

Neurogenetics of the Human Adenosine Receptor Genes: Genetic Structures and Involvement in Brain Diseases

Vincent Huin, Claire-Marie Dhaenens, Mégane Homa, Kévin Carvalho, Luc Buée, Bernard Sablonnière

▶ To cite this version:

Vincent Huin, Claire-Marie Dhaenens, Mégane Homa, Kévin Carvalho, Luc Buée, et al.. Neurogenetics of the Human Adenosine Receptor Genes: Genetic Structures and Involvement in Brain Diseases. Journal of Caffeine and Adenosine Research, 2019, 9 (3), pp.73-88. 10.1089/caff.2019.0011 . inserm-02460598

HAL Id: inserm-02460598 https://inserm.hal.science/inserm-02460598

Submitted on 30 Jan 2020 $\,$

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Neurogenetics of the human adenosine receptor genes: Genetic structures and involvement in brain diseases

Vincent Huin, MD, PhD^{a,b}, Claire-Marie Dhaenens, PharmD, PhD^{a,b}, Mégane Homa, MSc^a, Kévin Carvalho, MSc^a, Luc Buée, PhD^a, Bernard Sablonnière, MD, PhD^{a,b}

^a Univ. Lille, Inserm, CHU Lille, UMR-S 1172 - JPArc - Centre de Recherche Jean-Pierre AUBERT Neurosciences et Cancer, F-59000, Lille, France

^b CHU Lille, Institut de Biochimie et Biologie moléculaire, Centre de Biologie Pathologie et Génétique, F-59000, Lille, France

Running title: Neurogenetics of adenosine receptors

Correspondence to:

Vincent Huin: vincent.huin@inserm.fr

University of Lille, Inserm UMR-S 1172, JPArc, rue Polonovski, F-59045, Lille, France.

Telephone: +33 (0)3 59 89 96 05 / Fax: +33 (0)3 20 53 85 62

Keywords:

Adenosine receptor, Genetics, Single nucleotide polymorphism, Neurology, Psychiatry, Neurodegeneration

ORCID ID:

Vincent Huin=https://orcid.org/0000-0001-8201-5406

Claire-Marie Dhaenens=https://orcid.org/0000-0003-4719-4124

Mégane Homa=https://orcid.org/0000-0002-3904-6888

Kévin Carvalho=https://orcid.org/0000-0002-6432-7622

Luc Buée=https://orcid.org/0000-0002-6261-4230

Bernard Sablonnière=https://orcid.org/0000-0003-0384-4076

Total word count of the manuscript: 6550

(excluding title, abstract, authors and affiliations, keywords, figure legends, tables, acknowledgements, authorship confirmation statement, author disclosure statements, acknowledgments, footnotes, or references)

Character count for the title: 105, 14 words

Character count for the running title, including spaces: 36

Total word count of the abstract: 168

Number of references: 136

Number of tables: 2

Number of figures: 1

Number of supplemental figures: 0

Number of supplemental tables: 0

Number of supplemental data: 1 (supplemental material)

Authorship Confirmation Statement

Authors' contributions:

VH wrote the manuscript.

- CMD, MH, KC and BS revised the manuscript for intellectual content.
- LB provided intellectual input and editorial suggestions.

Confirmation statement:

All of the authors have reviewed and approved the manuscript prior to submission. We confirm that the manuscript has been submitted solely to this journal and is not published, in press, or submitted elsewhere.

Abstract

Adenosine receptors are G-protein-coupled receptors involved in a wide range of physiological and pathological phenomena in most mammalian systems. All four receptors are widely expressed in the central nervous system, where they modulate neurotransmitter release and neuronal plasticity. A large number of gene association studies have shown that common genetic variants of the adenosine receptors (encoded by the *ADORA1*, *ADORA2A*, *ADORA2B* and *ADORA3* genes) have a neuroprotective or neurodegenerative role in neurologic/psychiatric diseases. New genetic studies of rare variants and few novel associations with depression or epilepsy subtypes have receptors in neurologic and/or psychiatric diseases in humans, and discuss perspectives for further genetic research. We also provide an update on the genetic structures of the four human adenosine receptor genes and their regulation - a topic that has not been extensively addressed. Our review emphasizes the importance of (i) better characterizing the genetics of adenosine receptor genes and (ii) understanding how these genes are regulated.

Introduction

Adenosine exerts a variety of physiological effects in most mammalian systems through four subtypes of G-protein-coupled adenosine receptors (A₁R, A_{2A}R, A_{2B}R, and A₃R), each of which is encoded by their own gene (ADORA1, ADORA2A, ADORA2B, and ADORA3, respectively). The four adenosine receptors are widely expressed in the central nervous system (CNS) in general and in neurons, glial cells and endothelial cells in particular.^{1,2} In the CNS, A₁R and A_{2A}R modulate synaptic neurotransmission through their action on the release of various neurotransmitters (such as glutamate, y-aminobutyric acid, acetylcholine and dopamine) and on the activation of postsynaptic receptors. Excitatory transmission is inhibited by A1R activation and stimulated by A_{2A}R activation. The interplay between these two receptors helps to fine-tune synaptic plasticity.³ In several regions of the brain, the two receptors are co-expressed and can form heteromers together⁴ or with other Gcoupled receptors, such as dopamine receptors.^{5,6} Neuroprotective and neurodegenerative effects of A1R and A2AR have been observed in various neurologic diseases (Parkinson's disease, Huntington's disease, migraine, stroke, and epilepsy) and psychiatric diseases (schizophrenia, panic disorders, depression, and autism).^{7,8} A_{2B}R is widely expressed in many tissues, and is involved in ischemiareperfusion injury, fibrosis, cancer, inflammation, and the immune system.⁹ Lastly, A₃R is involved in the modulation of ischemic diseases, inflammatory diseases, cancer, autoimmune diseases, and chronic neuropathic pain.¹⁰ Until recently, A_{2B}R and A₃R were considered to have a minor physiological role because they have substantially lower affinities for adenosine and thus require higher extracellular adenosine concentrations for maximal activation. Only two studies have assessed them as genetic factors in human neurologic diseases.^{11,12}

Various aspects of adenosine receptor research and the receptors' roles in the pathophysiology of the neural system, widely discussed in the current literature,⁸ lead to numerous publications assessing the role of genetic variants in adenosine receptor genes and neurological or psychiatric diseases. Recent advances in these fields (including new topics with the studies of rare genetic variants^{12,13} and new associations with epilepsy and depression^{14,15} highlight the importance of genetic variants in adenosine receptor variants in adenosine receptor gene in CNS disorders.

Previous reviews or book chapters focused on the genes structure,¹⁶ on neurobiology,¹⁷ or on one specific gene and its neuroprotective/neurodegenerative effects.^{18,19} However, these previous works do not integrate and discuss all the genetic associations reported so far. It remains difficult to have a critical overview of the neurogenetics of adenosine receptor genes with the heterogeneity of the genetic studies (in diseases, sample size, methodologies and level of evidence). This prompted us to review the current state of knowledge regarding genetic associations of variants in adenosine receptor genes with human brain diseases and to confront them to functional assessment of the variants.

The criteria and databases used for this literature search are given as supplementary material.

ADORA1

Gene structure (Figure 1.A)

The human *ADORA1* gene was cloned in 1992 from a hippocampal cDNA library.^{20,21} It spans 77 kb, and is located on chromosome 1q32.1.²² The gene comprises six exons that undergo alternative splicing and alternative promoter usage.^{23,24} The Genotype-Tissue Expression (GTEx) database²⁵ indicates that *ADORA1* mRNAs are found in all tissues, with the highest expression levels in the spinal cord, brain (the frontal cortex, substantia nigra, and all the basal ganglia) and testis.

According to the GENCODE database,²⁶ the human ADORA1 gene encodes six different mRNAs (Figure 1.A). Three transcripts with different 5' untranslated regions (UTR) share the same coding exons (5 and 6), and yield a full-length 326-aa protein. The fourth transcript does not include exon 5, but contains an open reading frame (ORF) and might result in an N-terminal-truncated 210-aa protein. The two last transcripts contain exon 6 with an alternative 3' splice site, and are thought to encode a C-terminal-truncated 125-aa protein. Ren & Stiles showed that transcripts including exon 4 were widely expressed, whereas transcripts with exon 3 were present in the testis, kidney, fetal brain, and the frontal cortex and cerebellum of the adult brain but not in fat, skeletal muscle or heart.²³ Two alternative promoters with nonclassical TATA boxes have been found upstream of exons 3 and 4.^{24,27} The promoter upstream of exon 3 is constitutively expressed at low levels, and contains a binding site for nuclear factor-kappa B (NF-kB),²⁸ serum response factor and activator protein 1.29 The promoter upstream of exon 4 is more active but shows tissuespecific regulation and contains a site for an AGG element binding protein.³⁰ It is noteworthy that ADORA1 has only one upstream 262 bp CpG island, which overlaps with exon 4. Interestingly, there are two upstream AUG codons in exon 4, both of which are crucial for mRNA processing and translation. Indeed, mutations in these AUG codons are associated with greater levels of translation.²⁴ We hypothesized that an upstream ORF (uORF) exerts post-transcriptional control on ADORA1 transcripts including exon 4 in the same way that uORF5 regulates ADORA2A expression.³¹

Lastly, the ENCODE chromatin state segmentation from the University of California Santa Cruz Genomics Institute (UCSC) genome browser suggests that a third promoter is located upstream of exon 1.³²

Genetic risk factor in human diseases

Psychiatric diseases

In 2009, Gotoh et al. published a case control study including 200 schizophrenic patients and 210 controls. The researchers looked for an association between 29 single-nucleotide polymorphism (SNPs) in *ADORA1* and the presence of schizophrenia in a Japanese population.³³ None of the individual SNPs was significantly associated with the disease and the small number of samples did not allow the identification of linkage disequilibrium blocks. However, a multivariate analysis evidenced an association with three haplotypes built from two or three SNPs. An important limitation concerned the control population who did not perfectly matched to the schizophrenic population. Moreover, our literature database search did not found a replication study in other Japanese population.

A case-control study focused on the association between *ADORA1* and *ADORA2A* SNPs and Tourette syndrome including 162 patients and 270 controls.³⁴ Patients with a GG or GT genotype at rs2228079 were underrepresented. This silent SNP (located in exon 5) was associated with earlier age at tic onset and a greater prevalence of comorbidities (the concomitant presence of depression or obsessive compulsive disorders). However these comorbidities concerned only a small number of patients

with particular psychiatric disorder. Moreover, the data about the earlier age at tic onset and comorbidities in some adult patients were retrospective.

In a study of 806 Caucasian Americans, Clark et al. compared the number of alcohol dependence symptoms with the presence of variants in 71 candidate genes (including the coding, flanking and regulatory regions).¹³ Although the researchers did not find a significant association with the common variants considered in isolation, alcohol dependence was linked to an aggregate of eight variants (five rare variants and three common) within intron 5 of the *ADORA1* gene. This study focused on common and rare variants and their aggregation on putative regulatory regions in the genes is original, but lacks functional validation and still need to be replicated.

These three studies lack functional data and are balanced by negative results provided similar case-control studies evidencing no genetic association between *ADORA1* SNPs and bipolar affective disorders³⁵ or methamphetamine dependence or psychosis.³⁶ Moreover, the role of *ADORA1* common variant in anxiety disorders does not seem to be important regarding the study of Rogers et al. who did not found an association between seven *ADORA1* SNPs and effects of caffeine (an antagonist of A₁R) on anxiety.³⁷ Lastly, recent genome-wide association studies (GWAS) on schizophrenia,^{38,39} Tourette syndrome⁴⁰ and alcohol dependence⁴¹ did not replicate these associations. The genetic associations of *ADORA1* common variants in psychiatric diseases remain controversial and should then be interpreted with caution.

Neurologic and neurodegenerative disorders

ADORA1 mutations are thought to have a causal role in familial Parkinson's disease.⁴² In a study of a large consanguineous family in which two members were affected by early-onset Parkinsonism and cognitive dysfunction, Jaberi et al. used homozygosity mapping and exome sequencing to detect a homozygous missense mutation c.835G>A, p.(Gly279Ser) in exon 6 of the ADORA1 gene. Segregation analyses, bioinformatics predictions and the variant's rarity made this missense a good candidate. However, the Gly279Ser variant did not have a functional effect on the interaction between A₁R and the dopamine D1 receptor. Based on these genetic studies, and given A1R's role in neurodegenerative diseases, Jaberi et al. hypothesized that ADORA1 was a reasonable candidate disease-causing gene. Moreover, the researchers suggested that ADORA1 was located within the Parkinson's disease locus PARK16, which has been identified in several populations in earlier GWAS.^{43,44} However, it was later demonstrated that ADORA1 was not in the PARK16 locus, and that mutations in ADORA1 could not drive the PARK16 association signal.⁴⁵ Moreover, Jaberi et al. identified another homozygous variant in PTRHD1 that was later reported as a more likely candidate disease-causing gene. Indeed, Khodadadi et al. reported a different mutation in PTRHD1 in their study of another Iranian family with a similar clinical presentation.^{46,47} ADORA1 variants are not a common cause of Parkinson's disease or dementia with Lewy bodies⁴⁵ and the role of the Gly279Ser variant in early-onset Parkinsonism with cognitive dysfunction is subject to debate.

Variants in *ADORA1* are associated with susceptibility to epilepsy after severe traumatic brain injury.⁴⁸ In a retrospective study of 206 patients with severe traumatic brain injury (excluding penetrating head injury), Wagner et al. genotyped 6 SNPs in *ADORA1*. There was an association between the AA genotype at rs3766553 and an

elevated incidence of early post-traumatic seizures, whereas the GG genotype at rs3766553 and the CT genotype at rs10920573 were associated with an elevated incidence of late-onset seizures. However, the study was not informative regarding the risk to develop recurrent seizures.

To conclude, whereas the role of *ADORA1* in rare Mendelian cause of Parkinsonism has been questioned if not refuted; the role of *ADORA1* common variants in epilepsy after traumatic brain injury seems promising and might lead up to a better risk stratification and maybe to the development of new therapeutic strategies.

ADORA2A

Gene structure (Figure 1.B)

Of the four human adenosine receptor genes, *ADORA2A* has the most complex structure. It was first cloned from a hippocampal library in 1992.⁴⁹ It is 24 kb long, and is located on chromosome 22q11.23.^{50–52} Gene expression for *ADORA2A* mRNA has been evidenced in many tissues, including the liver, heart, lung, immune system, and brain.⁵³ In the GTEx project,²⁵ *ADORA2A* mRNAs were found to be expressed in all tissues; the highest expression levels were observed in the basal ganglia (the nucleus accumbens, putamen, and caudate), blood, spleen, lung, and cerebellum.

ADORA2A is composed of two invariant coding exons and several alterative exons involved in the formation of a 5'-UTR. Using molecular and bioinformatics analyses, Yu et al. identified new exons encoding the 5'-UTR of *ADORA2A* in the mouse and in the human.⁵⁴ In the human, the researchers evidenced six variants of non-coding exon 1, and referred to the invariant coding exons as exons 2 and 3. All transcripts encoded the same protein but displayed tissue-specific expression patterns. Kerth et

al. identified eight 5'-UTR splice variants of *ADORA2A* produced by the alternative transcription and/or splicing of five noncoding exons upstream of the two invariant coding exons.⁵⁵ In the Ensembl database, *ADORA2A* mRNAs have eight different transcription start sites (TSSs) (Figure 1.B), and non-coding RNAs give rise to three other TSSs. Not all the transcripts have been fully sequenced, and levels of evidence are low for some. In the GENCODE database,²⁶ *ADORA2A* has four transcripts with an alternative 5'-UTR encoding the same 412-aa protein (Figure 1.B).

Accordingly, the several putative promoter regions have not yet been extensively characterized. Most studies to date have been performed in the rat^{56–59} or the mouse.⁵⁴ A few researchers have sought to characterize the human promoters^{60,61}, mostly by focusing on a 2000 bp region of upstream of exon 2. By considering the histone code and the chromatin state segmentation data⁶² from the ENCODE project,⁶³ we observed five non-overlapping promoter regions. As determined by the CpG Islands Track Settings tool (UCSC), only one promoter region comprises a 1272 bp CpG island encompassing different alternative first non-coding exons (Figure 1.B). Methylation of this CpG island and the surrounding CpG shore was assessed in human brain samples from healthy individuals,^{64–66} patients with Huntington's disease (HD),⁶⁷ and schizophrenic patients⁶⁸ - demonstrating that *ADORA2A*'s promoters are under epigenetic control. It is very likely that each promoter region has different transcription factor binding sites; these might control the expression of the *ADORA2A* gene under various conditions (e.g. as a function of the cell type, the development stage and/or the response to different stimuli).

This complexity is heightened by the existence of different 5'-UTRs, which might recruit different RNA-binding proteins and thus regulates the degradation, storage, and/or translation of the corresponding *ADORA2A* mRNAs.⁶⁹ In the rat, the different

5'-UTRs can repress translation via the use of a conserved ORF (notably in humans) located 65 nucleotides upstream of the usual translation initiation site.⁷⁰ Moreover, the rat uORF leads to the translation of a 134-aa protein called uORF5.⁷¹ Hence, the *ADORA2A* gene is a rare example of a human dual-coding gene.

Yu et al. reported on a novel transcript that reads in the opposite direction to the ADORA2A gene.⁵⁴ Indeed, there appears to be an antisense non-coding RNA gene (ADORA2A-AS1) at the same locus. Three isoforms of ADORA2A-AS1 have at least one exon that complements part of the ADORA2A UTR. Lastly, read-through transcription between the upstream SPECC1L gene (coding for sperm antigen with calponin homology and coiled-coil domains 1-like) and ADORA2A has been reported. This SPECC1L-ADORA2A transcript results in a non-coding RNA that probably induces nonsense-mediated mRNA decay and is then unlikely to produce a protein.⁷² The functions of SPECC1L-ADORA2A and ADORA2A-AS1 are unknown but as other long non-coding RNAs (IncRNA), they might regulate different genes in the same locus, including ADORA2A. Indeed, the IncRNAs are known as regulator of gene expression modulating the transcription, splicing, and translation of other mRNA by RNA-DNA or RNA-RNA interaction.⁷³ They have been implicated in numerous psychiatric/neurologic diseases, in neurodevelopment and neurodegeneration.^{74,75} Then, genetic variants in these non-coding genes can (i) be in linkage disequilibrium with other more relevant variant in ADORA2A, (ii) be also located in an important region for the regulation of ADORA2A, like an enhancer and then modulate its transcription, but it is likely that (iii) some genetic variants affecting the expression of SPECC1L-ADORA2A or ADORA2A-AS1 could indirectly modify the expression of genes regulated by these IncRNAs. For example, ADORA2A-AS1 might form RNA

duplexes with ADORA2A mRNA whereas SPECC1L-ADORA2A could induce epigenetic changes in ADORA2A.

Genetic risk factor in human diseases

Psychiatric diseases

From 1998 onwards, ADORA2A variants were suspected to confer susceptibility to various anxiety disorders (e.g. panic disorder).⁷⁶ In an initial comparison between 89 German patients with panic disorder and matched controls, researchers observed a significant association of the disease with the T allele and the TT genotype of a silent SNP (rs5751876) located in ADORA2A exon 3. Later, Hamilton et al. highlighted a genetic linkage between panic disorder (especially the form with agoraphobia) and a haplotype encompassing rs5751876.77 All the significantly associated haplotypes comprised the C allele of rs5751876; this contradicted Deckert et al.'s report that the rs5751876 T allele was associated with an elevated risk of anxiety disorder. This discrepancy suggests that rs5751876 is not a causal variant conferring an elevated risk of anxiety disorder. Furthermore, it has also been reported that rs5751876 is tvpes⁷⁶ anxiety-related personality associated with various and (ii) (i) psychophysiological markers of anxiety-related arousal of the sympathetic nervous system in blood-injury phobia.⁷⁸ The same research group replicated the association between rs5751876 (the TT genotype) and panic disorder.⁷⁹ Again, the researchers found a stronger association in the agoraphobia subgroup - prompting them to suggest that the association with the panic disorder population as a whole might originate in this subgroup alone. Hohoff et al. found an association between an anxious personality and a haplotype composed of eight SNPs in ADORA2A

(including rs2298383 and rs5751876). In one study rs2298383 was not associated with panic disorders in the absence of agoraphobia.⁸⁰ However, a *post hoc* power analysis showed that the study including 71 patients and 100 controls was not powered to reliably detect a true association. Studies of different anxiety arousal paradigms in healthy subjects (such as caffeine-induced anxiety^{81,82} and anxiety response to amphetamine⁸³) also highlighted an association with ADORA2A variants. Recent studies have highlighted a complex interaction between ADORA2A genetic variants and anxiety/attentional responses linked to caffeine.^{37,84-86} Taken as a whole, these data provide strong evidence for a role of the ADORA2 gene in anxiety disorders, and fit with the abnormally high level of anxiety in Adora2a-knockout mice.⁸⁷ Most of these studies were conducted in Caucasian cohorts, and the association between rs5751876 and anxiety disorders was not replicated in two case control studies conducted in Chinese⁸⁸ or in Japanese⁸⁹ populations (respectively, 104 cases versus 192 controls; and 91 cases versus 100 controls) - suggesting that the association is ethnicity-dependent. These findings also suggest that rs5751876 (the most widely studied ADORA2A polymorphism) is not the causal variant conferring an elevated risk of anxiety; in fact, it is probably a tag SNP for the true causal variant. The latter might therefore be less frequent and/or not in linkage disequilibrium with rs5751876 in Asian populations. Another hypothesis is that ADORA2A confers susceptibility to anxiety in some patient subgroups (e.g. patients with agoraphobia).^{77,79} Another example was provided in 2012 by Subaran et al., who described an association with rs5751876 and rs4822492 in a subset of patients with panic disorders and bladder syndrome (n=92) but not in the whole cohort of patients with panic disorder (n=351).⁹⁰

Molero et al. studied the relationship between six *ADORA2A* polymorphisms in strong linkage disequilibrium and attention-deficit hyperactivity disorder traits.⁹¹ After correction for multiple comparisons, the rs35320474 SNP was still significantly associated with inattention symptoms but not with hyperactivity-impulsivity. Very recently, Fraporti et al. confirmed the association between the TT genotype of rs2298383 and the presence of anxiety disorders in children with attention-deficit hyperactivity disorder.⁹²

Janik et al. reported a link between *ADORA1*, *ADORA2A* and Tourette syndrome.³⁴ The T allele of rs5751876 was overrepresented among patients, and was associated with earlier age at onset. However, in contrast to previous studies, the T allele was underrepresented in patients with concomitant attention-deficit hyperactivity disorder and patients with anxiety but not obsessive-compulsive disorder. The later result was more questionable as it was obtained on small samples including only 128 non anxious patients with Tourette syndrome versus 34 anxious patients.

Freitag et al. linked several *ADORA2A* SNPs to autism spectrum disorders (ASD) in a genetic case-control analysis of 98 patients and 234 controls.⁹³ The CC genotype of rs2236624 (located in intron 3) was overrepresented in this group of patients. Increased current social interaction, nonverbal communication and repetitive behavior scores were associated with rs3761422 (CT and TT genotypes), rs5751876 (CT and TT genotypes), and rs35320474 (T/del and del/del genotypes). Lastly, the researchers evidenced a significant association between a haplotype of eight SNPs (including rs5751876) and anxiety in the group of 98 patients with ASD. The most frequent haplotype ACTCCCTG (built with the rs5751862, rs5760405, rs2298383, rs3761422, rs2236624, rs5751876, rs35320474 and rs4822492 variants) was carried by 73.3% of the patients with severe anxiety disorders but only 33.9% of the patients without a history anxiety. However, it should be noted that although the allele C of rs5751876 was significantly less frequent in autism patients, it was overrepresented in the subset of individuals with a history of anxiety.

The genetic link between *ADORA2A* and psychosis is more subject to debate. In a study of small number of schizophrenic patients (*n*=18) and healthy controls (*n*=33), Jagannathan et al. evaluated the relationship between gray matter volume (as measured using structural MRI) and 367 different SNPs in candidate genes.⁹⁴ The researchers found an inverse correlation between a lack of gray matter in the frontal, temporal and thalamic regions (previously reported to be altered in schizophrenia) and 18 SNPs in 16 genes, including rs3761422 in *ADORA2A*. However, it had previously been reported that *ADORA2A* variants (including rs5751876) were not involved in the development of schizophrenia.^{95–97} The absence of a relationship was confirmed by two recent GWASs of patients with schizophrenia.^{38,39}

Recently, Oliveira et al. published the results of a cross-sectional population-based study of 1253 participants, of whom 228 were experiencing current depression.¹⁵ The TT genotype of rs2298383 was less frequent in patients experiencing current depression than in controls. After adjusting for confounding variables, this association remained significant (odds ratio (OR) [95% confidence interval (CI)]=0.631 [0.425–0.937]; p=0.022). The TT genotype was associated with protection against sleep dysfunction and attentional impairment - two common symptoms observed in patients with depression. However, an earlier case-control study performed in an Asian population and including 192 mood disorder patients and 216 controls did not evidence an association between rs5751876 and major depression or bipolar disorder.⁹⁸

Neurologic and neurodegenerative disorders

In 2007, *ADORA2A* was linked to migraine with aura.⁹⁹ A haplotype block spanning the rs5760405, rs5751876, rs35320474, rs5751862, rs3761422 and rs4822492 SNPs in *ADORA2A* was tested in 265 patients with migraine (of whom 122 had migraine with aura) and 154 controls. A GCCCTG haplotype was more frequent in patients suffering from migraine with aura than in controls. However, there was no significant difference between patients with migraine without aura and controls.

Genetic variants in the *ADORA2A* gene are known to be genetic modifiers in the pathogenesis of HD, a hereditary neurodegenerative disorder caused by the expansion of CAG repeats within the *HTT* gene. The broad range of ages at onset is mostly explained by the number of CAG repeats but is also related to various environmental and genetic factors - including *ADORA2A* polymorphisms. A study published in 2009 reported an earlier age at onset (by 3.8 years) in patients with the TT genotype at rs5751876 than in patients with the CC genotype (p=0.02).¹⁰⁰ The impact of *ADORA2A* genotype on residual variation was modest; however, these results have been replicated in two other studies.^{101,102} One of the latter found that rs2298383 in intron 1 (in linkage disequilibrium with rs5751876) to be the most strongly associated *ADORA2A* variant.¹⁰¹ These findings are supported by our knowledge of other environmental modifiers of HD (such as caffeine intake¹⁰³) and data from animal models on the roles of A₁R and A_{2A}R.¹⁰⁴

Horgusluoglu-Moloch et al. used the dataset from the Alzheimer's Disease Neuroimaging Initiative cohort and selected 1,563 non-Hispanic Caucasian participants to correlate different quantitative phenotypes (including hippocampal volume, metabolic activity, cerebrospinal fluid total tau level, amyloidosis in the hippocampus, and composite memory) with 18407 SNPs in 81 genes from a targeted neurogenesis pathway.¹⁰⁵ The researchers showed that participants with at least one copy of the minor allele T of rs9608282 (located in the first intron of *SPECC1L*, upstream of *ADORA2A*) had a greater mean hippocampal volume and a greater neurogenesis-related hippocampal sub-region volume than participants bearing the other genotype. Horgusluoglu-Moloch et al. replicated these results in a meta-analysis that included two other cohorts (adding 277 additional participants) and evidenced an association with better episodic memory performance, and with a low cerebrospinal fluid total tau level. These results suggest that *ADORA2A* has a role in neurogenesis in adults. However, an analysis of the BRAINEAC brain tissue microarray-based gene expression database (http://www.braineac.org/) found that rs9608282 is not associated with *ADORA2A* expression levels in the hippocampus. Future research will have to validate this *SPECC1L* SNP and assess its potential function in the adenosine-related pathway.

Shinohara et al. identified an *ADORA2A* haplotype "A" (built from a C allele at rs2298383, a T allele at rs5751876, a deletion at rs35320474 and a C allele at rs4822492) as a risk factor for a severe form of encephalopathy in children following severe febrile seizures: acute encephalopathy with biphasic seizures and late reduced diffusion.¹⁴ The haplotype A was significantly more frequent in patients (*n*=85) than in controls (*n*=184). The most significant association was obtained in a recessive model (OR [95%CI]=2.32 [1.32-4.08]; *p*=0.003). The level of *ADORA2A* mRNA was 1.4-fold higher in the brain tissue of individuals with a homozygous haplotype A (*i.e.* diplotype AA), relative to individuals with AB or BB diplotypes. This finding was confirmed by the mRNA and protein expression levels measured in

lymphoblasts. Lastly, the researchers evidenced greater cAMP accumulation in response to adenosine in lymphoblasts with the AA diplotype than in cells with the BB diplotype (p=0.0006). These results support the hypothesis whereby elevated expression of *ADORA2A* in children may alter the intracellular adenosine/cAMP signal cascade, enhance excitatory neurotransmitter release, and thus favor excitotoxicity.

Neurologic traits

Two studies have suggested that a TT genotype at rs5751876 is associated with greater sensitivity to sensory stimuli. Firstly, *ADORA2A* variants were associated with differences in the efficiency of pre-attentive sensory memory subprocesses.¹⁰⁶ The researchers examined the results of a memory task after a visual stimulus in 199 healthy participants. Although the initial visual availability of stimulus information was greater in participants carrying the rarer homozygous genotype (*i.e.* TT genotypes for rs5751876 and rs2298383), less information was transferred to the working memory. This finding suggests that pre-attentive sensory memory subprocesses are modulated by *ADORA2A*. Secondly, Geiger et al. correlated functional MRI and genotyping results in healthy human subjects.¹⁰⁷ A TT genotype at rs5751876 (associated with an elevated anxiety risk) was found to modulate a fronto-insular network involved in attention and in the processing of interoceptive and exteroceptive information; it was suggested that this modulation increases the likelihood of anxiety and anxiety disorders.

In a pilot study of 19 healthy volunteers, Rupp et al. assessed the relationship between sleep restriction resistance and *ADORA2A* and *PER3* genotypes.¹⁰⁸ An

rs5751876 TT genotype group (n=9) showed lower behavioral resiliency after sleep restriction than a group with the CT genotype (n=9). Several studies has confirmed the relationship between *ADORA2A* variants and sleep, with notably an association between genetic variants in *ADORA2A* (especially rs5751876) and individual sensitivity to sleep disturbance caused by caffeine consumption.^{109–112}

To conclude, the genetic variants in *ADORA2A* are the most studied among adenosine gene receptor. They are associated with a lot of CNS diseases or traits, but sometimes provided contradictory results.^{76,77} The most likely genetic associations concern HD, epilepsy, depression and anxiety disorders because of functional assessment of the variants, prospective design and/or studies on larger samples. However, efforts are needed to identify the true functional variant(s).

ADORA2B

Gene structure (Figure 1.C)

The human *ADORA2B* gene was cloned from a human hippocampal cDNA library in 1992.¹¹³ It spans 31 kb, and is located on chromosome 17p12.^{114,115} Although only a single copy of *ADORA2B* is present in the genome, a pseudogene (adenosine A2B receptor pseudogene 1, *ADORA2BP1*) was found on chromosome 1q32.2.¹¹⁵ *ADORA2BP1* is a processed pseudogene that lacks an intronic region. It may have been created during evolution by integration of *ADORA2B* mRNA into a chromosome break site, followed by DNA synthesis and repair. *ADORA2B* is composed of two coding exons and an intron. The gene comprises a 1756 bp CpG island encompassing the whole of exon 1 and the core promoter. The *ADORA2B* promoter

contains binding sites for hypoxia-inducible factor 1-alpha,^{116,117} hypoxia-inducible factor 2-alpha,¹¹⁸ and Myb-related protein B.¹¹⁹ According to the GTEx database,²⁵ *ADORA2B* mRNAs are found in all tissues, with the highest expression levels in the skin, esophagus, vagina, colon, and anterior cingulate cortex.

Genetic risk factor in human diseases

ADORA2B has been highlighted as a potential modifier gene in prodromal HD with a variation in gray matter volume.¹¹ After correcting for the effect of the disease burden effect (*i.e.* the number of *HTT* gene CAG repeats, and age), Liu et al. searched for genetic factors that influenced prodromal HD in a multicenter study including 715 prodromal individuals. A GWAS did not reveal an association that remained significant after false discovery rate correction. However, a separate analysis of 3,404 SNPs in 310 HD pathway genes evidenced a correlation between two SNPs in strong linkage disequilibrium (rs71358386 in *NCOR1*, and rs78804732 in *ADORA2B* intron 1) and the gray matter volume in the cuneus lobe; the presence of the minor alleles was correlated with a low gray matter volume. The major limitation consisted in the absence of correlation between SNPs and motor or cognitive function in prodromal individuals. Consequently, functional studies and longitudinal study in HD patients will be required to assess a modifier role of *ADORA2B*.

ADORA3

Gene structure (Figure 1.D)

The human ADORA3 gene was first cloned in 1992.^{120,121} It is a short (4.3 kb) gene and is located on chromosome 1p13.2.^{122,123} It is composed of two exons and one intron,¹²⁴ and encodes a single 318-aa protein. According to the GTEx database,²⁵ *ADORA3* mRNAs expression levels are highest in the testis, spinal cord, adrenal gland, subtantia nigra, and spleen.

The entire *ADORA3* gene is encompassed by the first intron of the isoforms 2 to 4 of transmembrane and immunoglobulin domain containing 3 gene (*TMIGD3*). Moreover, isoform 1 of *TMIGD3* shares its 5' terminal exon with almost all of the first exon of *ADORA3*. This alternative TSS results in a fusion product that includes the 116 N-terminal amino acids of A₃R. Data on *TMIGD3* are scarce, although one study revealed a role for *TMIGD3* isoform 1 as a suppressor of NF-κB and osteosarcoma progression.¹²⁵ The *ADORA3* gene's promoter region has not been extensively studied but reportedly does not contain a TATA box; however, the existence of a CCAT box is subject to debate.^{126,127} Our analysis of the *ADORA3* promoter region evidenced a CAAT box located 98 bp upstream of the TSS. Functional binding of Octamer-Binding Protein 1 and NF-κB transcription factors was found at the proximal part of the *ADORA3* promoter region, and binding was inhibited by the rs1544224 polymorphism (c.-564T>C, p.?) located in the 5'-UTR (encoded by exon 1).¹²⁸

Genetic risk factor in human diseases

Prompted by A_3R 's known role in the neuromodulation of serotonin (a neurotransmitter involved in many neuropsychiatric phenotypes, including autism),

Campbell et al. examined a putative genetic link between rare coding variants of ADORA3 and autism. Using allelic association testing in 958 families (1,649 probands; 4,150 samples), the researchers found no evidence supporting an association between a common variant at ADORA3 locus and ASD. However, in a case-control study including 185 unrelated ASD cases and 305 controls followed by a subsequent replication analysis of exome sequence data from 339 cases and pairmatched controls,¹² two rare coding variants (Leu90Val and Val171lle) were significantly enriched in cases vs. controls. They were then assessed in vitro for their functional impact but showed incomplete penetrance and had opposite effects on antidepressant-sensitive 5-HT transporter activity. The relationship with disease was significant in the combined analysis using whole-exome sequencing data but not in the replication analysis. Taking into account replication data, only Leu90Val remain statistically significant (p=0.040). In total, Leu90Val was found in 4 patients among 524 ASD cases and was absent in 644 control. The subjects in this study were not matched based on genome-wide genotype data. This limitation could constitute a bias, especially for the frequency of rare variants which can be very different between subgroups of a population with similar ethnicity. Such limitation is important regarding the small number of carriers. Although the researchers provided functional data of the two rare variants in cell culture, the association of rare ADORA3 variants and ASD should be interpreted with caution. Further studies in larger samples are needed to confirm this genetic association.

Which genetic variants in adenosine receptors genes are functionally relevant? Despite their associations in several diseases, the functional effect of variants in adenosine receptors genes has been poorly described. The three more frequently studied ADORA2A SNPs (i.e. rs5751876, rs2298383 and rs35320474) and their associations with human brain diseases or traits are summarized in Table 2. The most-studied variant (rs5751876, c.1083T>C, p.(=)) is a synonymous variant that lacks a validated functional impact on protein function or synthesis. It has been suggested that several ADORA2A polymorphisms in linkage disequilibrium with rs5751876 are causal variants. For example, rs35320474 (c.*458dup, p.?, located in the 3'-UTR region of exon 4) has been discussed as a variant that might change A_{2A}R expression.⁸¹ Indeed, there is no data in favor of a transcriptional effect of rs35320474 in GTEx, but it modifies a U-rich motif. As these motifs can recruit RNAbinding proteins, rs35320474 might modify the translation of ADORA2A mRNA. However, such functional effect has not been published for rs35320474. The second most intensively studied polymorphism in ADORA2A is the rs2298383 SNP (c.-3206C>T, p.?), located in a regulatory triplex-forming oligonucleotide target sequence that has been conserved in mammals and is involved in the regulation of gene expression.¹²⁹ It is also noteworthy that rs2298383 variant is located in the terminal 3' exon of one of the isoforms of ADORA2A-AS1 and is associated with ADORA2A-AS1 mRNA expression.²⁵ It is well known that antisense RNAs regulate gene expression on the opposite strand. Hence, it would be interesting to study the effect of rs2298383 on the mRNA expression of ADORA2A and ADORA2A-AS1. In line with a putative transcriptional effect, rs2298383 is also an expression quantitative locus associated with ADORA2A mRNA expression in diverse tissues including different regions of the brain (i.e. nucleus accumbens, amygdala, cortex and

hippocampus) (https://gtexportal.org/home/snp/rs2298383).²⁵ More precisely, the TT genotype is associated with increased *ADORA2A* mRNA. But these results are in contradiction with the functional studies reported by Shinohara et al. who found elevated A_{2A}R mRNA and protein expression levels and protein activity with associated *ADORA2A* haplotype A, including the C allele of rs2298383.¹⁴ It is then probable that rs2298383, like the other variants in *ADORA2A*, only tag the true causal variant(s). In 2014, Hohoff et al. used positron emission tomography to investigate the influence of *ADORA1* and *ADORA2A* SNPs on the *in vivo* distribution and availability of cerebral A₁Rs in healthy humans.¹³⁰

Indeed, $A_{1A}Rs$ and $A_{2A}Rs$ are frequently co-expressed, and interact directly by heteromerization. Hohoff et al. observed that *ADORA2A* SNPs are associated with increased binding and availability of A_1Rs in various regions of the brain. The CT and TT genotypes at rs2236624 are associated with greater distribution and availability of A_1R in the superior frontal gyrus and dorsolateral prefrontal cortex, whereas the CT and TT genotypes at rs5751876 are associated with greater distribution and availability on and availability in the entorhinal cortex, hippocampus, and cortex as a whole. The brain regions associated with *ADORA2A* SNPs are involved in emotion processing, which might contribute to their effects on anxiety and panic disorder. The SNPs rs10920568 and rs3766566 in *ADORA1* also correlated with the *in vivo* distribution and availability of cerebral A_1Rs .

None of *ADORA1* common variants rs3766553 and rs10920573 (associated with epilepsy after traumatic brain injury⁴⁸) has been assessed during functional studies. We noted that rs10920573, located in intron 5, reported in the GTEx database,²⁵ is associated with variation in expression of *ADORA1* mRNA in the whole blood (p=8.8 e⁻¹⁵). Rarer allele T, and genotype TT of rs10920573 are associated with an

increase of ADORA1 mRNA, but Wagner et al. found that it was the heterozygotes genotype CT that was at greatest risk of epilepsy which is not in favor of protective effect of either allele. Functional impact of the two SNPs reported by Liu et al. is not established and we even don't know which gene between ADORA2B and NCOR1 is implied in prodromal HD.¹¹ NCOR1 and ADORA2A (but not ADORA2B) are part of the HD pathway, but it is noteworthy that SNP rs71358386 in NCOR1 regulates expression of ADORA2B (and not NCOR1) in various tissues including the brain cortex.²⁵ Minor allele G of rs71358386 is negatively correlated to grey matter volume in the cuneus lobe and is associated with low ADORA2B mRNA.²⁵ How a decrease in ADORA2B mRNA (and then probably in A_{2B}R too) induces a decrease in grey matter and potentially favors HD progression is not fully understood. However, recent studies demonstrated the existence of brain A_{2A}R-A_{2B}R heteroreceptor complexes, that might inhibit neurodegeneration in models of brain diseases by A2AR inactivation.¹³¹ The functional effect of the rare coding variant Leu90Val in ADORA3 was demonstrated in cell culture by Campbell et al. The researchers showed elevated basal cGMP levels and response to an A_{3A}R agonist compared with the wild type receptor. Moreover, the cells transfected with ADORA3 and 5-HT transporter gene showed an increase in antidepressant-sensitive 5-HT transporter activity.¹²

Conclusions

The results of a large number of genetic studies suggest that variants in adenosine receptor genes are associated with neurologic or psychiatric diseases. However, most of the research performed to date has been based on small case-control studies. Without any functional evidence and replication in other populations, such

studies do not provide enough evidences and must be interpreted with cautions because smaller sample size increase the risk that observations will be due to chance. The multiplication of genetic studies of recurrent genetic variants in multiple disorders, as seen for rs5751876 in psychiatric disorders, increases the risk of false-positive too. It may explain the heterogeneity of the findings and for example why the incriminated allele of rs5751876 associated with panic disorder varies depending the studies^{76,77} or why *ADORA2A* is absent in larger studies like GWAS on anxiety disorder.¹³² However, there are too many genetic associations between *ADORA2A* SNPs and anxiety disorders to disregard them all. Moreover, a growing body of evidence suggested that A_{2A}R influence the fear network¹³³ and bring rational explanations to these genetic associations. This is in line with the recent report of increased A_{1A}R availability in regions of the fear network, in *ADORA2A* risk allele carriers.¹³⁰

The development of new study designs in genetic research has prompted renewed interest in specific phenotypes,¹⁴ cross-sectional population-based analysis¹⁵ and rare variants.^{12,13} The study of rare variant is of particular interest as it is based on the common disease-rare variant hypothesis, according an increasing role of rare variant in human complex diseases or traits.¹³⁴ A screen for rare variants and/or the aggregation of variants in regulatory regions could also be interesting for *ADORA2A* because none of the SNPs reported to date have proved to be functional and may only be tags for one several rare variants in linkage disequilibrium. Such rare variants restricted to smaller groups of populations could also explain the ethnicity-dependent and/or heterogeneity of the findings concerning *ADORA2A* SNPs.

In the future, genetic research should take account of a number of confounding factors (e.g. ethnicity and gender) and common gene-environment interactions (e.g.

caffeine intake). Further studies are particularly needed to integrate the role of caffeine, a widely consumed antagonist of adenosine receptors, in HD,¹⁰³ depression¹³⁵ and neuropsychiatric disorders.¹⁸

Another major task relates to the characterization of the genes' structure and regulatory mechanisms. Indeed, most of the structural specificities and genetic variants in the adenosine receptor genes do not lead to changes in the protein sequences or biochemical properties. However, adenosine receptor expression is frequently deregulated in a context of neurodegenerative disease. Thus, there is great interest in the role of adenosine receptor genes and their posttranslational regulation as it could be the key to understand the deregulations of adenosine receptors in many common neurodegenerative diseases.

Lastly, there is a growing body of evidence suggesting a modulation of the neurodegenerative effects of $A_{2A}R$ through its interaction with other adenosine receptors.¹³¹ Further studies should include all robust associations in adenosine receptors genes, in related non-coding RNAs and in other relevant genes. For example, it would be interesting to study together variants in *ADORA2A* and in different genes implicated in adenosine metabolism or nucleoside transporter, such as the variants in *SLC29A3* which are associated with altered adenosine transport and predispose to depression with disturbed sleep.¹³⁶

Acknowledgments

The research leading to these results received funding from the French government's LabEx program ("Development of Innovative Strategies for a Transdisciplinary approach to Alzheimer's disease"- DISTALZ), the University of Lille, and the Institut National de la Santé et de la Recherche Médicale (INSERM).

Author disclosure statements

No competing financial interests exist.

References

1. Fredholm BB, Chen J-F, Cunha RA, et al. Adenosine and Brain Function. In: International Review of Neurobiology. Elsevier; pp. 191–270.

2. Burnstock G, Fredholm BB, Verkhratsky A. Adenosine and ATP receptors in the brain. Curr Top Med Chem 2011;11:973–1011.

3. Cunha RA. How does adenosine control neuronal dysfunction and neurodegeneration? J Neurochem 2016;139:1019–1055.

4. Ciruela F. Presynaptic Control of Striatal Glutamatergic Neurotransmission by Adenosine A1-A2A Receptor Heteromers. J Neurosci 2006;26:2080–2087.

5. Gines S, Hillion J, Torvinen M, et al. Dopamine D1 and adenosine A1 receptors form functionally interacting heteromeric complexes. Proc Natl Acad Sci 2000;97:8606–8611.

6. Hillion J, Canals M, Torvinen M, et al. Coaggregation, Cointernalization, and Codesensitization of Adenosine A $_{2A}$ Receptors and Dopamine D $_2$ Receptors. J Biol Chem 2002;277:18091–18097.

7. Cunha RA. Neuroprotection by adenosine in the brain: From A1 receptor activation to A2A receptor blockade. Purinergic Signal 2005;1:111–134.

8. Gomes CV, Kaster MP, Tomé AR, et al. Adenosine receptors and brain diseases: Neuroprotection and neurodegeneration. Biochim Biophys Acta BBA - Biomembr 2011;1808:1380–1399.

9. Vecchio EA, White PJ, May LT. The adenosine A2B G protein-coupled receptor: Recent advances and therapeutic implications. Pharmacol Ther 2019;198:20–33.

10. Borea PA, Varani K, Vincenzi F, et al. The A3 Adenosine Receptor: History and Perspectives. Pharmacol Rev 2014;67:74–102.

11. Liu J, Ciarochi J, Calhoun VD, et al. Genetics Modulate Gray Matter Variation Beyond Disease Burden in Prodromal Huntington's Disease. Front Neurol;9 . Epub ahead of print March 29, 2018. DOI: 10.3389/fneur.2018.00190.

12. Campbell NG, Zhu C-B, Lindler KM, et al. Rare coding variants of the adenosine A3 receptor are increased in autism: on the trail of the serotonin transporter regulome. Mol Autism 2013;4:28.

13. Clark SL, McClay JL, Adkins DE, et al. Deep Sequencing of 71 Candidate Genes to Characterize Variation Associated with Alcohol Dependence. Alcohol Clin Exp Res 2017;41:711–718.

14. Shinohara M, Saitoh M, Nishizawa D, et al. ADORA2A polymorphism predisposes children to encephalopathy with febrile status epilepticus. Neurology 2013;80:1571–1576.

15. Oliveira S, Ardais AP, Bastos CR, et al. Impact of genetic variations in ADORA2A gene on depression and symptoms: a cross-sectional population-based study. Purinergic Signal 2019;15:37–44.

16. Fredholm BB, Arslan G, Halldner L, et al. Structure and function of adenosine receptors and their genes. Naunyn Schmiedebergs Arch Pharmacol 2000;362:364–374.

17. Chen J-F, Lee C, Chern Y. Adenosine Receptor Neurobiology: Overview. In: International Review of Neurobiology. Elsevier; pp. 1–49.

18. Ferré S, Díaz-Ríos M, Salamone JD, et al. New Developments on the Adenosine Mechanisms of the Central Effects of Caffeine and Their Implications for Neuropsychiatric Disorders. J Caffeine Adenosine Res 2018;8:121–130.

19. Chen J-F. Adenosine Receptor Control of Cognition in Normal and Disease.In: International Review of Neurobiology. Elsevier; pp. 257–307.

20. Townsend-Nicholson A, Shine J. Molecular cloning and characterisation of a human brain A1 adenosine receptor cDNA. Brain Res Mol Brain Res 1992;16:365–370.

21. Libert F, Van Sande J, Lefort A, et al. Cloning and functional characterization of a human AI adenosine receptor. Biochem Biophys Res Commun 1992;187:919–926.

22. Townsend-Nicholson A, Baker E, Schofield PR, et al. Localization of the adenosine A1 receptor subtype gene (ADORA1) to chromosome 1q32.1. Genomics 1995;26:423–425.

23. Ren H, Stiles GL. Characterization of the human A1 adenosine receptor gene. Evidence for alternative splicing. J Biol Chem 1994;269:3104–3110.

24. Ren H, Stiles GL. Posttranscriptional mRNA processing as a mechanism for regulation of human A1 adenosine receptor expression. Proc Natl Acad Sci 1994;91:4864–4866.

25. Carithers LJ, Ardlie K, Barcus M, et al. A Novel Approach to High-Quality Postmortem Tissue Procurement: The GTEx Project. Biopreservation Biobanking 2015;13:311–319.

26. Harrow J, Frankish A, Gonzalez JM, et al. GENCODE: The reference human genome annotation for The ENCODE Project. Genome Res 2012;22:1760–1774.

27. Ren H, Stiles GL. Separate promoters in the human A1 adenosine receptor gene direct the synthesis of distinct messenger RNAs that regulate receptor abundance. Mol Pharmacol 1995;48:975–980.

28. Nie Z, Mei Y, Ford M, et al. Oxidative Stress Increases A $_1$ Adenosine Receptor Expression by Activating Nuclear Factor κ B. Mol Pharmacol 1998;53:663–669.

29. Ren H, Stiles GL. Dexamethasone Stimulates Human A ₁ Adenosine Receptor (A ₁ AR) Gene Expression through Multiple Regulatory Sites in Promoter B. Mol Pharmacol 1999;55:309–316.

30. Ren H, Stiles GL. A Single-Stranded DNA Binding Site in the Human A ₁ Adenosine Receptor Gene Promoter. Mol Pharmacol 1998;53:43–51.

31. Lee C, Lai H-L, Lee Y-C, et al. The A _{2A} Adenosine Receptor Is a Dual Coding Gene: A NOVEL MECHANISM OF GENE USAGE AND SIGNAL TRANSDUCTION. J Biol Chem 2014;289:1257–1270.

32. OUP accepted manuscript. Nucleic Acids Res . Epub ahead of print 2017. DOI: 10.1093/nar/gkx1020.

33. Gotoh L, Mitsuyasu H, Kobayashi Y, et al. Association analysis of adenosine A1 receptor gene (ADORA1) polymorphisms with schizophrenia in a Japanese population: Psychiatr Genet 2009;19:328–335.

34. Janik P, Berdyński M, Safranow K, et al. Association of ADORA1 rs2228079 and ADORA2A rs5751876 Polymorphisms with Gilles de la Tourette Syndrome in the Polish Population. PLOS ONE 2015;10:e0136754.

35. Deckert J, Nöthen MM, Albus M, et al. Adenosine A1 receptor and bipolar affective disorder: systematic screening of the gene and association studies. Am J Med Genet 1998;81:18–23.

36. Kobayashi >, Ujike H, Iwata N, et al. Association Analysis of the Adenosine A1 Receptor Gene Polymorphisms in Patients with Methamphetamine Dependence/Psychosis. Curr Neuropharmacol 2011;9:137–142.

37. Rogers PJ, Hohoff C, Heatherley SV, et al. Association of the Anxiogenic and Alerting Effects of Caffeine with ADORA2A and ADORA1 Polymorphisms and Habitual Level of Caffeine Consumption. Neuropsychopharmacology 2010;35:1973–1983.

38. Schizophrenia Working Group of the Psychiatric Genomics Consortium. Biological insights from 108 schizophrenia-associated genetic loci. Nature 2014;511:421–427.

39. Goes FS, McGrath J, Avramopoulos D, et al. Genome-wide association study of schizophrenia in Ashkenazi Jews. Am J Med Genet B Neuropsychiatr Genet 2015;168:649–659.

40. Yu D, Sul JH, Tsetsos F, et al. Interrogating the Genetic Determinants of Tourette's Syndrome and Other Tic Disorders Through Genome-Wide Association Studies. Am J Psychiatry 2019;176:217–227.

41. Treutlein J, Frank J, Streit F, et al. Genetic Contribution to Alcohol Dependence: Investigation of a Heterogeneous German Sample of Individuals with Alcohol Dependence, Chronic Alcoholic Pancreatitis, and Alcohol-Related Cirrhosis. Genes 2017;8:183.

42. Jaberi E, Rohani M, Shahidi GA, et al. Mutation in *ADORA1* identified as likely cause of early-onset parkinsonism and cognitive dysfunction: *ADORA1* Mutation Associated with Parkinsonism. Mov Disord 2016;31:1004–1011.

43. Satake W, Nakabayashi Y, Mizuta I, et al. Genome-wide association study identifies common variants at four loci as genetic risk factors for Parkinson's disease. Nat Genet 2009;41:1303–1307.

44. Simón-Sánchez J, Schulte C, Bras JM, et al. Genome-wide association study reveals genetic risk underlying Parkinson's disease. Nat Genet 2009;41:1308–1312.

45. Blauwendraat C, Nalls MA, Federoff M, et al. *ADORA1* mutations are not a common cause of Parkinson's disease and dementia with Lewy bodies: Mutational Analysis In *ADORA1*. Mov Disord 2017;32:298–299.

46. Khodadadi H, Azcona LJ, Aghamollaii V, et al. *PTRHD1* (C2orf79) mutations lead to autosomal-recessive intellectual disability and parkinsonism: *PTRHD1* Mutation In ID and Parkinsonism. Mov Disord 2017;32:287–291.

47. Elahi E. *PTRHD1* and possibly *ADORA1* mutations contribute to Parkinsonism with intellectual disability. Mov Disord 2018;33:174–174.

48. Wagner AK, Miller MA, Scanlon J, et al. Adenosine A1 receptor gene variants associated with post-traumatic seizures after severe TBI. Epilepsy Res 2010;90:259–272.

49. Furlong TJ, Pierce KD, Selbie LA, et al. Molecular characterization of a human brain adenosine A2 receptor. Brain Res Mol Brain Res 1992;15:62–66.

50. MacCollin M, Peterfreund R, MacDonald M, et al. Mapping of a Human A2a Adenosine Receptor (ADORA2) to Chromosome 22. Genomics 1994;20:332–333.

51. Le F, Townsend-Nicholson A, Baker E, et al. Characterization and Chromosomal Localization of the Human A2a Adenosine Receptor Gene: ADORA2A. Biochem Biophys Res Commun 1996;223:461–467.

52. Peterfreund RA, MacCollin M, Gusella J, et al. Characterization and Expression of the Human A2a Adenosine Receptor Gene. J Neurochem 2002;66:362–368.

53. Fredholm BB, Chern Y, Franco R, et al. Aspects of the general biology of adenosine A2A signaling. Prog Neurobiol 2007;83:263–276.

54. Yu L, Frith MC, Suzuki Y, et al. Characterization of genomic organization of the adenosine A2A receptor gene by molecular and bioinformatics analyses. Brain Res 2004;1000:156–173.

55. Kreth S, Ledderose C, Kaufmann I, et al. Differential expression of 5'-UTR splice variants of the adenosine A _{2A} receptor gene in human granulocytes: identification, characterization, and functional impact on activation. FASEB J 2008;22:3276–3286.

56. Chu Y-Y, Tu K-H, Lee YC, et al. Characterization of the Rat A2a Adenosine Receptor Gene. DNA Cell Biol 1996;15:329–337.

57. Lee YC, Lai HL, Sun CN, et al. Identification of nuclear factor 1 (NF1) as a transcriptional modulator of rat A(2A) adenosine receptor. Brain Res Mol Brain Res 2003;111:61–73.

58. Chiang M-C, Lee Y-C, Huang C-L, et al. cAMP-response Element-binding Protein Contributes to Suppression of the A _{2A} Adenosine Receptor Promoter by Mutant Huntingtin with Expanded Polyglutamine Residues. J Biol Chem 2005;280:14331–14340.

59. Brown ST, Reyes EP, Nurse CA. Chronic hypoxia upregulates adenosine 2a receptor expression in chromaffin cells via hypoxia inducible factor-2α: Role in modulating secretion. Biochem Biophys Res Commun 2011;412:466–472.

60. Morello S, Ito K, Yamamura S, et al. IL-1β and TNF-α Regulation of the Adenosine Receptor (A $_{2A}$) Expression: Differential Requirement for NF-κB Binding to the Proximal Promoter. J Immunol 2006;177:7173–7183.

61. Ahmad A, Ahmad S, Glover L, et al. Adenosine A2A receptor is a unique angiogenic target of HIF-2 in pulmonary endothelial cells. Proc Natl Acad Sci 2009;106:10684–10689.

62. Ernst J, Kellis M. Discovery and characterization of chromatin states for systematic annotation of the human genome. Nat Biotechnol 2010;28:817–825.

63. The ENCODE Project Consortium. An integrated encyclopedia of DNA elements in the human genome. Nature 2012;489:57–74.

64. Buira SP, Albasanz JL, Dentesano G, et al. DNA methylation regulates adenosine A _{2A} receptor cell surface expression levels. J Neurochem 2010;112:1273–1285.

65. Buira SP, Dentesano G, Albasanz JL, et al. DNA methylation and Yin Yang-1 repress adenosine A2A receptor levels in human brain: ZBP-89 and Yin Yang-1 control ADORA2A expression. J Neurochem 2010;115:283–295.

66. Villar-Menéndez I, Nuñez F, Díaz-Sánchez S, et al. Striatal adenosine A2A receptor expression is controlled by S-adenosyl-L-methionine-mediated methylation. Purinergic Signal 2014;10:523–528.

67. Villar-Menéndez I, Blanch M, Tyebji S, et al. Increased 5-Methylcytosine and Decreased 5-Hydroxymethylcytosine Levels are Associated with Reduced Striatal A2AR Levels in Huntington's Disease. NeuroMolecular Med 2013;15:295–309.

68. Villar-Menéndez I, Díaz-Sánchez S, Blanch M, et al. Reduced striatal adenosine A2A receptor levels define a molecular subgroup in schizophrenia. J Psychiatr Res 2014;51:49–59.

69. Hinnebusch AG, Ivanov IP, Sonenberg N. Translational control by 5'untranslated regions of eukaryotic mRNAs. Science 2016;352:1413–1416.

70. The 5' Untranslated Regions of the Rat A2A Adenosine Receptor Gene Function as Negative Translational Regulators. J Neurochem 2002;73:1790–1798.

71. Lee C, Lai H-L, Lee Y-C, et al. The A _{2A} Adenosine Receptor Is a Dual Coding Gene: A NOVEL MECHANISM OF GENE USAGE AND SIGNAL TRANSDUCTION. J Biol Chem 2014;289:1257–1270. 72. Djebali S, Lagarde J, Kapranov P, et al. Evidence for Transcript Networks Composed of Chimeric RNAs in Human Cells. PLoS ONE 2012;7:e28213.

73. Guennewig B, Cooper AA. The Central Role of Noncoding RNA in the Brain.In: International Review of Neurobiology. Elsevier; pp. 153–194.

74. Briggs JA, Wolvetang EJ, Mattick JS, et al. Mechanisms of Long Non-coding RNAs in Mammalian Nervous System Development, Plasticity, Disease, and Evolution. Neuron 2015;88:861–877.

75. Salta E, De Strooper B. Noncoding RNAs in neurodegeneration. Nat Rev Neurosci 2017;18:627–640.

76. Deckert J, Nöthen MM, Franke P, et al. Systematic mutation screening and association study of the A1 and A2a adenosine receptor genes in panic disorder suggest a contribution of the A2a gene to the development of disease. Mol Psychiatry 1998;3:81–85.

77. Hamilton SP, Slager SL, de Leon AB, et al. Evidence for Genetic Linkage Between a Polymorphism in the Adenosine 2A Receptor and Panic Disorder. Neuropsychopharmacology 2004;29:558–565.

78. Hohoff C, Domschke K, Schwarte K, et al. Sympathetic activity relates to adenosine A2A receptor gene variation in blood-injury phobia. J Neural Transm 2009;116:659–662.

79. Hohoff C, Mullings EL, Heatherley SV, et al. Adenosine A2A receptor gene: Evidence for association of risk variants with panic disorder and anxious personality. J Psychiatr Res 2010;44:930–937. 80. Gugliandolo A, Gangemi C, Caccamo D, et al. The RS685012 Polymorphism of ACCN2, the Human Ortholog of Murine Acid-Sensing Ion Channel (ASIC1) Gene, is Highly Represented in Patients with Panic Disorder. NeuroMolecular Med 2016;18:91–98.

81. Alsene K, Deckert J, Sand P, et al. Association Between A2a Receptor Gene Polymorphisms and Caffeine-Induced Anxiety. Neuropsychopharmacology 2003;28:1694–1702.

82. Childs E, Hohoff C, Deckert J, et al. Association between ADORA2A and DRD2 Polymorphisms and Caffeine-Induced Anxiety. Neuropsychopharmacology 2008;33:2791–2800.

83. Hohoff C, McDonald JM, Baune BT, et al. Interindividual variation in anxiety response to amphetamine: Possible role for adenosine A _{2A} receptor gene variants.
Am J Med Genet B Neuropsychiatr Genet 2005;139B:42–44.

84. Domschke K, Gajewska A, Winter B, et al. ADORA2A Gene Variation, Caffeine, and Emotional Processing: A Multi-level Interaction on Startle Reflex. Neuropsychopharmacology 2012;37:759–769.

85. Gajewska A, Blumenthal TD, Winter B, et al. Effects of ADORA2A gene variation and caffeine on prepulse inhibition: A multi-level risk model of anxiety. Prog Neuropsychopharmacol Biol Psychiatry 2013;40:115–121.

86. Renda G, Committeri G, Zimarino M, et al. Genetic determinants of cognitive responses to caffeine drinking identified from a double-blind, randomized, controlled trial. Eur Neuropsychopharmacol 2015;25:798–807.

87. Ledent C, Vaugeois J-M, Schiffmann SN, et al. Aggressiveness, hypoalgesia and high blood pressure in mice lacking the adenosine A2a receptor. Nature 1997;388:674–678.

88. Lam P, Hong C-J, Tsai S-J. Association study of A2a adenosine receptor genetic polymorphism in panic disorder. Neurosci Lett 2005;378:98–101.

89. Yamada K, Hattori E, Shimizu M, et al. Association studies of the cholecystokinin B receptor and A2a adenosine receptor genes in panic disorder. J Neural Transm 2001;108:837–848.

90. Subaran RL, Talati A, Hamilton SP, et al. A survey of putative anxietyassociated genes in panic disorder patients with and without bladder symptoms: Psychiatr Genet 2012;22:271–278.

91. Molero Y, Gumpert C, Serlachius E, et al. A study of the possible association between adenosine A _{2A} receptor gene polymorphisms and attention-deficit hyperactivity disorder traits: Adenosine A _{2A} receptor gene and ADHD traits. Genes Brain Behav 2013;12:305–310.

92. Fraporti TT, Contini V, Tovo-Rodrigues L, et al. Synergistic effects between ADORA2A and DRD2 genes on anxiety disorders in children with ADHD. Prog Neuropsychopharmacol Biol Psychiatry 2019;93:214–220.

93. Freitag CM, Agelopoulos K, Huy E, et al. Adenosine A2A receptor gene (ADORA2A) variants may increase autistic symptoms and anxiety in autism spectrum disorder. Eur Child Adolesc Psychiatry 2010;19:67–74.

94. Jagannathan K, Calhoun VD, Gelernter J, et al. Genetic Associations of Brain Structural Networks in Schizophrenia: A Preliminary Study. Biol Psychiatry 2010;68:657–666.

95. Deckert J, Nöthen MM, Rietschel M, et al. Human adenosine A2a receptor (A2aAR) gene: systematic mutation screening in patients with schizophrenia. J Neural Transm 1996;103:1447–1455.

96. Hong C-J, Liu H-C, Liu T-Y, et al. Association studies of the adenosine A2a receptor (1976T > C) genetic polymorphism in Parkinson's disease and schizophrenia. J Neural Transm 2005;112:1503–1510.

97. Luu S-U, Liao H-M, Hung T-W, et al. Mutation analysis of adenosine A2a receptor gene and interaction study with dopamine D2 receptor gene in schizophrenia: Psychiatr Genet 2008;18:43.

98. Tsai S-J, Hong C-J, Hou S-J, et al. Association study of adenosine A2a receptor (1976C>T) genetic polymorphism and mood disorders and age of onset: Psychiatr Genet 2006;16:185.

99. Hohoff C, Marziniak M, Lesch K-P, et al. An Adenosine A _{2A} Receptor Gene Haplotype is Associated with Migraine With Aura. Cephalalgia 2007;27:177–181.

100. Dhaenens C-M, Burnouf S, Simonin C, et al. A genetic variation in the ADORA2A gene modifies age at onset in Huntington's disease. Neurobiol Dis 2009;35:474–476.

101. Taherzadeh-Fard E, Saft C, Wieczorek S, et al. Age at onset in Huntington's disease: replication study on the associations of ADORA2A, HAP1 and OGG1. neurogenetics 2010;11:435–439.

102. Vital M, Bidegain E, Raggio V, et al. Molecular characterization of genes modifying the age at onset in Huntington's disease in Uruguayan patients. Int J Neurosci 2015;1–4.

103. Simonin C, Duru C, Salleron J, et al. Association between caffeine intake and age at onset in Huntington's disease. Neurobiol Dis 2013;58:179–182.

104. Blum D, Chern Y, Domenici MR, et al. The Role of Adenosine Tone and Adenosine Receptors in Huntington's Disease. J Caffeine Adenosine Res 2018;8:43–58.

105. Horgusluoglu-Moloch E, Nho K, Risacher SL, et al. Targeted neurogenesis pathway-based gene analysis identifies ADORA2A associated with hippocampal volume in mild cognitive impairment and Alzheimer's disease. Neurobiol Aging 2017;60:92–103.

106. Beste C, Stock A-K, Ness V, et al. Differential effects of ADORA2A gene variations in pre-attentive visual sensory memory subprocesses. Eur Neuropsychopharmacol 2012;22:555–561.

107. Geiger MJ, Domschke K, Homola GA, et al. ADORA2A genotype modulates interoceptive and exteroceptive processing in a fronto-insular network. Eur Neuropsychopharmacol 2016;26:1274–1285.

108. Rupp TL, Wesensten NJ, Newman R, et al. *PER3* and *ADORA2A* polymorphisms impact neurobehavioral performance during sleep restriction. J Sleep Res 2013;22:160–165.

109. Rétey JV, Adam M, Khatami R, et al. A Genetic Variation in the Adenosine A2A Receptor Gene (ADORA2A) Contributes to Individual Sensitivity to Caffeine Effects on Sleep. Clin Pharmacol Ther 2007;81:692–698.

110. Bodenmann S, Hohoff C, Freitag C, et al. Polymorphisms of *ADORA2A* modulate psychomotor vigilance and the effects of caffeine on neurobehavioural performance and sleep EEG after sleep deprivation: *ADORA2A* haplotypes, vigilance and CNS stimulants. Br J Pharmacol 2012;165:1904–1913.

111. Byrne EM, Johnson J, McRae AF, et al. A Genome-Wide Association Study of Caffeine-Related Sleep Disturbance: Confirmation of a Role for a Common Variant in the Adenosine Receptor. Sleep 2012;35:967–975.

112. Nunes RA, Mazzotti DR, Hirotsu C, et al. The association between caffeine consumption and objective sleep variables is dependent on ADORA2A c.1083T>C genotypes. Sleep Med 2017;30:210–215.

113. Pierce KD, Furlong TJ, Selbie LA, et al. Molecular cloning and expression of an adenosine A2b receptor from human brain. Biochem Biophys Res Commun 1992;187:86–93.

114. Townsend-Nicholson A, Baker E, Sutherland GR, et al. Localization of the adenosine A2b receptor subtype gene (ADORA2B) to chromosome 17p11.2-p12 by FISH and PCR screening of somatic cell hybrids. Genomics 1995;25:605–607.

115. Jacobson MA, Johnson RG, Luneau CJ, et al. Cloning and Chromosomal Localization of the Human A2b Adenosine Receptor Gene (ADORA2B) and Its Pseudogene. Genomics 1995;27:374–376.

116. Kong T, Westerman KA, Faigle M, et al. HIF-dependent induction of adenosine A2B receptor in hypoxia. FASEB J 2006;20:2242–2250.

117. Eckle T, Kewley EM, Brodsky KS, et al. Identification of Hypoxia-Inducible Factor HIF-1A as Transcriptional Regulator of the A2B Adenosine Receptor during Acute Lung Injury. J Immunol 2014;192:1249–1256.

118. Yang M, Ma C, Liu S, et al. HIF-dependent induction of adenosine receptor A2b skews human dendritic cells to a Th2-stimulating phenotype under hypoxia. Immunol Cell Biol 2010;88:165–171.

119. St. Hilaire C, Yang D, Schreiber BM, et al. B-Myb regulates the A2B adenosine receptor in vascular smooth muscle cells. J Cell Biochem 2008;103:1962–1974.

120. Salvatore CA, Jacobson MA, Taylor HE, et al. Molecular cloning and characterization of the human A3 adenosine receptor. Proc Natl Acad Sci 1993;90:10365–10369.

121. Sajjadi FG, Firestein GS. cDNA cloning and sequence analysis of the human
A3 adenosine receptor. Biochim Biophys Acta BBA - Mol Cell Res 1993;1179:105–
107.

122. Wilkie TM, Chen Y, Gilbert DJ, et al. Identification, Chromosomal Location, and Genome Organization of Mammalian G-Protein-Coupled Receptors. Genomics 1993;18:175–184.

123. Monitto CL, Levitt RC, DiSilvestre D, et al. Localization of the A3 adenosine receptor gene (ADORA3) to human chromosome 1p. Genomics 1995;26:637–638.

124. Murrison E, Goodson SJ, Harris CA, et al. The Human A3 Adenosine Receptor Gene. Biochem Soc Trans 1995;23:270S-270S.

125. Iyer SV, Ranjan A, Elias HK, et al. Genome-wide RNAi screening identifies TMIGD3 isoform1 as a suppressor of NF-κB and osteosarcoma progression. Nat Commun;7. Epub ahead of print December 2016. DOI: 10.1038/ncomms13561.

126. Murrison EM, Goodson SJ, Edbrooke MR, et al. Cloning and characterisation of the human adenosine A3 receptor gene. FEBS Lett 1996;384:243–246.

127. Atkinson MR, Townsend-Nicholson A, Nicholl JK, et al. Cloning, characterisation and chromosomal assignment of the human adenosine A3 receptor (ADORA3) gene. Neurosci Res 1997;29:73–79.

128. Kim S-H, Nam E-J, Kim Y-K, et al. Functional variability of the adenosine A3 receptor (ADORA3) gene polymorphism in aspirin-induced urticaria: ADORA3 gene polymorphism in aspirin-induced urticaria. Br J Dermatol 2010;163:977–985.

129. Goni JR. Triplex-forming oligonucleotide target sequences in the human genome. Nucleic Acids Res 2004;32:354–360.

130. Hohoff C, Garibotto V, Elmenhorst D, et al. Association of Adenosine Receptor Gene Polymorphisms and In Vivo Adenosine A1 Receptor Binding in The Human Brain. Neuropsychopharmacology 2014;39:2989–2999.

131. Borroto-Escuela DO, Hinz S, Navarro G, et al. Understanding the Role of Adenosine A2AR Heteroreceptor Complexes in Neurodegeneration and Neuroinflammation. Front Neurosci;12 . Epub ahead of print February 6, 2018. DOI: 10.3389/fnins.2018.00043.

132. Otowa T, Hek K, Lee M, et al. Meta-analysis of genome-wide association studies of anxiety disorders. Mol Psychiatry 2016;21:1391–1399.

133. Wei CJ, Augusto E, Gomes CA, et al. Regulation of Fear Responses by Striatal and Extrastriatal Adenosine A2A Receptors in Forebrain. Biol Psychiatry 2014;75:855–863.

134. Bodmer W, Bonilla C. Common and rare variants in multifactorial susceptibility to common diseases. Nat Genet 2008;40:695–701.

135. Wang L, Shen X, Wu Y, et al. Coffee and caffeine consumption and depression: A meta-analysis of observational studies. Aust N Z J Psychiatry 2016;50:228–242.

136. Gass N, Ollila HM, Utge S, et al. Contribution of adenosine related genes to the risk of depression with disturbed sleep. J Affect Disord 2010;126:134–139.

Table 1. Adenosine receptor genes and proteins

Official gene Name*	Old Name	Reference mRNA transcript [#]	Protein name	Protein length (aa)
ADORA1	RDC7	NM_000674.2	A ₁ R	326
ADORA2A	RDC8	NM_001278497.1	A _{2A} R	412
ADORA2B	-	NM_000676.2	$A_{2B}R$	332
ADORA3	-	NM_000677.3	A ₃ R	318

* HUGO nomenclature; [#] the reference mRNA transcripts are the longest transcripts listed in the NCBI RefSeq database (GRCh37/Hg19) and encode the longest protein isoform.

Accession discourse or traits	Studied population			Risk	Risk	Def
Associated brain diseases of traits	Patients	Ethnicity	Country	allele	genotype	Ref
	rs5751876					
Panic disorder	99 patients and 99 controls	Caucasian	Germany	Т	TT	76
Panic disorder	70 multiplex families and 83 triad families	Caucasian*	USA	С		77
Sympathetic measures during venipuncture	17 patients with blood-injury phobia	Caucasian	Germany		тт	78
Panic disorder	457 patients and 457 controls	Caucasian	Germany	Т		19
Panic disorder and agoraphobia	341 patients and 457 controls	Caucasian	Germany	С		19
Anxious personality trait	424 healthy participants	Caucasian	UK	С		19
Self-reported anxiety after caffeine administration	94 healthy participants	Caucasian*	USA		тт	81
Self-reported anxiety after d-amphetamine administration	99 healthy participants	n.a.	USA		TT	83
Self-reported anxiety after caffeine administration	416 healthy participants	Caucasian	UK		тт	37
Startle reflex after caffeine administration	110 healthy participants	Caucasian	Germany		тт	84
Prepulse inhibition/facilitation after caffeine administration	114 healthy participants	Caucasian	Germany		тт	85
Decreased attention after caffeine administration	106 healthy participants	Caucasian	Italy		СС	86
Decreased motor executive control after caffeine administration	106 healthy participants	Caucasian	Italy		тт	86
Panic disorder patients with BS	92 patients BS+ and 259 BS-	Caucasian	USA	Т		90
Tourette syndrome	162 patients and 270 controls	Caucasian	Poland	Т		34
Tourette syndrome with concomitant ADHD	63 patients ADHD+ and 99 patients ADHD+	Caucasian	Poland	С		34
Tourette syndrome with concomitant anxiety	34 patients anxiety+ and128 patients anxiety-	Caucasian	Poland	С		34
Increased autism-specific impairment	98 patients	Caucasian	Germany	Т		93
Migraine with aura <i>vs</i> without aura	122 patients aura+ and 143 aura-	Caucasian	Germany	С		99
Earlier age of onset HD	791 HD patients	Caucasian	France		TT	100

Table 2. Significant associations in brain diseases or traits of rs5751876, rs2298383 and rs35320474 in ADORA2A

Earlier age of onset HD	419 HD patients	Caucasian	Germany		TT	101
Earlier age of onset HD	18 HD patients	n.a.	Uruguay	т		102
Variation in pre-attentive sensory memory sub-processes	199 healthy participants	Caucasian	Germany		TT	106
AESD	85 patients and 184 controls	Asian	Japan	т		14
Attention and processing of information	65 healthy participants	Caucasian	Germany		тт	107
Lower behavioral resiliency after sleep restriction	20 healthy participants	n.a.	USA		тт	108
Insomnia-like EEG pattern after caffeine administration	19 healthy participants	Caucasian	Switzerland		CC	109
Sustained vigilant attention after prolonged wakefulness	45 healthy participants	Caucasian	Switzerland	Т		110
Lower effect of caffeine on attention after prolonged wakefulness	45 healthy participants	Caucasian	Switzerland	Т		110
Sensitive to caffeine's effects on sleep	2402 healthy participants (1470 families)	n.a.	Australia	С		111
Sleep disturbance caused by caffeine consumption	926 healthy participants	n.a.	Brazil	Т		112
A _{1A} R availability in brain regions of fear network	28 healthy participants	Caucasian	Germany	Т		130
	rs2298383					
Anxious personality trait	424 healthy participants	Caucasian	UK	Т		79
Self-reported anxiety after caffeine administration	102 healthy participants	Caucasian*	USA		CC	82
Anxiety disorder in ADHD patients	478 ADHD patients	Caucasian*	Brazil		TT	92
Autism spectrum disorder	98 patients and 234 controls	Caucasian	Germany		CC	93
Earlier age of onset HD	419 HD patients	Caucasian	Germany		TT	101
Variation in pre-attentive sensory memory sub-processes	199 healthy participants	Caucasian	Germany		CC	106
AESD	85 patients and 184 controls	Asian	Japan	С		14
Sustained vigilant attention after prolonged wakefulness	45 healthy participants	Caucasian	Switzerland	С		110
Lower effect of caffeine on attention after prolonged wakefulness	45 healthy participants	Caucasian	Switzerland	С		110
	rs35320474					
Self-reported anxiety after caffeine administration	94 healthy participants	Caucasian*	USA		Tins/Tins	81

Self-reported anxiety after d-amphetamine administration	99 healthy participants	n.a.	USA		Tins/Tins	83
ADHD traits	1747 twins	Caucasian	Sweden	Tins		91
Increased autism-specific impairment	98 patients	Caucasian	Germany	del		93
Migraine with aura vs without aura	122 patients aura+ and 143 aura-	Caucasian	Germany	Tins		99
Earlier age of onset HD	419 HD patients	Caucasian	Germany		del/del	101
AESD	85 patients and 184 controls	Asian	Japan	del		14
Sustained vigilant attention after prolonged wakefulness	45 healthy participants	Caucasian	Switzerland	del		110
Lower effect of caffeine on attention after prolonged wakefulness	45 healthy participants	Caucasian	Switzerland	del		110

ADHD: attention deficit/hyperactivity disorder patients; AESD: Acute encephalopathy with biphasic seizures and late reduced diffusion; BS: bladder syndrome; HD: Huntington's disease; Ref: references; del: deletion; ins: insertion; n.a.: not available; *: mostly Caucasian.

Figure legend

Figure 1. Adenosine receptor genes in the human.

Schematic representation of the (**A**) *ADORA1*, (**B**) *ADORA2A*, (**C**) *ADORA2B*, and (**D**) *ADORA3* genes. Exons and introns are shown as boxes and horizontal lines, respectively. Untranslated regions are indicated as grey boxes, and coding exons are marked in black. The different TSSs are indicated by black perpendicular arrows. Transcripts with dotted lines correspond to mRNAs not fully sequenced or with low levels of evidence. Dotted boxes represent the CpG islands. Hatched boxes mark the relative location of the promoter region, according to the histone code and chromatin state segmentation data from the ENCODE project.



Figure 1. Adenosine receptor genes in the human.

Supplementary material

Methods

We searched the literature published in English from MEDLINE PubMed, Science Direct, Web of Science, Google Scholar, OMIM and the GWAS catalog (https://www.ebi.ac.uk/gwas/home) until 1 April 2019. We used the search terms ("ADORA" or "Adenosine Receptor" or "ADORA1" or "RDC7" or "ADORA2A" or "A2aR" or "RDC8" or "ADORA2B" or "ADORA2" or "ADORA3" or "A3AR") AND ("Variant*" or "SNP" or "Polymorphism*" or "Genotype" or "Haplotype" or "Allel*", or "Neurology" or "Psychiatry" or "Pharmacogenetics" or "Promoter" or "Exon" or "Intron" or "Splicing" or "Translation" or "Transcript*" or "Epigen*"). Bibliographies of the publications that met these criteria and of each relevant review were also manually screened to ensure no potential articles were missed. Publications included in this review involved at least one of the following criteria: (a) Adenosine receptor gene variation implied in a human genetic trait or pathology involving the central nervous system; (b) Adenosine receptor gene cloning, structure and transcription regulation. Exclusion criteria were defined as follows: papers addressing nonneurological diseases or involving the peripheral nervous system, papers addressing only non-human genes. We excluded animal studies, conference proceedings, and editorials.