

Commentary

## Activation of AMPK for a break in hepatic lipid accumulation and circulating cholesterol

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Dysregulation of lipid metabolism leading to excessive lipids within the liver constitutes a major risk factor for life-threatening diseases, such as non-alcoholic fatty liver disease (NAFLD) as well as obesity and diabetes, and atherosclerotic cardiovascular disease. NAFLD represents the most common chronic liver disease and encompasses a wide range of liver abnormalities, from benign simple steatosis (ectopic accumulation of lipids), potentially progressing to an intermediate form, nonalcoholic steatohepatitis with inflammation (NASH), and to advanced stages of liver disease, including fibrosis, cirrhosis, and, in some cases, hepatocellular carcinoma. Currently, therapeutic approaches are limited and no pharmacological treatments approved by regulatory agencies are available. Accumulation of hepatic triglycerides (TGs) in NAFLD was estimated to derive for ~60% from fatty acids released from adipose tissue and for ~25% from *de novo* lipogenesis (DNL) (Donnelly et al., 2005). A valuable treatment option is targeting of the evolutionarily conserved energy sensor AMP-activated protein kinase (AMPK) originally identified as the kinase that phosphorylates and inhibits both acetyl coenzyme A carboxylase (ACC) and 3-hydroxy 3-methylglutaryl coenzyme A reductase (HMGCR), which are the rate-limiting enzymes for fatty acid and cholesterol biosynthesis, respectively (Lin and Hardie, 2018). Activation of AMPK in the liver by a number of pharmaceutical/nutraceutical compounds has been reported to inhibit fatty acid synthesis and promote fatty acid oxidation (FAO) via phosphorylation and inactivation of ACC (Boudaba et al., 2018; Fullerton et al., 2013; Smith et al., 2016). Because of these effects, AMPK hold promise as an attractive therapeutic target to rescue the liver from excessive lipid accumulation and limit possible risk of NAFLD progression.

Mammalian AMPK exists as a heterotrimeric complexes comprising a catalytic  $\alpha$  subunit and regulatory  $\beta$  and  $\gamma$  subunits that occurs as multiple isoforms ( $\alpha 1$ ,  $\alpha 2$ ,  $\beta 1$ ,  $\beta 2$ ,  $\gamma 1$ ,  $\gamma 2$ ,  $\gamma 3$ ) with unique tissue-specific expression profiles. AMPK plays a key role in controlling cellular energy balance and metabolism by turning off ATP-consuming anabolic pathways (e.g., lipid synthesis) and switching on catabolic pathways (e.g., fatty acid oxidation) in response to metabolic stresses (Lin and Hardie, 2018). Since AMPK is sensitive to changes in cellular

nucleotide levels (AMP:ATP and ADP:ATP ratios), AMP mimetics and compounds modulating cellular energy charge (e.g., by inhibition of ATP synthesis via inhibition of mitochondrial function) were attractive AMPK-activating treatments. However, most of these compounds activate AMPK indirectly and have AMPK-independent off-target pharmacological effects, thus limiting enthusiasm for clinical application and development (Smith et al., 2016). Recent crystallographic studies helped to identify a novel allosteric site, termed as allosteric drug and metabolite (ADaM) site, corresponding to a pocket formed between the  $\beta$  subunit and the kinase domain of the  $\alpha$  subunit (Lin and Hardie, 2018). Identification by Abbott Laboratories of the first direct AMPK activator A-769662 binding the ADaM site was an important pharmacological breakthrough that encouraged large screening programs to identify novel direct AMPK activators (Cameron and Kurumbail, 2016; Cool et al., 2006).

In this issue of EBioMedicine, Esquejo and colleagues (Esquejo et al., 2018) describe the therapeutic effect of a novel direct AMPK  $\beta$ 1 biased activator (PF-06409577) on NAFLD progression in mouse and rat models. In rodent liver,  $\alpha$ 1 $\beta$ 1 $\gamma$ 1 and  $\alpha$ 2 $\beta$ 1 $\gamma$ 1 heterotrimers are predominantly expressed and it was not surprising that PF-06409577 suppresses DNL in wild type but not AMPK $\alpha$ 1/ $\alpha$ 2-deficient primary mouse hepatocytes, as previously shown with A-769662 specific for  $\beta$ 1-containing complexes (Boudaba et al., 2018). However, it was important to demonstrate the ability of this AMPK  $\beta$ 1 biased activator to activate AMPK in human hepatocytes because  $\alpha$ 1 $\beta$ 2 $\gamma$ 1 has been identified as the major heterotrimer (Boudaba et al., 2018). Despite low levels of AMPK  $\beta$ 1 subunit in human hepatocytes, the authors show a dose-dependent inhibition of DNL with an EC<sub>50</sub> of 128 nM compared to 49 nM in rat hepatocytes (Esquejo et al., 2018). Similar results were found with previous AMPK  $\beta$ 1 biased activator A-769662 (Boudaba et al., 2018). In addition, consistent with AMPK-dependent phosphorylation and inactivation of ACC, PF-06409577-induced suppression of fatty acid synthesis was severely blunted in hepatocytes isolated from mice with alanine knock-in mutations (ACC-DKI) that render ACC activity insensitive to AMPK (Esquejo et al., 2018). Chronic PF-06409577 treatment in obese rodents resulted in reduction of liver TG content in wild type but not liver AMPK $\alpha$ 1/ $\alpha$ 2 KO mice, excluding significant contribution of AMPK activation in non-hepatic tissues. This lipid-lowering effect was accompanied by an increase in circulating  $\beta$ -hydroxybutyrate levels in wild type but not liver AMPK $\alpha$ 1/ $\alpha$ 2 KO mice, suggesting a contribution of FAO in addition to the inhibition of DNL. In support of the importance of AMPK activation and ACC phosphorylation in the stimulation of FAO, recent studies reported that a large panel of AMPK activators failed to induce FAO in AMPK $\alpha$ 1/ $\alpha$ 2 KO and ACC-DKI hepatocytes (Boudaba et al., 2018; Fullerton et al., 2013).

Interestingly, the authors find that regulation of hepatic lipogenesis by AMPK activation mainly resides in the phosphorylation and inactivation of ACC but not in the control of lipogenic gene expression, in accordance with previous study (Boudaba et al., 2018). Overall, these findings suggest therapeutic potential of direct targeting ACC to lower hepatic fat. This was recently tested in a small human trial where administration of a pharmacological inhibitor of ACC to subjects with hepatic steatosis led to a significant reduction in liver TGs, but this was also associated with hypertriglyceridemia, precluding further clinical development. (Kim et al., 2017). An alternative approach to lower DNL in humans could be through the activation of AMPK which is not accompanied with a rise in plasma TGs in preclinical animal models (Cool et al., 2006; Esquejo et al., 2018).

There has been increasing interest in finding new therapeutic strategies that could prevent or even reverse established hepatic fibrosis and progression to NASH. Encouraging findings from this study is the alteration of the hepatic fibrosis/ stellate cell activation pathways in response to chronic AMPK activation. Although, these data are interesting, the impact of chronic AMPK activation should be confirmed in relevant models of fibrosis. In addition, the mode of action of AMPK activation is elusive and remains to be elucidated.

The study from Esquejo *et al.* also addresses the consequence of AMPK activation on the regulation of cholesterol metabolism. The authors show that PF-06409577 treatment lowers plasma mevalonic acid levels, a direct product of HMGCR and resulted in the reduction of plasma cholesterol and increase of plasma HDL cholesterol in ZSF1 rat, a model mimicking human hypercholesterolemia. Although clinical relevance was provided by a reduction of circulating plasma cholesterol and LDL in cynomolgus monkeys treated for 6 weeks with PF-06409577, these data raise important questions about the role for AMPK and the potential target proteins and tissues involved in these beneficial effects. Analysis of mice with alanine knock-in mutation of the AMPK phosphorylation site in HMGCR could be an interesting area of future study. Lastly, long-term studies addressing the efficacy of AMPK  $\beta$ 1 biased activators for the prevention of atherosclerotic cardiovascular disease, alone or when used in combination strategies with current LDL-cholesterol lowering drugs (e.g., statins, PCSK9 inhibitors), are warranted.

Collectively, these results provide compelling evidence for the therapeutic potential of AMPK activation in the treatment of hepatic lipid disorders. The authors not only confirmed that AMPK activation improves hepatic lipid content in obese rodents but also showed a reduction of cholesterol plasma concentration in a hypercholesterolemic rat model and in non-human primates. Therefore, these findings open interesting therapeutic perspectives for the use of novel generation of small molecule AMPK activators for the treatment of dyslipidemia. In particular, the insights afforded by this study are truly exciting and will pave the way to new therapeutic strategies to mitigate or prevent the development of NAFLD, hyperlipidemia and the risk of associated complications (e.g. NASH, atherosclerotic cardiovascular disease). Thus, direct AMPK activators warrant further testing as a treatment of NAFLD in humans. However, before consideration, benefits must be balanced against potential cardiac hypertrophy, as recently reported after long-term use of a pan- $\beta$  AMPK activator in rats and rhesus macaques (Myers *et al.*, 2017). Although PF-06409577 entered a phase I clinical trial for the treatment of diabetic nephropathy, this study was not completed (Cameron and Kurumbail, 2016) and there is currently no available data on the possibility of safe therapeutics with biased or isoform-selective AMPK activators.

### **Disclosures**

The authors declare no conflicts of interests.

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### **References**

Boudaba, N., Marion, A., Huet, C., Pierre, R., Viollet, B., and Foretz, M. (2018). AMPK Re-Activation Suppresses Hepatic Steatosis but its Downregulation Does Not Promote Fatty Liver Development. *EBioMedicine* 28, 194-209.

Cameron, K.O., and Kurumbail, R.G. (2016). Recent progress in the identification of adenosine monophosphate-activated protein kinase (AMPK) activators. *Bioorg Med Chem Lett* 26, 5139-5148.

Cool, B., Zinker, B., Chiou, W., Kifle, L., Cao, N., Perham, M., Dickinson, R., Adler, A., Gagne, G., Iyengar, R., et al. (2006). Identification and characterization of a small molecule AMPK activator that treats key components of type 2 diabetes and the metabolic syndrome. *Cell metabolism* 3, 403-416.

Donnelly, K.L., Smith, C.I., Schwarzenberg, S.J., Jessurun, J., Boldt, M.D., and Parks, E.J. (2005). Sources of fatty acids stored in liver and secreted via lipoproteins in patients with nonalcoholic fatty liver disease. *J Clin Invest* 115, 1343-1351.

Esquejo, R.M., Salatto, C.T., Delmore, B., Reyes, A.R., Shi, Y., Moccia, R., Cokorinos, E.C., Peloquin, M., Barricklow, J., Bollinger, E., et al. (2018). Activation of Liver AMPK with PF-06409577 Corrects NAFLD and Lowers Cholesterol in Rodent and Primate Preclinical Models. *EBioMedicine*.

Fullerton, M.D., Galic, S., Marcinko, K., Sikkema, S., Pulinilkunnil, T., Chen, Z.P., O'Neill, H.M., Ford, R.J., Palanivel, R., O'Brien, M., et al. (2013). Single phosphorylation sites in Acc1 and Acc2 regulate lipid homeostasis and the insulin-sensitizing effects of metformin. *Nat Med* 19, 1649-1654.

Kim, C.W., Addy, C., Kusunoki, J., Anderson, N.N., Deja, S., Fu, X., Burgess, S.C., Li, C., Ruddy, M., Chakravarthy, M., et al. (2017). Acetyl CoA Carboxylase Inhibition Reduces Hepatic Steatosis but Elevates Plasma Triglycerides in Mice and Humans: A Bedside to Bench Investigation. *Cell Metab* 26, 394-406 e396.

Lin, S.C., and Hardie, D.G. (2018). AMPK: Sensing Glucose as well as Cellular Energy Status. *Cell Metab* 27, 299-313.

Myers, R.W., Guan, H.P., Ehrhart, J., Petrov, A., Prahalada, S., Tozzo, E., Yang, X., Kurtz, M.M., Trujillo, M., Gonzalez Trotter, D., et al. (2017). Systemic pan-AMPK activator MK-8722 improves glucose homeostasis but induces cardiac hypertrophy. *Science* 357, 507-511.

Smith, B.K., Marcinko, K., Desjardins, E.M., Lally, J.S., Ford, R.J., and Steinberg, G.R. (2016). Treatment of nonalcoholic fatty liver disease: role of AMPK. *Am J Physiol Endocrinol Metab* 311, E730-E740.