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► To cite this version:

Jérémie Babonneau, Christian Bernard, Estelle Marion, Annick Chauty, Marie Kempf, et al.. Development of a dry-reagent-based qPCR to facilitate the diagnosis of *Mycobacterium ulcerans* infection in endemic countries. *PLoS Neglected Tropical Diseases*, 2015, 9 (4), pp.e0003606. 10.1371/journal.pntd.0003606 . inserm-01818058

HAL Id: inserm-01818058

<https://inserm.hal.science/inserm-01818058>

Submitted on 18 Jun 2018

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RESEARCH ARTICLE

Development of a Dry-Reagent-Based qPCR to Facilitate the Diagnosis of *Mycobacterium ulcerans* Infection in Endemic Countries

J r mie Babonneau^{1*}, Christian Bernard², Estelle Marion³, Annick Chauty³, Marie Kempf⁴, Raymond Robert², Laurent Marsollier¹, Franco-Beninese Buruli Research Group[¶]

1 ATOMyCA, Inserm Avenir Team, CRCNA, Inserm U892, 6299 CNRS and LUNAM, CHU and Universit  d'Angers, Angers, France, **2** SR²B, Avrill , France, **3** Centre de diagnostic et de traitement de l'ulc re de Buruli, Pob , B nin, **4** Laboratoire de Bact riologie, Institut de Biologie en Sant —PBH, CHU d'Angers, Angers, France

¶ Membership of Franco-Beninese Buruli Research Group is provided after the Acknowledgments.

* jeremie_babonneau@hotmail.fr



OPEN ACCESS

Citation: Babonneau J, Bernard C, Marion E, Chauty A, Kempf M, Robert R, et al. (2015) Development of a Dry-Reagent-Based qPCR to Facilitate the Diagnosis of *Mycobacterium ulcerans* Infection in Endemic Countries. *PLoS Negl Trop Dis* 9(4): e0003606. doi:10.1371/journal.pntd.0003606

Academic Editor: Pamela L. C. Small, University of Tennessee Knoxville, UNITED STATES

Received: October 31, 2014

Accepted: February 7, 2015

Published: April 1, 2015

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Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: Financial support for this work was provided by INSERM-Avenir, the Raoul Follereau Foundation and the Region Pays de la Loire (ARMINA project) and University Hospital of Angers and university of Angers. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

Abstract

Background

Buruli ulcer is a neglected tropical disease caused by *Mycobacterium ulcerans*. This skin disease is the third most common mycobacterial disease and its rapid diagnosis and treatment are necessary. Polymerase chain reaction (PCR) is considered to be the most sensitive method for the laboratory confirmation of Buruli ulcer. However, PCR remains expensive and involves reagents unsuitable for use in tropical countries with poor storage conditions, hindering the development of reliable quantitative PCR (qPCR) diagnosis. We aimed to overcome this problem by developing a ready-to-use dry qPCR mix for the diagnosis of *M. ulcerans* infection.

Methodology/Principal Findings

We compared the efficiency of three different dry qPCR mixes, lyophilized with various concentrations of cryoprotectants, with that of a freshly prepared mixture, for the detection of a standard range of *M. ulcerans* DNA concentrations. We evaluated the heat resistance of the dry mixes, comparing them with the fresh mix after heating. We also evaluated one of the dry mixes in field conditions, by analyzing 93 specimens from patients with suspected Buruli ulcers. The dry mix was (i) highly resistant to heat; (ii) of similar sensitivity and efficiency to the fresh mix and (iii) easier to use than the fresh mix.

Conclusions

Dry qPCR mixes are suitable for use in the diagnosis of *M. ulcerans* infection in endemic countries. The user-friendly format of this mix makes it possible for untrained staff to perform diagnostic tests with a limited risk of contamination. The possibility of using this mix in either vial or strip form provides considerable flexibility for the management of small or large

amounts of sample. Thus, dry-mix qPCR could be used as a reliable tool for the diagnosis of Buruli ulcer in the field.

Author Summary

Buruli ulcer is a neglected tropical disease caused by *Mycobacterium ulcerans* that may result in severe skin ulceration if left untreated. Quantitative PCR (qPCR) is the most sensitive method for diagnosing Buruli ulcer, but is difficult to perform in endemic areas. We overcame these problems by developing an easy-to-use, heat-resistant dry-reagent qPCR device.

Introduction

Buruli ulcer, caused by *Mycobacterium ulcerans*, is a necrotizing skin disease mostly affecting children under the age of 15 years, and often leading to permanent disabilities[1]. It is one of the 17 neglected tropical diseases identified by the WHO and is the third most common mycobacterial disease affecting humans, after tuberculosis and leprosy [2,3]. Buruli ulcer is endemic to swampy rural areas of tropical and subtropical countries, principally in Africa, although it has been reported in more than 30 countries [2,3].

The early stages of the disease are characterized by a painless nodule, papule, plaque or edema. If left untreated, these lesions progress to the characteristic painless ulcer with undermined edges. Small lesions are treated with a combination of rifampicin and streptomycin for eight weeks [1,4,5]. Surgery may be required in addition to this antibiotic treatment, particularly if the lesions are extensive.

The diagnosis of Buruli ulcer can be confirmed by several methods: (i) direct smear examination, to detect acid-fast bacilli, is a rapid test that is simple to perform but of low sensitivity; (ii) culture on Lowenstein Jensen medium at 32°C; this is the most discriminatory method, but is not very sensitive and takes more than eight weeks, rendering it of little use to clinicians; (iii) histopathological examination is sensitive but expensive and requires a sophisticated laboratory, well trained personnel and invasive procedures (biopsy); (iv) polymerase chain reaction (PCR)-based IS2404 detection is the most widely used method. It is highly sensitive and specific and is also reasonably rapid, but it is expensive and requires trained personnel with specific equipment [6,7,8].

The WHO recommends that PCR confirmation should be performed for at least 70% of the cases reported for any district or country [9,10]. African molecular biology laboratories have recently been equipped with real-time thermal cyclers for the management of many diseases, including avian influenza. This material could be used to diagnose many neglected tropical diseases, including Buruli ulcer. However, due to the lack of a ready-to-use PCR tool, molecular diagnosis is often delayed, because the samples must be sent to reference laboratories, which are frequently located far from Buruli ulcer treatment centers. There may also be significant difficulty obtaining and storing expensive reagents.

In this context, as already done for conventional PCR [11,12], we developed a ready-to-use dry mix, containing all the reagents (master mix, primer and probes) required for quantitative PCR (qPCR) and a mixture of collagen and trehalose as cryoprotectants (dry qPCR mix), for the diagnosis of *M. ulcerans* infection, suitable for use in the countries in which this bacterium is endemic. The resistance of this dry mix to heat and its stability for long-term storage

overcome the shipping and storage constraints, and the ready-to-use format simplifies the handling of the test and facilitates qPCR diagnosis, potentially leading to direct improvements in the medical care of Buruli ulcer patients.

Materials and Methods

Ethics statement

Ethical approval for the analysis of specimens from patients was obtained from the institutional review boards of the CDTLUB of Pobè and Angers University Hospital. The specimens used for this study were anonymized.

DNA extraction and purification

Tissue specimens were finely chopped and homogenized in sterile water with a Tissue Lyser II (Qiagen). Swabs were homogenized in sterile water, and fine-needle aspirates (FNAs) were used directly. For DNA extraction, 400 μ l of sample was centrifuged at 3000 \times *g* for 10 minutes. The supernatant was discarded and the pellet was washed twice with 800 μ l of DNase-free water per wash, with centrifugation at 3000 \times *g* for 10 minutes to recuperate the pellet.

The supernatant was discarded and the pellet was suspended in 50 μ l of 50 mM NaOH and heated at 95°C for 20 minutes to lyse the bacteria. DNA was purified with a QIAquick purification kit (Qiagen), after adjusting the pH by adding 10 μ l of 3 M sodium acetate pH 5.2, according to the manufacturer's recommendations.

PCR conditions

Fresh qPCR mix composition. Oligonucleotide primers and *TaqMan* probes were selected based on the IS2404 sequence ([S1 Table](#)) [7,13]. The qPCR mixture contained 0.3 μ M of each primer, 0.1 μ M probe, a glycerol-free Master Mix (SuperHotTaq Master Mix 2X, BIORON) containing 5 mM MgCl₂ and 5 μ l of template DNA, in a total volume of 25 μ l. Amplification and detection were performed with a thermal cycler (Chromo4, Bio-Rad) using the following program: heating at 95°C for 10 minutes, followed by 40 cycles of 95°C for 15 s and 60°C for 1 minute. The results were analyzed with Opticon Monitor software (Bio-Rad).

Dry qPCR mix preparation. Various dry PCR mixtures were prepared and evaluated for efficiency and accuracy. The dry mixes contained 0.3 μ M of each primer, 0.1 μ M probe, and a glycerol-free Master Mix (SuperHotTaq Master Mix 2X, BIORON) containing 5 mM MgCl₂. They were freeze-dried as described in patent no. EP1629118 (A2), using two cryoprotectants, collagen (cryo P) and trehalose (cryo T). The minimum and maximum concentrations of the cryoprotectants compatible with the chosen qPCR technique were determined during a preliminary evaluation. We used concentrations of 5 to 20% for cryo T and 0.25 to 1% for cryo P. These two products were tested in pairwise comparisons ([Fig. 1](#)).

Two packaging formats were prepared: (i) vials: 850 μ l of the mix was dispensed into an 8 ml amber glass (VFA, Nanterre, France) vial and freeze-dried. The lyophilized mixture was reconstituted by adding 680 μ l of sterile distilled water. The reconstituted solution (20 μ l per well) was added to the test plate, together with 5 μ l of purified DNA per well, giving a total volume of 25 μ l per well; (ii) strips: 20 μ l of the mixture was added to each well of strips suitable for use in a real-time thermal cycler and freeze-dried. The lyophilized mixture was reconstituted by adding 20 μ l of sterile distilled water and 5 μ l of purified DNA per well.

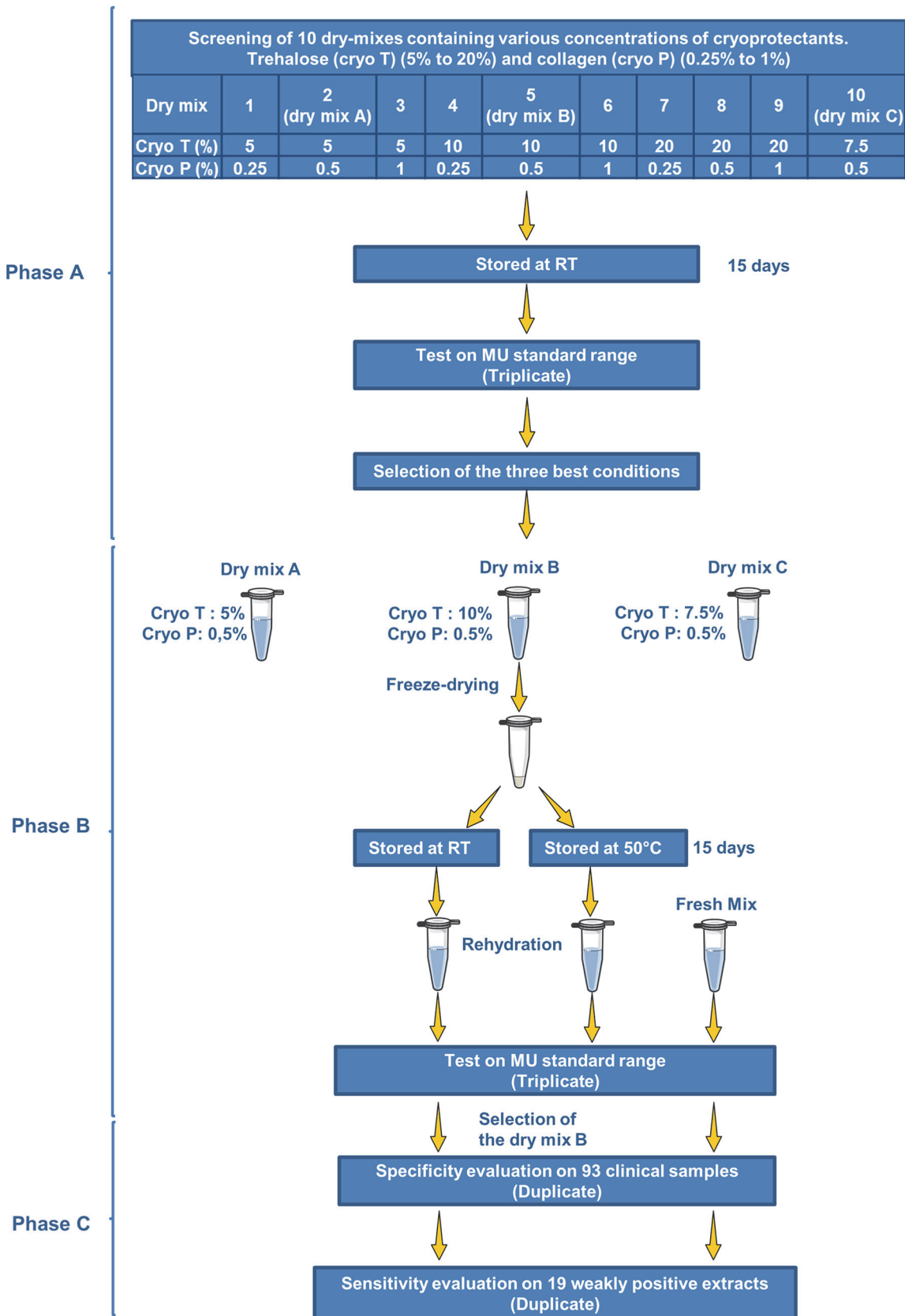


Fig 1. Diagram of the study.

doi:10.1371/journal.pntd.0003606.g001

Range of external *M. ulcerans* DNA standards

The range of external standards used in this study was generated from a mouse tail spiked with a known quantity of *M. ulcerans* (strain 1G897), extracted with the same extraction and purification protocol as was used for the tissue samples and serially diluted over 5 or 7 orders of magnitude, to concentrations of up to 1×10^6 U/ml.

Validation and characterization of dry qPCR mixes

The efficiency (i.e. yield), analytical sensitivities and accuracy of the various dry mixes were compared with those of fresh IS2404 qPCR mix, in quantitative assays on a range of external *M. ulcerans* DNA standards, serially diluted from 1×10^6 to 1×10^0 U/ml. Each mix was tested in triplicate, as previously described [14,15]. Dry mixes were considered unsuitable for diagnostic purposes if the threshold cycle (C_t) was 0.15 log (0.5 C_t) below the results obtained with the fresh mix, as described by the *Association Française de Normalisation* (NF T90-471).

Evaluation of stability at high temperature

The stability of the various dry qPCR mixes at high temperature was evaluated, by exposing these mixes to a temperature of 50°C for two weeks. The detection efficiencies obtained were compared with that for the fresh mix, for the standard range of bacterial concentrations, from 1×10^6 to 1×10^2 U/ml.

Diagnosis specimens

The specimens were collected by the local medical staff at the Pobè CDTUB. FNAs were collected from non-ulcerative lesions, swabs were taken from the undermined edges of the lesions and tissue specimens were obtained from surgically excised tissues, as described elsewhere [4,5,9,10,14,16,17].

Clinical validation

The diagnosis sensitivities of the fresh mix and dry mix B were compared, on human samples, by reanalyzing 19 purified specimens (S2 Table) of various types from a retrospective study after their storage at -80°C . The specimens used were selected on the basis of their low bacterial content, as previously determined at Angers University Hospital (10 U/ml to 1000 U/ml, corresponding to $\approx 37.5 C_t$ and $\approx 30 C_t$, respectively). Diagnosis sensitivity was evaluated by comparing the results obtained with dry mix B and the reference method for these specimens.

During this study (January 2013—December 2013), analyses were carried out on 93 specimens suspected to contain *M. ulcerans* DNA and sent from the Pobè CDTUB (swabs, $n = 48$; tissue specimens, $n = 27$; FNAs, $n = 18$; S3 Table) for diagnosis. The specificity of dry mix B was assessed by comparing the results obtained with this mix with those obtained for the reference method, at the molecular biology laboratory of Angers University Hospital, which has been responsible for carrying out PCR diagnosis for Pobè since 2006.

Internal quality control

Several controls were included, to exclude the possibility of contamination during the preparatory steps: (i) Extraction and purification controls were performed, with sterile water rather than a specimen for the negative control, and with an extract of the tail of a mouse inoculated with *M. ulcerans* containing 10^3 bacteria for the positive control; (ii) As a negative qPCR mix control, 5 μl of elution buffer was added to the mixture in place of the template DNA; (iii) A

range of external standard *M. ulcerans* DNA (strain 1G897) concentrations was used in each experiment, for quantification and to control for PCR efficiency.

Inhibition control reactions were performed to rule out the presence of inhibitors in the purified DNA.

A DNA extract from the standard range (2 µl of a 1x10⁵ U/ml DNA extract) was added to the mixtures, replacing 2 µl water. Inhibitory effects were excluded if the C_t was less than 0.15 log (0.5 C_t) away from the expected results.

Results

Selection of the optimal cryoprotectant concentrations for the dry mix

The minimum and maximum concentrations of the cryoprotectants compatible with the chosen qPCR technique were first determined. We used concentrations of 5 to 20% for cryo T and 0.25 to 1% for cryo P. These two products were tested in pairwise comparisons, to determine the optimal concentrations, using the external standard range obtained from mouse tails spiked with *M. ulcerans*. After preliminary evaluations, based on the efficiency of the dry mixes after storage at room temperature for 15 days, the conditions with suboptimal concentrations of cryoprotectant were discarded. Three conditions were retained for further investigations (Fig. 1). They were evaluated on the basis of four criteria: a comparison of the efficiencies of the dry mixes with that of the fresh mix, analytical sensitivity, accuracy and the visual appearance of the dry mixes. In qPCR assays, the results obtained with the fresh mix were significantly correlated with those obtained with the three dry mixes (Fig. 2). The cryoprotectants therefore had no inhibitory effect. Dry mixes A, B and C had efficiencies of 104%, 98% and 98%, respectively. No difference in accuracy was observed between the three dry mixes and the fresh mix ($\Delta C_t < 0.5$).

The detection thresholds (mean C_t) of the dry mixes A, B and C for the 1x10¹ concentration of the DNA standard range were 37.47 (5 U/ml), 36.19 (12 U/ml) and 36.35 (11 U/ml), respectively, whereas that for the fresh mix was 36.95 (8 U/ml). No copy was detectable in any mix

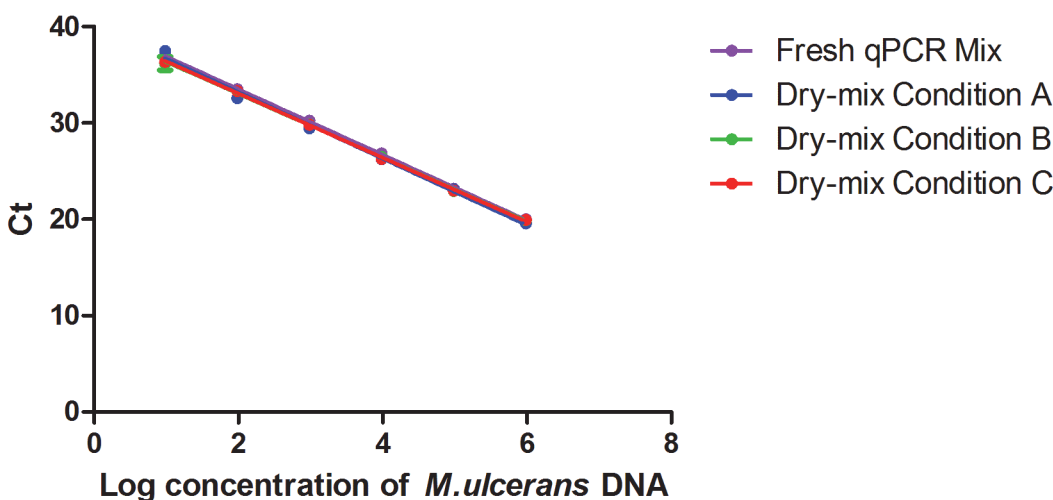


Fig 2. Comparison between 3 dry-reagent conditions and the standard reference method, based on an external standard curve. Serial 10-fold dilution from 1x10⁶ U/ml to 1x10⁰U/ml of *M. ulcerans* DNA. The serial curve was generated by a linear regression of the threshold cycle (C_t) against the logarithm of *M. ulcerans* DNA concentration. Each 10-fold dilution was performed in triplicate. One standard deviation to either side of the mean is shown. Standard QPCR Mix refers to the reference method with fresh mix ($y = -3.4107x + 40.357$); dry-mix condition A refers to the dry mix containing 5% cryo T and 1% cryo P ($y = -3.2333x + 39.087$); dry-mix condition B refers to the dry mix containing 10% cryo T and 1% cryo P ($y = -3.3727x + 39.942$); dry-mix condition C refers to the dry mix containing 7.5% cryo T and 0.5% cryo P ($y = -3.357x + 39.851$). For each assay, the coefficient of correlation was greater than 0.99.

doi:10.1371/journal.pntd.0003606.g002

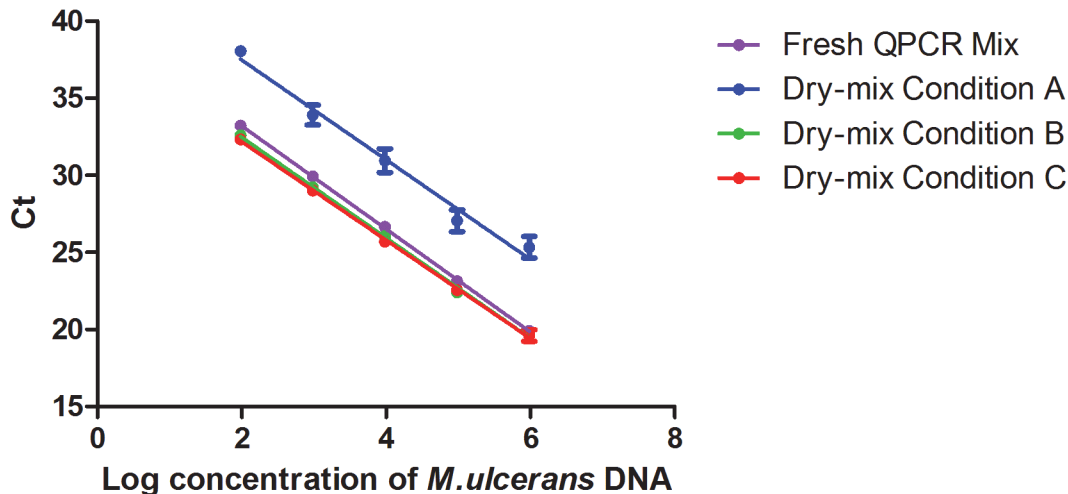


Fig 3. Comparison of the 3 dry-reagents conditions after exposure to a temperature of 50°C for 15 days and the standard reference method, based on an external standard curve. Serial 10-fold dilutions, from 1×10^6 U/ml to 1×10^2 U/ml, of *M. ulcerans* DNA. The serial curve was generated by linear regression of the threshold cycle (C_t) against the logarithm of *M. ulcerans* DNA concentration. Each 10-fold dilution was performed in triplicate. One standard deviation on either side of the mean is shown. “Standard QPCR Mix” corresponds to the reference method with fresh mix ($y = -3.34x + 39.88$); dry mix A is the dry mix containing 5% cryo T and 1% cryo P ($y = -3.23x + 43.94$); dry mix B is the dry mix containing 10% cryo T and 1% cryo P ($y = -3.26x + 38.98$); dry mix C is the dry mix containing 7.5% cryo T and 0.5% cryo P ($y = -3.19x + 38.55$).

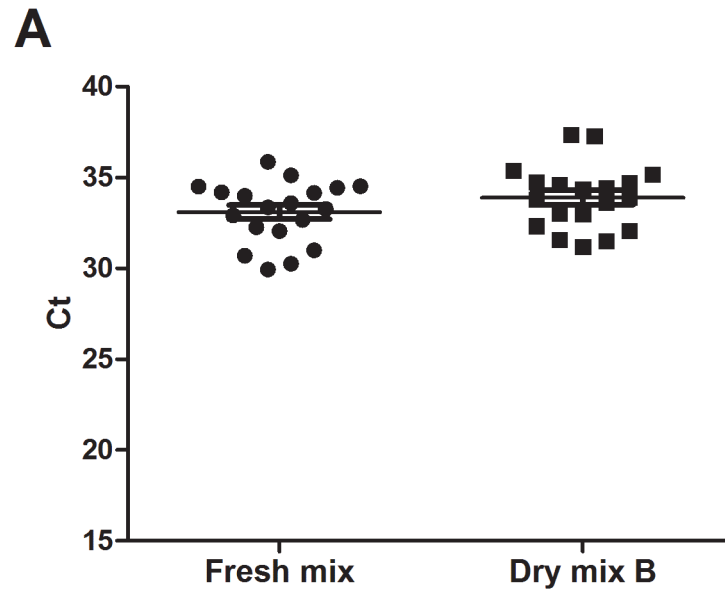
doi:10.1371/journal.pntd.0003606.g003

for the 1×10^0 concentration. The analytical sensitivity was thus 10 U/ml and concentrations below 10 U/ml were considered detectable but inconsistent for quantification. The correlation coefficients were greater than 0.99 and the visual aspect of the dry products were conforming.

Resistance to heat was therefore evaluated. The dry mixes were incubated at 50°C for two weeks, and their visual appearance was then assessed. Dry mix A looked shiny and retracted, whereas dry mix B still appeared to be dry, like the original freeze-dried product, and dry mix C appeared to have shrunk slightly in comparison to dry mix B. Dry mixes A, B and C had efficiencies of 104%, 103% and 106%, respectively, and correlation coefficients of 0.98, 0.99 and 0.99, which is acceptable. The qPCR results showed a lower accuracy for dry mix A after exposure to 50°C for 15 days than for the fresh mix, with a ΔC_t (dry mix A – fresh mix) between 3.93 and 5.42 (Fig. 3). After exposure to the same conditions, dry mixes B and C behaved similarly to the fresh mix ($\Delta C_t < 0.5$). These results, indicating that dry mix A was not stable at high temperatures, resulted in the exclusion of this dry mix from subsequent experiments. Dry mixes B and C both gave good results after incubation at 50°C, but the slightly shrunken appearance of dry mix C reflected an inappropriate concentration of cryoprotectants. We therefore decided to use only dry mix B in subsequent studies.

Validation of the analytical sensitivity of the selected dry mix against that of the fresh mix, for 19 selected weakly positive extracts

Nineteen purified DNA samples (S2 Table) with low concentrations were selected from a retrospective study for evaluation of the sensitivity of dry mix B. No differences in analytical sensitivity were found between the results obtained with the reference method and those obtained with the dry mixes, with 100% detection rates obtained with both methods. The ΔC_t value of the mean recorded for the quantification with the dry mix was more than 0.5 C_t (ΔC_t mean = 0.81) away from value for the fresh mix (Fig. 4), due to the imprecision introduced by the low copy number. No inhibitors were detected in the purified DNA during this experiment. The



	Fresh Mix (Ct)	Dry mix B (Ct)
Mean	33.09	33.9
Std. deviation	1.688	1.75
Δ Ct Mean	0	0.81
Δ Ct Std. deviation	0	0.062

Fig 4. CT values for the 19 weakly positive samples, for 2 conditions. (A) C_t results for each sample obtained by the fresh mix and the dry mix B, Horizontal bars indicate mean values \pm SD. (B) Mean values \pm SD and average $\Delta C_t \pm$ SD between fresh mix and dry mix B.

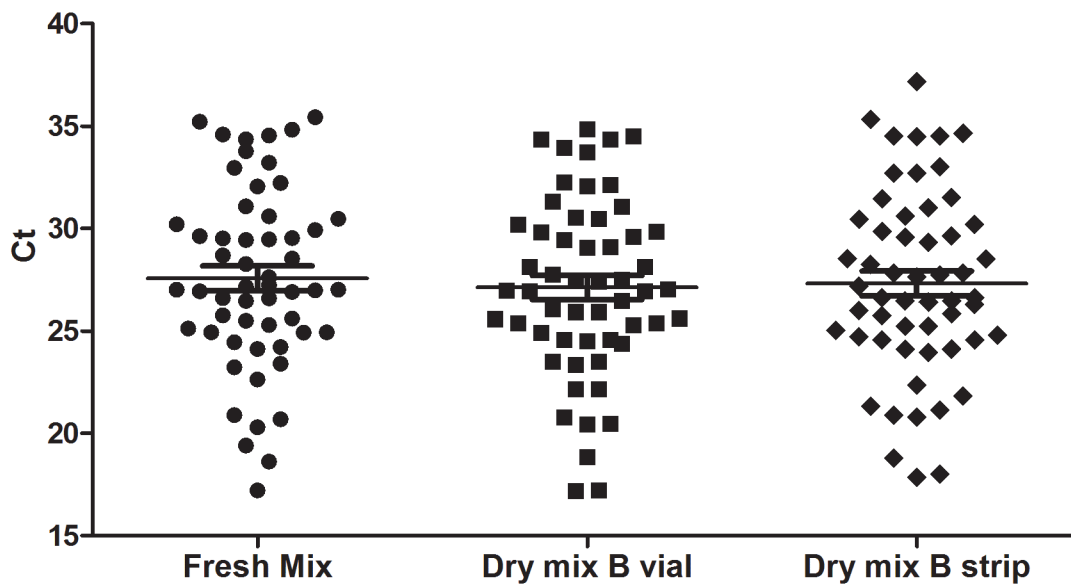
doi:10.1371/journal.pntd.0003606.g004

sensitivity of the dry mix was therefore similar to that of the fresh mix for use on clinical samples, even for the detection of small amounts of DNA.

Clinical evaluation, comparing the selected dry mix with the fresh mix, for 93 patients with infection suspected on clinical grounds

For validation of the dry mix in field conditions, the selected dry mix was evaluated on 48 swabs, 27 tissue samples and 18 FNAs from 93 patients with suspected Buruli ulcer. Two

A



B

	Fresh Mix (Ct)	Dry mix B vial (Ct)	Dry mix B strip (Ct)
Mean	27.57	27.12	27.31
Std. deviation	4.438	4.352	4.533
Δ Ct Mean	0	0.45	0.26
Δ Ct Std. deviation	0	0.086	0.095

Fig 5. CT values for the 55 positive samples, for 3 mix conditions. (A) C_t results for each sample obtained by the fresh mix and the dry mix B in vial or in strip format, Horizontal bars indicate mean values \pm SD. (B) Mean values \pm SD and average $\Delta C_t \pm$ SD between fresh mix and dry mix B in vial or in strip format.

doi:10.1371/journal.pntd.0003606.g005

different formats of the dry mix were also evaluated, for greater flexibility: (i) vials containing a volume sufficient for 32 reactions, for diagnostic tests on normal to large series of samples and (ii) eight-well strips, to decrease costs and facilitate the analysis of small series of samples. The results obtained with dry mix B, in the vial and strip formats, were compared with those

obtained with the fresh mix. Identical results were obtained for all three types of test (S3 Table): 55 specimens were positive for *M. ulcerans* DNA and 38 were negative. The ΔC_t values recorded for the quantification with the dry mix were less than 0.5 C_t away from the values recorded with the fresh mix (Fig. 5). No inhibitors were detected in the purified DNA used. All cases testing positive were treated for Buruli ulcer. All subjects testing negative for Buruli ulcer were treated in accordance with the alternative diagnosis reached by the clinician. The diagnosis specificity of the two methods (dry mix and fresh mix) was 100%.

Discussion

The WHO recommends IS2404 qPCR amplification for the confirmation of Buruli ulcer diagnosis, because this technique is both the fastest and the most sensitive [9,10]. Many African countries have recently obtained qPCR equipment, for avian influenza surveillance. This equipment could also be used for the diagnosis of other diseases, including neglected diseases. However, the handling of sensitive reagents can be critical in tropical countries. This problem is jeopardizing the development of efficient diagnostic tools.

In this context, we developed a dry qPCR mix, overcoming local problems relating to transport, storage, cost and freezing/thawing issues, which can substantially lower the sensitivity of qPCR. Our dry-qPCR mix presents several major advantages over fresh qPCR mix. Its resistance to high temperatures should ensure its stability during handling in tropical conditions.

Furthermore, the number of new Buruli ulcer cases per week in African countries in which this disease is endemic is generally between 0.7 (Congo: 38 new cases in 2012) and 26.7 (Ivory Coast: 1386 new cases in 2012) [10]. Molecular diagnosis is therefore often delayed while additional samples are collected, so that molecular diagnostic tests can be carried out in batches, to decrease costs and the time spent on tests. The ready-to-use format simplifies handling by laboratory technicians, and the possibility of using either vials or microtube strips maximizes flexibility, enabling the laboratory to optimize time/sample and reagent/sample ratios. This simplification of handling decreases the time required for manipulations and minimizes the risk of contamination.

The dry mix qPCR approach could be adapted for other sets of primers and probes, such as the ketoreductase-B (KR) domain of the *M. ulcerans* mycolactone polyketide synthase genes. This would facilitate the diagnosis of many other neglected tropical diseases, such as rabies, trachoma and dengue.

Supporting Information

S1 Table. Primers and probe used to detect *M. ulcerans* DNA sequences by TaqMan real-time PCR.

(DOCX)

S2 Table. Listing of the weakly positive control samples and Ct results for the dry mix vs. gold standard method. “FNA” = fine needle aspiration, “U” = ulcerative forms, “E” = edematous forms, “Q” = plaques, “N” = nodules and two or more letters indicate mixed forms.

(DOCX)

S3 Table. Listing of the 93 diagnosis specimens and Ct results for the two dry mixes vs. the gold standard method. “FNA” = fine needle aspiration, “U” = ulcerative forms, “E” = edematous forms, “Q” = plaques, “N” = nodules, “OS” = osteomyelitis and two or more letters indicate mixed forms.

(DOCX)

S1 Dataset. Raw data of the QPCR with the different tested mixes.
(XLSX)

S1 Text. Document explaining [S1 Dataset](#).
(TXT)

Acknowledgments

We thank the staff at the *Centre de Dépistage et de Traitement de l'Ulcère de Buruli* (CDTUB), Pobè, Bénin; Catherine Quinqueneau and the staff of the *Laboratoire de Bactériologie*, CHU, Angers, France; and the administrative staff of the Regional Delegation (ADR Inserm grand Ouest) for technical assistance.

Members of the Franco-Beninese Buruli Research Group:

France:

Paris: Quentin B. Vincent^{1, 2}, Laurent Ab^{1, 2, 4}, Christian Johnson³, Alexandre Alcaïs^{1, 2, 4, 5}
Angers: Estelle Marion⁶, Marie Kempf⁷, Jane Cottin⁷, Jean-Paul Saint-André⁸, Laurent Marsollier⁶

Benin:

Pobè: Marie-Françoise Ardant^{3, 9}, Ambroise Adeye⁹, Aimé Goundote⁹, Annick Chauty^{3, 9}
Cotonou: Didier Agossadou¹⁰

(1) Laboratory of Human Genetics of Infectious Diseases, Necker Branch, Institut National de la Recherche Médicale U1163 (INSERM), Paris, France

(2) Université Paris Descartes, Sorbonne Paris Cité, Institut Imagine, Paris, France

(3) Fondation Raoul Follereau, Paris, France

(4) St Giles Laboratory of Human Genetics of Infectious Diseases, Rockefeller Branch, Rockefeller University, New York, NY, USA

(5) Unité de Recherche Clinique, Paris Centre Descartes Necker Cochin, Assistance Publique-Hôpitaux de Paris, Paris, France

(6) Institut National de la Recherche Médicale U892 (INSERM) et CNRS U6299, équipe 7, Université et CHU d'Angers, Angers, France

(7) Laboratoire de Bactériologie, CHU d'Angers, Angers, France

(8) Laboratoire d'Anatomie Pathologique, CHU d'Angers, Angers, France

(9) Centre de Dépistage et de Traitement de l'Ulcère de Buruli (CDTUB), Pobè, Bénin

(10) Programme de Lutte Contre la Lèpre et l'Ulcère de Buruli, Ministère de la Santé, Cotonou, Bénin

Author Contributions

Conceived and designed the experiments: JB CB RR LM. Performed the experiments: JB CB. Analyzed the data: JB EM LM. Contributed reagents/materials/analysis tools: CB AC MK RR LM. Wrote the paper: JB CB EM RR LM.

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