

Molecular Actions of PPAR*α* **in Lipid Metabolism and Inflammation**

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Abstract

 Peroxisome proliferator-activated receptor alpha (PPARα) is a nuclear receptor of clinical interest as a drug target in various metabolic disorders. PPARα also exhibits marked anti-inflammatory capacities. The first generation PPARα agonists, the fibrates, have however been hampered by drug-drug interaction issues, statin drop-in and ill-designed cardiovascular intervention trials. Notwithstanding, understanding the 35 molecular mechanisms by which PPAR α works will enable control of its activities as a drug target for metabolic diseases with an underlying inflammatory component. Given its role in reshaping the immune system, the full potential of this nuclear receptor subtype as a versatile drug target with high plasticity becomes increasingly clear, and a novel generation of agonists may pave the way for novel fields of applications.

I. Introduction to PPAR family members

 The peroxisome proliferator-activated receptors (PPARs) are ligand-inducible transcription factors that belong to the nuclear receptor (NR) superfamily. Three PPAR subtypes have been identified: PPARα (NR1C1), PPARβ/δ (NR1C2) and 98 PPAR_γ (NR1C3) (1-5).

 The first member of the PPAR family, PPARα, was identified in 1990 and named by its ability to become activated by chemicals known to induce peroxisome proliferation in rodents (6). Peroxisomes are subcellular organelles found in most plant and animal cells and harbor a complex set of enzymes acting on various metabolic functions including β-oxidation of fatty acids (FA), bile acid and cholesterol metabolism (7).

 Peroxisome proliferators cover a various set of synthetic and structurally diverse industrial and pharmaceutical chemicals, including herbicides, industrial solvents and hypolipidemic drugs, that cause significant increases in the size and number of peroxisomes in liver, liver hypertrophy, liver hyperplasia and hepatocarcinogenesis in 108 rodents (8). Following the initial discovery of mouse PPAR α , the two other PPAR subtypes, PPARβ/δ and PPARγ were identified by homology screens (2,9). Contradicting their name, the latter two subtypes do not cause peroxisome 111 proliferation in mice (8). The PPAR subtypes are encoded by distinct genes located on 112 different chromosomes (reviewed in (10)) and share a high degree of structural and sequence homology, but each subtype displays a divergent pattern of tissue-specific expression and ligand-binding specificity (9,11).

115 PPARs were originally described as orphan nuclear receptors, but shortly thereafter a plethora of potential endogenous ligands were described (12), acknowledging PPARs 117 to be activated by a wide range of endogenous ligands including fatty acids and fatty acid metabolites, such as hydroxylated eicosanoids, prostaglandins and leukotrienes, and by many synthetic compounds including fibrates and antidiabetic 120 thiazolidinedione drugs (10,13).

121 Most of the physiological functions of PPARs can be explained by their activity as 122 transcription factors that modulate the expression of specific target genes (14-17). 123 PPARs can bind to sequence-specific target elements in the promoter region of target genes following heterodimerization with the retinoid receptor to drive several aspects of normal cellular physiology as well as pathology (15,18). This includes lipid metabolism, glucose homeostasis, cell differentiation, proliferation, obesity, cancer and inflammation (19,20). In addition to positively regulating gene expression, activated PPARs can inhibit gene expression by negatively interfering with the activity of pro-inflammatory transcription factors including signal transducer and activator of transcription (STATs), activator protein-1 (AP-1), and nuclear factor-κB 131 (NF-κB) (21-23). Such transrepression mechanisms are likely to participate in the 132 anti-inflammatory actions of PPARs (24).

133 In this review, we will focus mainly on the molecular biology of the PPAR α subtype and its role in inflammation control and inflammation-related disorders. Hereby, the focus is on liver, adipose, cardiovascular and immune system as prime tissues/organ systems of interest.

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144 **II. PPAR expression profile, gene and protein structure**

145 PPAR α controls the expression of numerous genes involved in a plethora of lipid 146 metabolic pathways, including microsomal, peroxisomal and mitochondrial fatty acid 147 oxidation (FAO), fatty acid binding and activation, fatty acid elongation and 148 desaturation, synthesis and breakdown of triglycerides (TGs) and lipid droplets, 149 lipoprotein metabolism, gluconeogenesis, bile acid metabolism, and various other 150 metabolic pathways and genes (25). Consistent herewith, PPAR α is widely expressed 151 in tissues with high fatty acid oxidation rates, such as heart, liver and skeletal muscle, 152 and functions as a major regulator of fatty acid homeostasis (26-28). PPAR α 153 expression is also significant in brown adipose, adrenal and kidney tissue, as well as 154 in several inflammatory/immune cell types such as monocytes/macrophages 155 (26,27,29).

156 It is generally agreed that active PPAR α is predominantly localized in the nucleus 157 (30). However, PPARα was reported to be localized in the cytoplasm of rat 158 chondrocyte primary cultures (31). Similarly, in differentiated human macrophages, 159 PPAR α was also much more localized in the cytoplasm (32). Hence, as with other 160 nuclear receptors, PPAR α displays a dynamic shuttling between the cytosol and the 161 nucleus. This shuttling is influenced by Ca^{2+} signaling and ligand binding, the latter 162 enhancing nuclear import of PPAR α (33). In this way, internal and/or external cell 163 signals can be translated into changes in subcellular localization and turnover/stability 164 of PPAR α and, as a consequence, affect its regulation of cellular functions (33). The 165 observation of a nucleo-cytoplasmic shuttling is not only restricted to PPAR α , but 166 also applies to PPAR γ . MEK-dependent export of PPAR γ from the nucleus reduces its 167 transcriptional activity and therefore represents a mechanism by which nuclear 168 activities of PPAR γ and possibly also other NR are downregulated (34). Further investigations are required to fully elucidate the underlying molecular mechanism of nucleo-cytoplasmic shuttling as well as the subcellular functions of this receptor. 171 Moreover, a better understanding of where and how PPARs are activated, turned over and regulated is of the utmost importance.

173 The human and mouse PPAR α genes, located on chromosome 22 and chromosome 15 respectively, encode 468 amino acid (AA) polypeptides with 91% homology (35). As 175 in the mouse, the human PPAR α gene encoded mRNA is derived from 8 exons with a 5' untranslated region encoded by exons 1, 2, and part of exon 3. The remainder of exon 3 and exons 4-8 contribute to the coding region of PPARα (35,36) (**Fig. 1A**). The last 232 bp of exon 8 contribute to the 3'-untranslated region (37). In correspondence with other nuclear receptors and PPARs, the encoded PPARα protein displays four distinguishable functional domains whose cooperation modulate 181 receptor activity (38). These domains are: an N-terminal region (A/B domain), a central DNA-binding domain (DBD, C domain), a flexible hinge region (D domain) and a C-terminal ligand-binding domain (LBD, E domain) (39,40) (**Fig. 1B**) (**Table 1)**.

185 The N-terminal A/B domain, which is the least conserved domain among PPARs, harbors a ligand-independent activation function-1 (AF-1) region. This region, enriched with acidic amino acids, has a low level of basal transcriptional activity and can function independently of ligand-binding (tested via GAL4-fusion proteins in one-hybrid assays) (35,40). Although the A/B domain suffers from a poor structural organization, secondary structure formation in this domain is suggested as an 191 important step towards AF-1-mediated transactivation (38,40). The importance of the A/B domain is emphasized by the fact that its deletion results in a gene-dependent alteration in PPARα transcriptional activity. For example, A/B domain deletion

 interrupts PPARα-mediated transactivation of the acyl-CoA oxidase (ACOX) promoter, however, transactivation of the cytochrome P450 4A6 (CYP4A6) promoter is unaffected (40,45). In addition, AF-1 has also been proposed to be a key determinant of subtype-selective target gene expression amongst the PPAR subtypes. For example, addition of the PPARα A/B domain to PPARγΔAB (A/B domain 199 truncated) enhances its ability to activate $PPAR\alpha$ -specific target genes (38). Vice 200 versa, addition of the PPAR_Y A/B domain to the non-adipogenic PPAR β /δ Δ AB was sufficient to provide adipogenic potential to the resulting PPARβ/δ chimeric protein (44) . Post-translational modifications (PTMs) further fine-tune PPAR α functionality. PPARα AF-1 domain phosphorylation leads to changes in transcriptional activity and even ligand binding of the receptor (43). For example, MAPK-mediated 205 phosphorylation of serine 12 and 21 in the human PPAR α A/B domain increases the transcriptional activity in response to mitogenic stimuli, such as high concentration of insulin (58). On the other hand, glycogen synthase kinase 3 (GSK3)-mediated 208 phosphorylation of serine 73 increases the ubiquitination and degradation of PPAR α 209 (42). Concerning its relation/association with other domains in the protein, PPAR α 210 AF-1 synergistically activates transcription together with AF-2, as found for other 211 nuclear receptors (40).

212 The DBD, which is highly conserved and rich in cysteine and basic amino acids, 213 consists of two zinc-finger binding motifs (45). This domain is responsible for 214 physical interaction with DNA. Like other PPAR subtypes, PPAR α binds DNA as an 215 obligate heterodimer with retinoid X receptors (RXR) $(NR2B)$ (49) . The receptor can 216 be activated not only by phosphorylation of the A/B domain or via ligand binding to 217 PPARs themselves, but also via associating with RXR, which for example binds 9-cis 218 retinoic acid. The latter activation mechanism is referred to as the permissive effect. 219 Ligand binding of either receptor thus activates the PPAR-RXR complex; however, 220 simultaneous binding of both ligands can lead to a more potent or even synergistic 221 activation (59-65). These different activation mechanisms allow fine-tuning of PPAR 222 activity.

223 The PPAR α -RXR heterodimer recognizes and binds to PPAR response elements 224 (PPREs), which are localized in gene regulatory regions and organized as a direct 225 repeat type 1 (DR-1). DR-1 consists of two copies of the hexameric nucleotide 226 recognition motif 5'-AGGTCA-3' separated by a single nucleotide (47,48). An 227 additional consensus AACT motif is positioned 5' to the DR1 (66,67). Binding to the 228 PPRE occurs in such a way that PPAR α binds to the 5' extended half-site of the 229 response element, while RXR occupies the 3' half-site $(5,66,68)$. The PPAR α -RXR 230 heterodimers can form independently of the PPARα ligand. These unliganded 231 heterodimers recruit the corepressor protein complex and inhibit target gene 232 transcription (69). Upon ligand binding, the corepressor complex is released from the 233 PPAR α -RXR heterodimer and the coactivator complexes will be recruited to the 234 promoter region of target genes, thereby initiating transcription (70) (**Fig. 2A**).

235 The first natural PPRE was found in the promoter region of the ACOX gene (36,71). 236 In addition, recent genome-wide profiling of $PPAR\alpha$ binding sites revealed about 237 46% of PPAR α -RXR binding sites within intronic regions (72). Using PPAR γ as 238 study case, chromatin accessibility in general was identified as a major determinant 239 for transcriptional activity, as PPAR-RXR heterodimers were found to primarily bind 240 in open chromatin regions (73-75).

241 While PPAR α binds DNA only as a heterodimer and not as a monomer, deletion of its 242 N-terminal A/B domain generates a truncated protein that binds DNA non-specifically 243 as a monomer in *in vitro* assays (45). Although the physiological significance of such

244 binding is not completely understood, it may serve as evidence of an interdomain 245 communication. The DBD of PPAR γ was shown to form an interface with its own 246 LBD as well as with the LBD of its heterodimeric partner, hereby influencing ligand 247 binding (76). More crystallization efforts are needed to challenge or validate this 248 interface concept in the case of PPAR α . The DBD of PPAR α (and PPAR γ) is also 249 subject to phosphorylation, which serves to further modulate the transcriptional 250 activity of the receptor (46).

251 Like PPARs, many other nuclear receptor members also need RXRs as their 252 obligatory heterodimerization partner, raising therefore the possibility of a 253 competition between particular NRs for their common binding partner (77). This 254 competition may have important biological outcomes. For instance, liver X receptor α 255 (LXR α) activation by either overexpression of LXR α or ligand activation suppresses 256 PPAR α signaling through reduction of active PPAR α -RXR heterodimers in the liver 257 and thus diminished PPRE occupancy. RXRα supplementation restored the LXR-258 mediated inhibition of PPAR α activity (78). Conversely, excess PPAR α in the 259 presence of its ligands suppresses the sterol regulatory element-binding protein-1c 260 (SREBP-1c) promoter that contains two LXR response elements (79), indicating that 261 the mechanism of competition for RXR α between PPAR α and LXR may be reciprocal 262 (78). This relationship between PPAR α , LXR and RXR could be crucial for a mutual 263 regulation of both LXR and PPARα activities, and thus for the nutritional regulation 264 of their downstream target genes. However, the physiological validation of these 265 mechanisms still remains to be demonstrated. Another example of competition for 266 RXRα between RXR-partnering NRs was described for the thyroid hormone receptor 267 (TR) with LXR (80).

268 Of note, Silent information regulator 1 (SIRT1) was also shown to bind PPAR α , 269 through a region homologous to the PPAR α binding domain in RXR α . Interestingly, 270 the balance between SIRT1 and RXR appears to determine whether PPAR α is a 271 positive or negative regulator of genes involved in fatty acid metabolism (81). 272 Additionally, the PPARα-SIRT1 complex would directly bind a single hexameric 273 motif embedded in estrogen-related receptor (ERR) elements (ERREs), thus 274 competitively downregulating ERR target genes, at least in the heart. These findings 275 suggest a PPAR α -dependent transcriptional repression mechanism in the control of 276 ERR-driven mitochondrial respiration and cardiac contraction (82,83). In addition, 277 exposure to a high-fat diet (HFD) also induces the formation of a PPARα-SIRT1 278 complex, impairing an association of PPARα with the pro-inflammatory transcription 279 factor p65. Collectively, this shift is expected to lead to the repression of PPARα-280 target genes involved in fatty acid oxidation and enhancement of pro-inflammatory 281 pathways in the heart (84). Albeit intriguing, more experimental evidence is necessary 282 to prove that PPAR α can bind DNA as an RXR-independent monomer and whether 283 this mechanism is also operative in other tissues such as the liver.

284 Immediately adjacent to the DBD is the highly flexible hinge domain (D), connecting 285 the DBD and the LBD (35). This region binds corepressor proteins, such as the 286 nuclear receptor corepressor (NCoR), to the receptor in its inactive, unliganded state 287 (51) and serves as a target for PTMs including phosphorylation on serines 179 and 288 230 catalyzed by PKC and the SUMOylation at lysine 185 that promote NCoR 289 recruitment (52,53). In addition, mutation of a potential phosphorylation site within 290 the PPAR α hinge was shown to prevent dimerization capacities (46). Another study 291 on truncated PPAR α lacking part of the hinge region and LBD suggested that the 292 hinge region might contain a nuclear localization signal (NLS) (50). Consistently, a series of truncation mutants revealed that PPAR nuclear transport is mediated by at least two nuclear localization signals, i.e. one in the DBD-hinge region and one in the 295 AF-1 domain (33). It was suggested that the hinge region also interferes with PPAR α binding to DNA and hence takes up an important suppressor role (85). More specifically, the ribosomal protein L11 (rpL11) was shown to interact with the D-298 domain of PPAR α and to inhibit PPRE-driven reporter activity (85). Furthermore, heat shock protein 90 (Hsp90) was reported to associate with both the hinge and LBD of PPARα. Using several approaches to disrupt Hsp90 complexes within the cell, 301 Hsp90 was confirmed as a repressor of PPAR α (and PPAR β /δ) activity (54).

302 Next, the PPAR α -LBD harbors a ligand-dependent transactivation function (AF-2) 303 and has a structure consisting of an α-helical sandwich and a four-stranded β-sheet, 304 forming a Y-shaped cavity of 1400 Å called the ligand-binding pocket (LBP) (55). 305 The volume of the PPAR α -LBP is quite comparable to other PPAR subtypes, but is 306 substantially larger as compared to most other NRs. Among the PPARs, The PPAR α 307 LBP is more lipophilic which potentially explains the greater affinity of PPAR α to 308 bind the more saturated fatty acids (55). Following ligand-binding, the AF-2 domain 309 undergoes conformational changes, which allows for the interaction with various 310 coactivators carrying LXXLL motifs (L–leucine, X–any amino acid) (55), such as 311 cAMP-response element-binding protein (CBP)/p300 and steroid receptor 312 coactivator-1 (SRC-1/NCoA-1) (55). In addition, the LBD is crucial for dimerization 313 with RXR; this was confirmed in studies involving deletion of PPAR α helix 10-12 as 314 well as a L433R mutation in PPARα, which caused impaired heterodimerization with 315 RXR (51,57). PPARα furthermore possesses two nuclear export signals (NES) in the 316 DBD and LBD regions, which are recognized by the export receptors calreticulin and 317 chromosomal maintenance 1 (CRM1), respectively. A recent study confirmed the

318 presence of a functional NES in the LBD, i.e. at AA 300-308 in human PPAR α (86,87).

320 As mentioned earlier, human and murine PPAR α show 85% identity at the nucleotide level and 91% identity at the amino acid level, however evidence exists for a genetic 322 heterogeneity in the functional coding sequence of human PPAR α that translates into functional differences in receptor activity. For example PPARα L162V, a variant of 324 the human PPAR α gene resulting in the introduction of an amino acid substitution 325 within the PPAR α DBD, is found at an allelic frequency of ~0.025–0.073 in ethnically diverse populations (88-90), while being more frequent (0.745) in North 327 Indians (91). Functional studies on the L162V gene variant demonstrate enhanced ligand-induced activity when compared to the wild-type (88,91). Importantly, in 329 various studies the L162V polymorphism is correlated with dyslipidaemia, coronary ischemic events (37,88-90,92-94) and adiposity (95,96). However, other studies have not replicated these correlations (97,98). These inconsistencies could be due to differing environments, genetic background and disease status between the studied populations. In this context, the effect of the L162V polymorphism could be modulated via gene-drug and gene-nutrient interactions (94,99,100). For example, the association of the L162V genotype with a particular lipoprotein profile may be modified by dietary fats (94,101).

337 V227A, a natural variant in which the PPAR α hinge region is affected, was identified in a Japanese population with a frequency of 5.0% (102-104) and in a Chinese 339 population with a frequency of approx. 4% (105). This variant attenuates the PPAR α - mediated transcription of CYP4A6 and the mitochondrial 3-hydroxy-3- 341 methylglutaryl-CoA (mHMG-CoA) synthase genes in the presence of fibrates. This transcriptional repression was ascribed, at least partially, to increased corepressor

343 binding to PPAR α and defective release of NCoR from chromatin (106). Interestingly, the V227A variant was associated with perturbations in plasma lipid levels and modulated the association between dietary polyunsaturated fatty acid intake and high-346 density lipoprotein (HDL) cholesterol concentrations (105). Of note, the PPAR α V227A polymorphism may be involved in the pathogenesis of non-alcoholic fatty liver disease (NAFLD) and may play a protective role in obesity (107). Although epidemiological and interventional studies demonstrate a link between PPARα polymorphisms and metabolic parameters, more studies are required to understand 351 whether and how the response to fibrate drugs and other PPARα-regulated pathways are affected by the polymorphisms.

 In addition to polymorphic variants, a truncated splice variant of PPARα, generated by alternative splicing resulting in the skipping of exon 6, has been detected in human 355 PPAR α (Fig. 1A) (50,108). This variant lacks the hinge region and the entire LBD and is unable to bind to PPREs. As a consequence, it exerts a dominant negative transcriptional repressive activity on full-length PPARα through titration of the coactivator CBP by competing for coactivators. It was speculated that differences in 359 the expression level of truncated hPPAR α might be implicated in the heterogeneity in response to fibrates among different patients (50). Of interest, genome-wide correlation analysis with subsequent pathway enrichment analysis indicated a 362 selective role for truncated PPAR α as an anti-proliferative and anti-inflammatory factor (109). Further experimental manipulation of human and mouse hepatocytes by specific knock-down and overexpression analysis substantiated this hypothesis (109). *In vivo* evidence for a dominant negative transcriptional repressive activity is lacking so far. Therefore, whether this variant has any specific physiological significance with

- regard to metabolic processes, inflammation as well as its relevance for
- hepatocarcinogenesis still needs to be revealed and requires further investigation.

370 **III. Modulating factors of PPAR activity**

371 **PPARα gene and protein expression**

372 The expression of the PPAR α gene is regulated by various physiological conditions 373 such as stress, glucocorticoids (GCs), growth hormones, insulin and leptin. As an 374 important physiological response, PPARα expression is induced during fasting 375 resulting in an enhanced expression of PPAR α target genes in liver to operate fatty 376 acid oxidation and to form substrates that can be metabolized by other tissues. Vice 377 versa, PPAR α also controls, directly or indirectly, lipogenic pathways (which will be 378 further discussed in detail in section V). Liver lipogenesis particularly allows FA 379 synthesis in a fed state when dietary carbohydrates are abundant (110,111). 380 Adipocyte-derived factors may further influence the expression level and subsequent 381 PPAR α activity. Consistent with this, administering leptin, considered pro-382 inflammatory, correlated with an increased PPARα hepatic expression in ob/ob mice 383 (112). The mechanisms underlying the induction of PPAR α transcription by leptin 384 may be connected to changes in fatty acid flux mediated by leptin signaling (112). 385 Leptin-stimulated PPAR α gene expression was also observed in muscle cells, through 386 the activation of and changes in the subcellular localization of AMPK- α 2 (113). 387 Additionally, leptin was found to enhance PPARα gene transcription in pancreatic β-388 cells (114) but not in adipose tissue (115). The basis for this discrepancy is not 389 known, but findings generated using isolated model systems will definitely require 390 further confirmation *in vivo*.

391 On the other hand, the anti-inflammatory adipokine adiponectin also increased 392 PPARα-responsive promoter activity in H4IIEC3 cells, an effect that required co-393 expression of PPAR_Y coactivator-1 α (PGC-1 α), suggesting that adiponectin-induced 394 PPAR α activity may be mediated through up-regulation of PGC-1 α expression (116).

395 In contrast, growth hormone (GH) has been shown to rather decrease $PPAR\alpha$ mRNA in cultured rat hepatocytes (117,118). A similar effect was shown in 397 hypophysectomized rats (119) . In addition, GH also decreases PPAR α transcriptional 398 activity (120) as well as the expression level of PPAR α (121). Similarly, treatment of primary hepatocytes for 3 days with another anabolic signal, insulin, also led to 400 reduced PPAR α levels (122), whereas short-term exposure to this hormone increased PPARα transactivation via phosphorylation of the ligand-independent AF-1 domain (123).

403 Next to tissue-specific differences in PPAR α levels, the expression of PPAR α in a given tissue further oscillates under the control of the circadian rhythm. For example, PPAR α expression peaks during daytime in mice, which correlates with the timing of fasting in these nocturnal rodents (124). These diurnal patterns thus correspond well 407 with known functions of PPAR α in physiology but also pathophysiology (125). PPAR α is under the transcriptional control of the GC-activated glucocorticoid receptor (GR) in rat hepatocytes (126). As such, PPARα expression follows a diurnal 410 rhythm, which parallels that of circulating corticosterone and the levels of PPAR α are 411 also up-regulated during times of stress (127). Moreover, the circadian expression of PPAR α was reported to be directly regulated by the core-clock transcription factors brain and muscle Arnt-like protein (BMAL) and circadian locomotor output cycles kaput (CLOCK) both *in vivo* and *in vitro (128)*. In another study, the proline- and acidic amino acid-rich basic leucine zipper (PARbZip) proteins were also shown to 416 modulate the rhythmic expression and activity of PPAR α . The mechanism was 417 proposed to be indirect and involved a cyclic generation of PPAR α ligands which initiate a feed-forward loop, in which PPARα enhances transcription from its own gene (129). In addition, enhanced PPARα transcript levels were observed in both B

420 and T cells upon GCs treatment (130), indicative of a function of PPAR α in the 421 immune system (see further below under section 6).

422 Additionally, PPAR α expression is also reported to be related to aging. In this line of 423 reasoning, examination of mRNA expression in mouse spleens revealed an age- 424 associated decline in the levels of mRNA encoding PPAR α and its concomitant target 425 genes ACOX and catalase, which was accompanied by a constitutive pro-426 inflammatory cytokine production (131). Similarly, a recent study has shown that 427 aging inhibits PPARα activation as assessed by diminished mRNA levels of its target 428 genes ACOX and carnitine palmitoyl transferase 1 (CPT1) in aged rat livers (132).

429 Metabolite levels also affect PPAR α expression; intriguingly, glucose and metabolites 430 (glucose-1-phosphate and glucose-6-phosphate) were shown to bind PPARα with high 431 affinity resulting in a significantly altered PPAR α secondary structure (133). In this 432 report, in vitro addition of glucose alone reduced the interaction of PPAR α with its 433 coactivator SRC-1. Conversely, in the presence of the activating ligands arachidonic 434 acid and clofibric acid, glucose rather increased PPAR α interaction with SRC-1, DNA 435 binding and activation of the β-oxidation pathway (133). In addition, high glucose 436 levels regulate PPAR α expression in human macrophages. Incubation of murine J774 437 macrophages with high glucose concentrations increased the expression of PPAR α at 438 the mRNA and protein levels and enhanced its binding to the PPRE of the lipoprotein 439 lipase (LPL) promoter (134). In contrast, glucose decreases PPARα expression in 440 pancreatic β-cells leading to a reduced FAO and increased phospholipids and TG 441 synthesis, a mechanism believed to underlie β -cell glucolipotoxicity (135).

442 NF- κ B-driven cytokines such as TNF α , IL-1 β and IL-6 also affect the expression of 443 PPARα. In TNFα-treated male Sprague Dawley-rats, PPARα mRNA and 444 corresponding PPAR α protein levels were significantly reduced, likely contributing to

445 the decrease of peroxisomal β-oxidation (136). IL-1β and IL-6 both repress the induction of PPAR α expression exerted by the combined action of clofibric acid and 447 the GC dexamethasone in fetal rat hepatocytes (137).

 The presence or absence of the activity of other transcription factors also impacts PPARα levels and activity. In hepatocytes, Krüppel-like factor 6 (KLF6) deficiency reduces PPAR α protein expression and downstream targets resulting in attenuation of 451 lipid and glucose abnormalities associated with a HFD. KLF6 mediated induction of PPARα protein expression through miRNA 10b repression (138). PPARα is also regulated at the transcriptional level by other nuclear receptors such as hepatocyte nuclear factor 4 (HNF4), chicken ovalbumin upstream promoter-transcription factor II 455 (COUP-TFII), and others. While HNF4 positively regulates hPPAR α expression via a DR1 element present in the hPPARα promoter, COUP-TFII also bound this site but 457 antagonized this effect (139). The regulation of PPAR α by HNF4, consistent with the tissue-expression pattern of PPARα, was also demonstrated *in vivo* using HNF4- 459 deficient mice that display significantly lower PPAR α mRNA levels compared with the wild-type mice (140). Exemplary of another metabolite, bile acids stimulate hPPAR α expression in a species-specific manner through a mechanism involving the 462 binding of Farnesoid X receptor (FXR) on its promoter (141). Moreover, PPAR α expression in the mouse small intestine was shown to be under the control of LXR and the pregnane X receptor (PXR) (142).

 Cross-talk with other nuclear receptor signaling pathways may also modulate PPAR 466 activity, as illustrated by the direct control of the PPAR β promoter by the vitamin D receptor (143). In this respect, and as mentioned earlier, nuclear receptors that function as heterodimers with RXR can potentially compete with PPAR α signaling in tissues in which the amount of RXR is limiting (77).

 Furthermore, PPARα turnover is controlled by ligand specificity and availability (**Fig. 3**). In this context, Blanquart *et al.* demonstrated that synthetic PPARα ligands such as Wy14,643, GW7647 or fibrates prolonged half-life and increased stability of the receptor, preventing thereby its ubiquitination and its subsequent degradation via the proteasome (41). Besides, PPARα agonists, such as fenofibrate, markedly up- regulated hPPAR α mRNA levels in primary cultures of human hepatocytes as well as hPPAR α promoter activity suggesting that PPAR α positively autoregulates its own expression (139).

478 Of note, hPPAR α mRNA levels in the liver vary among individuals (50,109), 479 suggesting that PPAR α is strongly regulated at the gene level in human liver by genetic and/or environmental factors (50).

481 Last but not least, the function of PPAR α as a transcription factor is controlled by its PMTs and cofactor recruitment (**Fig. 3**), which will be further discussed in detail in the next sections.

484 Together, regulation of the PPAR α expression levels efficiently modulates PPAR α activator responses and thus may affect downstream pathways involved in metabolism and inflammatory processes.

PPARα cofactors

489 As with other nuclear receptors, PPAR α transactivation activity is regulated by the presence of coregulators, which may function as corepressors or coactivators depending on the context. In the absence of a specific ligand, PPARα binds co- repressors, such as NCoR and silencing mediator for retinoid and thyroid hormone 493 receptors (SMRT) (26). The receptor interacting protein 140 (RIP140) is a PPAR α corepressor, which is known to repress the activity of NRs by competing with coactivators and by recruiting downstream effectors such as histone deacetylases (HDACs) (144). Interestingly, mice devoid of the corepressor protein RIP140 show resistance to HFD-induced obesity and hepatic steatosis in association with upregulation of genes involved in energy dissipation. Such phenotype may be 499 suggestive of a role for this corepressor in PPAR α signaling (145). Whether or not 500 PPAR α is directly involved remains to be shown. On the other hand, known 501 coactivators that have been found in a large complex with $PPAR\alpha/RXR$ heterodimers include PGC-1α, PGC-1β, CBP/p300, SRC-1, PPAR-interacting protein (PRIP), PPAR-binding protein (PBP/TRAP220/DRIP205/Med1) as well as coactivator- associated proteins PIMT (NCoA6IP) and coactivator-associated arginine methyltransferase-1 **(**CARM-1**)**. Some of the coactivators, including the SRC-1 family and CBP/p300, contain intrinsic histone acetyltransferase (HAT) activities 507 required to facilitate chromatin remodeling (Reviewed in (146)). The PPAR α - interacting cofactor (PRIC) complex isolated from rat liver nuclear extracts reveals 509 the presence of many of these coregulators, likely forming one mega complex (147). Most cofactors are not receptor-exclusive, yet, emerging gene knockout mouse 511 models show that some of the coactivators are essential for PPAR α signaling. For example, conditional deletion of PBP/MED1 gene in liver results in the abolition of PPARα ligand-induced pleiotropic effects, indicating that PBP/MED1 is essential for 514 the function of PPAR α (148).

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PPARα post-translational modifications

 As for other nuclear receptors, PPARα activity is finely tuned by various PTMs including phosphorylation, ubiquitination and SUMOylation (41,53,149).

 PPAR α is a phosphoprotein of which the transcriptional activity is affected by cross-522 talk with several kinases, including MAPK (ERK1/2), c-Jun N-terminal kinases (JNK)/p38), AMPK, protein kinase A and C (PKA, PKC) and GSK3 (**Table 2**). The 524 effects of phosphorylation on PPAR α activity depend on a number of features such as the nature of the modified residue, as well as the stimulus and the kinase involved. PPARα phosphorylations may also have different functional outcomes depending on the context, with effects on receptor stability, susceptibility to proteosomal degradation, enhanced recruitment of cofactors and transactivation. PPARα was first 529 shown to be phosphorylated in primary rat adipocytes in culture in response to insulin, resulting in the stimulation of basal as well as ligand-dependent transcriptional 531 activity (149). Along the same line of interest, insulin treatment of human HepG2 hepatoma cells increases PPARα phosphorylation enhancing thereby its transcriptional activity. This insulin-mediated phosphorylation of PPARα via the ERK1/2 MAPK pathway involves amino acids Ser12 and Ser21 in the AF-1 domain resulting in transcriptional activation probably by relieving inhibition by corepressors (42,58). In line with this, in rat neonatal cardiac myocytes, stress-activated p38 537 MAPK phosphorylates Ser 6, 12 and 21 residues located within the N-terminal A/B domain of the PPARα protein leading to an increase in ligand-dependent transactivation of targets and enhanced cooperativity with the transcriptional coactivator PGC-1. It was speculated that PPARα phosphorylation and the subsequent specific recruitment of PGC-1 favor cardiac mitochondrial fatty acid β-oxidation during periods of stress (150). In contrast, during cardiac hypertrophic growth, the ERK-mediated phosphorylation of PPARα was shown to have an opposite effect to 544 that of p38 kinase resulting in a reduced capacity for myocardial lipid and energy 545 homeostasis (151) demonstrating therefore the importance of cellular context. In FAO 546 hepatic-derived cells PPARα phosphorylation level is increased by ciprofibrate, 547 subsequently enhancing its transcriptional activity as demonstrated by a rise in ACOX 548 gene expression (152).

549 Given that substrates for MAPK phosphatase 1 (MKP-1, also known as DUSP1) 550 include many members of the MAPK family including ERK1/2 and p38 MAPK, 551 overexpression of MKP-1 in turn affected the constitutive PPAR α activity, thus acting 552 in a feedback control mechanism (153). Taken together, these studies hint at pivotal 553 activating roles of MAPK on PPAR α activity, thereby placing this receptor under the 554 control of growth factors and various cellular stressors, as already mentioned above.

 In vivo phosphorylation studies show that the level of phosphorylated PPARα is increased by treatment with its agonist Wy-14,643 as well as the PKC activator phorbol myristoyl acetate (PMA) (46). Particular modifications can increase a subset of PPAR-mediated functions, e.g. phosphorylation at serine 179 and 230 in the 559 PPARα hinge region by PKC leads to enhanced transactivation potential, but impairs its transrepression activity (52). Consistently, PKC inhibition impairs ligand-activated PPARα transcriptional activity and decreases PPARα ligand-induction of its target genes including PPARα itself and CPT1, but enhances PPARα transrepression properties. Thus, the PKC signaling pathway acts as a molecular switch dissociating 564 the transactivation and transrepression functions of PPAR α (52). In support, it was 565 proposed that the capacity of simvastatin to mitigate PKC action indirectly stimulates 566 PPARα transrepression activities towards NF- κ B and as such explains statin-mediated acute anti-inflammatory effects (154). As indicated earlier, PKC phosphorylation is 568 believed to be important for PPARα and RXR interaction as mutation of the PKC

569 phosphorylation sites T129 and S179 in the hinge region of PPARα reduces 570 heterodimerization (46).

571 The fasting-responsive kinase PKA enhances $PPAR\alpha$ activity converging in the 572 stimulation of PPARα target genes. It was speculated that under stress, fasting or 573 exercise, PPARα phosphorylation in response to PKA leads to an enhancement of β-574 oxidation, production of glucose and ketone bodies, which serve as fuel for muscles 575 and brain (155). Thus, physiological cAMP modulators such as fasting, stress or 576 exercise potentially modulate PPARα activity. Of note, PKA-mediated PPARα 577 phosphorylation occurs mainly in the DBD, while the enhancement of activity 578 generally involves the AF-2 domain (155), suggesting an effect of phosphorylation on 579 interdomain communication. As another starvation-linked kinase, AMPK activity may 580 influence the expression of PPAR α and PPAR α target genes in certain tissues. In the 581 murine muscle cell line C2C12, for example, leptin stimulates PPAR α gene 582 expression and subsequent FAO through the activation of α 2-subunit containing 583 AMPK (113). Moreover, AMPK activation increases mRNA expression of PPAR α 584 and its target genes such as ACOX and CPT1 as well as PGC-1; this effect was 585 correlated with an increased FAO in skeletal muscle (156). In addition, AMPK and 586 p38 MAPK were involved in the activation of PPAR α by adiponectin in muscle cells 587 (157). However, whether and which residues within PPAR α may be phosphorylated, 588 remains an unresolved issue. Next to adiponectin, AMPK was also described as a 589 modulator of PPARα expression in response to glucose levels (158,159) suggesting 590 that cellular energy levels might affect the PPAR α -regulated signaling network in a 591 ligand-independent manner. Recently, a novel mechanism was uncovered by which 592 AMPK controls PPARα-driven gene expression in murine liver subjected to a 593 prolonged fasting. Activated AMPK was recruited to promoters of glucose and lipid 594 metabolism genes that are co-controlled by PPAR α and GR α (160). For the latter 595 example in liver, it also remains to be discovered which residue(s) of involved 596 transcription factors, PPAR α , GR or both, may be phosphorylated by AMPK.

 PPARα phosphorylation, besides affecting target gene regulation, also affects the stability and turnover of PPARα through ubiquitination (41,161). Ubiquitination involves the conjugation of a small protein, ubiquitin, to the target protein via a sequential action of particular enzymes which primarily leads to degradation of the 601 receptor by the 26S proteasome. PPAR α is targeted by the E3 enzyme murine double 602 minute 2 (MDM2), which leads, depending on the cellular PPAR α levels, to stabilization of receptor levels or degradation via the ubiquitin-proteasome pathway (162). The effect of degradation on the transcriptional activity is still under debate; Blanquart *et al.* showed an inverse correlation between PPARα degradation and activity, which is in contrast to results obtained by the group of Vanden Heuvel (41,42). Recently, a muscle ring finger-1 (MuRF1)-mediated monoubiquitination of cardiomyocyte PPARα was discovered that enhances nuclear export of the receptor 609 and consequently inhibits $PPAR\alpha$ -induced FAO in cardiac tissue (87).

610 GSK3 is a key enzyme of glycogen metabolism and able to inactivate glycogen 611 synthase. Its activation phosphorylates PPAR α predominantly at serine 73 in the A/B 612 domain. This modification negatively affects $PPAR\alpha$ stability and promotes ubiquitin 613 proteasome-mediated degradation (42). PPAR α ubiquitination may thus constitute a 614 regulatory pathway that may contribute to the duration and the magnitude of the 615 response to PPAR α activators (41,161). Function-wise, when blocking GSK3 by 616 biliverdin reductase A (BVRA) in the liver, a protective effect against steatosis is 617 inferred, due to a preservation of hepatic PPAR α activity (163).

 Next to phosphorylation and ubiquitination, PPARs can be modified by SUMOylation. SUMOylation is a reversible modification that entails the covalent attachment of small ubiquitin-related modifier (SUMO) proteins via specific enzymes 621 (164). Murine PPAR α was shown to be SUMOylated on lysine 358 (56). This 622 particular SUMOylation triggers the interaction with GA-binding protein α bound to the 7α-hydroxylase cytochrome P450 7b1(*cyp7b1*) promoter resulting in specific down-regulation of this gene. Hence, SUMO modification can be regarded as a 625 positive regulator of mPPAR α transrepressive activity (56). Likewise, human PPAR α undergoes SUMOylation by SUMO-1 on lysine 185 in the hinge region, leading to down-regulation of its transactivation capacity through the selective recruitment of NCoR (53) (**Table 3**). Both SUMOylation sites are targeted by the E3 enzyme protein inhibitor of activated STAT (PIAS)y (78,164).

630 Finally, another means of regulating $PPAR\alpha$ activity indirectly includes the phosphorylation of RXR and of various associated cofactors such as PGC-1α, SMRT and NCoR (reviewed in (26)).

633 PTMs play a key role in modulating PPAR α activity in (patho)physiological contexts. The modifying enzymes themselves are modulated by the cellular metabolic status 635 and environment, hereby positioning PPAR α into a very complex regulatory network. 636 The level of complexity is even intensified knowing that PPAR α forms part of a physical assembly for variable corepressor or coactivator complexes, which are also on their turn subjected to PTMs (26). Full understanding of these complex patterns of 639 regulation requires more high-throughput technologies to characterize the PPAR α interactome, its dynamic interactions with modifying enzymes as well as the 641 mechanisms by which these interactions modulate $PPAR\alpha$ activity. These might point

IV. PPARα **natural and synthetic ligands**

PPARα natural ligands

 Adoption of orphan NRs through the identification of their endogenous ligands is key 669 towards understanding their biology (13) . The promiscuity of PPAR α for its ligands 670 can be attributed to the large size of the LBP. PPAR α is activated by a wide range of endogenous or naturally occurring biological molecules encompassing a variety of FAs and FA derivatives, including acyl-CoAs, oxidized FAs, eicosanoids, endocannabinoids, phytanic acids (165-167) and long chain dietary FAs (166,168) 674 (listed in Table 4). As such, FAs and their metabolites that interact with PPAR α can be derived from the diet or obtained via *de novo* synthesis. Many of those FAs exhibit 676 an affinity for PPAR α within a nanomolar range (166). This supports the fact that PPAR α serves as an intracellular lipid sensor. It was proposed that PPAR α senses certain endogenous lipid metabolism intermediates as ligands to regulate the expression of target genes involved in their metabolism (169). *In vivo* evidence for this came from studies using ACOX1 knockout mice. The disruption of this first and rate-limiting enzyme of the peroxisomal FAO pathway caused accumulation of long chain fatty acyl-CoA coinciding with an increased peroxisome proliferation, hepatocarcinoma and profound activation of PPARα in rodents (169,170). These 684 findings suggested that substrates of the ACOX1 enzyme may possibly be $PPAR\alpha$ 685 agonists. Numerous arachidonic acid metabolites serve as ligands for PPAR α ; in this context, 8(*S*)-HETE, an intracellular hydroxy FA derived from the oxygenation of arachidonic acid, mainly by lipoxygenases and monooxygenases, was shown to be a strong activator of PPARα (171). In addition, 20-hydroxyeicosatetraenoic acid (20- HETE), a main product of cytochrome P450 (CYP4A and CYP4F) activity that 690 catalyzes the conversion of arachidonic acid, can bind to and activate PPAR α in

691 HepG2 cells leading to enhanced expression of the PPAR α target gene, apolipoprotein 692 A-I (APOA1) (172). Leukotriene B4 (LTB4) is another putative PPAR α ligand, which 693 connects the receptor to inflammation and immune responses. Its ability to act as a 694 PPAR α agonist has been reported in several studies (165,173,174), and the binding 695 affinity is found to be within the nanomolar range (174). Stimulation of 5- 696 lipoxygenase activity leads to activation of PPAR α via production of intracellular 697 LTB_{4 (165,175)}. In endothelial cells, oxidized phospholipids were found to activate 698 PPAR α in a phospholipase A2-dependent manner suggesting that phospholipids may 699 be precursor of endogenously generated PPARα ligands (176).

 Mice with liver-specific inactivation of fatty acid synthase (FAS), a key lipogenic 701 enzyme, developed hypoglycemia, fatty liver and defects in expression of $PPAR\alpha$ target genes when subject to a fat-depleted diet. These effects were reversed by a synthetic PPARα agonist, identifying thereby the *de novo* synthesized end products of 704 FAS as endogenous activators of PPAR α in adult liver (177). This phenomenon is not limited to the liver. Indeed, selective inactivation of FAS in the hypothalamus is found to affect PPARα-dependent gene expression and feeding behavior, which are 707 corrected after hypothalamic infusion of a PPAR α activator (178). These observations suggest that FAS, which is known to be regulated by nutrition, is involved in the generation of endogenous PPARα ligands. The same research group demonstrated that FAS generates 1-palmitoyl-2-oleoyl-sn-glycerol-3-phosphocholine (16:0/18:1- 711 GPC), which seems to be a putative PPAR α ligand (179).

 Other natural compounds such as polyphenols have been identified as ligands of 713 PPAR α (180,181). PPAR α activation with resveratrol, a natural polyphenol, and some of its derivatives and analogs, results in a brain protection against stroke (182,183). Resveratrol was identified as an AMPK activator, linking this sensor of the nutritional

716 status again to PPAR α functionality (184) (see chapter 3). Since PPAR α acts as an 717 intracellular sensor of FAs levels, of which an alteration is associated with the development of several diseases such as diabetes, obesity, atherosclerosis and inflammation, finding a way of regulating the receptor activity through modulating (levels of) endogenous ligands may have the potential to generate new therapies.

PPARα synthetic ligands

 Synthetic PPARα ligands include fibrates such as clofibrate, fenofibrate, and bezafibrate (listed in **Table 4**). Fibrates are used in the pharmacological treatment of hypertriglyceridaemia (48,185) despite the low potencies of these first generation agonists on PPARα. Fibrates act by decreasing TG-rich lipoproteins in serum through 727 an increase in genes involved in FAO and a decrease in apolipoprotein C-III (APOC3) 728 gene expression (186,187). See further chapter 6 for a current status of PPAR α -targeted therapies in a clinical setting.

 Other synthetic compounds that are not approved for clinical use, but are often used for research purposes to activate PPARα include GW7647, GW9578 and Wy-14,643 (also known as pirinixic acid) (29). In addition, other chemicals with environmental impact, particularly plasticizers (example DEHP-used in the manufacturing of 734 plastics) and herbicides have been shown to function as $PPAR\alpha$ ligands (28). As their 735 potential impact in human health is unclear, further research is necessary to address 736 this with respect to lifetime health.

Selective PPARα modulators (SPPARMs)

739 The development of a selective PPAR α agonist may provide a good approach to maximize the therapeutic potential of these drug targets. Accordingly, a new

V. PPARα target genes and their function

 Numerous studies with PPARα-deficient mice during the past years have demonstrated the critical roles played by this receptor in energy metabolism, hepatic functions, inflammation, cardiac pathophysiology, cell cycle alterations and rodent hepatocarcinogenesis (27). We will next discuss functionalities of PPARα target genes and proteins per tissue type. **Table 5** summarizes the many genes that have been identified as PPARα target genes in different organs since the end of the 1990s. 771 We have indicated whether the regulation was found to be direct or indirect.

PPARα target genes in liver

 The PPAR α subtype exerts its main functions in the liver, where the receptor is involved in various aspects of lipid metabolism. These functions range from FA transport, binding, uptake, synthesis, degradation through mitochondrial or peroxisomal oxidation, storage, lipoprotein metabolism and ketogenesis under fasting 778 conditions (111). Hence, although the phenotype of $PPARα$ -deficient mice is normal, 779 fasting results in increased plasma free FA (207) levels, accumulation of lipids in the liver, hypoketonaemia, hypoglycaemia and hypothermia (110,208-210). The wide range of genes that are expressed after PPARα activation indicates that this receptor 782 has a central regulatory role in lipid metabolism (211). Recently, in a hepatocyte- specific PPARα knockout mouse model impairment in liver and whole-body FA homeostasis was observed, resulting in hepatic lipid accumulation and hypercholesterolemia during aging (212). Still today, novel mechanisms are discovered. The molecular pathway that leads to the upregulation of PPARα during 787 fasting was recently investigated by Maus and coworkers. They found that FFAs bind 788 to PLC γ -coupled cell surface receptors, such as the fatty acid transporter CD36, in the

789 liver and heart. This binding activates a specific pathway that leads to Ca^{2+} influx 790 across the plasma membrane, called store-operated Ca^{2+} entry (SOCE). Subsequently, 791 increased intracellular Ca^{2+} levels induce in a cAMP-dependent manner PGC-1 α and 792 PPAR α expression, ultimately leading to a transcriptional reprogramming of the cell 793 to FA utilization (213).

794 Altogether these data demonstrate that $PPAR\alpha$ is a master regulator of FA catabolism 795 by governing the regulation of key enzymes involved in peroxisomal, mitochondrial 796 and microsomal FAO.

797 Additionally, PPARα also plays an important role in ketogenesis by regulating 798 expression of HMG-CoA synthase 2 (Hmgcs2), a rate-limiting enzyme of 799 ketogenesis, which catalyzes condensation of acetyl-CoA and acetoacetyl-CoA to 800 generate HMG-CoA (214). Accordingly, the expression of mHMG-CoA synthase 801 gene, harboring a functional PPRE (214) in its promoter, is impaired in PPAR α null 802 mouse liver (215). Of note, HMG-CoA synthase specifically stimulates its own 803 transcription rate by acting as a coactivator of PPARα. A recent report identified 804 liver-specific PPAR α as a critical factor for ketogenesis. It was shown that PPAR α 805 protects from fish oil-induced oxidative stress leading to acute liver failure through its 806 regulatory actions on ketone body levels, hence suggesting a potential therapeutic 807 advantage of ketone bodies such as β-hydroxybutyrate in preventing lipid 808 peroxidation-induced liver damage (216). Of particular importance in regulating 809 ketogenesis, in addition to FAO, is the metabolic fibroblast growth factor 21 (FGF21). 810 Its hepatic expression is PPAR α -dependent and is induced by fasting, a ketogenic diet 811 and fenofibrate (217). Moreover, upon fasting, FGF21 production required c-AMP-812 responsive element-binding protein H (CREBH), the liver-enriched transcription 813 factor. Interestingly, it was reported that CREBH and PPAR α act as interactive trans-

814 activators that regulate each other's expression and act in synergy to activate the 815 expression of their target gene FGF21 to regulate lipolysis, FAO and ketogenesis 816 upon fasting or under an atherogenic HFD (218). Of note, CREBH acetylation is 817 required for the fasting-induced CREBH-PPARα interaction *in vivo* (219)*.* These 818 findings on synergistic cross-talk between PPAR α and CREBH might provide a 819 regulatory mechanism for nuclear receptors or transcription factors to extend their 820 biological actions and thereby coordinate adaptive physiological responses.

821 As far as lipogenesis is concerned, besides examples listed in **Table 5**, Oosterveer *et* 822 *al.* showed that induction of some lipogenic genes upon fibrate treatment is regulated transcriptionally by SREBP-1c. Consistently, in SREBP-1*-/-* 823 mice, chronic PPARα 824 activation with fenofibrate failed to induce lipogenic genes such as stearoyl-CoA 825 desaturase (Scd1) and FA desaturase 2 (Fads2) suggesting an indirect regulatory 826 mechanism (220). In line, in primary hepatocytes, the PPARα agonists Wy-14,643 827 and GW7647 increased SREBP-1c activity by triggering PPARα activity on a DR-1 828 element in the human SREBP-1c promoter (221). Moreover, in PPARα-deficient 829 mice, a disturbance in the normal regulation of SREBP-1-sensitive lipogenic genes 830 such as acetyl-CoA carboxylase (ACC) and FAS was observed (222). The effect of 831 PPAR α agonists on SREBP targets has been ascribed to increased activation of 832 SREBP-1c via enhanced proteolytic cleavage (223). PPARα agonists increase the 833 protein levels of the mature form of SREBP-1c, by increasing the rate of proteolytic 834 cleavage of its membrane-bound precursor form, without changing its mRNA levels. 835 Of note, PPARα can also indirectly modulate SREBP-1c transcription via cross-836 regulation with LXRα signaling pathway. In this line, Hebbachi *et al.* reported that the 837 intact PPAR α is required to mediate the LXR α -dependent response of Scd-1 and FAS 838 gene expression to insulin and that an exogenous non-steroidal synthetic ligand of
839 LXRα, TO-901317, was effective in rescuing PPARα deficiency, suggesting 840 participation of PPAR α in generating endogenous LXR α ligands (224).

841 In addition, malonyl-CoA decarboxylase (MCD) is transcriptionally upregulated 842 following PPAR α activation. MCD degrades the lipogenic intermediate malonyl-843 CoA, which is both a precursor of FA biosynthesis and an inhibitor of the 844 mitochondrial fatty acid transporter CPT-1 (225). PPARα hence might inhibit the 845 lipogenic pathway indirectly by inducing MCD, in line with its role in promoting 846 energy expenditure (226,227).

847 Evidence exists for a functional role of PPARα in bile acid homeostasis; however the 848 mechanism is not understood. While various studies have reported on the expression 849 modulation of Cyp7A1, a microsomal cytochrome P450 isoenzyme that regulates *de* 850 *novo* hepatic biosynthesis of bile acids by PPARα, contradictory observations were 851 made (listed in **Table 5**). In transfected cells, both murine and human cholesterol 7α-852 hydroxylase (Cyp7a1) gene promoters were stimulated by FA or Wy-14,643 853 treatments, however, the murine promoter harboring an additional PPRE displayed a 854 greater activity (228). In addition, the expression level of Cyp7a1 diminished in 855 PPARα-deficient mice, thus revealing the involvement of PPARα (229). In contrast, 856 bezafibrate treatment decreases liver bile acids and Cyp7a1 expression in wild-type 857 mice and PPARα-deficient mice (230). Moreover, a Cyp7a1 decrease was observed in 858 wild-type mice upon ciprofibrate treatment but not in PPARα-deficient mice (231). 859 Besides, other genes involved in bile acid synthesis are regulated by PPAR α including 860 cholesterol 27α -hydroxylase (Cyp27a1) whose activity was downregulated by 861 ciprofibrate treatment in a PPARα-dependent manner (231) and sterol-12a-862 hydroxylase (rat Cyp8B1), which was reported to be directly upregulated by PPAR α 863 through a functional PPRE (229).

864 PPARα*-*deficient mice display severe hypoglycemia after fasting (110), illuminating 865 the central role of PPARα in glucose metabolism. Plasma glucose is maintained 866 during fasting by combination processes including *de novo* glucose synthesis 867 (gluconeogenesis), glycogen breakdown (glycogenolysis), and control of peripheral 868 glucose utilization. Fasting-induced hypoglycemia could be caused by inhibition of 869 either of these pathways. Interestingly, genes involved in gluconeogenesis have been 870 identified as PPAR α targets including phosphoenolpyruvate carboxykinase (Pck1), 871 pyruvate carboxylase (Pcx), and lactate dehydrogenase A (Ldha) (232). Although it 872 was originally proposed that Pck1 may be specifically regulated by PPAR α in human 873 but not mouse hepatocytes (233), PPARα was shown to activate Pck1 promoter 874 activity in murine hepatoma Hepa1c1c7 cells (234), whereas in the livers of rats 875 treated with clofibrate a reduced expression level of Pck1was observed (235). In 876 addition, PPAR α promotes the metabolic conversion of glycerol into glucose through 877 the induction of several hepatic enzymes such as glycerol-3-phosphate dehydrogenase 878 (GPDH) and glycerol kinase, and of glycerol transporters aquaporins 3 and 9; all of 879 these genes are identified as targets of PPAR α (233). Moreover, PPAR α contributes to 880 gluconeogenesis by inducing the expression of pyruvate dehydrogenase kinase 881 isoform 4 (Pdk4) (236) Pdk4 phosphorylates and thereby inactivates the pyruvate 882 dehydrogenase complex, resulting in a conservation of pyruvate as gluconeogenic 883 substrate (237). Besides, PPAR α targets glycogen synthesis in liver by regulating its 884 direct target, glycogen synthase 2 (Gys-2), which is a rate-limiting enzyme in the 885 storage of glycogen in liver and adipose tissue. In consistence, Wy-14,643 was shown 886 to stimulate Gys-2 expression in wild-type but not in PPAR α -/- hepatocytes (238). 887 TRB3, another PPARα-target gene, is induced in liver during fasting and in diabetic 888 mice in a PGC-1-dependent manner. Upregulation of TRB3 expression through direct

 transcriptional control by PPARα inhibits insulin signaling by binding to and 890 preventing activation of Akt/PKB. The latter kinase is a positive regulator of the 891 cellular response to insulin (239,240). How this can be reconciled with the profile of 892 PPAR α agonists in other models or clinical, in man, is as yet unclear.

 PPARα has also a profound impact on amino acid metabolism; specifically, it downregulates genes involved in major pathways of amino acid metabolism including 895 transamination, deamination, urea cycle, oxidation of α keto acids and synthesis of amino acid-derived-products. In support of a role of PPARα in amino acids gene expression, plasma urea levels are elevated in PPARα-deficient mice. The molecular mechanisms underlying downregulation of amino acid metabolism by PPARα remain elusive; however, some studies proposed that PPARα may modulate the activity of 900 other transcription factors that are directly involved in amino acid homeostasis, including HNF4*α* and C*/*EBPα (241).

PPAR target genes in heart

904 During the transition from fetal to adult cardiomyocytes, both PPAR α and PGC-1 α expression are increased, corresponding with the primary use of FAO as an energy source (242). In the context of a pathological cardiac hypertrophy, the expression of PPARα itself is also downregulated (151,243). Conversely, PPARα activity is increased in the diabetic heart, which is characterized by increased lipid oxidation. Correspondingly, overexpression of PPARα leads to a cardiomyopathy mimicking a 910 diabetic heart (244-246). To dissect the *in vivo* role of PPAR α in myocardial 911 metabolism, transgenic mice with cardiac-specific expression of $PPAR\alpha$ were 912 generated (245,247). Interestingly, Myosin Heavy Chain (MHC)-PPAR α mice, which developed hepatic insulin resistance, also displayed enhanced IL-6 levels in the

914 circulation (245). Consistent with the metabolic phenotype of myocardial lipid 915 accumulation in MHC-PPAR α mice, the expression of genes involved in cellular FA 916 transport was activated by PPAR α . Conversely, cardiac glucose transport and 917 glycolytic genes were repressed in MHC-PPAR α mice (247), likely occurring via a 918 transcriptional mechanism, since fenofibrate also downregulated GLUT4-promoter-919 driven luciferase reporter gene expression upon cotransfection with PPAR α 920 expression plasmid into rat neonatal ventricular cardiomyocytes (247). As mentioned 921 also above, Oka *et al.* have recently demonstrated that SIRT1, which plays an 922 important role in metabolic adaptation to starvation conditions, suppresses FAO in 923 failing hearts possibly by inhibiting the heterodimerization of PPAR α and RXR α , 924 suggesting thereby that preventing PPARα-SIRT1 or restoring PPARα-RXR 925 heterodimerization may improve the progression of heart failure through modulation 926 of metabolic remodeling (81). Together, these findings highlight the complexity of a 927 balanced cardiac energy metabolism and the necessity of distinguishing the etiology 928 of heart failure to be able to predict the outcome of $PPAR\alpha$ agonists.

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930 **PPAR target genes in lipid metabolism**

931 PPAR α agonists have a pronounced effect on lipoprotein metabolism, which is 932 involved in lipid and cholesterol transport. In this regard, PPAR α ligands reduce very 933 low-density lipoprotein (VLDL) production and enhance the catabolism of TG-rich 934 particles, which indirectly decreases small dense LDL (sdLDL) particles and enhance 935 the formation of HDL particles and hepatic elimination of excess cholesterol (27). 936 More specifically, PPAR α activators decrease plasma TG-rich particles levels by 937 increasing the activity of LPL (48,248), which is the key enzyme in the hydrolysis of 938 TGs. PPARα controls LPL gene expression directly through a functional PPRE

939 located in its promoter region (249), and indirectly by reducing the levels of APOC3, 940 a natural inhibitor of LPL activity (250), likely via a mechanism involving the 941 transcription factors Rev-erbα, HNF4α or (Forkhead box protein O1) FOXO1 (251-942 253). Rev-erbα, itself induced by fibrates via a PPRE in the Rev-erbα gene promoter, 943 negatively interferes with the HNF4-mediated activation of APOC3 gene promoter 944 (253). FOXO1 stimulates APOC3 expression via specific binding to its target site 945 (CCAAACA) in the APOC3 promoter. The association of FOXO1 with the APOC3 946 promoter was significantly reduced in response to $PPAR\alpha$ activation, in line with the 947 inhibition of APOC3 promoter activity in PPARα-expressing HepG2 cells (252). 948 Besides APOC3, LPL activity is tightly modulated by other factors such as 949 apolipoprotein A-V (APOA5), angiopoietin-like protein 3 (ANGPTL3) and 950 angiopoietin-like protein 4 (ANGPTL4). In line, $PPAR\alpha$ increases expression levels 951 of APOA5, which is critical for sustaining normal plasma TG levels, through 952 functional PPREs located in its promoter (254,255). Moreover, PPARα upregulates 953 the expression levels of ANGPTL4 (256), which acts as an inhibitor of LPL activity 954 by converting active LPL dimers to inactive monomers (257), which may play a role 955 in tissue-specific directing of FA. PPARα activators affect also the metabolism of 956 HDL by regulation the expression of its major apolipoproteins, AI (APOA1) and AII 957 (APOA2). In humans, it increases the expression of these two genes by binding to 958 PPREs in their promoters, contributing to increased plasma HDL levels (258,259). In 959 rodents, however, gene expression of APOA1 is not promoted by PPARα because of 960 disabling mutations within the PPRE. In fact, the murine APOA1 gene is even 961 negatively regulated by PPAR α agonists through an indirect mechanism involving the 962 PPARα-dependent induction of the NR Rev-erbα, a negative regulator of transcription 963 (260).

964 **PPAR target gene expression in immune cells**

965 The functional expression of PPAR α by several immune cell types suggests that this 966 receptor may play an important role in regulation of immune responses. The exact 967 role of PPARα in, for instance, innate host defense mechanisms is however largely 968 unknown. PPAR α was shown essential for anti-mycobacterial responses in murine 969 bone marrow-derived macrophage cells, involving gene activation of transcription 970 factor EB (TFEB) and induction of lipid catabolism (261) . PPAR α deficiency resulted 971 in an increased bacterial load and exaggerated inflammatory responses during 972 mycobacterial infection (261). In human macrophages, PPAR α activation upregulates 973 the expression ABCA1, a member of the ATP-binding cassette-transporter family, by 974 an indirect mechanism involving the induction of $LXR\alpha$ expression. The increase in 975 ABCA1 expression promotes APOA1-mediated cholesterol efflux (262). Adiponectin 976 receptor 2 (AdipoR2), an LXR target gene, is positively regulated by PPAR α both in 977 primary and THP-1 differentiated macrophages (263). Given that AdipoR2 is 978 expressed in human atherosclerotic lesions and macrophages, modulating the 979 expression of this gene by PPAR α and LXR α may provide beneficial effects in the 980 treatment of cardiovascular disease. This finding highlights a mechanism of crosstalk 981 between adiponectin and PPAR α -LXR α signaling pathways (263). In murine 982 macrophages, PPAR α activation was also shown to upregulate the expression of the 983 classic PPAR α target gene CPT1 (154). Further in mice, PPAR α activation in hepatic 984 Küpffer cells or monocyte-recruited macrophages results in anti-inflammatory effects 985 via the down-regulation of IL-15 and IL-18 (264).

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PPAR targets linked to the rodent-only hepatocarcinoma side effect

 Sustained induction of the peroxisomal FAO system accounts for the massive proliferation of peroxisomes, which may contribute to hepatocarcinoma, a property 992 intrinsic to mouse and rat PPAR α but not observed in humans (25,27). By using hepatocyte-specific PPARα-deficient mice, PPARα agonist treatment was shown to 994 induce hepatomegaly via hepatocyte PPAR α activation. Interestingly, the proliferative 995 response was mediated exclusively by $PPAR\alpha$ activation in hepatocytes (264).

 Several mechanisms have been proposed to elucidate the species difference in response to peroxisome proliferators. For example, in humans, lower expression 998 levels of PPAR α or the presence of splice variants was suggested to contribute to the resistance of humans to peroxisome proliferation upon treatment with fibrate drugs (50,108). However, most of the proposed mechanisms have been met with reasonable criticism (reviewed in (265)). The work of Cheung *et al.* with "humanized" PPARα mice suggests that there are inherent species differences in the receptor that may be 1003 more critical. In this regard, PPAR α -humanized mice that express human PPAR α in a 1004 mouse PPAR α null background exhibit, unlike wild-type mice, a diminished hepatocellular proliferation while the induction of mitochondrial and peroxisomal FAO enzymes was comparable to wild-type (266). Moreover, it was proposed that the mechanism by which peroxisome proliferators lead to mitogenic and hepatocarcinogenic effects in mouse liver is due to murine PPARα-dependent 1009 downregulation of let7c microRNA, which in turn releases the repression of c-myc, an 1010 important oncogene that is related to liver cancer development. Interestingly, this 1011 effect does not occur in the humanized PPAR α mice (267), thus the induced expression of c-myc protein through species-specific regulation of a microRNA, contributes to PPARα ligand-induced liver hyperplasia in mice (267). Still, some

- questions arise regarding the fact that the PPARα-humanized mouse does not precisely model humans because there could be species differences in coactivators,
- corepressor and target gene response elements.

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VI. PPAR agonists in a clinical setting

 PPARα is widely expressed and serves to activate FAO pathways in the liver in response to fasting. This re-tunes energy metabolism to permit efficient use of mobilized lipids in the fasted state. The resulting changes may lower secretion of 1041 TGs from the liver and promote TG clearance from plasma, and thereby also reduce atherogenic lipoprotein particles. It was this activity that prompted therapeutic development to reduce cardiovascular risk in patients with raised plasma lipids. In clinical trials, a marked reduction in circulating TGs with a rise in HDL-C was 1045 consistently documented, with reductions in LDL cholesterol (LDL-C) depending on 1046 the pre-treatment TG levels (reviewed in (27)).

 By virtue of their TG-lowering capacity, the official indication for fibrates is hypertriglyceridaemia. For hypertriglyceridaemia, fibrates are the first choice when a change of diet is not sufficient. For dyslipidaemia and hypercholesterolemia, statins 1050 remain the first choice because of a higher potency in lowering LDL-C (329).

 Fibrates are generally well-tolerated drugs and have a good safety profile irrespective of concomitant therapy (330). However, changes in biochemical markers, some of which related to pathological states, may occur. Among these are elevations in homocysteine, creatinine levels, alanine aminotransferase and aspartate aminotransferase. Of note, lowering of homocysteine with folic acid did not affect cardiovascular diseases (CVD) (331,332). Along the same line, even after prolonged treatment, the increase of creatinine is rapidly reversible upon treatment stop 1058 indicating absence of irreversible tissue damage (333). Muscle-related adverse effects, such as an increase in creatine kinase and myalgia may occur but rhabdomyolysis has only been observed upon combining gemfibrozil and cerivastatin, an effect due to

 classical P450-mediated drug interaction. Rhabdomyolysis was found to be rare with 1062 (single) fibrate therapies (334).

PPAR agonists and effects on lipid and lipoprotein metabolism in humans

1065 The prominent role of PPAR α in governing lipoprotein and lipid metabolism in humans was demonstrated by the fact that fibrates, widely prescribed drugs to treat 1067 dyslipidaemia, act by binding to PPAR α and induce PPAR α -dependent gene 1068 transcription (6,335).

 A plethora of clinical studies have shown that fibrate treatment lowers serum TG concentration by about 30% (-35% P<0.01 (336); -30% P<0.045 (337); -40-20% P<0.001 (338); -37% (339); -38% (329)), reduces VLDL TG by about 40% (-41% P<0.01 (336); -44-20% P<0.001 (338)) and reduces (in combination with statins) the atherogenic sd-LDL by about 20% (-23% (329)). Additionally, HDL increases slightly (+19% P<0.02 (336); +82-106% P<0.005 (338); +14% (329)) and serum concentration of APOA1 and APOA2 increase by about 20% (+19% P<0.02 (258,336)). In obese patients with type 2 diabetes, fenofibrate decreased serum TG concentrations, while glycated haemoglobin increased (340). In contrast, the Fenofibrate Intervention and Event Lowering in Diabetes (FIELD) and Action to Control Cardiovascular Risk in Diabetes (ACCORD) studies show glucose-neutral effects of fenofibrate, supporting the notion that the first generations of fibrates are 1081 not antidiabetic drugs (330,333). In line, insulin sensitivity was not affected by fenofibrate administration (337,340). Nevertheless, yet controversial, a more recent study reported that fenofibrate may increase insulin sensitivity by inhibiting adipose tissue expression of retinol-binding-protein-4 (RBP-4), an adipocytokine (341). For LPL, in patients on gemfibrozil (6 weeks), there was no significant change in LPL in adipose tissue (342), possibly due to pharmacokinetic effects. Nevertheless, plasma 1087 TG levels decreased by 46% and several patients demonstrated increases in muscle LPL activity, though these were inconsistent and not statistically significant (342). 1089 Previous studies, using other PPAR α agonists did report a significant increase of skeletal muscle LPL activity (+39% P<0.05 (336); +50% P<0.005 (339)) and even adipose tissue LPL activity (343). It was originally proposed that gemfibrozil predominantly decreases TG levels by decreasing hepatic VLDL secretion (337,342,344). However increased lipolysis may rather be caused by a primary liver effect (e.g. APOC3 decreased and APOA5 increased). Additionally, although fenofibrate reduces plasma TG levels, it does not seem to lower concentrations or turnover rates of non-esterified FAs in adipose tissue (337), correlating with a lack of 1097 effect on insulin sensitivity.

PPAR agonists and effects on liver in humans

1100 Dyslipidaemia and insulin resistance are known risk factors for NAFLD. As $PPAR\alpha$ 1101 activation reduced hepatic steatosis in mice, applicability of fibrates was suggested as a potential treatment for NAFLD in humans (111). In a relatively small (16 patients) and phenotypically heterogeneous cohort study, patients completed 48 weeks of therapy with fenofibrate without adverse events (345). Significant decreases in TG, glucose, alkaline phosphatase and gamma-glutamyl transpeptidase, and an increase of APOA1 levels were found, yet, effects on fibrosis as addressed by liver histology 1107 were minimal (345). Similar studies with fenofibrate for 3 months confirmed results of a lowered TG yet unchanged hepatic fat content (346). Short-term treatment (2-8 weeks) with bezafibrate combined with diet and exercise of steatosis-suffering donors for liver transplantation was found to decrease macrovesicular steatosis (347),

1111 although in this particular study the impact of bezafibrate alone was not addressed. 1112 Treatment of non-alcoholic steatohepatitis (348) patients with gemfibrozil (for 4 1113 weeks) lowers ALT, AST and γ GT plasma levels (349), but not in a pilot study with 1114 clofibrate (for 12 months) (350). The relatively weak potency of these fibrates 1115 combined with lower expression level of PPAR α in human as compared to mouse 1116 liver was presented as an explanation (351). However, in all studies, plasma TG levels 1117 effectively decreased. Novel PPAR α agonists with greater potency and efficacy, such 1118 as pemafibrate, or with dual (elafibranor) or triple (lanifibranor) agonism, may prove 1119 to be more useful in the treatment of NASH/NAFLD (352). As mentioned also in 1120 section 5.5, it is important to emphasize that humans are resistant to the induction of 1121 neoplasms by fibrate drugs in contrast to rodents (25,353). This implies that following 1122 ligand activation, the PPARα-mediated pathways controlling lipid metabolism work 1123 independent from those controlling cell proliferation pathways (266).

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1125 **PPAR agonists and effects on the cardiovascular system in humans**

1126 As in rodents, PPAR α is downregulated in the failing human heart (354). PPAR α 1127 protein levels were compared between homogenates from left ventricular biopsies 1128 from five control donor hearts vs biopsies from five patients with compensated end-1129 stage heart failure at the time of transplantation (354). Whether augmenting PPAR α 1130 activity would improve the progression of heart failure is an interesting, but to date 1131 unanswered question. Sub-group analysis of the Helsinki Heart Study (HHS) trial 1132 suggested potential benefit in preventing heart failure with gemfibrozil compared to 1133 placebo (13.3% vs. 10.6%, HR 0.76; P=0.04) (355). Potential mechanisms may relate 1134 to direct cardiac and/or on PPAR α 's lipid lowering effects. PPAR α agonists may also 1135 improve cardiac performance by decreased transcription of pro-inflammatory

1136 cytokines as shown in vitro (356) and/or as in rodent studies, by reduced activation of aldosterone-mediated myocardial apoptosis (357) and/or by decreased endothelin-1- mediated cardiac fibrosis (358). Further research is needed to assess whether or not fibrates, and especially the novel SPPARMs (see further **section 6.5**) can prevent or ameliorate chronic heart failure in man.

1141 Gemfibrozil was shown to decrease the incidence of coronary heart disease in both HHS and Veterans Affairs HDL Intervention Trial (VA-HIT). Treatment was 1143 concomitant with a marked increase in HDL-C and reductions in serum levels of total, 1144 low-density lipoprotein and non-HDL-C and TGs (199,355), yet there is no definite proof that these events are the mode of action leading to a decrease incidence of 1146 coronary heart disease.

 Investigating a role for fibrates in the prevention of cardiovascular disease, a pooled meta-analysis of long-term randomized placebo-controlled clinical trials pointed out that fibrates significantly reduce the odds of nonfatal myocardial infarction, but not of cardiovascular mortality, fatal myocardial infarction or stroke, compared to placebo 1151 (359). Fibrates can reduce the risk of major cardiovascular events predominantly by prevention of coronary events, and might have a role in individuals at high risk of cardiovascular events and in those with combined dyslipidaemia (360,361). Further, in a small-scale pilot study, bezafibrate was found to induce plaque regression in thoracic and abdominal aortas with marked TG reduction and HDL-C increase (362).

Human clinical trials and clinical trial endpoints

 Primary endpoint results from the HHS and VA-HII studies as well as the post-hoc and analysis of the ACCORD and FIELD trials suggest that fibrates may benefit 1160 patients with high TG (\geq 200mg/dl) and low HDL-C (330,333). This hypothesis is

 now formally being tested in a phase 3 study with pemafibrate, called Pemafibrate to Reduce Cardiovascular OutcoMes by Reducing Triglycerides IN patiENts With 1163 diabetes (PROMINENT).

 Clinical trials have found that fibrate treatment of atherogenic dyslipidaemia reduces vascular atherosclerosis and CV risk (363). In the Bezafibrate Coronary Atherosclerosis Intervention Trial (BECAIT), bezafibrate treatment decreased angiographic evidence of coronary atherosclerosis (364,365). Moreover, in the HHS, gemfibrozil decreased cardiovascular events, especially among patients with diabetes (355). Data from the VA-HIT (366), and subgroup analyses from the Bezafibrate Infarction Prevention Trial (BIPT) (367) have demonstrated that fibrate therapy 1171 significantly reduces the risk of cardiovascular events in particular in patients with diabetes with raised TGs and low HDL-C. However, large, well-powered interventional studies resulted in disappointing outcomes. The FIELD trial saw no benefit of fenofibrate, with the intervention group showing only non-significant differences compared to the controls (330). One potential explanation for this failure was the relatively high drop-in usage of statins in the placebo group and the lower 1177 overall rates of cardiovascular events. The ACCORD study looked at addition of fenofibrate to simvastatin, but enrolled patients with little dyslipidaemia (368-371). The currently running phase 3 trial PROMINENT will test the efficacy of pemafibrate in diabetic patients with high baseline TG levels (372). Meanwhile, fibrates are 1181 mainly prescribed for the management of raised TGs in patients at risk of pancreatitis. 1182 One clinical feature noted through many of the fenofibrate studies was reduction in 1183 markers of systemic inflammation, including C-reactive protein (CRP) and IL-6. These reductions occurred even in populations lacking a significant change in lipid parameters, suggesting that the effect was direct, and not mediated through changes in

1186 lipid-mediated vascular damage (373). This phenomenon may be the physiological 1187 consequence of immune cell PPAR activation and cross-talk/inhibition of NF- κ B 1188 signaling. The combination of anti-inflammatory efficacy along with promotion of 1189 hepatic FAO suggested that PPAR α activation may prevent the progression of liver 1190 steatosis to NASH and ultimately cirrhosis. The dual PPARα/δ agonist elafibranor 1191 was tested for efficacy in patients with liver steatosis. A reduction in histological end-1192 points of acute NASH was seen, although the trial did not achieve its originally set 1193 target of NASH resolution (206). Therapeutic efficacy of elafibranor is currently 1194 assessed in the phase 3 trial RESOLVE-IT, enrolling patients with active NASH and 1195 fibrosis stage F1-3 (374).

1196

1197 **Selective PPAR modulators (SPPARMs) in humans**

1198 Overall it is surprising that there are so few therapeutic applications for fibrates, 1199 considering the important role that $PPAR\alpha$ has in regulating hepatic energy 1200 metabolism in response to the fasted state. Furthermore, although we have good 1201 evidence for cross-talk between $PPAR\alpha$ and inflammatory signaling pathways, 1202 including NF- κ B, the fibrates have not found a role in the management of chronic 1203 inflammatory diseases.

1204 In an attempt to maximize the therapeutic potential of $PPAR\alpha$ as a drug target, SPPARMα are being developed to gain greater pharmacological effect (375,376). A restricted activation potential can be explained at the molecular level by a general decrease in affinity for cofactors due to partial activation, binding to different target gene promoters and, but not limited by, the induction of a differential receptor 1209 conformation, which leads to a changed PTM status and the recruitment of a specific

1210 subset of PPAR cofactors, also according to their expression level in different organs 1211 (26,375,377).

1212 Pemafibrate is an example of a high affinity PPAR α agonist with SPPARM properties 1213 which is currently in phase 3 clinical development (203). Pemafibrate was designed to 1214 possess higher PPARα activity and selectivity over existing PPARα agonists. 1215 Pemafibrate monotherapy induced atheroprotective effects in several murine models 1216 of atherosclerosis. As a multi-level strategy compound, pemafibrate acts on 1217 inflammation, lipid metabolism and RCT (190,378). Promising results were achieved 1218 with pemafibrate for the treatment of a severe complication of type 2 diabetes, i.e. 1219 diabetic nephropathy, by targeting the pathways leading to renal oxidative stress 1220 (379). The first-line treatment for hypercholesterolaemia, a common feature of 1221 atherosclerotic cardiovascular disease (ASCVD), is statins, which lower the levels of 1222 LDL-C. However, remaining residual risks, such as elevated TG levels and decreased 1223 HDL-C, underscore the necessity for complementary therapies. Pemafibrate was 1224 found superior to fenofibrate with regard to serum TG-lowering effect and hepatic and 1225 renal safety, when used over 24 weeks in adults with dyslipidaemia (203). 1226 Consequently, in recent randomized, double-blind, placebo-controlled clinical trials in 1227 patients with dyslipidaemia, statins were combined with the SPPARM α pemafibrate, 1228 which showed a favorable benefit-to-risk ratio during phase 2 clinical trials for the 1229 treatment of combined dyslipidaemia (380,381). The compound was used as an add-1230 on therapy in combination with statin treatment to treat the residual CV risk. Next to 1231 decreasing TG levels, this drug combination targets also several other atherogenic 1232 lipoproteins, hereby improving apolipoprotein ratios and lowering LDL-C levels 1233 (380,381). In patients with type 2 diabetes with hypertriglyceridaemia, pemafibrate 1234 was also shown to ameliorate lipid abnormalities, and well tolerated (382).

 Next to SPPARMs, combining selective RXR agonists with statins can be an alternative option for the treatment of atherosclerosis. Suboptimal doses of both treatments not only minimized dose-related adverse effects, but also resulted in synergistic beneficial effects on endothelial function, compared to each drug alone. 1239 The anti-inflammatory effects were mediated in part by increased RXRα-PPARα and 1240 RXRα-PPARγ interactions (383).

1241 Finally, dual or pan-PPAR modulators with partial or selective activation profiles may 1242 be good alternatives, depending on the clinical application.

1243 Metabolic disorders such as diabetes, atherosclerosis, hyperlipidaemia and obesity, 1244 rarely occur in isolation, but usually arise in the same individuals. This complex 1245 phenotype is called the metabolic syndrome or syndrome X (4,375,384). Treatment of 1246 this aggregation of metabolic abnormalities, usually accompanied by an inflammatory 1247 context, would benefit from a more holistic approach, such as the application of a 1248 pan-PPAR agonist, to target the multiple facets of this complex disease phenotype.

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VII. Inflammation-priming and inflammation-resolving molecular actions of PPARα

1261 Evidence suggests that PPAR α can counter inflammation via multiple, distinct mechanisms (23,385,386). Both *in vitro* and *in vivo* studies identified PPARα 1263 influences on both acute and chronic inflammatory processes (154,165,387,388).

1264 The first indication of a beneficial role for $PPAR\alpha$ in suppressing inflammation came from studies on LTB4 action in PPARα*-*deficient mice. Compared to WT mice, the duration of LTB4-induced ear-swelling responses was prolonged. This anti-1267 inflammatory effect correlated with LTB₄ activation of PPAR α , thereby identifying a 1268 feedback loop through PPAR α to limit LTB4 action. These findings implied that 1269 some of the anti-inflammatory actions of PPAR α occur through up regulation of enzymes involved in ω and β-oxidation pathways and thereby accelerated degradation of signaling lipids including LTB4 (165). Subsequently, Poynter *et al.* showed that 1272 PPARα-deficient murine splenocytes had NF-κB activity, and thereby produced more IL-6 and IL-12 in response to lipopolysaccharide (LPS), supporting PPARα- dependent anti-inflammatory effects in non-metabolic tissues (131). Most recent research has focused on the anti-inflammatory actions of PPARα. Hence, the majority of the subchapters below discuss the many and diverse molecular mechanisms accounting for the anti-inflammatory effect of PPARα in various models (**Fig. 4 and Fig. 5**). However, under certain conditions PPARα induces target genes that are more consistent with a pro-inflammatory profile (see further below), suggestive of inflammation-priming (**Fig. 6)** or -sensitization, the phenomenon of sequential signals leading to cellular adaptation and affecting subsequent responses, specifically in a 1282 context of inflammation. The augmented second response may well be gene-specific 1283 and transient, as has been described for the response of synoviocytes to a repeated

 treatment with pro-inflammatory cytokines (389). These consecutive signals may encompass pro-inflammatory mediators, as in the study above, but may also combine signals of a different nature. For example, the anti-inflammatory glucocorticoid hormones transcriptionally upregulate several components of the innate immune system and thus prime pro-inflammatory responses (390-392), a circuit thought to contribute to more effective resolution of inflammation. We hypothesize similar 1290 mechanisms may also apply to $PPAR\alpha$ agonist action. Hence, an overall anti-1291 inflammatory network may also comprise a component which is "pro-inflammatory" 1292 dependent on the specific inflammatory and cellular context. More work is needed to 1293 substantiate these speculations. Data that may support this concept, however limited, are discussed in the last subsection.

Effector cells mediating the anti-inflammatory activity of PPAR

1297 The extent to which PPAR α plays a decisive role in various immune cells in humans as compared to rodents remains a point of contention. Literature and public resources 1299 such as the human immune genome project indicate that $PPAR\alpha$ expression is lower in human immune cells than in hepatocytes. This originally supported the concept that 1301 cell types like hepatocytes with high PPAR α are likely contributing to the immune regulatory effects by providing secondary signals to the immune cells, but data is 1303 accumulating on functional PPAR α effects in different immune cells. In addition, it is 1304 not inconceivable that during an inflammatory response, a change in the PPAR α expression profile of immune cells may turn a dormant factor into an active contributor to assist in blocking the inflammatory response. In support of that assumption, the well-known adrenal stress hormones GCs enhance PPARα transcript 1308 levels in T and B lymphocytes (130), at least in the mouse.

1309 PPARα is not only expressed in undifferentiated human monocytes but also in 1310 differentiated human monocyte-derived macrophages and is transcriptionally active 1311 upon ligand binding. In TNF and IFN- γ activated macrophages PPAR α can induce 1312 apoptosis (32). All three PPAR subtypes play important roles in human macrophages. 1313 For example, the anti-inflammatory properties of PPAR γ have been connected to its 1314 ability as a driver of M2 polarization in human atherosclerosis (393).

1315 Brocker *et al.* recently compared hepatocyte- to macrophage-specific PPARα null 1316 mice and found that the downregulation of the inflammatory cytokines IL-15 and IL-1317 18 depended on macrophage PPAR α , identifying Küpffer cell PPAR α as an important 1318 mediator of the anti-inflammatory effects of PPAR α agonists (264). Functional human 1319 PPAR α was demonstrated following a global gene expression profiling experiment on 1320 human primary macrophages treated with specific PPAR α agonists (263). The role of 1321 PPARs, fatty acid signaling, and fatty acid oxidation within primary human immune 1322 cell types in a variety of inflammatory conditions requires further experimental study. 1323 Linking inflammation to metabolism, murine macrophage LXR is critical for PPAR α 1324 to promote cholesterol efflux and reverse cholesterol transport (RCT) *in vivo*, 1325 contributing to anti-atherogenic effects and hence indirectly resolving inflammatory 1326 responses within the blood vessels. In support, PPARα activation was demonstrated to 1327 inhibit the development of atherosclerosis by promoting macrophage RCT in 1328 humanized hypercholesterolemic mice expressing the human APOA1 gene (394).

1329 Connecting metabolic and immune responses to cancer, in murine and human 1330 melanoma models, an increase in free FA species during tumor progression was 1331 shown to activate PPAR α signaling in CD8⁺ tumor-infiltrating lymphocytes. 1332 Consequently, CD8⁺ tumor-infiltrating lymphocytes switched toward FA catabolism 1333 to preserve their effector functions. Promoting FA catabolism with fenofibrate

1334 markedly improved the capacity of $CD8⁺$ tumor-infiltrating lymphocytes to delay 1335 tumor growth, via a synergistic mechanism involving a Programmed cell Death 1336 protein-1 (PD-1) blockade to efficiently boost the efficacy of melanoma 1337 immunotherapy (395). Together, these data highlight the interplay between the 1338 metabolic programs of T lymphocytes and tumor cells, with potential impact on 1339 melanoma immunotherapy (396). Ligand activation of PPAR α also antagonizes NF-1340 κB in murine T and B lymphocytes, and conversely, T cell activation results in 1341 reduced PPAR α expression. Of note, treatment of T lymphocytes with specific 1342 PPAR α activators can upregulate the expression of endogenous PPAR α -controlled 1343 genes in combination with histone deacetylase inhibitors. The presence of functional 1344 PPARα within T and B lymphocytes again emphasizes a role for this nuclear receptor 1345 in cells of the immune system (130). In addition to targeting NF- κ B, PPAR α also 1346 counters proinflammatory signaling in T lymphocytes at the level of IL-2 and c-Jun 1347 activation (397). In evidence, genetic ablation of the PPAR α gene resulted in the loss 1348 of antagonism of NF-κB, increased production of Th1 and decreased production of 1349 Th2 cytokines by T cells (397). In another study, the PPAR α agonist Wy-14,643 was 1350 shown to increase IL-4 secretion and to inhibit IFN-γ expression in murine mitogen-1351 activated splenocytes, and to reduce the proliferation of human T cell lines (398). 1352 Similarly, stimulation of isolated CD4-positive human T cells with PPAR α agonists at 1353 clinically relevant concentrations decreased IFN- γ , TNF α and IL-2 production. 1354 However, in this study no effect on viability was noted (399).

1355 Epidermal Langerhans cells (LCs) play a pivotal role in initiating and maintaining 1356 primary immune responses in the skin. PPAR α is expressed in immature LC but 1357 downregulated in mature LC. Pharmacological PPAR α activation inhibits LC 1358 maturation, migratory capacity, cytokine expression, and the ability to drive T-cell

 proliferation. Moreover, PPARα in LC also inhibits NF-*κ*B activity. Taken together, 1360 PPAR α activation by endogenous ligands may provide a molecular signal that allows 1361 LC to remain in an immature state (400). The activation of PPARs by drugs, and 1362 products of intermediary energy metabolism within the diverse cells of the immune system may reveal new opportunities to target inflammatory process or to advance 1364 adjunctive therapies to enhance anti-cancer treatment.

Interactions between PPARα and other transcription factors

1367 The best-known mechanism by which PPAR α (as well as the other PPAR subtypes) inhibits many inflammatory genes is transrepression. In this tethering mechanism, PPAR α acts directly to negatively regulate the expression of pro-inflammatory genes by antagonizing the activities of other regulators of inflammation such as members of 1371 the NF-κB, AP-1 and STAT families, as also already alluded to in the section above (27) (**Fig. 4A**).

 In this regard, it was found that fibrate-activated PPARα inhibits IL-1-stimulated secretion of IL-6 and 6-keto-prostaglandin by decreasing IL-6 and COX-2 gene 1375 transcription in smooth-muscle cells (SMCs). Inhibition of COX-2 induction occurred 1376 transcriptionally as a result of PPAR α interference with NF- κ B signaling (23). This 1377 effect of PPAR α in the vascular wall may influence the progress of atherosclerosis. Accordingly, Delerive *et al.* have demonstrated that aortas from PPARα-null mice display an exacerbated inflammatory response to LPS, as assessed by enhanced IL-6 secretion, and that fibrate treatment represses IL-6 mRNA levels in LPS-stimulated aortas of PPARα wild-type, but not of PPARα-null mice, confirming that the observed 1382 anti-inflammatory properties of fibrates are PPARα-dependent. This vascular anti-inflammatory response is a result of PPARα interference with NF-κB and AP-1

1384 transactivation capacity involving direct protein-protein interaction with p65 and c-1385 Jun. Of note, a bidirectional antagonism between the PPAR α and NF- κ B signaling 1386 pathway exists (387).

1387 In human vascular endothelial cells, $PPAR\alpha$ inhibits the thrombin-mediated activation of endothelin-1, a vasoactive peptide. This inhibition occurs at the transcriptional level and occurs via interference with the AP-1 signaling pathway by preventing the binding of AP-1 proteins to their target sequence (401). Consistently, PPARα 1391 activators prevent $TNF\alpha$ -induced vascular cell adhesion molecule 1 (VCAM-1) expression in human endothelial cells, partly through inhibition of the NF-κB pathway. This PPARα action, with a consequent decrease in monocyte adherence to endothelial cells, may have important implications on early stage of atherosclerosis (402) . PPAR α agonists decrease cytokine-induced tissue factor in monocytes most likely by interfering negatively with AP-1 and/or NF-κB activity (403,404). Because these pathways are involved in the activation of inflammatory response genes 1398 implicated in atherogenesis, PPAR α activation may be beneficial in combatting the pathogenesis of atherosclerosis. In addition, oxidized eicosapentaenoic acid (EPA) attenuates monocyte chemotactic protein 1 (MCP-1) and IL-8 expression in 1401 endothelial cells, by inhibiting NF- κ B in a PPAR α -dependent manner (405).

 The transcription factor, specificity protein 1 (Sp1), is also involved in the anti- inflammatory mechanism of PPARα. PPARα activation inhibits vascular endothelial growth factor receptor-2 (VEGFR-2) in endothelial cells, which occur at the transcriptional level via decreasing Sp1 site-dependent binding to the promoter 1406 sequence through an increased interaction with the Sp1 transcription factor (406). 1407 Thus, PPAR α activators may control inflammatory responses in part by targeting

1408 endothelial VEGFR2 signaling, which is critical for angiogenic responses during 1409 chronic inflammation.

1410 Further work has demonstrated that the mechanism of PPAR interference with NF-κB 1411 action additionally involves a PPARα-mediated induction of the inhibitory protein 1412 IkBa expression. Interestingly, fibrates increase IkBa expression in primary smooth 1413 muscle cells and hepatocytes, thus preventing nuclear p50/p65 NF-κB translocation 1414 and thereby arresting p65-mediated gene activation, to oppose development of the 1415 inflammatory response. PPARα-mediated IκBα upregulation did not require a 1416 functional PPRE in the I κ B α promoter, but rather involved functional Sp1 and NF- κ B 1417 sites, in a cross-talk mechanism (407). Cytosolic retention of p50/p65 NF-κB extends 1418 the list of candidate anti-inflammatory $PPAR\alpha$ mechanisms from those mainly 1419 characterized by direct binding and inactivation of transcription factors, such as p65/ 1420 NF-KB and c-Jun (23,387). In further, detailed molecular studies fibrates were found 1421 to decrease basal and IL-6-stimulated expression of the human fibrinogen gene in 1422 human primary hepatocytes and hepatoma HepG2 cells by interfering with the pro-1423 inflammatory transcription factor CAATT/enhancer-binding protein β (C/EBP-β) via 1424 sequestration of the coactivator glucocorticoid receptor-interacting protein 1425 1/transcriptional intermediary factor 2 (GRIP1/TIF2) (408) (**Fig. 4A**). This anti-1426 inflammatory action of PPAR α is not restricted to fibrinogen but also applies to other 1427 acute phase genes containing a C/EBP response element in their promoters (408). 1428 In addition, fibrate-activated PPARα negatively regulates IL-1-stimulated CRP gene

1429 expression in isolated primary human hepatocytes by two mechanisms, the inhibition 1430 of p50-NF- κ B translocation and the down-regulation of basal p50-NF- κ B and C/EBP-1431 β expression in a PPARα-dependent fashion (409). Such observations reveal a direct 1432 contribution of PPAR α in controlling the inflammatory gene response in the liver.

 Furthermore, chronic PPARα agonist treatment inhibited the expression of IL-6 induced acute phase response (APR) gene expression *in vivo*. This suppression occurs transcriptionally due to the down-regulation of the IL-6 receptor components gp80 and gp130 in the liver, thereby reducing the phosphorylation and activation of the 1437 downstream transcription factors STAT3 and c-Jun that transduce the IL-6 signal; and to the reduction of the basal expression of the transcription factors CCAAT enhancer- binding protein-α, -β, -δ (410). Related to STAT signaling, Van der Meer *et al*. 1440 proposed that activated PPAR α may downregulate gene expression in HepG2 cells by 1441 causing the loss of STAT1 and STAT3 binding to DNA. Therefore, interfering with 1442 STAT-dependent transcriptional activity, which has been previously well documented for other PPAR subtypes, may also contribute to the anti-inflammatory action of PPARα activation (72) (**Fig. 4A**). Inhibitory effects of PPARα agonists on IL-1- and IL-6-induced expression of APR proteins including CRP, serum amyloid A (SAA), fibrinogen and haptoglobin, were also observed in mice displaying liver-restricted 1447 expression of PPAR α (411).

 PPAR α reduced the stimulation of the APR mediated by LPS and prevented the LPS-1449 induced increase of proinflammatory mediators IL-1, IL-6 and TNF in plasma via a liver-specific transcriptional mechanism. This resulted in decreased expression of the 1451 adhesion molecules, intercellular adhesion molecule 1 (ICAM-1) and VCAM-1 in the murine aorta, suggesting that PPARα can control the systemic and vascular inflammatory response via a liver-specific action (411). Importantly, this implies that 1454 the spectrum of PPAR α -controlled genes may be vastly larger in a more physiological 1455 context than in isolated cells (412). Further evidence of this comes from analysis in human liver slices where there was suppression of inflammatory genes, but not in

1457 primary hepatocytes. The most highly repressed genes included chemokines, IFN- γ -1458 induced genes and other immune-related genes (412).

1459 Fibrate activation of PPARα inhibits hepatic inflammatory responses, which 1460 unimpeded lead to liver injury and progression to fibrosis, by a mechanism relying on 1461 transrepressive activity, rather than binding to cognate PPREs and activation of lipid 1462 metabolism genes (388). These findings indicate that $PPAR\alpha$ agonists with 1463 dissociated activity, selectively modulating $PPAR\alpha$ transrepression, might have the 1464 potential to prevent the progression of NASH to liver fibrosis through a direct 1465 counteraction of inflammatory responses independent of effects on intrahepatic lipid 1466 accumulation. In the same line, a recent report has shown that PPAR α gene and 1467 protein expression are decreased in NASH (413). The PPARα levels were restored by 1468 pharmacological inhibition or genetic suppression of miRNA-21, a micro-RNA 1469 upregulated in the liver of patients with NASH, decreasing thereby liver injury, 1470 inflammation and fibrosis (413). These findings suggest that inhibition of the miRNA-1471 21 pathway might be an interesting new therapeutic strategy to combine with some of 1472 the current PPARα pharmacological agonists in NASH (413).

 A PPRE-dependent mechanism of PPARα-NF-κB interference in the regulation of complement C3 transcription has been observed in a human hepatoma cell line, 1475 identifying transactivation as another mechanism for the anti-inflammatory effects of PPARα (**Fig. 2B**). Ligand-dependent activation of PPARα abolishes TNFα-mediated upregulation of complement C3 through a physical interaction of PPRE-bound 1478 PPAR α with p65 within the C3 promoter (414).

1479 Finally, the transcription factor Sp1 is also involved in the anti-inflammatory 1480 mechanism of PPAR α . PPAR α activation inhibits vascular endothelial growth factor 1481 receptor-2 (VEGFR-2) in endothelial cells, which occur at the transcriptional level via 1482 decreasing Sp1 site-dependent binding to the promoter sequence through an increased

1483 interaction with the Sp1 transcription factor (406).

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1485 **Cooperation between PPARα and other anti-inflammatory nuclear receptors**

1486 Simultaneous activation of PPAR α and GR α , a well-characterized NF- κ B repressor, 1487 enhances transrepression of NF- κ B-driven gene expression and additively represses 1488 proinflammatory cytokine production, including MCP-1 and IL-6 in various cell lines 1489 (385) (**Fig. 4B**). This finding offers a rationale for the control of inflammation by 1490 GR α and PPAR α by inflammatory gene repression and suggests new approaches for 1491 the treatment of inflammatory diseases where combinations of PPAR α and GR α 1492 activation could offer a more effective anti-inflammatory intervention.

1493 This phenomenon is not restricted to $PPAR\alpha$ since a similar proof of concept was 1494 recently also shown for PPAR γ agonists in skin and lung. Indeed, house-dust mite 1495 induced skin sensitization, which aggravates asthmatic responses in mice via a mixed 1496 Th2/Th17 response, was prevented by treatment combining glucocorticoids with 1497 rosiglitazone (415). Similarly, the role of $PPAR\alpha$ in estrogen-mediated anti-1498 inflammatory activity in lungs was shown by the demonstration of a weakened 1499 response to estrogen in PPARα-deficient mice (416).

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1501 **Regulation of MAPK and PKC kinase activities**

1502 Given that inflammatory responses are driven in part by the activation of MAPKs,

1503 PPARα ligands were also shown to modulate the degree of macrophage inflammation

- 1504 not only by reducing NF-κB activation, but also by reducing MAPK phosphorylation,
- 1505 hence effectively decreasing the expression of the proinflammatory mediator COX-2
- 1506 and inducible nitric oxide synthase (163,417).

1507 PPARα was further shown to polarize T-cell differentiation processes, also via a 1508 modulation of MAPK activities. In CD4(+) T lymphocytes, unliganded PPAR α 1509 suppressed p38 MAPK phosphorylation. Ligand activation reversed this inhibition, 1510 augmenting the expression of the transcription factor of T cells (T-bet), a lineage-1511 determinant of Th1 inflammatory responses (418) which drives IFN-γ expression. 1512 PPAR α thus reduced the level of activation-induced IFN- γ produced by CD4⁺ T cells. 1513 Mechanistically, the control of PPARα over T-bet expression occurs via a DNA-1514 binding independent mechanism, mediated through the ability of PPAR α to repress 1515 the phosphorylation of p38-MAP kinase following T cell activation. These data 1516 suggested a novel and important function for PPARα within T cells. Both in 1517 transformed T and B lymphocytes, PPARα ligands effectively repress NF-κB activity 1518 (130). Additionally, the anti-inflammatory effects of PPAR α in macrophages involve 1519 an inhibition of the PKC signaling pathway (52,154). Little is known about the exact 1520 molecular mechanisms underlying the PPARα-mediated modulation of (potentially 1521 other) kinase activities and to what extent this may contribute to an overall anti-1522 inflammatory action.

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1524 **Anti-inflammatory activity of PPARα in a PPRE-dependent manner**

1525 In addition to down-regulating the expression of pro-inflammatory genes, $PPAR\alpha$ also suppresses the inflammatory response by direct up-regulation of genes with anti- inflammatory properties, suggesting a possible cooperation between PPARα- dependent transactivation and transrepression to govern anti-inflammatory pathways. Besides the upregulation of IκBα, discussed above, as a mechanism contributing to its anti-inflammatory actions (409,419), activated PPARα strongly up-regulated the hepatic expression of IL-1 receptor antagonist (IL-1ra), an inhibitor of cytokine signaling, during LPS-induced inflammation, while numerous inflammatory genes including STAT3, SAA and IL-18 were found to be down-regulated (420). Transactivation and chromatin immunoprecipitation studies identified IL-1ra as a 1535 direct positive target gene of PPAR α with a functional PPRE present in the promoter. 1536 Moreover, up-regulation of IL-1ra by PPARα was conserved in human HepG2 hepatoma cells and the human monocyte/macrophage THP-1 cell line (420).

 In the same context, Heme Oxygenase-1 (HO-1), a PPARα target gene with a 1539 functional PPRE, is also directly upregulated by $PPAR\alpha$ ligands and contributes to the 1540 anti-inflammatory effects in human vascular cells (421).

1541 Interestingly, the gene coding for vanin-1 (Vnn1), a protein with anti-inflammatory 1542 potential, is robustly activated in liver by PPAR α agonists, such as Wy-14,643, fenofibrate, and clofibrate (211), and functional PPRE sites have been identified in the Vnn1 promoter (422). Vanin activity prevents steatosis development in response to 1545 fasting, while its deficiency affects multiple genes involved in inflammation/immunity (423).

 Considering the above studies, it is clear that PPARα may affect markers of inflammation in a direct or indirect manner. Below, we continue the discussion on recent findings regarding the role of PPARs in some specific examples of inflammatory disease models.

PPARα in the metabolic syndrome as an inflammatory disease

 Inflammation is a critical component of the metabolic syndrome, which represents a cluster of risk factors that include dyslipidemia, insulin resistance, hypertension, 1555 inflammation and coagulation disorders. Consequently, PPAR α , being on the crossroad between inflammation and metabolism, is an important regulator of the

 metabolic syndrome (27,424). It is now well-accepted that pro-inflammatory cytokines and specific immune cell populations promote metabolic disease progression. Accordingly, a chronic low-grade inflammatory condition, which is characterized by increased circulating inflammatory cytokines and acute phase 1561 proteins and which often accompanies the metabolic syndrome, has been implicated as a major factor both in the establishment of the metabolic syndrome and its 1563 associated pathophysiological consequences (207). PPAR α , and other PPAR isoforms, have all demonstrated anti-inflammatory effects in these conditions (425). 1565 In hyperlipidemic patients and patients with metabolic syndrome PPAR α activation 1566 using fenofibrate decreases plasma levels of several acute phase and inflammatory 1567 proteins that are normally increased during inflammatory conditions, such as high-1568 sensitivity CRP and IL-6 (23,373,410). In addition, fenofibrate treatment also reduces 1569 plasma concentrations of the cytokines interferon-γ (IFN- γ) and TNF α in hyperlipoproteinemia type IIb patients (426). Addition of fenofibrate to simvastatin- treated patients with atherosclerosis and early glucose metabolism disturbances 1572 further reduces lymphocyte release of IL-2, IFN- γ and TNF- α , accompanied by a reduction in plasma CRP levels, compared to statin alone (427). It was also found that fibrates have an anti-inflammatory effect in the interaction between adipocytes and macrophages in obese adipose tissue, thereby improving the dysregulated adipocytokine production and obesity-related metabolic syndrome (428).

PPARα in age-related inflammation

 Enhancing PPARα levels and activity was shown to ameliorate age-related inflammatory processes. Increased inflammatory marker levels during aging correlate 1581 with a decreased PPAR α activity. Administration of PPAR α activators improved the splenocyte cellular redox balance and reduced the age-associated elevations in NF-κB activity and constitutive pro-inflammatory cytokine production (131,429). The PPAR pan agonist MHY2013 alleviated age-related hepatic lipid accumulation, in part by upregulating β-oxidation signaling and suppressing inflammation in the liver (430). In a similar context, a recent study has demonstrated that LPS-induced liver 1587 inflammation and lipid accumulation were associated with a chronically overactive inflammasome/IL-1β pathway in aged rat livers and that these effects were due to the 1589 dysregulation of PPAR α and SREBP1c. Indeed, in these aged rat livers, PPAR α was consistently inactivated and the levels of SREBP1c were chronically increased (132). 1591 Therefore, targeting PPAR α signaling might reduce the age-related sensitivity to endotoxin-induced liver inflammation.

1593 These data, positioning PPAR α at the crossroad of aerobic metabolism and inflammation, fit the observations that link non-aerobic glycolytic ATP generation to the production of pro-inflammatory cytokines (80). To illustrate, complex host responses to the TLR9 agonist CpGA were found to be mediated by plasmacytoid dendritic cell (pDC) production of IFNs, operating in an autocrine fashion. FAO was 1598 necessary for full pDC activation, pointing to PPAR α as a potential target to modulate downstream effects of type I IFNs (431).

PPARα in inflammatory bowel disease

 Considering the intestine as an organ contributing to immune system, a link between PPARα signaling and the control of colitis was found upon studying the IL-10 gene deficient mouse as a model for human inflammatory bowel disease. In this model 1605 anti-inflammatory IL-10 gene expression changes were related to PPAR α activation although the mechanisms responsible remain undefined (432). Infectious disease control in intestine also involves PPARα activation. As such, clearance of a Giardia 1608 muris infection was found to coincide not only with IL-17 production, contributing to 1609 protection, but also with an early PPAR α induction (433). Along similar lines, 1610 fenofibrate represses IL-17 and IFN- γ expression and improves colitis in IL-10-1611 deficient animals, suggesting a possible therapeutic potential in Crohn's disease (434). 1612 Of note, PPARα protects the intestine from colitis-induced permeability (435)

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1614 **PPARα in inflammatory skin disease**

 PPARα activation impaired epidermal Langerhans Cell (LCs) maturation and 1616 migratory capacity and alters cytokine production through the inhibition of NF-κB 1617 phosphorylation (400), suggesting a role for PPAR α in combatting inflammatory skin diseases (436). In this regard, PPARα-deficient mice exhibit impaired recruitment of neutrophils and monocytes/macrophages to the wound bed and present a transient delay in healing, which coincides with the inflammatory phase (437). These data support the concept that PPARα favors skin healing via a modulation of the 1622 inflammatory phase (437). In addition, PPAR α activation is beneficial in mouse models with a hyperproliferative epidermis (438) and in models of irritant and allergic 1624 dermatitis (439). The anti-inflammatory properties of PPAR α agonists, coupled with their anti-proliferative and pro-differentiating effects, suggest that they could be beneficial for the treatment of a variety of cutaneous diseases (439). In a model of 1627 atopic dermatitis, topical application of the PPAR α agonist WY14643 significantly decreased antigen-induced skin inflammation, providing *in vivo* evidence for the role 1629 of PPAR α in controlling skin inflammation and pointing to a potential benefit in combating systemic inflammatory responses in patients with atopic dermatitis, a disease characterized by inflammatory infiltrates and barrier dysfunction (440). 1632 Interestingly, PPAR α may be the molecular mediator of the anti-allergic and anti1633 inflammatory effects of palmitoylethanolamide, a natural FA derivative present in 1634 murine skin (441). Taken together, the many observations above underscore the 1635 implications of PPAR α in inflammatory skin disorders and paves the way for trials of 1636 topical PPAR α agonist application.

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1638 **PPARα in other inflammatory diseases**

1639 Expanding the applicability of PPARα agonists to other inflammatory diseases, anti-1640 inflammatory properties of activated PPARα were observed in primary murine 1641 astrocytes, a cell type implicated in the pathology of experimental autoimmune 1642 encephalomyelitis (EAE), an animal model of multiple sclerosis (MS). Fibrates 1643 decreased the LPS-induced secretion of pro-inflammatory cytokines TNF α , IL-1 β and 1644 IL-6 and NO production, likely via interference with the DNA binding of NF-κB 1645 (442) , suggesting a putative function of PPAR α in controlling neurodegenerative 1646 diseases. In the same line, PPAR α activation significantly inhibited radiation-induced 1647 microglial production of pro-inflammatory cytokines such as TNFα, IL-1β, Cox-2 and 1648 iNOS by inhibiting nuclear translocation of the NF-κB/p65 subunit and reducing 1649 phosphorylation of nuclear c-Jun/AP-1 (443). Genetic ablation of PPAR α in an 1650 experimental model of autoimmune encephalomyelitis increased clinical symptoms in 1651 male but not female SV.129 H^{2b} mice, consistent with the finding that male mice of 1652 this strain express higher levels of PPAR α than female mice (397). Recently, PPAR α 1653 was reported to contribute to the anti-inflammatory activity of melatonin in Spinal 1654 Cord Injury (SCI), again by modulating the NF-κB pathway. It was demonstrated that 1655 melatonin was less effective in PPAR α null, compared to WT mice, as evaluated by 1656 inhibition of the degree of spinal cord inflammation and tissue injury, neutrophil

 infiltration, pro-inflammatory cytokine expression, NF-κB activation and iNOS expression (444).

The paradox of PPARα as a pro-inflammatory mediator

1661 The above studies (**Fig. 5**) are in line with the well-accepted notion that PPAR α is involved primarily in anti-inflammatory signaling. As stated in the introduction to this chapter, PPARα has also been ascribed potential "pro-inflammatory" effects (**Fig. 6**). Lee *et al.* suggested that oxidized phospholipids induce MCP-1 and IL-8 expression through a PPARα-dependent signaling cascade in human aortic endothelial cells (445) . These data demonstrate that PPAR α can mediate the activation of endothelial cells by stimulating the production of monocyte chemotactic activity in response to oxidized phospholipids and lipoproteins (445)

 Additionally, PPARα activation by Wy-14,643 enhances IL-1β-induced pro- inflammatory cytokine release, including IL-6, IL-8 and TNF-α, in ocular cells and induces proangiogenic responses (446). In contrast, in the FIELD and ACCORD 1672 studies (330,333) fibrates showed a protection against diabetic retinopathy. In line, the antiangiogenic activity of PPARα ligands specifically depended on the activation of PPARα since PPARα ligands inhibited FGF2-corneal neovascularization in wild-type 1675 but not in PPAR α deficient mice (447). Consistent with these data fenofibrate was shown to improve diabetic retinopathy in type 1 diabetes animal models and ischemia-induced retinal neovascularization. Herewith, PPARα-dependency also formed the mechanistic basis for the therapeutic effects of fenofibrate (448). The anti-1679 inflammatory effects of fenofibrate-activated PPAR α were ascribed to the inhibition of NF-κB signaling in the diabetic retina (448). The basis for these differences

 remains unclear but might reflect a reduced complexity of *in vitro* studies versus *in vivo* studies.

 Then again, in a mouse model of endotoxemia, fenofibrate administration strongly increased circulating LPS-induced TNF levels (449). In a rat hypertensive model, bezafibrate was shown to sensitize for endotoxin effects, concomitant with low levels 1686 of HDL and increased levels of TG (450). In murine macrophage lines and in contrast to Wy-14,643, the natural PPARα ligands LTB4 and 8(S)-HETE showed a pro- inflammatory potential by increasing NOS activity (348). These observations are again not restricted to PPARα, since PPARγ activity in T-cells and dendritic cells promotes Th2 immunity (451,452). These are interesting findings suggesting that PPAR α activation may result in different responses depending on the inflammatory settings, an effect which may involve combinatorial signaling. Although the mechanisms by which such potentially "pro-inflammatory" effects occur is not understood, these data point to the need for a careful evaluation of the effects of PPARα agonists in the relevant cell types, and in different physiological contexts. 1696 How to reconcile these seemingly contradictory findings with the wealth of data on 1697 anti-inflammatory PPAR actions is as of yet unclear. For instance, reduced PPARα expression has also been linked to a lower overall survival and an enhanced tissue 1699 bacterial load in sepsis (453). The timing and nature of the pro-inflammatory stimulus and/or treatment may play a decisive role. It may well be that PPARα exerts a 1701 context-dependent immune-system priming role, as has been documented recently 1702 also for glucocorticoids (391). Given that GR and PPAR α are able to physically interact and able to co-control genes involved in both inflammatory and metabolic pathways (160,385), increases the likelihood of this assumption. More research is clearly necessary to disentangle and explain these paradoxical observations. This

- 1706 hitherto hidden role of PPAR α not only merits further mechanistic attention but also
- requires a better understanding at the level of physiological relevance.
1708 **VIII. Future perspectives**

1709 On account of their pleiotropic effects, PPARs may be targets in a number of disease 1710 conditions. It is clear there is still a lot of ground to cover, foremost by studying novel 1711 generations of more potent and better tolerated single, dual or pan PPAR agonists in 1712 the clinic. More basic research involving novel $PPAR\alpha$ agonists is important, as many 1713 mechanistic aspects differ from the first generation of $PPAR\alpha$ agonists. Plus, novel 1714 fields of application may still arise, for which the first generation of drugs did not 1715 show therapeutic benefit.

1716 Fibrates might be beneficial in the prevention of coronary heart disease. The highest 1717 benefit is achieved in cardiovascular disease patients with high TG and low HDL-C 1718 (454). A clinical benefit of PPAR α activation with novel agonists remains to be 1719 demonstrated in patients with high TG/low HDL and residual cardiovascular risk. 1720 Fibrates might be useful in the treatment and prevention of atherosclerosis, especially 1721 the new fibrates with greater potency open new perspectives. However, it remains 1722 unclear whether fibrates in general are useful in the treatment or prevention of heart 1723 failure.

1724 It is now evident that the effects of $PPAR\alpha$ are much more widespread and cover 1725 numerous aspects of inflammation and inflammatory related diseases, however many 1726 mechanistic fundamental details of its function still remain to be unraveled. For 1727 instance, nuclear receptor regulation of circadian rhythms in inflammatory metabolic 1728 diseases deserves more clinical investigation. There is newfound appreciation for the 1729 timing of treatment interventions. For example, a recent study shows a link between 1730 the circadian clock and perioperative myocardial injury in patients undergoing aortic 1731 valve replacement, whereby antagonism of Rev-erbα, another nuclear receptor family 1732 member and PPAR α target gene (260) seems to provide a benefit for cardioprotection

 (455). Moreover, the circadian clock regulates the expression and activity of genes 1734 and proteins involved in the progression of several autoimmune diseases (456,457). Future therapeutics relying on PPARs in a context of inflammation will therefore 1736 most probably have to take "the clock" into consideration. 1737 The role of PPAR α needs to be further evaluated in various pathophysiological

1738 conditions. For instance, it is unclear how $PPAR\alpha$ may influence immune cell (metabolic) control in immune/inflammatory diseases that are glucocorticoid resistant, 1740 such as sepsis.

1741 Defining the molecular mechanisms of PPAR α function that govern inflammatory programs will improve our understanding of its roles and will allow us to further exploit their potential in the therapeutic field.

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 Figure 1: Schematic presentation of the human PPAR gene and protein domain structure-function relationship. (A) The human PPAR α gene contains at least 8 3526 exons. Exons are shown in boxes with translated regions shaded. 5' UTR encoded by exons 1, 2 and the 5′-end of exon 3. The coding DNA sequence spans the 3′ region of exon 3, exons 4–7 and the 5′ extremity of exon 8. The 3′ UTR consists of the last 232 3529 base pairs of exon 8. The splicing events S_{wt} and S_{tr} generating hPPAR α_{wt} and hPPAR α_{tr} transcripts respectively are depicted. The variant transcript lacks exon 6 resulting in a frame shift generating a premature TGA termination codon. The 3532 corresponding protein products of the two splice isoforms are underlined. (B) PPAR α contains four distinct domains. The amino-terminal A/B domain contains the ligand- independent activation function-1 (AF-1), the C domain consists of a highly conserved DNA-binding domain (DBD), the D domain consists of a variable hinge region and the C terminal E domain includes ligand-binding domain (LBD) and a ligand-dependent activation function-2 (AF-2). Per domain, main functions are listed. Hsp90, Heat shock protein 90; PPRE, PPAR response element; UTR, untranslated region; wt, wild type*;* tr*,* truncated.

 Figure 2: PPRE-dependent transcription regulation. (A) PPRE-dependent activation. In the absence of specific ligands, PPARα/RXR heterodimers bind to the PPRE located in the promoter of target genes, or elsewhere in the genome, and recruit corepressor complexes thereby maintaining PPARα target genes in a silent state (ligand-independent repression). Upon ligand activation, the nuclear corepressors are 3545 released and the coactivator complexes are then recruited and bound to $PPAR\alpha/RXRa$ heterodimers, hence inducing expression of target genes involved in several biological processes including glucose and lipid metabolism. (B) PPRE-dependent inhibition of NF-κB transcriptional activity. Upon ligand activation, PPRE-bound PPARα directly

 interferes with the activity of p65 antagonizing thereby its binding to an NF-κB-RE in the complement promoter. Sirt1; Silent information regulator 1; PPRE, PPAR- response element; RXR, Retinoid X receptor; DR1, Direct repeat1; NF-κB-RE, NF-κB response element.

Figure 3: Levels of modulating PPAR α *activity.* PPAR α activity is controlled at several levels including gene expression, protein translation, ligand selectivity, posttranslational modifications affecting receptor stability and cofactor recruitment. Ub, Ubiquitinated; S, SUMOylated; P, Phosphorylated; PPRE, PPAR response element; GSK3, glycogen synthase kinase 3; AMPK, AMP kinase; PKC, protein kinase A; PKA, protein kinase A; PIASy, Protein inhibitor of activated STATy; RXR, retinoid X receptor.

 Figure 4: Models of PPARα transrepression activity. (A) PPARα directly interacts 3561 with pro-inflammatory transcription factors such as NF- κ B, AP-1 and STAT to negatively regulate their targets leading to anti-inflammatory effects through a DNA-3563 independent mechanism. (B) Simultaneous ligand-activation of PPAR α and GR leads to the enhanced repression of TNF-induced IL-6 transcriptional activity, likely via a 3565 mechanism involving a direct physical interference between PPAR α and GR. Ub, Ubiquitinated; S, SUMOylated; P, Phosphorylated; PPRE, PPAR response

element; GSK3, glycogen synthase kinase 3; AMPK, AMP kinase; PKC, protein

kinase A; PKA, protein kinase A; PIASy, Protein inhibitor of activated STATy; RXR,

retinoid X receptor; C/EBP-β, CAATT/enhancer-binding protein β; STAT, signal

 transducer and activator of transcription; AP-1, activator protein-1; NF-κB, nuclear factor-κB.

 Figure 5: Anti-inflammatory effects of PPARα activation. (A) Effect of human 3573 PPAR α activation, by various synthetical ligands on inflammatory mediators. (B)

 Effect of rodent PPARα on inflammatory mediators. Note that for some of these effects, conclusions on an involvement of PPARα were reached by using PPARα -/- mice, in which an aggravated inflammatory effect was observed. hASMCs, human aortic smooth muscle cells; hUVEC, human vascular endothelial cells; COX-2, cyclooxygenase; PG, prostaglandin; VCAM, vascular cell adhesion molecule; MCP- 1, Monocyte chemotactic protein-1; VEGFR2, vascular endothelial growth factor 3580 receptor-2; CRP, C-reactive protein; IFN- γ , interferon- γ ; APP, acute phase response proteins; LPS, lipopolysaccharide; SAA, serum amyloid A; IL-1ra, IL-1 receptor antagonist; LTB4, Leukotriene B4; iNOS, inducible nitric oxide synthase. *Figure 6: Stimulatory effects of human and murine PPARα activation on inflammatory mediators*. Note here that mostly natural ligands seem able to trigger PPARα-dependent pro- inflammatory effects. MCP-1, Monocyte chemotactic protein- 1; LPS, lipopolysaccharide; 8(*S*)-HETE, 8(*S*)- hydroxyeicosatetraenoic acid; LTB4, Leukotriene B4; NOS, nitric oxide synthase.

Table 1: PPARα functional domains and their relative functions. PPARα displays a classical nuclear receptor canonical structure. Per domain, main PPARα functions are listed (details in the text). Hsp90, Heat shock protein 90; PPRE, PPAR response element.

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Table 2: Phosphorylation modifications affecting PPARa activity. PPARa undergoes several phosphorylation modifications that markedly impact receptor function (details in the text).

Table 3: SUMOylation modifications affecting PPARα activity

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Table 4: PPARα Natural and Synthetic Ligands

Table 4: The natural and synthetic PPAR ligands and their physiological roles.

(h), EC50 for human PPAR α ; (m), EC50 for murine PPAR α

Table 5: List of selected PPARα target genes

Table 5: Selected PPARα target gene. PPARα regulates the expression of its target genes involved in different biological process via distinct mechanisms (more details for some examples are discussed in the text). \uparrow , represents an increase upon fibrate treatment; \blacklozenge , represents a decrease upon fibrate treatment; $=$, represents no change upon fibrate treatment; **?**, no information available on a change; **#**, represents conflicting results; WT, wild-type mouse; KO, knockout PPARα mouse.

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PPARα/Sirt1 takes place on imperfect DR1s, thereby causing downregulation of many known PPARα target genes mediating FA utilization (56). (C) PPRE-dependent inhibition of NF-κB transcriptional activity. Upon ligand activation, PPRE-bound PPAR α directly interferes with the activity of p65 antagonizing thereby its binding to an NF-κB-RE in the complement promoter (56). Abbreviations used in the figure: Sirt1; Silent information regulator 1; PPRE, PPAR-response element; RXR, Retinoid X receptor; DR1, Direct repeat1; NF-κB-RE, NF-κB response element.

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