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**Three-hybrid screens:
inducible third-party systems**

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25 - Three-hybrid screens: *inducible third-party systems*

1. INTRODUCTION

Recent studies have brought up the existence of several molecular complexes either stable, like the RNA polymerase II holoenzyme, transcription factor TFIID and mediators, or transient as could be observed in the various steps of the transcription process e.g. initiation, elongation and termination. These observations point out the fact that regulation of different cellular mechanisms is orchestrated by interactions between molecular species either to modify proteins or to position one of them within a complex that then will be functional. Several techniques such as affinity precipitation, glycerol gradient sedimentation and the yeast two-hybrid system are currently used to study protein-protein interactions (1). These various methods, usually investigating the connection between two partners, keep in account only the strong (and stable) interactions and neglect the weaker ones. Considering the complexes studied so far, it is suspected and sometimes shown that interactions often occur between more than two proteins, e.g. to stabilize the complex. In an effort to understand the various biological mechanisms -and in our case gene expression regulation-, we were interested in developing the yeast three-(or tri-)hybrid system.

The three-hybrid system, as illustrated in figure 1, is based on the reconstitution of a transcriptional activator complex either to search for or to study a protein that interacts with two others and to acquire information about ternary complex assembly (2). This technique detects direct or mediated interactions between two fusion proteins that contain either a DNA binding domain (DBD; the DBD-X protein) or an activation domain (AD; the AD-Y protein). In some cases, when these two hybrid proteins interact weakly or not at all, a third partner (the protein Z) is necessary to promote (to induce) the formation of the transcriptional activator allowing the transcription of the reporter genes. Thus, specific and stable protein-protein interactions between X, Y and Z lead to the activation of the reporter genes that are integrated in the yeast genome. The HIS3 reporter gene contains a specific DNA sequence which can be recognized by the DBD of the transcriptional activator. Activation of the HIS3 gene permits the endogenous synthesis of histidine allowing the yeast to grow on histidine-lacking media. Activation of the LacZ gene, another reporter gene containing the same DNA binding element, will lead to the synthesis of the β -galactosidase (β -Gal) that catalyses the transformation of either X-Gal (5-bromo-4-chloro-3-indolyl β -D-galactopyranoside) or ONPG (o-nitrophenyl- β -D-galactopyranoside) into a detectable blue or yellow product, respectively.

There are several options to reconstitute the transcriptional activator: (i) the third partner Z can act as a bridging factor (Fig.1A) by interacting with both the DBD-X and AD-Y, thus allowing the DNA-binding protein X to target the basal transcription machinery; (ii) In case of weak interactions, Z may function as a stabilising factor (Fig.1B) that strengthens the interaction between X and Y; (iii) In some other cases, reconstitution of the transcriptional activator requires some post-transcriptional modifications of one of the two partners, in order to allow interaction between X and Y. The third partner Z will then be a regulating factor (Fig.1C), an enzyme that will not necessarily be part of the reconstituted transcriptional activator.

When DBD-X and AD-Y are sufficient to reconstitute a stable transcriptional activator, the three-hybrid system can also be used for the search of inhibitors. In this case, the Z partner can act as an inhibitor (Fig.1D) by interacting with, or enzymatically modifying, one of the two main partners, thus preventing their interactions. This will result in the inhibition of the expression of the reporter genes followed by growth restriction on histidine-lacking media as well as repression of the β -Gal activity.

In order to control the specificity of the ternary complex formation that depends on the expression of one of its components, we have set up an inducible three-hybrid system, in which the expression of the third partner Z can be easily switched "on" or "off". Therefore we have used the already characterized yeast Met25 promoter (3). Excess of methionine in the culture medium leads to the transcriptional repression of a cDNA cloned downstream of this promoter. Thus, we have designed the two plasmids, pBridge and pLexA9-3H (Fig.2, left panel), containing first the DNA of either GAL4 or second LexA DBD, which can be fused to the cDNA of the X protein, and second the methionine regulated expression cassette, containing the Met25 promoter, which can be fused to the cDNA of the third partner Z.

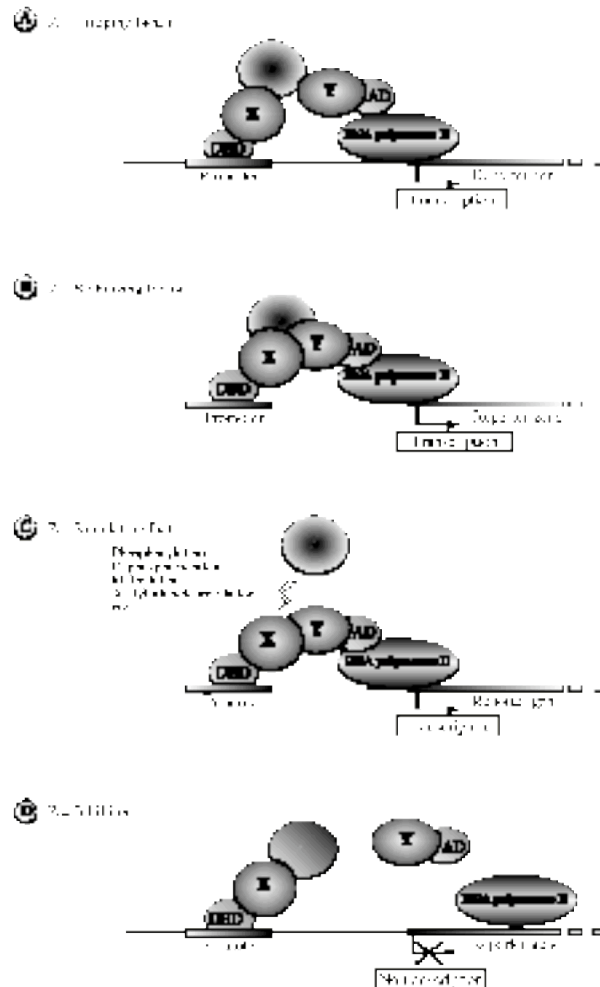


Figure 1: **The three-hybrid system**

The three-hybrid system can be used to characterise or detect new factors such as (A) a bridging factor Z, which interacts with both proteins, X and Y, that do not interact each other, (B) a stabilising factor Z that strengthens a weak interaction between the two proteins X and Y, (C) a regulating factor Z that stimulates the interaction between the two fused proteins X and Y by modifying one of them, (D) an inhibitor Z that prevents the interaction between two interacting proteins X and Y. In the three first cases, upon expression of the third partner (Z) the transcriptional activator formed stimulates the transcription of the reporter genes whereas in the last case, the third polypeptide inhibits the transcriptional activator formation thus preventing the transcription of the reporter genes.

1.1 The inducible third partner Z as Activator

In order to reconstitute the transcriptional activator, the third partner Z (as shown in Figure.1: A,B,C) can be overexpressed in the absence of methionine. We have illustrated this feature with the formation of the cdk-activating kinase (CAK) that

contains three subunits: a cyclin dependent-kinase, cdk7, the cyclin H and MAT1, a stimulatory factor (4). Although the role of CAK, a subcomplex of the multiprotein complex TFIIH, is not well known in the various events that regulate the cell cycle cascade, it seems to play a role in protein coding gene transcription through the phosphorylation of the carboxy-terminal domain (CTD) of RNA polymerase II (5). It has been shown that cdk7 and cyclin H interact with each other but the role of MAT1 in the assembly of CAK was still unknown (6, 7). Using the three-hybrid system, we could demonstrate that the weak interaction between the two fused proteins DBD-cdk7 (X) and AD-MAT1 (Y) was stabilized upon expression of the third partner, cyclin H (Z), cloned in the Met25 expression cassette (8). This assembly does not occur in the presence of methionine that prevents the expression of cyclin H. This assay shows, that MAT1 contacts cdk7 or integrates the CAK complex in the presence of cyclin H.

The inducible three-hybrid system could be used to screen a cDNA library in order to search for some activating partners. Therefore two different screenings are possible: 1.) To identify proteins (Y) that may target an already formed binary complex (X-Z; see Fig.1A), the screening is made with a cDNA library inserted downstream of the activation domain (AD) sequence like most of the commercially available cDNA libraries. In this case, in order to isolate positive clones, yeast growth will be performed on a medium lacking methionine; the colonies will then be replica-plated on a medium containing both methionine and X-Gal on which they will no longer grow or stay white. 2.) To identify proteins (Z) that stabilize or regulate a weak interaction (X-Y; see Fig.1B,C), the cDNA-library must be cloned downstream of the Met25 promoter. The screening is then made on a histidine-lacking medium in the absence of methionine. Positive clones will be identified through a replica-plating on methionine-containing medium as mentioned above.

1.2. The inducible partner Z as Inhibitor

As an example to investigate Z components that target either X or Y proteins and consequently prevent the reconstitution of the transcriptional activator, we studied two oncogenic proteins, Raf and Ras (9), involved in signal transduction. The fused DBD-Ras (X) and AD-Raf (Y) proteins specifically interact each other in a two-hybrid system (9). An inhibition of the transcription of the reporter gene can be observed (8) in the absence of interaction between DBD-Ras (X) and AD-Raf (Y). This may occur when the cDNA of a nonfused cRaf is cloned downstream of the Met25 promoter and further expressed as the third partner (Z) in the absence of methionine. In this case, cRaf (Z) competes with AD-Raf (Y) for targetting DBD-Ras (X). Because the Met25 promoter is stronger than the hybrid gene promoter (3), its activation (in absence of methionine) leads to an excess of cRaf titrating out the DBD-Ras. Thus the transcriptional activator will not be reconstituted resulting in the repression of the reporter genes.

2. MATERIALS

2.1. Growth and maintenance of yeast

1. YPD Medium: 20 g/l Bacto™ Peptone (Difco); 10 g/l yeast extract (Difco); 20 g/l D(+)-Glucose (Riedel-deHaën, Germany); water to 1 l; autoclave at 121°C for 15 min (or add water to 950 ml; autoclave for 30 min, then add 50 ml sterile filtrated 40% D(+)-Glucose). For culture plates add 20 g/l Agar (Difco).

2. Yeast-Minimal Medium (MM): 6.7 g/l yeast nitrogen base without amino acids (Difco); 20 g/l D(+)-Glucose; water to 1 l; autoclave at 121°C for 15 min (or add water to 950 ml; autoclave for 30 min, then add 50 ml 40% D(+)-Glucose). For culture plates add 20 g/l Agar. Depending on the different strains the following amounts of specific

nutrients should be added (either as a tenfold stock for small volumes or directly as powder for higher volumes) except the one(s) for which the medium should select: 0.2 g/l L-Tryptophane; 0.2 g/l L-Histidine; 0.2 g/l L-Adenine; 1.0 g/l L-Leucine 0.3 g/l L-Lysine; 0.2 g/l Uracile (Sigma).

3. 90 mm (Greiner) and 245*245 mm (PolyLabo) culture plates.

2.2. Transformation

1. Stock solutions: 50% PolyEthylenGlycol 4000 (MERCK; sterile filtrated); 100% Dimethyl sulfoxide (DMSO; MERCK); 10x LiAc: 1 M Lithium acetate (Sigma), pH 7.5 (autoclave); 10x TE buffer: 0.1 M Tris-HCl, 10 mM EDTA, pH 7.5 (autoclave); 10 mg/ml Herring testis carrier DNA (Sigma; boil just prior use for 5 min, then cool on ice).

2. TE/LiAc: 1x TE, 1x LiAc.

3. PEG/LiAc: 40% PEG, 1x TE, 1x LiAc.

4. Appropriate minimal medium containing plate.

2.3. Methionine selection

1. Yeast minimal medium as described in **subheading 2.1**.

2. Methionine stock: We prepare a 0.2 μ M filtered 1 M L-methionine (Sigma) in deionized H₂O. This solution can be frozen to -20°C and kept for years.

Minimal medium is supplemented with 1 mM L-methionine.

3. 3-Amino-1,2,4-triazole (3AT; Sigma).

2.4 Protein extraction and immunopurification

2.4.1. Protein extraction

1. Extraction buffer: 50 mM Tris-HCl pH 7.8, 10% Glycerol, 0.1 mM EDTA, 100-300 mM KCl, 1 mM PhenylMethylSulfonylFluoride (PMSF; Sigma) and 1x protease inhibitor cocktail (Roche Diagnostics; contains Leupeptine, Pepstatin, Aproptinin, Antipain and Chimostatin).

2. Acid treated glass beads (Sigma).

3. A 0.45 μ m microspin-column (Ultrafree-MC; Millipore) is used to remove the biggest particles.

2.4.2. Immunopurification

1. Antibodies are crosslinked to protein A-Sepharose CL-4B (AP Biotech).

2. Stock solutions: 5 M NaCl; 1M Sodium Borate pH 9; 0.2 M Ethanolamine pH 8; 10 mg/ml BSA.

3. Sodium Azide (Sigma), Dimethyl Pimelidate (DMP; ICN Pharmaceuticals).

4. Buffers: TG₁₀EK_X: 20 mM Tris-HCl pH 7.8, 10% Glycerol, 0.1 mM EDTA, X mM KCl.

Crosslink wash buffer: 3 M NaCl, 50 mM Na Borate pH 9.

Crosslinking buffer: 3 M NaCl, 200 mM Na Borate pH 9, 20 mM DMP.

Elution buffer: TG₁₀EK₅₀; 0.01% NP40; 0.2 mg/ml insuline; 2 mg/ml peptide corresponding to the epitope.

2.5. β -Gal assays

Stock solution: Z-buffer: 21.5 g/l Na₂HPO₄·12 H₂O; 5.5 g/l NaH₂PO₄·H₂O; 0.75 g/l KCl; 0.246 g/l MgSO₄·7 H₂O; adjust to pH 7.0 and autoclave.

2.5.1. Colony lift filter assays

For LexA-transformants, use plates which contain MM with 2% D(+)-Galactose and 1% Raffinose instead of D(+)-Glucose.

1. Hybond™ Nylon membranes (Amersham); 3MM Whatman paper; liquid Nitrogen.

2. X-Gal (5-Bromo-4-chloro-3-indolyl- β -D-galactopyranoside; Merck) solution: 20 mg X-Gal/ml of N,N-Dimethylformamide (DMF; Sigma); store in the dark at -20°C .
3. Z-M-X-buffer: 100 ml Z-Buffer; 0.27 ml β -Mercaptoethanol; 1.67 ml X-Gal solution.

2.5.2. Liquid culture assay

1. Z-M-buffer: 100 ml Z-buffer; 0.27 ml β -Mercaptoethanol.
2. Z-O solution: dissolve 4 mg ONPG (o-nitrophenyl- β -D-galactopyranoside; Sigma)/ml Z-buffer (prepare ca. 1 hour before each use).
3. 1 M Na_2CO_3

2.5.3. On-plate assay

1. 10x S-buffer (for 500 ml): 68 g KH_2PO_4 , 9.9 g $(\text{NH}_4)_2\text{SO}_4$, 21 g KOH; pH \approx 8.0.
X-Gal: as described under **subheading 2.5.1**.
2. Minimal medium as described under **subheading 2.1**.

3. METHODS

3.1. Growth and maintenance of yeast

Working with yeast should be done under sterile conditions, because the different culture media do not contain antibiotics. Thus, contaminations should be avoided using sterile glass and plastic ware. It is recommended to work at a cleanbench or beside a burning flame.

3.1.1. Yeast strains

The strains used in the three-hybrid system are the same that the ones used for the two-hybrid. There are different yeast strains either more or less sensitive for the detection of protein-protein interactions, depending on the vectors, the transcriptional activator (Gal4 or LexA-VP16) and the protein used as the bait (see **note 4.1**). The most common yeast strains are the L40 strain (MATa, trp1, his3, leu2, ade2, $\text{LYS2}::(\text{LexAop})_4\text{-HIS3}$, $\text{URA3}::(\text{LexAop})_8\text{-LacZ}$) for the LexA-VP16 system and the HF7c strain (ura3, his3, lys2, ade2, trp1, leu2, $\text{LYS2}::\text{Gal1}_{\text{UAS}}\text{-Gal1}_{\text{TATA}}\text{-HIS3}$, $\text{URA3}::\text{GAL4}_{17\text{mersx3}}$, $\text{CyC1}_{\text{TATA}}\text{-lacZ}$) or the Y190 (ura3, his3, lys2, ade2, trp1, leu2, cyhr2, $\text{LYS2}::\text{Gal1}_{\text{UAS}}\text{-HIS3}_{\text{TATA}}\text{-HIS3}$, $\text{URA3}::\text{GAL1}_{\text{UAS}}\text{-GAL1}_{\text{TATA}}\text{-lacZ}$) when using the GAL4 system. These strains are commercially available.

3.1.2. Yeast amplification

Freshly streaked yeast grow at 28°C for 4-5 days until colonies are 1-2 mm in diameter. One such colony can then be used for amplification in liquid culture. For the maintenance of yeast strains, a colony resuspended in 0.5 ml of sterile water should be restreaked every 6-8 weeks. For long-time storage yeast strains can be stored in the appropriate medium containing 15% glycerol at -70°C .

3.2. Plasmid constructs

For the activation experiments we used the LexA-VP16 system. Thus, the bait (cdk7) is linked to the DBD of LexA in pLex9-3H (Fig.2; left panel) carrying the TRP1 selection gene that allows growth on tryptophane lacking MM. The second partner (MAT1), which has no significant interaction properties with the bait, is fused to the AD of VP16 in pVP16 (Fig.2; right panel), containing the LEU2 selection gene that allows growth on leucine lacking MM. In addition, the cDNA of the third partner (cyclin H) is cloned in the Met25 expression cassette (see **note 4.2**), which is on the same plasmid as the bait (pLex9-3H).

For the inhibition experiments we used the GAL4 system, in which the bait (H-Ras(V12)) is fused to the DBD of GAL4 in pBridge and the interacting partner (cRaf1)

is added behind the AD of GAL4 in pGAD-GH. The cDNA of the third partner (cRAF1) is cloned into the Met25 expression cassette, which is on the same plasmid as the bait (pBridge).

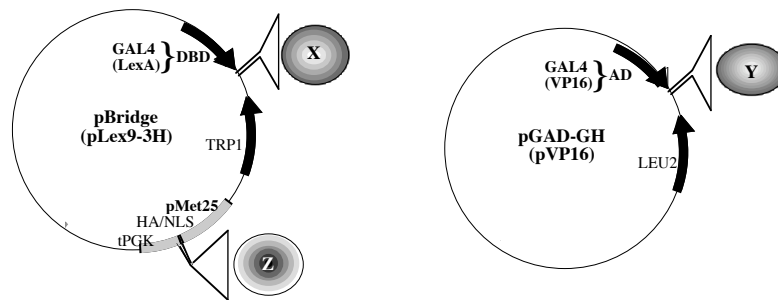


Figure 2: **The three-hybrid vectors:**

Left panel: pBridge, derived from the pGBT9 vector, contains the TRP1 selective gene, the DNA-binding domain of GAL4 and the Met25 expression cassette (see **note 4.2.**) with the NotI and BglII cloning sites. pLex9-3H, derived from the pBTM116 vector, contains the TRP1 selective gene, the DNA-binding domain of LexA and the Met25 expression cassette with the NotI and SrfI cloning sites. The Met25 cassette contains a hemagglutinine (HA) tag sequence, a nuclear localisation signal (NLS) sequence and the terminator of the phosphoglycerate kinase (tPGK). *Right panel:* The plasmids pGAD-GH and pVP16 contain the LEU2 selective gene and the activation domain of GAL4 and VP16, respectively.

3.3. Transformation

To introduce plasmid DNA into yeast, the LiAc-mediated transformation is very easy, fast and reproducible. The method described here allows single transformation or simultaneous cotransformation of two different plasmids carrying different selection genes. Our protocol can be used for up to 10 parallel transformations but can easily be scaled-up. Furthermore, with this method a cDNA library transformation can be performed; the culture or buffer volumes are indicated in brackets. It is important to mention that a positive and a negative control should be incorporated to demonstrate the efficiency of the transformation. This efficiency should be around 10^5 colonies/ μg plasmid for a single transformation.

1. Inoculate 10 ml of YPD or the appropriate MM with a single colony of about 2 mm in diameter and incubate overnight at 28°C. (For the library screening, further inoculate 150 ml of the same medium with the 10 ml of the preculture and let grow overnight.) Dilute the suspension to an OD₆₀₀ of 0.2 to 0.3 and incubate the culture for approximately 3 hours to get an OD₆₀₀ around 0.5±0.1 (the log phase).
2. Centrifuge 50 ml (1000 ml) of the culture for 5 min at 1000 g. Discard the supernatant and wash the cells in 30 ml (500 ml) of sterile water. After a second centrifugation step, resuspend the cells in 1 ml (8 ml) sterile TE/LiAc.
3. Mix 0.1 mg (20 mg) herring sperm DNA with 0.1 μg of each plasmid -for simultaneous cotransformation use 0.2 μg of each different plasmid- (0,5 mg cDNA library) in a sterile 1.5 ml (500 ml) tube and add 100 μl (8 ml) of the yeast solution. Mix vigorously and add 600 μl (60 ml) PEG/LiAc. Vortex and incubate for 30 min at 30°C with shaking (200 rpm).
4. Add 70 μl (7 ml) DMSO, mix carefully by inverting the tubes and incubate for 15 min at 42°C. Centrifuge for 20 sec (5 min) at 13000 rpm and remove the supernatant. Dissolve the yeast-pellet in 500 μl (10 ml) TE and plate 100 μl (200 μl) of this suspension on a 90 mm (245*245 mm) plate which allows selection for the desired transformant(s).
5. Incubate the plate(s) at 30°C until the colonies are 1-2 mm in diameter (4-5 days).

3.4 Methionine selection

The advantage of the three-hybrid system using the Met25 inducible promoter resides in the "on" or "off" switch for the expression of the third protein (see **note 4.3.**). A simple replica-plating experiment allows to quickly check the interaction specificity between the various partners:

1. Select at least three different clones from the leucine and tryptophane lacking plates selective for both plasmids. Scrape the whole colony and resuspend in 100 μ l of sterile H₂O.
2. Make one to ten dilutions in sterile H₂O up to 10⁻⁴.
3. Drop 10 μ l of each dilutions either on selective MM plates containing or lacking 1 mM methionine.
4. Let the yeast grow 4 days at 28°C. The duration of the growth may be dependent on the proteins expressed and can take up to 10 days.

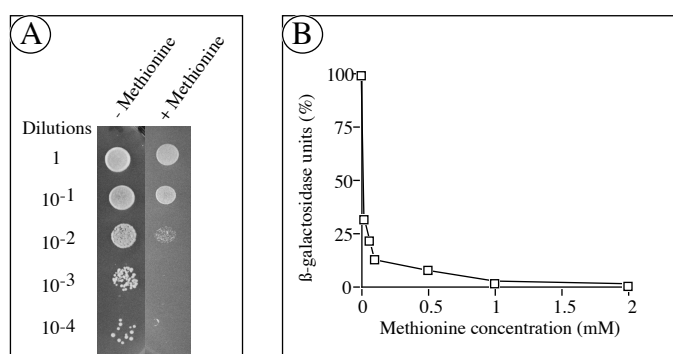


Figure 3: **A.** L40 yeast cells cotransformed with pLex9-3H, containing LexA-cdk7 and cyclin H under the dependence of the Met25 promoter, and with MAT1pVP16 were spotted at various dilutions, as indicated at the left of the figure, on a selective minimal medium lacking histidine. Yeast growth was performed during 4 days at 30°C, in absence (-) or presence (+) of 1 mM methionine, controlling the expression of the third partner, cyclin H. In presence of the third partner we can observe a yeast growth for the highest dilutions whereas in presence of methionine, and so in absence of the cyclin H, we can observe yeast growth only for the first dilutions. **B.** β -Galactosidase assay from liquid culture of pLex9-3H/cdk7/Met-cyclinH-pVP16/MAT1 cotransformed yeast was done as described in **subheading 3.6.2.** The β -Gal activity was measured as a function of the methionine concentration and so as a function of the presence of the third partner. The interaction, reflected by the β -Gal activity, is no more detectable in presence of 1 mM methionine. 100% corresponds to the β -Gal activity of the yeast grown on a methionine lacking medium and 0% to the activity of yeast grown on a 10 mM methionine containing medium.

An example of the methionine dependent reconstitution of the transcriptional activator is shown in figure 3A. The three proteins of the TFIIH CAK complex (constructions described in **subheading 3.2.**) were expressed in yeast. Cyclin H is conditionally expressed from the Met25 promoter. The complex is then formed only if the three proteins are present. Thus, the yeast are able to grow in the absence of methionine because the cyclin H is expressed and the ternary complex is reconstituted leading to the transcriptional activation of the reporter genes. The colonies that are plated on selective MM lacking 1 mM methionine grow faster than colonies plated on MM containing methionine. Thus, the methionine repression is observed for the 10⁻² to 10⁻⁴ dilutions. Because of a weak interaction between DBD-cdk7 and AD-MAT1, a background growth can be observed for the lower dilutions. Furthermore, non-specific interactions can take place between one of the hybrid proteins and the fused DBD directly. In order to remove these background, 3-Amino-1,2,4-triazole (3AT; between 5 and 50 mM) can therefore be added to the medium. Moreover, the Met25 promoter is known to be leaky, meaning that the expression of

the third partner, cyclin H, is not completely repressed, resulting in some detection of yeast growth on a methionine containing medium.

As shown above, the Met25 promoter is down regulated upon addition of methionine. To determine the optimum conditions of the methionine repression, several concentrations have been tested out using the liquid β -galactosidase assay (see **subheading 3.6.2**) with the system containing the three proteins of the CAK complex, as described above. Thus, the resulting β -Gal activity, corresponding to the concentration of the third partner, was measured as a function of the methionine concentration. As shown in figure 3B, the β -Gal activity, reflecting the cyclin H expression, is about 30% of the activity without methionine for 0.01 mM methionine, 10% for a concentration of 0.5 mM and completely repressed for 1 mM. Actually, this system can also be useful for the control of the expression of toxic proteins, in order to decrease the amount of these proteins in the cell.

3.5. Protein extraction and immunopurification

3.5.1. Protein extraction

To investigate whether or not the hybrid-proteins are well expressed, it is necessary to prepare yeast protein extracts and to perform western-blotting assays.

1. Inoculate 50 ml of selective medium for the plasmids (described in **subheading 2.1.**) to OD₆₀₀ of 0.05. Grow at 28°C with shaking until an OD₆₀₀ between 0.8 and 1. Depending on the protein that is expressed, the growth can take between over-night and 3 days.
2. Transfer the culture in a 50 ml tube and cool the cells by placing the tube on ice for few minutes. Centrifuge 5 min at 3000 rpm, 4°C.
3. Resuspend the cell pellet in 1 ml cold H₂O, transfer to a microfuge tube, centrifuge 30 sec. at maximum speed, remove the supernatant and add 300 μ l of extraction buffer. Different buffers can be used for the extraction but usually a high salt buffer (300 mM KCl) is recommended for a higher recovery. To analyse the *in vitro*-interaction between the hybrid proteins using immunoprecipitation or immunopurification assays, the cell pellet should be resuspended at this stage in a low salt extraction buffer (100 mM KCl), in order to retain potential weak interactions.
4. Add glass beads to 1-2 mm below the surface and keep on ice for 10 minutes.
5. Vortex vigorously in cold room 3 times 1 minute with intervals of 1 minute on ice.
6. Centrifuge at maximum speed (13000 rpm) for 15 minutes at 4°C.
7. Transfer the supernatant to a microspin-column (Ultrafree-MC; Millipore) in order to filter residual big particles (see **note 4.4.**). Centrifuge 30 sec at maximum speed.
8. Samples can be frozen at -80°C and kept for several months.
9. Estimate the total protein concentration by performing a Bradford assay.
10. Run 10 to 100 μ g of protein on a SDS-PAGE (8 to 15%) and perform a western blot.

3.5.2. Immunoprecipitation of HA tagged protein from yeast

The purification of a new protein, identified as an interacting partner during a cDNA library screening, is important for further analysis. It is therefore useful that the proteins expressed by the vectors containing the cDNA library are tagged.

We have introduced the hemagglutinine sequence corresponding to the HA-tag (SYPYDVPDYASLGGPSS, one amino acid code), upstream of the cloning site (Fig.2) in the methionine regulated expression cassette. Thus, the expressed protein can be immunoprecipitated or immunopurified directly from a yeast crude extract with a HA-tag specific antibody (commercially available). Instead of the HA-antibody, a specific antibody can as well be used in the following protocol.

1. Antibody crosslink to protein A-Sepharose via DMP
 - a. Wash the protein A-Sepharose 3 times with 10 volumes of crosslink wash buffer.
 - b. Add 0.7 volume of purified anti-HA monoclonal antibody with 4.3 volumes of crosslink wash buffer. Incubate at RT 1 h under gentle shaking.
 - c. Wash 2 times with 10 volumes of crosslink wash buffer
 - d. Add 10 volumes of crosslinking buffer. DMP should be added immediately before use. Incubate 30 min at RT under gentle shaking.
 - e. Wash once with 10 volumes of Ethanolamine 0.2 M and incubate with 10 volumes of Ethanolamine 0.2 M at RT for 2h30 or at 4°C over night.
 - f. Wash 2 times with 10 volumes of TG₁₀EK₃₀₀/ 0.05% NP40. The beads are then ready to use. If you want to keep the antibody crosslinked to protein A-Sepharose for several weeks, add 1 mg/ml BSA and 0.2% Na azide. Before use, wash then 3 times with 10 volumes of TG₁₀EK₃₀₀/ 0.05% NP40.

2. Immunoprecipitation of the HA tagged protein.

For a pull down assay, very little yeast extract is needed. Remember that if the strength of the interaction is not known, it is recommended to make the yeast extract at low salt concentration or to dialyse the extract against a low salt buffer.

- a. Incubate 300 μ l of the yeast extract with 50 μ l of the antibody sample crosslinked to the protein A-Sepharose for at least 4 hours at 4°C under gentle shaking.
- b. Wash the beads 3 times with 1 ml of TG₁₀EK₁₀₀₋₅₀₀ (usually 300 mM KCl is preferred, once more depending on the strength of the interaction).
- c. Add 50 μ l of Laemmli sample buffer to the beads removed of the supernatant and load (5-20 μ l) on a SDS-polyacrylamide gel.

3.5.3. Immunopurification of the HA tagged protein.

Using the three-hybrid system, it is possible to directly purify the protein under the control of the Met25 promoter as well as the tripartite complex. The immunoprecipitation procedure can be scaled up to 2 liters of yeast culture or even more, if necessary. We describe the method for a 20 ml culture:

1. Proceed as in **subheading 3.5.2.** until **step 2.b.** After the TG₁₀EK₁₀₀₋₅₀₀ washings, wash 2 times more with 1 ml of TG₁₀EK₅₀.
2. Remove the supernatant and add 50 μ l of elution buffer which contains the oligopeptide corresponding to the HA_x or the specific antibody epitope. Incubate at 4°C over-night with gentle shaking.
3. Centrifuge at low speed (2000 rpm) for few seconds and transfer the supernatant to a microfuge tube. Centrifuge and transfer once more to discard all the remaining beads. Most of the time the oligopeptide does not interfere with the activity of the purified protein; if necessary, dialyse against TG₁₀EK₅₀ for 4 hours at 4°C.
4. Verify by western blot the presence of the protein in the elution fraction using the HA or the specific antibody. Compare with a yeast extract transfected with a control expression vector.

3.6. β -Gal assays

Three different β -galactosidase assays are described. The colony lift filter assay with the substrate X-Gal (5-bromo-4-chloro-3-indolyl- β -D-galactopyranoside) is a qualitative method similar to the on-plate assay. Both techniques allow the analysis of a large number of transformants appearing on the plates. However they are less sensitive than the liquid culture assay with the substrate ONPG (o-nitrophenyl- β -D-galactopyranoside). The liquid culture assay allows the quantification of β -Gal activity. It can then be used to compare the relative strength of protein-protein interactions of different proteins or different mutations of the same protein.

The colony lift filter assay and the ONPG assay use the freeze/thaw cycle in liquid nitrogen for the lysis of the cell walls. This is a rapid step for the accurate analysis of β -galactosidase activity.

For the on-plate assay X-Gal is added directly to the selective MM leading to a slight growth restriction because of a pH shift.

3.6.1. Colony lift filter assay

It is recommended to use fresh colonies (1-2 mm in diameter) that are growing on the selective MM. When colonies grown in presence of methionine have to be analysed, it is necessary that these transformants have not been plated on a 3AT-containing medium (**see subheading 3.4.**).

1. For every plate to be assayed presoak a sterile 3MM Whatman paper in 2.5 ml (5 ml) Z-M-X-buffer in a 90 mm (150 mm) plate. Lay, under sterile conditions, a dry nylon membrane filter on the yeast plate for few seconds to transfer the colonies onto the filter. Poke holes through the filter into the agar using hot needles in three asymmetric locations in order to orientate the filter compared to the plate for later assignments of positive clones.
2. Carefully remove the filter from the surface and lay it colony side-up on an aluminium boat, which floats on liquid nitrogen. After 10 sec, submerge the boat with the filter for 1 min and thaw it at room temperature. Place the filter colony side-up on the presoaked Whatman filter avoiding air bubbles under the filter.
3. Incubate at 30°C until blue colonies appear (from 30 min to 12 hours).

3.6.2. Liquid culture assay

In order to compare the results of different experiments in a statistical manner, we suggest the analysis of at least three separate colonies of each transformant.

1. Inoculate one colony in 10 ml of the appropriate MM with or without 1 mM methionine and incubate at 30°C under gentle shaking until the stationary phase (**see note 4.5.**). For further comparative studies it is important to have all cultures in the same growth phase.
2. Add approximately 16 ml of MM (+/- methionine) to 2 ml of each culture to achieve an OD₆₀₀ of 0.2-0.3 and incubate for 3 hours at 30°C until the OD₆₀₀ is in the range of 0.5-0.8. Note the OD₆₀₀ and the volume of the culture.
3. After centrifugation (5 min, 1000 g), the cells are resuspended in 1 ml of Z-buffer. Transfer to a 1.5 ml microfuge tube and centrifuge at high speed for 20 sec. Discard the supernatant and dissolve the pellet in 250 μ l of Z-buffer. Divide the cell suspension into two 100 μ l aliquots and set up a standard reaction tube with 100 μ l Z-buffer.
4. Freeze the tubes 3 times for 1 min in liquid nitrogen and thaw them each time in a water bath.
5. Add to each tube 700 μ l Z-M-buffer and 160 μ l of freshly prepared Z-O-solution, incubate at 30°C until a yellow color develops. It is important to note the time of this reaction.
6. Stop the reaction by adding 400 μ l 1 M Na₂CO₃ solution. Centrifuge for 10 min at maximum speed to clear the supernatant and measure the OD₄₂₀ against the standard.

The linear range of this assay lays in the area from 0.02 to 1.0 for an OD₄₂₀. Calculate the β -Gal units using the following formula: β -Gal = $1000 \cdot OD_{420} / (OD_{600} \cdot V \cdot t)$, V=Volume of the culture and t=reaction time. With both V and t constant you can simplify the formula to: β -Gal = $1000 \cdot OD_{420} / OD_{600}$

Using this technique, we could demonstrate that the formation of the CAK-complex (Fig.4, lanes 1 and 2) is highly dependent on cyclin H. After

cotransformation of the L40 strain with both pLex9-3H, containing LexA-cdk7 and Met25-cyclinH, and pVP16-MAT1, the β -Gal activity of such clones grown in a methionine-containing medium is less than 20% of the activity of colonies grown in a methionine-lacking one, due to the repression of the cyclin H. The observed basal activity could be due to e.g. the leakiness of the Met25 promoter as discussed under **subheading 3.4**. In control experiments, yeast cotransformed with pLex9-3H, containing LexA-cdk7 and Met25-cyclin H, and native pVP16 (lane 3 and 4) or with pLex9-3H, containing only Met25-cyclin H, and pVP16-MAT1 (lanes 5 and 6) does not present any β -Galactosidase activity.

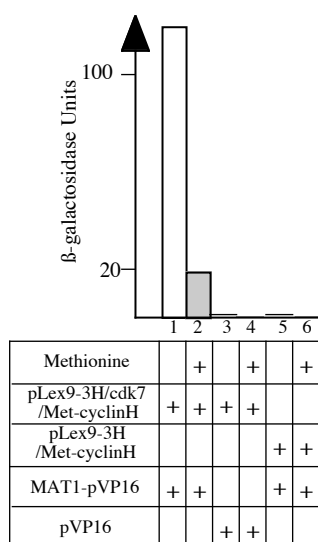


Figure 4: **Liquid culture assay**. L40 cells were cotransformed with the plasmids as indicated and analysed as described in **subheading 3.6.2**. with V=20 ml and t=1 min. In addition, the controls with vectors without fused protein were performed (lanes 3 to 6).

3.6.3. On-plate assay

β -Gal activity can be directly detected on the plates if the colonies are plated or replica-plated on a selective MM containing 1x S-buffer and 50 to 250 μ l X-Gal solution and incubated at 28°C. Blue color will appear with the colonies, after 4-8 days.

3.7. Inducible three-hybrid system:

Three verifications for positive clones during the library screening

The three-hybrid system described here possesses an inducible promoter for the conditional expression of the third partner. This allows to perform an additional control of an interaction specificity, especially in a screening experiment for an activator. Once the positive clones are identified first by the appearance of the histidine prototrophy and second by their β -galactosidase activity (see **note 4.6.**), a third control can be done by transferring these positive clones onto a methionine containing medium as described under **subheading 3.4**. (see **note 4.7.**). Only the clones whose growth will be repressed by methionine will be considered as true positives. For the screening of a cDNA library to look for an inhibitor of a stable binary complex see **note 4.8.**

4. NOTES

4.1. Different systems for different tasks

It is difficult to recommend a particular yeast strain for a random system. Because of the different properties concerning the activator system (GAL4 or LexA) and the promoter strength of the HIS3 or the other reporter genes, the appropriate strain has to be tested. The choice of the plasmids is also important because there are many differences in the number of copies per cell (high or low copy number). In addition, the promoter length of the alcohol-dehydrogenase I gene (ADH1), which controls the expression of the GAL4, LexA or VP16 fusion polypeptides, is varying between most of the commonly used plasmids. This leads to a different expression level of the fused proteins(10). There are also other vectors expressing the protein of interest fused to the N-terminus of LexA (11).

4.2. Generation of the Met25-expression vectors

The Met25-expression cassette, which contains the region from -474 to -1 of the Met25 promoter (3, 12),, the hemagglutinine epitope (HA), the nuclear localisation sequence and two rare restriction sites (NotI and SrfI for pLexA; NotI and BglII for pBridge) was cloned in the unique PvuII site of the plasmid vectors pBTM116 and pGBT9 to give the plasmids pLexA and pBridge, respectively. The transcription arrest signal is provided by addition of the phosphoglycerate kinase termination region corresponding to the BglII-HindIII terminator fragment from plasmid pEMBLYe30/2 (Fig.2, left panel)(8)..

It should be taken in account that the Met25 promoter is a strong promoter in the absence of methionine, compared to the ADH1 promoter. Therefore, the third polypeptide is much more expressed than the two other fused proteins.

4.3. Methionine and other inducible promoters are possible

There are several other three-hybrid systems using various inducible promoters :

- Galactokinase/galactose epimerase promoter (GAL1/GAL10), which is repressed by glucose and induced by galactose (13, 14). This promoter cannot be used in a GAL4 based two-hybrid system because the GAL4 activator binds the sequence between GAL1 and GAL10 and induces transcription.

- Acid Phosphatase promoter (PHO5), regulated by the presence or absence of inorganic phosphate (15).

- Metallothionein protein promoter (CUP1), induced by addition of copper to the medium (16, 17).

Some other promoters can be used for the regulated expression of the third partner in a three-hybrid system:

- Sorbitol dehydrogenase protein (SOR1), an enzyme responsible for sugar metabolism, is up regulated in presence of sorbitol. McGonigal et al. described some inducible protein expression vectors that can be used in a two or three-hybrid system (18).

- Aromatic aminotransferase II (ARO9) that catalyses the first step of tryptophan, phenylalanine and tyrosine catabolism in *S. cerevisiae* is upregulated by those aminoacids (19).

4.4. Advantage of Microspin-columns

Sometimes yeast extract immunoblot could be "smear-like" independently of the antibodies used. For optimum detection we found that filtering the yeast extract on a 0.45 μ m spin column can efficiently overcome this problem. Several techniques can be used such as affinity chromatography; however the advantages of the filter method reside mainly in the low cost and in time saving.

4.5. β -Galactosidase assay in function of the culture volume

To analyse the strength of an interaction or the amount of hybrid proteins in the liquid ONPG-assay, it can be useful to test different volumes of yeast culture. We have noticed that, using low culture volume, the β -galactosidase activity measured was not in the linear range (OD_{600} between 0.02 to 1.0). Thus, further analyses should be done under the conditions found to be the most suitable. It has to be noticed that in the liquid ONPG-assay, weak (or background) interactions yield no yellow colour. The reaction can then be stopped if no yellow color has appeared after 30 min.

4.6. β -Galactosidase assays in cDNA library screening

During the selection of positive clones by performing the on-plate and the colony lift filter assays in a yeast three-hybrid screening, only strong interactions are identified using the Y190 and L40 strains. Thus, the liquid culture assay is more sensitive for the identification of the weakest interactions in these strains.

4.7. One more hybrid - one more control

The two-hybrid system is used for detection of interaction between two proteins. Many interactions could not be detected because of the lack of specificity or because the right positioning of the interacting domains was not achieved. The three-hybrid system can overcome these problems by increasing the specificity of an interaction.

Thus, the presence of the third polypeptide is necessary to detect ternary complexes. One can imagine that a four-hybrid system can be used to detect quaternary complexes. Since many complexes involved in the different cellular processes contain more than three proteins, adding one more polypeptide to the bait means an increased specificity.

4.8. cDNA library screening for an inhibitor

To screen a cDNA library to search for a binary complex inhibitor, the methionine regulated system presented here is not appropriate. In presence of methionine, the repression of the inhibitor will lead to the formation of the transcriptional activator complex. Since each clone will express the reporter genes and will then be able to grow, replica-plating of all the corresponding colonies (approximately 2×10^6) will not be very practical. Therefore, a reverse three-hybrid system can be adapted from the system presented here (see **chapter 24**). But once a candidate is identified as an inhibitor, our system can be used to rapidly verify the inhibitory properties of the new protein.

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