

# A complementary tool for management of disseminated Histoplasma capsulatum var. capsulatum infections in AIDS patients

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## A complementary tool for management of disseminated *Histoplasma capsulatum var. capsulatum* infections in AIDS patients

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#### 30 **ABSTRACT** 31

In South America, disseminated histoplasmosis due to Histoplasma capsulatum var. 32 33 capsulatum (H. capsulatum), is a severe and frequent opportunistic infection in AIDS 34 patients. In areas outside the USA where specific-Histoplasma antigen detection is not available, the diagnosis is difficult. With the galactomannan antigen (GM) detection, a test 35 36 commonly used for invasive aspergillosis diagnosis, there is a cross-reactivity with H. *capsulatum* that can be helpful for the diagnosis of histoplasmosis. The aim of this study was 37 to evaluate the GM detection for the diagnosis of disseminated histoplasmosis in AIDS 38 39 patients. The performance of the GM detection was evaluated with serum collected in French Guiana where H. capsulatum is highly endemic. Sera from AIDS patients with disseminated 40 histoplasmosis occurring from 2002 to 2009 and from control HIV-positive patients without 41 42 histoplasmosis were tested with the GM detection and Histoplasma-specific antibody 43 detection (IEP). In 39 AIDS patients with proven disseminated histoplasmosis, the sensitivity of the Histoplasma IEP was only 35.9% and was linked to the TCD4+ lymphocyte level. For 44 45 the GM detection, the sensitivity (Se) was 76.9% and specificity (Sp) was 100% with the recommended threshold for aspergillosis diagnosis (0.5). The test was more efficient with a 46 threshold of 0.4 (Se: 0.82 [95% CI: 0.66-0.92], Sp: 1.00 [95% CI: 0.86-1.00], LR+: >10, LR-: 47 0.18). This study confirms that the GM detection can be a surrogate marker for the diagnosis 48 of disseminated histoplasmosis in AIDS patients in endemic areas where Histoplasma EIA is 49

50 not available.

**KEYWORDS**: histoplasmosis; AIDS; *Histoplasma capsulatum*; Diagnosis; Platelia; galactomannan; antibody; cross-reactivity

## **INTRODUCTION**

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7 8 9 Histoplasmosis caused by Histoplasma capsulatum var. capsulatum (H. capsulatum) is 10 endemic in the United States, in several countries of Central and South America and in 11 scattered areas of Asia and Africa (Kauffman, 2007). Disseminated histoplasmosis is a severe 12 illness that occurs almost exclusively in immunosuppressed patients, particularly in patients with acquired immunodeficiency syndrome (AIDS). In French Guiana, histoplasmosis is the 13 most frequent opportunistic infection in HIV-infected patients and the first cause of AIDS-14 15 related death (Couppie et al., 2004; Huber et al., 2008; Lewden et al., 2004). Even if isolation of Histoplasma from cultures is the reference procedure for histoplasmosis diagnosis 16 (Kauffman, 2008), it can take weeks and is positive in only 50-70% of cases 17 18 (Sathapatayavongs et al., 1983). Serological methods based on specific antibody detection can 19 be performed rapidly but usually give false negative results in AIDS patients (Tobon et al., 2005). In this context, circulating specific-Histoplasma antigen detection (Histoplasma EIA) 20 21 represents a useful option for diagnosis but this method is often unavailable in the majority of 22 endemic areas outside the USA (Connolly et al., 2007; Hage et al., 2011).

The Platelia Aspergillus enzyme immunoassay (EIA) is a ready-to-use test which detects 23 galactofuranose-containing side chains of galactomannan. This test is commonly used for the 24 25 diagnosis of invasive aspergillosis, particularly in solid organ transplant recipients or patients with haematological malignancies (Aquino et al., 2007), but cross-reactivity exists with many 26 27 other fungi (Aquino et al., 2007; Dalle et al., 2005; Desoubeaux et al., 2014; Giacchino et al., 2006; Huang et al., 2007; Van Der Veer et al., 2012; Xavier et al., 2009) in particular H. 28 29 capsulatum (Narreddy and Chandrasekar, 2008; Wheat et al., 2007). Galactomannan antigen (GM) detection can lead to a false-positive diagnosis of aspergillosis in these 30 immunocompromised patients who have histoplasmosis (Jones et al., 2009; Vergidis et al., 31 2012). In areas outside the USA where Histoplasma EIA is not available, this cross-reactivity 32 with the galactomannan antigen could be helpful for the diagnosis of disseminated 33 34 histoplasmosis (Pineau et al., 2010; Ranque et al., 2007; Riviere et al., 2012). GM detection 35 could be particularly interesting in HIV-infected patients because of the low incidence of invasive aspergillosis in this population compared to patients with organ transplantation or 36 37 hematological malignancies (Desoubeaux et al., 2014).

Given the rarity of histoplasmosis in Europe, preliminary studies on GM detection for this 38 diagnosis were only conducted on a limited number of samples (Ranque et al., 2007). Thus, 39 the aim of our study was to evaluate more widely the GM detection for the diagnosis of 40 disseminated histoplasmosis in AIDS patients. This evaluation was carried out in a H. 41 42 capsulatum endemic area (French Guiana) to obtain a larger cohort of patients.

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#### 45 **PATIENTS AND METHODS** 46

#### 47 **Study population**

Disseminated histoplasmosis cases occurring in AIDS patients followed in the Cayenne 48

- hospital (French Guiana) were selected from 2002 to 2009. Among this cohort, all patients for 49
- whom sera were collected at the time of diagnosis of histoplasmosis ( $\pm 10$  days=baseline) and 50 51
- available for analysis (stored at -20°C) were included in the study. HIV-patients were

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1 considered as histoplasmosis cases, if there was microbiological evidence of *Histoplasma* 2 infection (positive direct examination and/or culture and/or polymerase chain reaction (Simon 3 et al., 2010) for *Histoplasma capsulatum*). Thirty HIV-positive patients who had never been 4 in an endemic area for *Histoplasma* were included as negative controls. For these controls, no 5 critical antibiotics (Piperacillin-tazobactam), polyvalent immunoglobulins or dialysis were 6 present when GM detection was performed.

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## 8 GM detection and *Histoplasma*-specific antibody detection

Sera stored at -20°C were tested with the Platelia Aspergillus EIA (BioRad, France) in accordance with the manufacturer's specifications. Briefly, after a pretreatment (6 min, 120°C), 50 µl of serum was added to 50 µl of conjugate before being incubated at 37°C for 90 min. After washing, chromogen substrate solution was added and the plates were darkincubated for 30min. After the addition of stop solution, the optical density (OD) was determined at 450nm (reference filter 620/630nm). Samples were considered as positive when the galactomannan index (GMI) was  $\geq$  0.5 (De Pauw et al., 2008).

In parallel, an immunoelectrophoretic assay for *Histoplasma*-specific antibody detection (IEP)
was performed according to manufacturer's instruction (Beckman Paragon, France). Briefly,
3μL of *Histoplasma* antigen (Laboratoire Méridian, France) were electrophoresed (20min,
100V; Beckman Paragon, France) on ready-to-use Hydragel-IEP-Plus gels (Sebia, France).
After a 24h-incubation with serum, the gel was washed, placed under a press, stained with
acid violet, washed again and finally dried out. The test was considered as positive if at least

- 22 one precipitation line was detected by visual observation.
- 23

#### 24 Analysis and Statistical Methods

25 Data were analyzed with SIGMA Stat software (2.03) using the Mann-Whitney rank sum test 26 for a two-group comparison and the  $\chi^2$  test for patient characteristic comparison. 27 Relationships between two variables were analyzed by Spearman rank order correlation test. 28 Values were reported as the median and interquartile range IQR [25%; 75%]. Analysis of 29 Receiver Operator Characteristic (ROC) curves was performed to determine the cut-off for 30 positivity. A comparison was considered statistically significant if the p value was  $\leq 0.05$ .

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# 33 **RESULTS** 34

### 35 **Patient Characteristics**

Between 2002 and 2009, 39 AIDS patients diagnosed for disseminated histoplasmosis and 36 37 with an available concomitant serum were included in the study (Table 1). There was no 38 significant difference in the median age or sex ratio between case and control patients. 39 Diagnosis of histoplasmosis was obtained mainly on hematology (48.7%) or digestive 40 (28.2%) samples. Thirty seven patients (94.9%) had a positive culture for Histoplasma capsulatum. For two patients, a rapid and extensive development of Candida albicans on the 41 42 culture did not allow the growth of Histoplasma but PCR and direct examination were both 43 positive, confirming the diagnosis of histoplasmosis. All the index case and control patients 44 were negative for the diagnosis of Aspergillus infection (culture and anti-Aspergillus antibody 45 detection).

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### 47 Anti-Histoplasma antibody detection

48 For AIDS-patients with disseminated histoplasmosis, the sensitivity (Se) of Histoplasma IEP

- 49 was only 35.9% [95% CI: 21.7-52.8] while the specificity (Sp) was 100% [95% CI: 85.9-100]
- 50 (Table 1). The sensitivity was linked to the TCD4+ lymphocyte level as the counts of these
- cells was statistically higher in patients with positive *Histoplasma*-specific antibody detection

than in patients with negative serology (26 [5; 37] *vs* 84 [77; 90]) (p=0.007; Mann-Whitney
rank sum test). On the contrary, there was no statistical difference for TCD8+ lymphocyte
levels (data not shown).

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## 5 GM detection for diagnosis of histoplasmosis

6 Galactomannan indexes (GMI) were significantly higher in HIV-positive patients with histoplasmosis compared to Histoplasma-uninfected ones (Table 1). With the recommended 7 threshold for invasive aspergillosis diagnosis (0.5), the sensitivity was 76.9% [95% 8 confidence intervals (95% CI): 60.3-88.3] and the specificity was 100% [95% CI: 85.9-100] 9 for histoplasmosis diagnosis. The TCD4+ or TCD8+ lymphocytes counts were not 10 statistically different between groups of patients with positive or negative GMI (data not 11 shown). Moreover, the GMI level was not correlated with TCD4+ or TCD8+ lymphocyte 12 counts in HIV-positive patients with histoplasmosis (data not shown). 13

14 The area under the curve was 0.963 on the ROC curve (Fig. 1). Two other thresholds (0.4 and

- 15 0.35) appeared to be potentially more interesting than the recommended threshold 0.5, with
- sensitivities of 82.1% [95% CI: 65.9-91.9] and 87.2% [95% CI: 71.8-95.1] and specificities of
   100% [95% CI: 85.9-100] and 93.3% [95% CI: 76.5-98.8] respectively. Positive likelihood
- ratios (LR+) were 13.08 [95% CI: 3.41-50.2],  $+\infty$  [95% CI: non-calculable],  $+\infty$  [95% CI: non-
- 19 calculable] for thresholds of 0.35; 0.4; 0.5 respectively and negative likelihood ratios (LR-)
- 20 were 0.14 [95% CI: 0.06-0.31], 0.18 [95% CI: 0.09-0.35] and 0.23 [95% CI: 0.13-0.41] for
- 21 thresholds of 0.35; 0.4; 0.5 respectively.

Taking account of this information, the threshold 0.4 seems to be the most relevant (Se: 82.1% [95% CI: 65.9-91.9], Sp: 100% [95% CI: 85.9-100], LR+:  $+\infty$  [95% CI: noncalculable], LR-: 0.18 [95% CI: 0.09-0.35]). Coupling galactomanann and anti-*Histoplasma* antibody detection did not significantly improve the overall diagnosis performance (Se: 82.1% [95% CI: 65.9-91.9], Sp: 100% [95% CI: 85.9-100].

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#### 29 30 DISCUSSION

31 In areas outside the USA where Histoplasma EIA is not available, the diagnosis of 32 disseminated histoplasmosis is often difficult to obtain in a timely manner consistent with the 33 life-threatening character of this disease: culture takes a long time, direct examination is of 34 poor sensitivity and *Histoplasma*-specific PCR is not available in most centers. As shown by others (Tobon et al., 2005; Wheat, 2006), we confirmed that detection of anti-Histoplasma 35 specific antibodies had a low sensitivity (35.9%) in AIDS patients. It is now well-established 36 that CD4+T-cell depletion due to HIV was responsible for an IL-7-dependant alteration of B-37 lymphocyte responses (Moir and Fauci, 2009). This phenomenon could explain the CD4+T 38 39 cell-dependent decrease in anti-Histoplasma antibody production observed in our study among AIDS patients with histoplasmosis. 40 In contrast, we found that the GM detection performed well (Se: 76.9%, Sp: 100%). However, 41

42 the threshold used for Aspergillus (0.5) could be lowered to 0.4 to increase the sensitivity (Se 43 82.1%) without decreasing the specificity. Keeping the recommended threshold for aspergillosis diagnosis (0.5) would lead to a decrease in sensitivity of 5.2%. Unlike antibody 44 45 levels, GMI were not influenced by TCD4+ lymphocyte counts. For GM detection, Ranque et al. already reported a sensitivity of about 73% in 11 patients with pulmonary histoplasmosis 46 and 100% in 6 HIV-positive patients (Rangue et al., 2007). Thus, in Histoplasma endemic 47 areas where Histoplasma EIA is not available, this quick and easy-to-perform technique might 48 be a powerful alternative for the diagnosis of disseminated histoplasmosis in HIV-positive 49 50 patients.

In two other studies (Wheat et al., 2007; Xavier et al., 2009), the sensitivity of the GM 1 detection was about 48% and 67% in histoplasmosis diagnosis but these studies were not only 2 3 limited to AIDS patients and the clinical presentation was not specified (e.g., fungemia versus non-disseminated disease). As the GM detection cross-reactivity seems to occur with high 4 levels of Histoplasma antigens (Wheat et al., 2007), the sensitivity could be better in AIDS 5 6 patients with disseminated disease because of a high Histoplasma burden (Ranque et al., 7 2007). For this reason, it seems to be essential to reserve the GM detection for disseminated histoplasmosis diagnosis in AIDS patients on the basis of epidemiological, clinical, and 8 laboratory arguments. Moreover, as the incidence of invasive aspergillosis in HIV-positive 9 patients is generally <0.5% (Tong et al., 2009), the risk of misdiagnosis with aspergillosis is 10 low. The positivity of the GM detection seems to be also very useful in the diagnosis and the 11 monitoring of African histoplasmosis due to Histoplasma capsulatum var duboisii (Therby et 12 al., 2014). Similarly, the cross-reactivity of the GM detection exists with Cryptocococcus 13 (Dalle et al., 2005) but the incidence of this infection was lower in French Guiana with about 14 15 0.25 per 100 HIV/AIDS patients-years (Debourgogne et al., 2011). Nevertheless, an important limitation of this test concerns its significant cost. Moreover, the test requires a large number 16 17 of controls which does not make its use consistent with small series. In South America, where histoplasmosis should be considered as neglected disease, the price of this test does not easily 18 19 allow its use outside the rich countries. GM detection could also be very helpful for histoplasmosis diagnosis outside endemic areas 20 in HIV-positive travellers but the PPV and NPV should be reconsidered because of a lower 21

22 prevalence of the disease in this context. However, the contribution of this test should be

23 lower for histoplasmosis diagnosis in immunocompetent patients, especially compared to the

- detection of specific anti-*Histoplasma* antibodies that immunocompetent patients are able to synthesize.
- The results of this study are somewhat limited by the retrospective design and the size of the cohort even if, to our knowledge, this is the largest study concerning GM detection specifically performed on AIDS patients with disseminated histoplasmosis. Moreover, the impact of serum storage at -20°C is unknown on the performance of the GM detection test. However, it is usually believed that long-term storage may rather decrease galactomannan levels (Aquino et al., 2007) which would imply a higher sensitivity with fresh serum.

In conclusion, this study confirms that GM detection can be very helpful for the diagnosis of
 disseminated histoplasmosis in AIDS patients, particularly in endemic areas where
 *Histoplasma* EIA is not available.

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#### **TABLES**

## Table 1: Patients baseline characteristics, galactomannan and Histoplasma-specific antibody detection in AIDS patients with or without histoplasmosis 3

	AIDS patients		
	with histoplasmosis	without histoplasmosis	P value
Total, No.	39	30	
Age, median IQR [25%;75%], years	<b>43</b> [38;50]	<b>46</b> [37;53]	0.565 <sup>a</sup>
Male / Female patients, No.	23/16	20/10	0.513 <sup>b</sup>
Samples for histoplasmosis diagnosis, No. (%):			
- Digestive (colon, liver, esophagus biopsies)	11 (28.2%)	-	-
- Pulmonary (BAL, bronchial aspiration)	<b>6</b> (15.4%)	-	-
- Cerebral (CSF)	1 (2.6%)	-	-
- Haematologic (Blood, Bone marrow)	<b>19</b> (48.7%)	-	-
- Others (ganglion biopsy)	<b>2</b> (5.1%)	-	-
Histoplasmosis diagnosis method, No./total available data (%):			
- Direct examination	<b>15/37</b> (40.5%)	-	-
- Culture	<b>37/39</b> (94.9%)	-	-
- Specific PCR	<b>19/20</b> (95.0%)	-	-
Patients with positive specific antibodies, No. (%):	14 (35.9%)	<b>0</b> (0.0%)	< <b>0.001</b> <sup>b</sup>
Patients with positive GMI $\ge 0.5$ (GM detection), No. (%):	<b>30</b> (76.9%)	<b>0</b> (0.0%)	< <b>0.001</b> <sup>b</sup>
GMI (GM detection), median IQR [25%;75%]	<b>1.38</b> [0.52;4.8]	<b>0.12</b> [0.08;0.21]	<b>&lt;0.001</b> <sup>a</sup>
Patients with positive GMI (GMI $\ge$ 0.5) or antibodies, No. (%):	<b>32</b> (82.1%)	<b>0</b> (0.0%)	<b>&lt;0.001</b> <sup>b</sup>

 $^a$  calculated by Mann-Whitney rank sum test;  $^b$  calculated by  $\chi^2$  test BAL: broncho-alveolar lavage, CSF: Cerebro-spinal fluid

#### **FIGURE LEGEND**

#### Fig 1: Performance of GM detection according to the threshold

Receiver Operating Characteristics (ROC) curve for determination of cut-off and assay 

sensitivity (Se) and specificity (Sp). The thresholds 0.35, 0.4 and 0.5 are shown on the ROC

curve with a triangle, circle and square, respectively.

