHD in MICB98 265 mismatched vs. matched transplantations were 18.9% vs. 12.5%, respectively 266 (Figure 1A). Matching *MICB* at position 98 decreased the risk of chronic GVHD by 267

4% (40.9% vs. 36.9%) at 4 years post-transplantation (HR, 1.21; 95% Cl, 1.10 to 1.33; P < 0.001) (Table 2 and Figure 1B). In addition, *MICB98* mismatches were associated with a higher rate of relapse (HR, 1.42; 95% Cl, 1.05 to 1.93; P = 0.024).

Knowing that amino acid position 98 is involved in the binding of MICB to the 271 UL16 protein of the CMV, we assessed the interaction between *MICB98* mismatches 272 and the CMV status in their effect on clinical outcomes. For this purpose, we 273 performed multivariate analyses and included an interaction factor in the model. 274 Table 3 represents the risks of various clinical outcomes associated with (1) MICB98 275 mismatches when donor and recipients are negative for CMV, (2) CMV positivity in 276 277 donor and/or recipients when MICB98 is matched and (3) the interaction of MICB98 matching with CMV status. A statistically significant value for the interaction factor 278 indicates that the effect of *MICB98* matching depends on the category of CMV status 279 280 and vice versa. When the hazard ratio of the interaction factor is < 1 or >1, the hazard ratio of a variable (here, MICB98 matching or CMV status) is, respectively, 281 lower or higher in the category at risk of its interacting variable compared to the 282 reference category. For example, when the hazard ratio of the interaction factor is < 283 1, the hazard ratio of MICB98 mismatches is lower when the donor and/or recipient 284 are positive for CMV (category at risk of the CMV status variable) and higher when 285 both the donor and recipient are negative for CMV (reference category of the CMV 286 status variable). 287

For acute GVHD III-IV, the hazard ratio of the interaction was < 1 and was statistically significant (hazard ratio for acute GVHD III-IV, 0.26; 95% CI, 0.17 to 0.40; P < 0.001), indicating that the effect of *MICB98* mismatching on acute GVHD is more important when both the donor and the recipient are negative for CMV (acute GVHD III-IV hazard ratio, 3.63; 95% CI, 3.15 to 4.18; P< 0.001) compared to when the donor

and/or the recipient are positive for CMV (acute GVHD III-IV hazard ratio, 3.63 x 0.26
= 0.94). This observation was confirmed by representing graphically cumulative
incidences of acute GVHD III-IV in the above mentioned two CMV subgroups (Figure
24 24 24 28).

For OS, the interaction between *MICB98* mismatching and CMV status was 297 statistically significant and was > 1 (hazard ratio, 1.53; 95% CI, 1.38 to 1.69; P < 298 0.001). CMV positivity in the donor and/or recipient was associated with a slightly 299 lower survival when *MICB98* was matched (hazard ratio, 1.16; 95% CI, 1.14 to 1.19; 300 P< 0.001). However, because of the positive interaction with *MICB98* mismatches, 301 302 this effect was higher when *MICB98* was mismatched (hazard ratio $1.16 \times 1.53 =$ 1.77) (Table 3). The Kaplan-Meier estimates showing the higher impact of the CMV 303 status on OS in *MICB98* matched and mismatched groups are presented in Figures 304 305 2C and 2D, respectively. In other words, the risk of death associated with CMV positivity in the donor and/or recipient is lower in MICB98 matched vs. mismatched 306 groups. 307

Finally, to assess whether MICB98 mismatches had a GVHD-independent 308 effect on CMV infections in donor/recipients pairs at risk for CMV reactivation (i.e., 309 the donor and/or recipient was positive for CMV pre-HCT), we performed a 310 multivariate Fine and Gray analysis that included MICB98 matching as well as the 311 presence/absence of acute GVHD grades III-IV and chronic GVHD as time-312 dependent covariates in the model (Table 4). In accordance with the higher risk of 313 death described above, MICB98 mismatches were associated with a higher 314 incidence of CMV infections (hazard ratio, 1.84; 95% CI, 1.34 to 2.51; P < 0.001) 315 (Table 4 and Figure 3). MICB98 mismatches were not associated with EBV or HHV6 316 infections (Supplemental Table 1). 317

319 DISCUSSION

This is the first study analyzing the role of *MICB* matching in transplantation (whether HCT or solid organ).

Here we report that HCT from a MICB98 mismatched, but otherwise fully HLA 322 10/10 and MICA matched donor, carries a significantly increased risk of acute and 323 chronic GVHD. Interestingly, the effect on GVHD was not accompanied by a 324 decreased relapse rate. This unusual observation may be attributed to the CMV 325 status that is not independent of the MICB98 matching status. The significant 326 327 interaction of *MICB98* matching with CMV status (P < 0.001) indicates that the CMV status has a strong positive impact on relapse when MICB98 is mismatched (HR, 328 $0.77 \times 2.61 = 2.01$) (Table 3). 329

CMV biology has been known to be linked to *MICB* for more than 15 years. 330 Initially, Cosman et al. demonstrated that CMV infected cells can evade the immune 331 system by the retention of MICB and ULBP-1 and -2 antigens in the cell via binding to 332 the CMV protein UL16³⁶. This interaction hampers the ability of newly synthesized 333 MICB proteins to mature and transit the secretory pathway ⁵³. By dissecting the 334 molecular basis of MICB binding to UL16, Spreu et al. reported that the UL16-MICB 335 interaction is dependent on helical structures of the MICB α 2 domain ⁵⁴. Finally, 336 more recently, it was shown that UL16 binding was not equivalent for all MICB 337 alleles. The MICB*008 allele in particular was shown to have a decreased binding 338 activity compared to other alleles that do not have a methionine at position 98 in the 339 MICB α2 domain ³⁷. Importantly, position 98 is the only polymorphic position of MICB 340 that is known to be in direct contact with UL16³⁸. It is therefore not surprising that 341 mismatches at this position have less impact on acute GVHD in the presence of CMV 342

than in its absence. In the absence of CMV, the *MICB98* polymorphism may indeed 343 not be able to modulate the expression of MICB at the cell surface through interaction 344 with UL16 and consequently is not able to influence the alloreactivity that remains 345 higher in the mismatch than in the matched situation. Another explanation for the 346 higher MICB-mediated alloreactivity in the absence of CMV may be the absence of T-347 cell exhaustion, that is known to be induced by CMV positivity ⁵⁵. Ultimately, this 348 observation demonstrates that to lower the risk of acute GVHD in the absence of 349 CMV (donor and recipient seronegative), a *MICB98* matched donor is a better choice 350 than a *MICB98* mismatched donor. 351

CMV causes mortality in two ways: (1) directly by causing viral diseases, such 352 as pneumonitis, a situation that is becoming rare (viral diseases represent less than 353 2% of deaths) thanks to preemptive therapies, or (2) indirectly by clinical events 354 associated with virus seropositivity or the development of viral infections that are 355 independent of the viral disease itself ⁵⁶. The indirect effects of CMV are recognized 356 as a major cause of adverse outcomes after HCT, including GVHD and overall 357 mortality ⁵⁶⁻⁵⁸. Our dataset showed that the CMV effect on overall survival is amplified 358 in *MICB98* mismatched HCT compared to *MICB98* matched HCT, indicating that 359 matching donors at this position could be a useful strategy to decrease the risk of 360 death related to CMV. Because MICB98 mismatches were further shown to be 361 associated with CMV infection episodes, and this independently of the occurrence of 362 GVHD, deaths related to CMV may be due to CMV infections. 363

Collectively, these results suggest that pre-transplantation *MICB98* typing may help in lowering the risk of both GVHD- and CMV-related mortality. In the absence of CMV, matching *MICB98* provides a means to lower the incidence of GVHD, whereas in the presence of CMV, it helps improve overall survival. Fortunately, the level of

MICB98 mismatching is only 5.9% in HLA 10/10 matched donor/patient pairs that are also matched for *MICA;* although in absolute terms, this represents several thousand patients per year. Therefore, finding a *MICB98*-matched donor should be relatively easy in clinical practice.

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385 AUTHOR CONTRIBUTIONS

RC performed the experiments, designed the study, analyzed the data and wrote the manuscript. SB designed the study, analyzed the data and wrote the manuscript. PS, AM, IK, CM, APi and VR performed the experiments and analyzed the data. IA performed the statistics. PAvdB, DB, AC, DC, FCI, KG, JK, JJ, MLaba, PL, MMi, NM, PM, MOu, APa, RP, CPi, GS, ES, RT, AT, IY and BvdH provided samples and clinical data, interpreted the clinical data and discussed the results. BL, MMo, AN and CPa interpreted the clinical data and discussed results. FB, YK, MMB, MOt, and

393 BvdH analyzed the data and reviewed statistics. All authors contributed to the writing

of the report and approved the final version of the manuscript.

395

396 **Ethics declarations**

397 **Conflict of interest**

398 SB is the scientific founder and a (minority) shareholder of BIOMICA SAS. JK is the 399 co-founder and chief scientific officer of Gadeta. He received personal fees from 400 Gadeta. In addition, JK has a patent issued/pending. ES is the inventor of a patent 401 application filed by the University Medical Center Utrecht on the prediction of an 402 alloimmune response against mismatched HLA (PCT/EPT2013/073386). All other 403 authors declare no competing interests.

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668 **FIGURE LEGENDS**

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670 Figure 1. Effect of *MICB98* matching on severe acute and chronic GVHD

- The cumulative incidences of grades III-IV acute (panel A) and chronic GVHD (Panel
- B) are shown for *MICB98* mismatched (1) versus matched (2) patients.
- 673
- Figure 2. Effect of *MICB98* matching and CMV status on GVHD and Overall
 Survival.
- Panels A and B represent the cumulative incidences of grades III-IV acute GVHD in
- 677 HCT with donors and recipients negative for CMV (A) and HCT with donors and/or
- recipients positive for CMV (B). Panels C and D show the Kaplan-Meier estimates of
- overall survival in *MICB98* matched (C) and mismatched (D) transplants.
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681 Figure 3. Effect of *MICB98* matching on CMV reactivation/infection

- 682 The cumulative incidences of post-transplant CMV infection episodes in MICB98
- mismatched (1) versus matched (2) patients are shown.

TABLES Table 1. Demographics of the Study Population

	Total transplants	MICB 98 matched transplants	MICB 98 mismatched transplants	P-value*
	(n= 943)	(n=887)	(n=56)	
Transplantation centers [†] 1 2 3 4 5 6 7 8 9	106 (11.2%) 158 (16.8%) 114 (12.1%) 157 (16.6%) 48 (5.1%) 99 (10.5%) 96 (10.2%) 49 (5.2%) 116 (12.3%)	100 (11.3%) 142 (16%) 109 (12.3%) 153 (17.2%) 47 (5.3%) 90 (10.1%) 91 (10.3%) 46 (5.2%) 109 (12.3%)	6 (10.7%) 16 (28.6%) 5 (8.9%) 4 (7.1%) 1 (1.8%) 9 (16.1%) 5 (8.9%) 3 (5.4%) 7 (12.5%)	0.16
Age at transplant (years) 0-17 18-49 50-64 65 or older	58 (6.2%) 360 (38.2%) 458 (48.6%) 67 (7.1%)	57 (6.4%) 333 (37.5%) 430 (48.5%) 67 (7.6%)	1 (1.8%) 27 (48.2%) 28 (50%) 0 (0%)	0.034
Year of transplantation 2005–2008 2009-2013	360 (38.2%) 583 (61.8%)	338 (38.1%) 549 (61.9%)	22 (39.3%) 34 (60.7%)	0.97
Patient-donor sex Male-Female Other combinations Missing	159 (16.9%) 779 (82.6%) 5 (0.5%)	150 (16.9%) 732 (82.5%) 5 (0.6%)	9 (16.1%) 47 (83.9%) 0 (0%)	1.00
Patient-donor CMV status negneg. posneg./negpos./pospos. Missing	357 (37.9%) 560 (59.4%) 26 (2.7%)	329 (37.1%) 533 (60.1%) 25 (2.8%)	28 (50%) 27 (48.2%) 1 (1.8%)	0.082
Source of cells Bone marrow Peripheral blood stem cells	195 (20.7%) 748 (79.3%)	183 (20.6%) 704 (79.4%)	12 (21.4%) 44 (78.6%)	1.00
Conditioning regimen Non-myeloablative/reduced-intensity Myeloablative without total-body irradiation Myeloablative with total-body irradiation Missing	635 (67.3%) 140 (14.8%) 167 (17.7%) 1 (0.1%)	598 (67.4%) 130 (14.7%) 158 (17.8%) 1 (0.1%)	37 (66.1%) 10 (17.9%) 9 (16.1%) 0 (0%)	0.79
GvHD prophylaxis Cyclosporin only Cyclosporin and Methotrexate Cyclosporin and Mycophenolate Other combinations Missing	183 (19.4%) 243 (25.8%) 360 (38.2%) 135 (14.3%) 22 (2.3%)	171 (19.3%) 231 (26%) 335 (37.8%) 130 (14.7%) 20 (2.2%)	12 (21.4%) 12 (21.4%) 25 (44.6%) 5 (8.9%) 2 (3.6%)	0.49
<i>In vivo</i> T-cell depletion ‡ No Yes Missing	231 (24.5%) 690 (73.2%) 22 (2.3%)	214 (24.1%) 653 (73.6%) 20 (2.3%)	17 (30.3%) 37 (66.1%) 2 (3.6%)	0.34
Disease Acute myeloid leukemia Chronic myeloid leukemia Acute lymphoblastic leukemia Myelodysplastic syndrome Non-Hodgkin lymphoma Others §	240 (25.5%) 34 (3.6%) 121 (12.8%) 161 (17.1%) 127 (13.5%) 260 (27.6%)	225 (25.4%) 32 (3.6%) 114 (12.9%) 152 (17.1%) 121 (13.6%) 243 (27.4%)	15 (26.8%) 2 (3.6%) 7 (12.5%) 9 (16.1%) 6 (10.7%) 17 (30.4%)	0.99
Disease stage at transplantation ¶ Early Late Not applicablell Missing	371 (39.3%) 507 (53.8%) 44 (4.7%) 21 (2.2%)	348 (39.2%) 477 (53.8%) 42 (4.7%) 20 (2.3%)	23 (41.1%) 30 (53.6%) 2 (3.6%) 1 (1.8%)	0.97
Time until treatment <12 months >12 months	440 (46.7%) 503 (53.3%)	416 (46.9%) 471 (53.1%)	24 (42.9%) 32 (57.1%)	0.65
Non-Permissive HLA-DPB1 matching** Matched Mismatched Missing	420 (44.5%) 394 (41.8%) 129 (13.7%)	392 (44.2%) 374 (42.2%) 121 (13.6%)	28 (50%) 20 (35.7%) 8 (14.3%)	0.42

The results are presented as the number of patients and corresponding percentages of the study population. HLA: Human Leukocyte Antigen. All clinical variables of the table were used for adjustment in the multivariate models.

P-values were determined with Pearson's Chi square test or Fisher's exact test for small
 sample sizes

⁶⁹³ † Patients received their transplant in six centers of the Francophone Society of Bone ⁶⁹⁴ Marrow Transplantation and Cell Therapies (SFGM-TC) (1 to 6; N =682) and in three Dutch ⁶⁹⁵ centers that are part of the Europdonor operated by the Matchis Foundation network (7 to 9; ⁶⁹⁶ N=261).

in vivo T-cell depletion was performed by the addition of anti-thymocyte globulin (ATG) or
Alemtuzumab to the conditioning regimen.

§ Other diseases include multiple myeloma, Hodgkin lymphoma, Fanconi anemia, aplastic
anemia, chronic lymphocytic leukemia, plasma cell leukemia, other acute leukemias, solid
tumors (not breast), hemophagocytosis and inherited disorders.

¶ Early corresponds to diseases in the first complete remission or in the chronic phase. Late corresponds to second or higher complete remissions, accelerated phases, partial remissions, progressions, primary induction failures, relapses or stable diseases. Not applicable corresponds to bone marrow failure (aplastic anemia, Fanconi anemia), inherited disorders, hemophagocytosis and solid tumors.

** *HLA-DPB1* matching was defined at the T-cell-epitope matching level ⁴⁹ with typing data at
 2nd field resolution following the World Health Organization official nomenclature .

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710 Table 2. Analysis of the Impact of *MICB* Mismatches at amino acid position 98 on

hazard ratio (95% CI) P-value Acute GVHD II-IV 1.20 (1.15-1.24) < 0.001 ۰ Acute GVHD III-IV 2.28 (1.56-3.34) < 0.001 Chronic GVHD 1.21 (1.10-1.33) < 0.001 -0-Relapse[†] 1.42 (1.05-1.93) 0.024 Overall survival 1.01 (0.84-1.20) 0.93 Relapse-free survival 0.98 (0.91-1.06) 0.63 Non-Relapse Mortality 0.62 (0.37-1.04) 0.071 0.0 0.5 1.0 1.5 2.0 2.5 3.0 3.5

711 Clinical Outcomes after Multivariate Modeling*

Results are presented as Hazard Ratios with 95% confidence intervals (CI). GVHD: Graftversus-host disease. * All models were adjusted for patient's age, patient-donor sex, patientdonor serological status for cytomegalovirus, year of transplantation, time to transplantation, transplantation center, source of stem cells, conditioning regimen, GVHD prophylaxis, treatment with anti-thymocyte globulin or Alemtuzumab, *HLA-DPB1* matching status, disease category and severity at transplantation. † Transplantations performed for non-malignant diseases were excluded from the analysis.

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Table 3. Analysis of the Impact of *MICB* Mismatches at position 98, CMV status and

Outcomes and risk factors	hazard ratio (95% CI)	<i>P</i> -value
Acute GVHD II-IV		
MICB98 matching (mismatches)	1.47 (1.05-2.07)	0.025
CMV status (D+/R- or D-/R+ or D+/R+)‡	1.18 (0.92-1.51)	0.2
Interaction: MICB98 matching X CMV status	0.57 (0.29-1.10)	0.095
Acute GVHD III-IV		
MICB98 matching (mismatches)	3.63 (3.15-4.18)	< 0.001
CMV status (D+/R- or D-/R+ or D+/R+)	1.50 (1.15-1.96)	0.003
Interaction: MICB98 matching X CMV status	0.26 (0.17-0.40)	< 0.001
Chronic GVHD		
MICB98 matching (mismatches)	1.26 (1.25-1.27)	< 0.001
CMV status (D+/R- or D-/R+ or D+/R+)	1.34 (1.15-1.56)	< 0.00
Interaction: MICB98 matching X CMV status	0.91 (0.70-1.18)	0.48
Relapse [†]		
MICB98 matching (mismatches)	0.89 (0.78-1.01)	0.073
CMV status (D+/R- or D-/R+ or D+/R+)	0.77 (0.70-0.84)	< 0.001
Interaction: MICB98 matching X CMV status	2.61 (1.79-3.82)	< 0.001
Overall survival		
MICB98 matching (mismatches)	0.80 (0.64-1.00)	0.054
CMV status (D+/R- or D-/R+ or D+/R+)	1.16 (1.14-1.19)	< 0.001
Interaction: MICB98 matching X CMV status	1.53 (1.38-1.69)	< 0.002
Relapse-free survival		
MICB98 matching (mismatches)	0.78 (0.70-0.86)	< 0.001
CMV status (D+/R- or D-/R+ or D+/R+)	1.09 (1.05-1.13)	< 0.00
Interaction: MICB98 matching X CMV status	1.57 (1.45-1.70)	< 0.001
Non-relapse mortality		
MICB98 matching (mismatches)	1.14 (0.46-2.86)	0.78
CMV status (D+/R- or D-/R+ or D+/R+)	1.38 (1.12-1.70)	0.003
Interaction: MICB98 matching X CMV status	0.41 (0.22-0.76)	0.005

722 their interaction on Clinical Outcomes after Multivariate Modeling*

723 Results are presented as Hazard Ratios with 95% confidence intervals (CI). GVHD: Graftversus-host disease. * All models were adjusted for patient's age, patient-donor sex, patient-724 donor serological status for cytomegalovirus, year of transplantation, time to transplantation, 725 726 transplantation center, source of stem cells, conditioning regimen, GVHD prophylaxis, 727 treatment with anti-thymocyte globulin or Alemtuzumab, HLA-DPB1 matching status, disease category and severity at transplantation. † Transplantations performed for non-malignant 728 diseases were excluded from the analysis. **‡** D and R stand for donor and recipient, 729 respectively. The reference category for the CMV status is D-/R-. 730

731 Table 4. Effect of GVHD and *MICB98* matching on CMV infections

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	hazard ratio (95% CI)*	<i>P</i> -value
GVHD		
Chronic		
Absent (n=307)	Ref.	-
Present (n=169)	0.99 (0.83-1.19)	1.05
Acute III-IV		
Absent (n=388)	Ref.	-
Present (n=78)	1.12997 (1.1290-1.13)	< 0.001
MICB98 matching		
Matched (n=437)	Ref.	-
Mismatched (n=19)	1.84 (1.34-2.51)	< 0.001
Only the pairs in which the donor and/o	or the recipient was/were positive for (CMV pre-
HCT were included in the analysis. Th	ne results are presented as Hazard Ra	atios with
95% confidence intervals (CIs). GVH	D: Graft-versus-host disease. Ref.: R	eference

category. * Multivariate Fine and Gray model including *MICB98* matching, acute
 GVHD III-IV and chronic GVHD as time-dependent covariates in the model. In

addition, the model was adjusted for patient's age, patient-donor sex, patient-donor

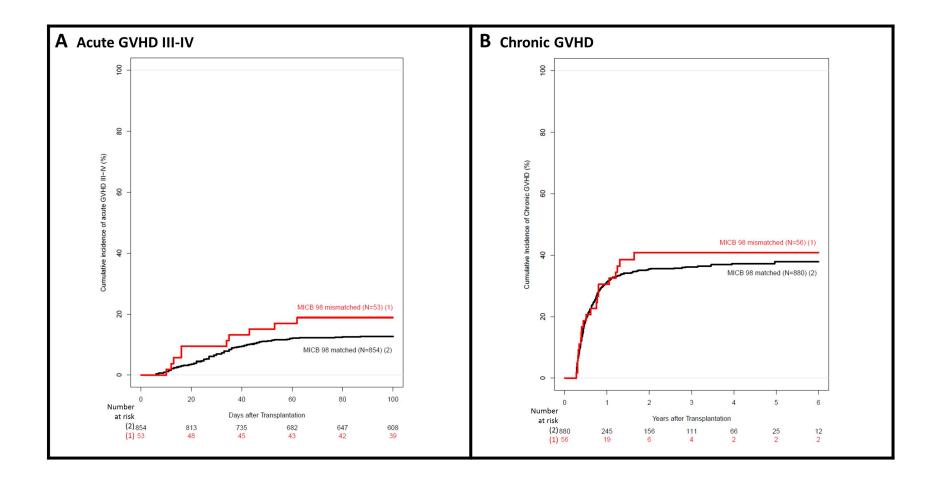
serological status for cytomegalovirus, year of transplantation, time to transplantation,

transplantation center, source of stem cells, conditioning regimen, GVHD prophylaxis,

treatment with anti-thymocyte globulin or Alemtuzumab, HLA-DPB1 matching status,

742 disease category and severity at transplantation.

Figure 1



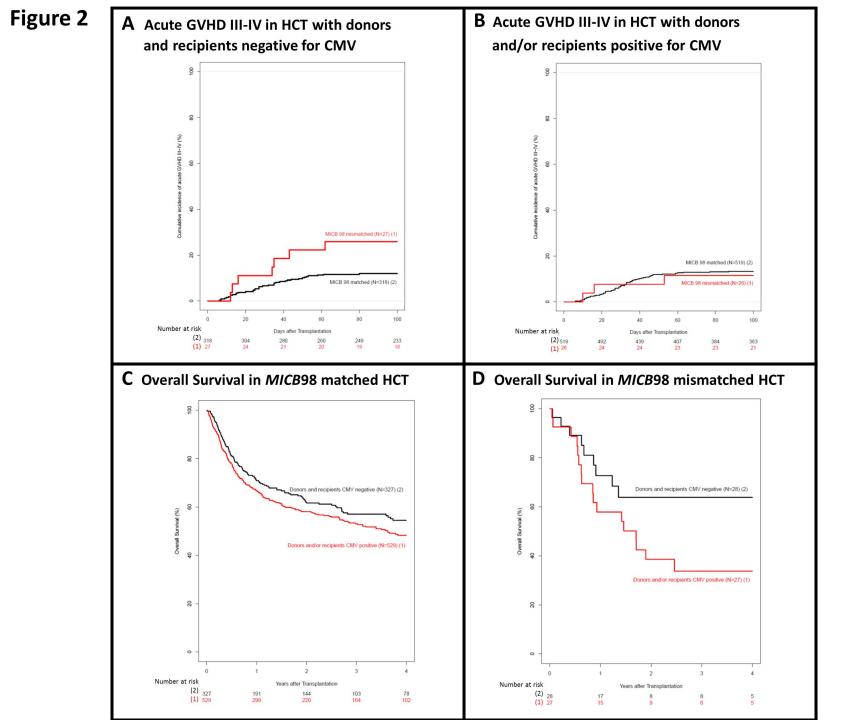


Figure 3

