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1 **Mesenchymal stem cells in regenerative medicine applied to**
2 **rheumatic diseases: role of secretome and exosomes**

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22

1 ABSTRACT

2 Over the last decades, mesenchymal stem cells (MSC) have been extensively studied with
3 regard to their potential applications in regenerative medicine. In rheumatic diseases, MSC-
4 based therapy is the subject of great expectations for patients who are refractory to proposed
5 treatments such as rheumatoid arthritis (RA), or display degenerative injuries without possible
6 curative treatment, such as osteoarthritis (OA). The therapeutic potential of MSCs has been
7 demonstrated in several pre-clinical models of OA or RA and both the safety and efficacy of
8 MSC-based therapy is being evaluated in humans. The predominant mechanism by which
9 MSCs participate to tissue repair is through a paracrine activity. Via the production of a
10 multitude of trophic factors with various properties, MSCs can reduce tissue injury, protect
11 tissue from further degradation and/or enhance tissue repair. However, a thorough *in vivo*
12 examination of MSC-derived secretome and strategies to modulate it are still lacking. The
13 present review discusses the current understanding of the MSC secretome as a therapeutic for
14 treatment of inflammatory or degenerative pathologies focusing on rheumatic diseases. We
15 provide insights on and perspectives for future development of the MSC secretome with
16 respect to the release of extracellular vesicles that would have certain advantages over
17 injection of living MSCs or administration of a single therapeutic factor or a combination of
18 factors.

19

1

2 *Highlights:*

3 - Mesenchymal stem cell-based therapy generates great interest for regenerative medicine

4 - Mesenchymal stem cells mainly participate to tissue repair via paracrine activity

5 - A multitude of trophic and survival factors are secreted by mesenchymal stem cells

6 - Extracellular vesicles exert similar protective and reparative properties as cells

7 - Extracellular vesicles represent a promising alternative to stem cell implantation

8

9

10 *Keywords:* mesenchymal stem cells, trophic factors, exosomes, microparticles, secretome,
11 regenerative medicine, rheumatic diseases

12

1 **1. Characteristics and functions of mesenchymal stromal cells**

2

3 Mesenchymal stem or stromal cells (MSCs) are multipotent adult stem cells capable of self-
4 renew and differentiation potential. They are found in large quantities in bone marrow (BM-
5 MSCs) or in adipose tissue (ASCs) but could reside in virtually all post-natal organs and
6 tissues [1]. MSCs are isolated as a heterogeneous cell population characterized by their
7 capacity to adhere to plastic, develop as fibroblast colony-forming-units (CFU-F) and
8 differentiate into three cell lineages of mesodermal origin: osteocytes, chondrocytes and
9 adipocytes. After culture expansion, they are positive for the cell surface markers CD73,
10 CD90 and CD105 and negative for CD11b, CD14, CD34, CD45 and human leukocyte antigen
11 (HLA)-DR [2]. While expanded BM-MSCs are negative for the CD34 marker, recent studies
12 report that freshly isolated BM-MSC are enriched in the CD34⁺ fraction of BM nucleated
13 cells [3]. Conversely, CD34 is expressed on native ASCs and during the first population
14 doublings but rapidly disappears upon cell proliferation *in vitro* [4, 5].

15 MSCs produce a large amount of secreted factors, such as cytokines, chemokines or growth
16 factors, which mediate diverse functions via a crosstalk between different cell types [6-8]. In
17 the BM niche, MSCs and osteoblasts constitute the stromal fraction in a complex network
18 formed by hematopoietic stem cells (HSCs), endothelial stem cells and their progeny. Within
19 the niche, MSCs control survival, proliferation and differentiation of stem cells. They also
20 play a role in tissue regeneration either locally or over large distances through the secretion of
21 trophic factors. These soluble mediators may act directly, triggering intracellular mechanisms
22 of injured cells, or indirectly, inducing secretion of functionally active mediators by
23 neighboring cells. Indeed, in case of injury, MSCs attenuate tissue damage, inhibit fibrotic
24 remodeling and apoptosis, promote angiogenesis, stimulate endogenous stem cell recruitment
25 and proliferation, and reduce immune responses (Figure 1).

26

1 **2. Choice of the best cell source for regenerative medicine**

2

3 BM-MSCs and ASCs are the best characterized and the most studied sources of adult MSCs.
4 However, new cell sources, in particular from the Wharton jelly, are also interesting for
5 therapeutic applications [1]. Thanks to their differentiation properties, their use in
6 regenerative medicine has been first tested in tissue engineering applications, for bone and
7 cartilage repair. These approaches require defining an optimal combination of scaffold,
8 growth factor and stem cells and, local delivery requiring surgical procedures [9]. More
9 recently, the capacity of MSCs to secrete a variety of trophic factors with diverse functions
10 has motivated the interest of evaluating local or systemic injection of MSCs to stimulate
11 tissue repair in different pathologies. However, the question of the best source of cells for a
12 particular therapeutic application is under evaluation.

13 Significant differences between BM-MSCs and ASCs have been reported. The cytokine
14 profile of ASCs and BM-MSCs differs [10]. ASCs secrete higher levels of pro-angiogenic
15 factors including vascular endothelial growth factor (VEGF), hepatocyte growth factor
16 (HGF), basic fibroblast growth factor (bFGF), angiopoietin (Ang)-1, Ang-2 or platelet derived
17 growth factor (PDGF) to promote angiogenesis [11-13]. Together with angiogenic activity
18 that makes them attractive cells for cardiovascular diseases, ASCs also represent a powerful
19 tool for neurodegenerative medicine. ASCs can stimulate the regeneration of nervous tissues
20 by promoting nerve healing and *de novo* axon growth, via the release of several neurotrophic
21 factors such as brain derived neurotrophic factor (BDNF), nerve growth factor (NGF) or glial
22 derived neurotrophic factor (GDNF) [14]. They protect neurons against apoptosis [15] and
23 slowed disease progression in models of Huntington disease [16].

24 Regarding the immunomodulation function of stem cells, a first report concluded that ASCs
25 share immunosuppressive properties with BM-MSCs [17]. Since then, the
26 immunomodulatory function of BM-MSCs and ASCs was compared with that of chorionic

1 placenta- or palatine tonsil-derived MSCs (CP-MSCs) and even some differences exist,
2 comparable effects were observed [18, 19].

3 ASCs and MSCs exhibit cell specific differences at transcriptional and proteomic levels as
4 well as functional differences in their differentiation processes towards adipocytes,
5 osteoblasts and chondrocytes [20]. MSCs demonstrate higher differentiation potential toward
6 chondrogenic and osteoblastic lineages whereas ASCs possess a better capacity to
7 differentiate into adipocytes [21, 22]. The best strategy for MSC-based therapy has therefore
8 to be determined according to their distinct characteristics associated with their tissue origin
9 for a particular therapeutic application.

10

11 **3. Secretome-based therapeutic efficacy of mesenchymal stem cells for rheumatic** 12 **diseases**

13

14 The therapeutic value of BM-MSCs or ASCs for rheumatic diseases including osteoarthritis
15 (OA) and rheumatoid arthritis (RA) has been evaluated during the last few years. Because the
16 interest of using MSCs for cartilage tissue engineering has been reviewed elsewhere [9], we
17 will focus here on the paracrine effect of MSCs for preventing cartilage degradation or
18 stimulating endogenous cartilage regeneration. Focusing on rheumatic diseases, it is likely
19 that the route of MSC administration will differ according to the pathology. In case of
20 systemic disease, such as rheumatoid arthritis (RA) where several joints may be affected,
21 systemic delivery via the bloodstream should be favored. On the contrary, for lesions that are
22 limited to a single joint, local delivery should be preferred because of better availability of
23 cells and safety.

24

25 ***3.1. Local delivery of mesenchymal stem cells for osteoarthritis treatment***

26

1 The rationale for using local injection of MSCs for inducing regeneration of OA cartilage is
2 based on a number of *in vitro* studies, when MSCs and chondrocytes are mixed in pellet- or
3 alginate-based co-cultures [23-25]. Whatever the source of MSCs (BM, adipose tissue or
4 synovium), factors secreted by MSCs increased cartilage matrix production by chondrocytes
5 [25]. However, neither the exact mechanism of action when ASCs or BM-MSCs are not in
6 direct contact with chondrocytes, nor the identification of possible mediators, had been
7 investigated. Such paracrine effect was recently demonstrated in our group, where proteins
8 secreted by ASCs were shown to protect OA chondrocytes against apoptosis and degeneration
9 towards hypertrophic or fibrotic phenotypes; HGF being involved in the anti-fibrotic effect
10 observed (Maumus et al., submitted). Although OA is not considered an inflammatory
11 disease, pro-inflammatory mediators, such as cytokines, metalloproteinases (MMP), reactive
12 oxygen species (ROS), are secreted by OA chondrocytes or synoviocytes and, participate to
13 joint tissue alterations. Several pro-inflammatory cytokines are significantly down-regulated
14 in chondrocytes when cultured with ASCs suggesting that ASCs may also be protective
15 through the down-regulation of inflammatory mediators [26]. Interestingly, paracrine factors
16 of BM-MSCs share the same anti-inflammatory effects on OA cartilage and synovial explants
17 *in vitro* [27].

18 Local injection of BM-MSCs or ASCs in the joint is likely to exert several roles: inhibition of
19 osteophyte formation, decrease of synovial inflammation, reduction of cartilage degeneration
20 with less fibrosis and apoptosis of chondrocytes or stimulation of chondrocyte proliferation
21 and extracellular matrix synthesis (Figure 2). Indeed, using the pre-clinical murine model of
22 collagenase-induced OA, a single ASC injection in the knee joint of mice inhibited synovial
23 activation and formation of chondrophyte/osteophyte in joint ligaments as well as cartilage
24 destruction, probably by suppressing synovial macrophage activation [28]. Intra-articular
25 injection of BM-MSCs can also prevent the development of post-traumatic arthritis [29].
26 Other pre-clinical studies using larger animal models of OA (rat, rabbit, guinea pig, sheep,

1 donkey and goat) revealed similar results with cartilage regeneration after injection of MSCs
2 in the damaged joint [30-35].
3 Finally, an Iranian phase I clinical trial recently reported that intra-articular injection of
4 autologous BM-MSCs in six patients with knee OA was safe and improved pain, functional
5 status of the knee. As important, magnetic resonance imaging (MRI) displayed increased
6 cartilage thickness and decrease of subchondral edemas in three out of six patients [36]. All
7 these data support the trophic action of MSCs for protecting cartilage from degradation and
8 stimulating regeneration.

9

10 ***3.2. Systemic delivery of mesenchymal stem cells for rheumatoid arthritis treatment***

11

12 The interest of using MSCs to reduce inflammation in various autoimmune and/or
13 inflammatory disorders has been investigated for many years (for review, see [37]). In the
14 collagen-induced arthritis (CIA) murine model, which is representative of RA in humans,
15 contrasted results have been reported (for reviews, see [38, 39]). Injection of primary murine
16 BM-MSCs was shown to inhibit occurrence of arthritis and even partially reverse clinical
17 signs when injected after disease onset [40, 41]. Besides reduced levels of pro-inflammatory
18 cytokines in mouse sera, mechanisms involved in reduction of clinical signs were suggested
19 to be through $CD4^+CD25^+Foxp3^+$ Treg cell induction as well as T-cell anergy [42-45]. The
20 therapeutic benefit of xenogeneic human MSCs, either from adipose tissue or umbilical cord,
21 has also been described [42, 46, 47]. In contrast, other studies failed to demonstrate any
22 improvement with MSC treatment. Systemic infusion of the allogeneic C3H10T1/2 cell line
23 did not decrease the clinical signs of arthritis [48]. Similar results were obtained using
24 primary murine MSCs isolated from different strains of mice suggesting that different genetic
25 backgrounds influence the immunosuppressive effect of BM-MSCs [49]. Alternatively, the
26 immunomodulatory role of BM-MSCs was reported to be dependent on the window of

1 injection, with therapeutic benefit only when two cell injections on day 18 and 24 were done
2 [41]. More recently, inhibition of TNF- α via infusion of a specific inhibitor resulted in
3 enhanced suppressive activity of MSCs, confirming previous report that exposure of MSCs to
4 TNF- α blocks their suppressive capacity [48, 50]. This hypothesis was further supported by
5 enhanced immunomodulatory activity of BM-MSCs when the anti-inflammatory Bortezomib,
6 a proteasome inhibitor, was injected before MSC infusion [51].
7 Despite these conflicting results, the safety and efficacy of allogeneic transplantation of
8 MSCs from BM or umbilical cord have been tested in four patients with refractory RA [52].
9 No serious adverse events were reported but no patient achieved the DAS-28-defined
10 remission in the follow-up period. Nevertheless, larger randomized studies are required to
11 address the interest of using MSCs for RA treatment.

12

13 **4. Role of extracellular vesicles released by mesenchymal stem cells in the treatment of** 14 **degenerative diseases**

15

16 *4.1. Biogenesis and characterization of extracellular vesicles*

17

18 Extracellular vesicles are released in the extracellular space by almost all cells and were
19 considered for long time, as inert cellular debris. They form a variety of complex structures
20 among which exosomes, microparticles (MP) and apoptotic bodies are the best described
21 vesicles. They are surrounded by a phospholipid bilayer and can be distinguished by their size
22 and composition. Exosome size ranges between 30 and 120 nm whereas MP diameter is
23 included between 100 nm and 1 μ m and apoptotic bodies between 1 and 5 μ m. They contain
24 numerous proteins, lipids as well as messenger and micro RNAs responsible for intercellular
25 signaling. Exosomes were first described 30 years ago as being released by sheep
26 reticulocytes [53] and then, by most cell types including immune cells (B cells, dendritic

1 cells, mast cells or T cells) [54, 55], cancer cells [56] and MSCs [57]. They are also found in
2 physiological fluids such urine, plasma or exsudates [58].
3 Exosomes originate from internal bud of multivesicular endosomes which fuse with the
4 plasma membrane and are released by exocytosis [59]. They are rich in tetraspanins (CD9,
5 CD63 and CD81), heat-shock proteins (Hsp60, Hsp70, Hsp90) and frequently expose Alix,
6 clathrin, Tsg101 and unique cell type specific proteins that reflect their cellular source. MPs
7 originate from the budding of small cytoplasmic protrusions which then detached from the
8 cell surface through a process dependent on calcium influx, calpain and cytoskeleton
9 reorganization. They expose high amounts of phosphatidylserine, contain protein associated
10 with lipid rafts and are enriched in cholesterol, sphingomyelin and ceramide [60].
11 Extracellular vesicles from MSCs additionally express the characteristic markers CD13,
12 CD29, CD44, CD73 and CD105 [61-63] and contain proteins, mRNA and microRNA which
13 have been characterized by proteomic or transcriptomic analyses [64, 65].

14

15 ***4.2. Isolation of extracellular vesicles and characterization of proteins***

16

17 . Most of the protocols rely on the purification of vesicles from supernatants of cells grown in
18 absence of serum [66]. The purification of the exosomal fraction for molecular and functional
19 analyses relies on three different methods which are most frequently used: ultracentrifugation
20 [67], ultrafiltration [68] and immunoprecipitation technologies using antibody loaded
21 magnetic cell beads [69]. Extracellular vesicles can then be efficiently separated from protein
22 aggregates by using their low buoyant velocity and differences in floatation velocity [70].
23 However, the methods for purification and analysis of the different extracellular vesicle
24 populations have to be improved and standardized

25 A number of biochemical techniques have been used to identify the protein, RNA species and
26 lipid content of vesicles (for review, see [71]). During the last years, western blot,

1 fluorescence-activated cell sorting or MS-based proteomic analyses have been performed.
2 More recently, high-throughput studies have been conducted and to date, several thousands of
3 proteins and RNAs have been described in extracellular vesicles purified from various cell
4 types or biological fluids. These studies allowed the identification of a common set of
5 components, mainly associated with the biogenesis or structure of vesicles or, proteins
6 specific for the cell origin or physiopathological status. Quantitative and comparative analyses
7 are still needed to a better understanding the function and role of the extracellular vesicles.

8

9 ***4.3. Biologic function of extracellular vesicles released by mesenchymal stem cells***

10

11 Extracellular vesicles are an integral component of the cell-to-cell crosstalk contributing to
12 tissue regeneration and likely taking part to the paracrine action of MSCs in regenerative
13 medicine. The paracrine role of MSCs is supported by the beneficial effect of conditioned
14 media (CM) that reproduce benefits reported with the direct injection of MSCs as exemplified
15 for myocardial infarction therapy in swine and hamster models [57, 72-75]. The
16 cardioprotective activity was contained in a >1000 kDa MW fraction and therefore mediated
17 by large complexes with a diameter of 50-100 nm [74]. In addition, extracellular vesicles
18 from MSCs can promote angiogenesis by increasing endothelial cell proliferation and
19 capillary network formation both *in vitro* and *in vivo* [76]. They can improve acute kidney
20 injury by decreasing apoptosis, fibrosis, lymphocyte infiltration, tubular atrophy and by
21 increasing tubular epithelial cell proliferation [63, 77-79]. In some reported cases,
22 pretreatment of extracellular vesicles with RNase abrogated their therapeutic properties
23 highlighting the important role of RNA species [77, 78]. The anti-fibrotic action of MSC-
24 derived exosomes was also shown on liver by the reduction of collagen I and III deposit as
25 well as TGF- β 1 expression and Smad2 phosphorylation leading to the inhibition of epithelial-
26 to-mesenchymal transition and protection of hepatocytes [61]. Finally, extracellular vesicles

1 released by MSCs were shown to inhibit auto-reactive lymphocyte proliferation and promote
2 secretion of the anti-inflammatory cytokines IL-10 and TGF- β [80]. While the role of MSC-
3 derived extracellular vesicles has not been addressed in rheumatic diseases, it is tempting to
4 speculate that they may improve outcomes of OA or RA via anti-fibrotic, anti-apoptotic, anti-
5 inflammatory and pro-regenerative properties.

6

7 **5. Conclusions**

8

9 Regenerative medicine is a subject of great expectations and gives rise to enormous hopes for
10 patients who are refractory to proposed treatments, or display severe forms of diseases
11 without possible treatment. MSC-based therapy might therefore be an advantageous
12 alternative to current approaches. In the last decade, the therapeutic potential of MSCs has
13 been demonstrated in numerous pre-clinical models of inflammatory and degenerative
14 pathologies and MSC-based therapy is being evaluated in clinics with promising results.
15 While MSCs are considered relatively safe, the development of therapeutic strategies that
16 may avoid administration of MSCs will attenuate the safety concerns relative to the use of
17 living stem cells. In this respect, extracellular vesicles would have certain advantages over
18 administration of a single factor that cannot mimic the actions of MSCs. Several questions
19 have however to be addressed before clinical use could be considered.

20

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22

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3

4 **References**

5

- 6 [1] L. da Silva Meirelles, P. C. Chagastelles, N. B. Nardi. Mesenchymal stem cells reside
7 in virtually all post-natal organs and tissues, *J Cell Sci* 119 (2006) 2204-13.
- 8 [2] M. Dominici, K. Le Blanc, I. Mueller, I. Slaper-Cortenbach, F. Marini, D. Krause, R.
9 Deans, A. Keating, D. Prockop, E. Horwitz. Minimal criteria for defining multipotent
10 mesenchymal stromal cells. The International Society for Cellular Therapy position
11 statement, *Cytotherapy* 8 (2006) 315-7.
- 12 [3] C. S. Lin, H. Ning, G. Lin, T. F. Lue. Is CD34 truly a negative marker for
13 mesenchymal stromal cells?, *Cytotherapy* 14 (2012) 1159-63.
- 14 [4] M. Maumus, J. A. Peyrafitte, R. D'Angelo, C. Fournier-Wirth, A. Bouloumie, L.
15 Casteilla, C. Sengenès, P. Bourin. Native human adipose stromal cells: localization,
16 morphology and phenotype, *Int J Obes (Lond)* 35 (2011) 1141-53.
- 17 [5] C. Sengenès, K. Lolmede, A. Zakaroff-Girard, R. Busse, A. Bouloumie. Preadipocytes
18 in the human subcutaneous adipose tissue display distinct features from the adult
19 mesenchymal and hematopoietic stem cells, *J Cell Physiol* 205 (2005) 114-22.
- 20 [6] S. Meirelles Lda, A. M. Fontes, D. T. Covas, A. I. Caplan. Mechanisms involved in
21 the therapeutic properties of mesenchymal stem cells, *Cytokine Growth Factor Rev* 20
22 (2009) 419-27.
- 23 [7] A. J. Salgado, R. L. Reis, N. J. Sousa, J. M. Gimble. Adipose tissue derived stem cells
24 secretome: soluble factors and their roles in regenerative medicine, *Curr Stem Cell
25 Res Ther* 5 (2010) 103-10.
- 26 [8] J. Doorn, G. Moll, K. Le Blanc, C. van Blitterswijk, J. de Boer. Therapeutic
27 applications of mesenchymal stromal cells: paracrine effects and potential
28 improvements, *Tissue Eng Part B Rev* 18 (2012) 101-15.
- 29 [9] C. Vinatier, C. Bouffi, C. Merceron, J. Gordeladze, J.-M. Brondello, C. Jorgensen, P.
30 Weiss, J. Guicheux, D. Noel. Cartilage engineering: Towards a biomaterial-assisted
31 mesenchymal stem cell therapy, *Curr Stem Cell Res Ther* 4 (2009) 318-329.
- 32 [10] H. Skalnikova, J. Motlik, S. J. Gadher, H. Kovarova. Mapping of the secretome of
33 primary isolates of mammalian cells, stem cells and derived cell lines, *Proteomics* 11
34 (2011) 691-708.
- 35 [11] C. Nakanishi, N. Nagaya, S. Ohnishi, K. Yamahara, S. Takabatake, T. Konno, K.
36 Hayashi, M. A. Kawashiri, T. Tsubokawa, M. Yamagishi. Gene and protein
37 expression analysis of mesenchymal stem cells derived from rat adipose tissue and
38 bone marrow, *Circ J* 75 (2011) 2260-8.
- 39 [12] S. T. Hsiao, A. Asgari, Z. Lokmic, R. Sinclair, G. J. Dusing, S. Y. Lim, R. J. Dillely.
40 Comparative Analysis of Paracrine Factor Expression in Human Adult Mesenchymal
41 Stem Cells Derived from Bone Marrow, Adipose, and Dermal Tissue, *Stem Cells Dev*
42 21 (2012) 2189-2203.
- 43 [13] S. Kachgal, A. J. Putnam. Mesenchymal stem cells from adipose and bone marrow
44 promote angiogenesis via distinct cytokine and protease expression mechanisms,
45 *Angiogenesis* 14 (2011) 47-59.

- 1 [14] T. Lopatina, N. Kalinina, M. Karagyaur, D. Stambolsky, K. Rubina, A. Revischin, G.
2 Pavlova, Y. Parfyonova, V. Tkachuk. Adipose-derived stem cells stimulate
3 regeneration of peripheral nerves: BDNF secreted by these cells promotes nerve
4 healing and axon growth de novo, *PLoS One* 6 (2011) e17899.
- 5 [15] X. Wei, L. Zhao, J. Zhong, H. Gu, D. Feng, B. H. Johnstone, K. L. March, M. R.
6 Farlow, Y. Du. Adipose stromal cells-secreted neuroprotective media against neuronal
7 apoptosis, *Neurosci Lett* 462 (2009) 76-9.
- 8 [16] S. T. Lee, K. Chu, K. H. Jung, W. S. Im, J. E. Park, H. C. Lim, C. H. Won, S. H. Shin,
9 S. K. Lee, M. Kim, J. K. Roh. Slowed progression in models of Huntington disease by
10 adipose stem cell transplantation, *Ann Neurol* 66 (2009) 671-81.
- 11 [17] B. Puissant, C. Barreau, P. Bourin, C. Clavel, J. Corre, C. Bousquet, C. Taureau, B.
12 Cousin, M. Abbal, P. Laharrague, L. Penicaud, L. Casteilla, A. Blancher.
13 Immunomodulatory effect of human adipose tissue-derived adult stem cells:
14 comparison with bone marrow mesenchymal stem cells, *Br J Haematol* 129 (2005)
15 118-29.
- 16 [18] S. Janjanin, F. Djouad, R. M. Shanti, D. Baksh, K. Gollapudi, D. Prgomet, L.
17 Rackwitz, A. S. Joshi, R. S. Tuan. Human palatine tonsil: a new potential tissue source
18 of multipotent mesenchymal progenitor cells, *Arthritis Res Ther* 10 (2008) R83.
- 19 [19] J. M. Lee, J. Jung, H. J. Lee, S. J. Jeong, K. J. Cho, S. G. Hwang, G. J. Kim.
20 Comparison of immunomodulatory effects of placenta mesenchymal stem cells with
21 bone marrow and adipose mesenchymal stem cells, *Int Immunopharmacol* 13 (2012)
22 219-24.
- 23 [20] D. Noel, D. Caton, S. Roche, C. Bony, S. Lehmann, L. Casteilla, C. Jorgensen, B.
24 Cousin. Cell specific differences between human adipose-derived and mesenchymal-
25 stromal cells despite similar differentiation potentials, *Exp Cell Res* 314 (2008) 1575-
26 84.
- 27 [21] J. I. Huang, N. Kazmi, M. M. Durbhakula, T. M. Hering, J. U. Yoo, B. Johnstone.
28 Chondrogenic potential of progenitor cells derived from human bone marrow and
29 adipose tissue: a patient-matched comparison, *J Orthop Res* 23 (2005) 1383-9.
- 30 [22] R. Vishnubalaji, M. Al-Nbaheen, B. Kadalmani, A. Aldahmash, T. Ramesh.
31 Comparative investigation of the differentiation capability of bone-marrow- and
32 adipose-derived mesenchymal stem cells by qualitative and quantitative analysis, *Cell*
33 *Tissue Res* 347 (2012) 419-427.
- 34 [23] X. T. Mo, S. C. Guo, H. Q. Xie, L. Deng, W. Zhi, Z. Xiang, X. Q. Li, Z. M. Yang.
35 Variations in the ratios of co-cultured mesenchymal stem cells and chondrocytes
36 regulate the expression of cartilaginous and osseous phenotype in alginate constructs,
37 *Bone* 45 (2009) 42-51.
- 38 [24] L. Wu, J. C. Leijten, N. Georgi, J. N. Post, C. A. van Blitterswijk, M. Karperien.
39 Trophic effects of mesenchymal stem cells increase chondrocyte proliferation and
40 matrix formation, *Tissue Eng Part A* 17 (2011) 1425-36.
- 41 [25] L. Wu, H. J. Prins, M. N. Helder, C. A. van Blitterswijk, M. Karperien. Trophic
42 Effects of Mesenchymal Stem Cells in Chondrocyte Co-Cultures are Independent of
43 Culture Conditions and Cell Sources, *Tissue Eng Part A* 18 (2012) 1542-51.
- 44 [26] C. Manfredini, E. Gabusi, M. Maumus, A. Piacentini, G. Filardo, J.-A. Peyrafiite, P.
45 Bourin, C. Jorgensen, A. Facchini, D. Noel, G. Lisignoli. ADIPOSE STROMAL
46 CELLS EXHERT ANTI-INFLAMMATORY EFFECTS ON CHONDROCYTES
47 AND SYNOVIOCYTES FROM OSTEOARTHRITIS PATIENTS via PGE2,
48 *Arthritis Rheum* (in press).
- 49 [27] G. M. van Buul, E. Villafuertes, P. K. Bos, J. H. Waarsing, N. Kops, R. Narcisi, H.
50 Weinans, J. A. Verhaar, M. R. Bernsen, G. J. van Osch. Mesenchymal stem cells

- 1 secrete factors that inhibit inflammatory processes in short-term osteoarthritic
2 synovium and cartilage explant culture, *Osteoarthritis Cartilage* 20 (2012) 1186-96.
- 3 [28] M. Ter Huurne, R. Schelbergen, R. Blattes, A. Blom, W. de Munter, L. C. Grevers, J.
4 Jeanson, D. Noel, L. Casteilla, C. Jorgensen, W. van den Berg, P. L. van Lent.
5 Antiinflammatory and chondroprotective effects of intraarticular injection of adipose-
6 derived stem cells in experimental osteoarthritis, *Arthritis Rheum* 64 (2012) 3604-13.
- 7 [29] B. O. Diekman, C. L. Wu, C. R. Louer, B. D. Furman, J. L. Huebner, V. B. Kraus, S.
8 A. Olson, F. Guilak. Intra-articular delivery of purified mesenchymal stem cells from
9 C57BL/6 or MRL/MpJ superhealer mice prevents post-traumatic arthritis, *Cell*
10 *Transplant* (2012).
- 11 [30] F. Toghraie, M. Razmkhah, M. A. Gholipour, Z. Faghih, N. Chenari, S. Torabi
12 Nezhad, S. Nazhvani Dehghani, A. Ghaderi. Scaffold-free adipose-derived stem cells
13 (ASCs) improve experimentally induced osteoarthritis in rabbits, *Arch Iran Med* 15
14 (2012) 495-9.
- 15 [31] M. Horie, H. Choi, R. H. Lee, R. L. Reger, J. Ylostalo, T. Muneta, I. Sekiya, D. J.
16 Prockop. Intra-articular injection of human mesenchymal stem cells (MSCs) promote
17 rat meniscal regeneration by being activated to express Indian hedgehog that enhances
18 expression of type II collagen, *Osteoarthritis Cartilage* 20 (2012) 1197-207.
- 19 [32] H. Al Faqeh, B. M. Nor Hamdan, H. C. Chen, B. S. Aminuddin, B. H. Ruszymah. The
20 potential of intra-articular injection of chondrogenic-induced bone marrow stem cells
21 to retard the progression of osteoarthritis in a sheep model, *Exp Gerontol* 47 (2012)
22 458-64.
- 23 [33] M. Sato, K. Uchida, H. Nakajima, T. Miyazaki, A. R. Guerrero, S. Watanabe, S.
24 Roberts, H. Baba. Direct transplantation of mesenchymal stem cells into the knee
25 joints of Hartley strain guinea pigs with spontaneous osteoarthritis, *Arthritis Res Ther*
26 14 (2012) R31.
- 27 [34] A. N. Mokbel, O. S. El Tookhy, A. A. Shamaa, L. A. Rashed, D. Sabry, A. M. El
28 Sayed. Homing and reparative effect of intra-articular injection of autologous
29 mesenchymal stem cells in osteoarthritic animal model, *BMC Musculoskelet Disord*
30 12 (2011) 259.
- 31 [35] J. M. Murphy, D. J. Fink, E. B. Hunziker, F. P. Barry. Stem cell therapy in a caprine
32 model of osteoarthritis, *Arthritis Rheum* 48 (2003) 3464-74.
- 33 [36] M. Emadedin, N. Aghdami, L. Taghiyar, R. Fazeli, R. Moghadasali, S. Jahangir, R.
34 Farjad, M. Baghaban Eslaminejad. Intra-articular injection of autologous
35 mesenchymal stem cells in six patients with knee osteoarthritis, *Arch Iran Med* 15
36 (2012) 422-8.
- 37 [37] S. Ghannam, C. Bouffi, F. Djouad, C. Jorgensen, D. Noel. Immunosuppression by
38 mesenchymal stem cells: mechanisms and clinical applications, *Stem Cell Res Ther* 1
39 (2010) 2.
- 40 [38] E. Schurgers, H. Kelchtermans, T. Mitera, L. Geboes, P. Matthys. Discrepancy
41 between the in vitro and in vivo effects of murine mesenchymal stem cells on T-cell
42 proliferation and collagen-induced arthritis, *Arthritis Res Ther* 12 (2010) R31.
- 43 [39] G. I. MacDonald, A. Augello, C. De Bari. Role of mesenchymal stem cells in
44 reestablishing immunologic tolerance in autoimmune rheumatic diseases, *Arthritis*
45 *Rheum* 63 (2011) 2547-57.
- 46 [40] A. Augello, R. Tasso, S. M. Negrini, R. Cancedda, G. Pennesi. Cell therapy using
47 allogeneic bone marrow mesenchymal stem cells prevents tissue damage in collagen-
48 induced arthritis, *Arthritis Rheum* 56 (2007) 1175-86.
- 49 [41] C. Bouffi, C. Bony, G. Courties, C. Jorgensen, D. Noel. IL-6-dependent PGE2
50 secretion by mesenchymal stem cells inhibits local inflammation in experimental
51 arthritis, *PLoS One* 5 (2010) e14247.

- 1 [42] M. A. Gonzalez, E. Gonzalez-Rey, L. Rico, D. Buscher, M. Delgado. Adipose-derived
2 mesenchymal stem cells alleviate experimental colitis by inhibiting inflammatory and
3 autoimmune responses, *Gastroenterology* 136 (2009) 978-89.
- 4 [43] E. Zappia, S. Casazza, E. Pedemonte, F. Benvenuto, I. Bonanni, E. Gerdoni, D.
5 Giunti, A. Ceravolo, F. Cazzanti, F. Frassoni, G. Mancardi, A. Uccelli. Mesenchymal
6 stem cells ameliorate experimental autoimmune encephalomyelitis inducing T-cell
7 anergy, *Blood* 106 (2005) 1755-61.
- 8 [44] S. T. Roord, W. de Jager, L. Boon, N. Wulffraat, A. Martens, B. Prakken, F. van Wijk.
9 Autologous bone marrow transplantation in autoimmune arthritis restores immune
10 homeostasis through CD4⁺CD25⁺Foxp3⁺ regulatory T cells, *Blood* 111 (2008) 5233-
11 41.
- 12 [45] M. J. Park, H. S. Park, M. L. Cho, H. J. Oh, Y. G. Cho, S. Y. Min, B. H. Chung, J. W.
13 Lee, H. Y. Kim, S. G. Cho. TGF beta transduced mesenchymal stem cells ameliorate
14 autoimmune arthritis through reciprocal regulation of T(reg) -Th17 cells and
15 osteoclastogenesis, *Arthritis Rheum* 63 (2011) 1668-80.
- 16 [46] Y. Liu, R. Mu, S. Wang, L. Long, X. Liu, R. Li, J. Sun, J. Guo, X. Zhang, J. Guo, P.
17 Yu, C. Li, X. Liu, Z. Huang, D. Wang, H. Li, Z. Gu, B. Liu, Z. Li. Therapeutic
18 potential of human umbilical cord mesenchymal stem cells in the treatment of
19 rheumatoid arthritis, *Arthritis Res Ther* 12 (2010) R210.
- 20 [47] B. Zhou, J. Yuan, Y. Zhou, M. Ghawji, Jr., Y. P. Deng, A. J. Lee, A. J. Lee, U. Nair,
21 A. H. Kang, D. D. Brand, T. J. Yoo. Administering human adipose-derived
22 mesenchymal stem cells to prevent and treat experimental arthritis, *Clin Immunol* 141
23 (2011) 328-37.
- 24 [48] F. Djouad, V. Fritz, F. Apparailly, P. Louis-Pence, C. Bony, J. Sany, C. Jorgensen, D.
25 Noel. Reversal of the immunosuppressive properties of mesenchymal stem cells by
26 tumor necrosis factor alpha in collagen-induced arthritis, *Arthritis Rheum* 52 (2005)
27 1595-603.
- 28 [49] C. Sullivan, J. M. Murphy, M. D. Griffin, R. M. Porter, C. H. Evans, C. O'Flatharta,
29 G. Shaw, F. Barry. Genetic mismatch affects the immunosuppressive properties of
30 mesenchymal stem cells in vitro and their ability to influence the course of collagen-
31 induced arthritis, *Arthritis Res Ther* 14 (2012) R167.
- 32 [50] C. C. Wu, T. C. Wu, F. L. Liu, H. K. Sytwu, D. M. Chang. TNF-alpha inhibitor
33 reverse the effects of human umbilical cord-derived stem cells on experimental
34 arthritis by increasing immunosuppression, *Cell Immunol* 273 (2012) 30-40.
- 35 [51] A. Papadopoulou, M. Yiangou, E. Athanasiou, N. Zogas, P. Kaloyannidis, I. Batsis, A.
36 Fassas, A. Anagnostopoulos, E. Yannaki. Mesenchymal stem cells are conditionally
37 therapeutic in preclinical models of rheumatoid arthritis, *Ann Rheum Dis* 71 (2012)
38 1733-40.
- 39 [52] J. Liang, H. Zhang, B. Hua, H. Wang, L. Lu, S. Shi, Y. Hou, X. Zeng, G. S. Gilkeson,
40 L. Sun. Allogenic mesenchymal stem cells transplantation in refractory systemic lupus
41 erythematosus: a pilot clinical study, *Ann Rheum Dis* 69 (2012) 1423-9.
- 42 [53] B. T. Pan, R. M. Johnstone. Fate of the transferrin receptor during maturation of sheep
43 reticulocytes in vitro: selective externalization of the receptor, *Cell* 33 (1983) 967-78.
- 44 [54] G. Raposo, H. W. Nijman, W. Stoorvogel, R. Liejendekker, C. V. Harding, C. J.
45 Melief, H. J. Geuze. B lymphocytes secrete antigen-presenting vesicles, *J Exp Med*
46 183 (1996) 1161-72.
- 47 [55] L. Zitvogel, A. Regnault, A. Lozier, J. Wolfers, C. Flament, D. Tenza, P. Ricciardi-
48 Castagnoli, G. Raposo, S. Amigorena. Eradication of established murine tumors using
49 a novel cell-free vaccine: dendritic cell-derived exosomes, *Nat Med* 4 (1998) 594-600.
- 50 [56] J. Wolfers, A. Lozier, G. Raposo, A. Regnault, C. Thery, C. Masurier, C. Flament, S.
51 Pouzieux, F. Faure, T. Tursz, E. Angevin, S. Amigorena, L. Zitvogel. Tumor-derived

- 1 exosomes are a source of shared tumor rejection antigens for CTL cross-priming, *Nat*
2 *Med* 7 (2001) 297-303.
- 3 [57] R. C. Lai, T. S. Chen, S. K. Lim. Mesenchymal stem cell exosome: a novel stem cell-
4 based therapy for cardiovascular disease, *Regen Med* 6 (2011
5) 481-92.
- 6 [58] C. Tetta, S. Bruno, V. Fonsato, M. C. Deregibus, G. Camussi. The role of
7 microvesicles in tissue repair, *Organogenesis* 7 (2011) 105-15.
- 8 [59] S. Pant, H. Hilton, M. E. Burczynski. The multifaceted exosome: biogenesis, role in
9 normal and aberrant cellular function, and frontiers for pharmacological and
10 biomarker opportunities, *Biochem Pharmacol* 83 (2012) 1484-94.
- 11 [60] L. Biancone, S. Bruno, M. C. Deregibus, C. Tetta, G. Camussi. Therapeutic potential
12 of mesenchymal stem cell-derived microvesicles, *Nephrol Dial Transplant* 27 (2012)
13 3037-42.
- 14 [61] T. Li, Y. Yan, B. Wang, H. Qian, X. Zhang, L. Shen, M. Wang, Y. Zhou, W. Zhu, W.
15 Li, W. Xu. Exosomes Derived from Human Umbilical Cord Mesenchymal Stem Cells
16 Alleviate Liver Fibrosis, *Stem Cells Dev* (2012).
- 17 [62] W. Zhu, L. Huang, Y. Li, X. Zhang, J. Gu, Y. Yan, X. Xu, M. Wang, H. Qian, W. Xu.
18 Exosomes derived from human bone marrow mesenchymal stem cells promote tumor
19 growth in vivo, *Cancer Lett* 315 (2012) 28-37.
- 20 [63] S. Bruno, C. Grange, F. Collino, M. C. Deregibus, V. Cantaluppi, L. Biancone, C.
21 Tetta, G. Camussi. Microvesicles derived from mesenchymal stem cells enhance
22 survival in a lethal model of acute kidney injury, *PLoS One* 7 (2012) e33115.
- 23 [64] H. S. Kim, D. Y. Choi, S. J. Yun, S. M. Choi, J. W. Kang, J. W. Jung, D. Hwang, K.
24 P. Kim, D. W. Kim. Proteomic analysis of microvesicles derived from human
25 mesenchymal stem cells, *J Proteome Res* 11 (2012) 839-49.
- 26 [65] F. Collino, M. C. Deregibus, S. Bruno, L. Sterpone, G. Aghemo, L. Viltono, C. Tetta,
27 G. Camussi. Microvesicles derived from adult human bone marrow and tissue specific
28 mesenchymal stem cells shuttle selected pattern of miRNAs, *PLoS One* 5 (2010)
29 e11803.
- 30 [66] C. Thery, S. Amigorena, G. Raposo, A. Clayton. Isolation and characterization of
31 exosomes from cell culture supernatants and biological fluids, *Curr Protoc Cell Biol*
32 Chapter 3 (2006) Unit 3 22.
- 33 [67] R. M. Johnstone, M. Adam, J. R. Hammond, L. Orr, C. Turbide. Vesicle formation
34 during reticulocyte maturation. Association of plasma membrane activities with
35 released vesicles (exosomes), *J Biol Chem* 262 (1987) 9412-20.
- 36 [68] H. G. Lamparski, A. Metha-Damani, J. Y. Yao, S. Patel, D. H. Hsu, C. Ruegg, J. B. Le
37 Pecq. Production and characterization of clinical grade exosomes derived from
38 dendritic cells, *J Immunol Methods* 270 (2002) 211-26.
- 39 [69] A. Clayton, J. Court, H. Navabi, M. Adams, M. D. Mason, J. A. Hobot, G. R.
40 Newman, B. Jasani. Analysis of antigen presenting cell derived exosomes, based on
41 immuno-magnetic isolation and flow cytometry, *J Immunol Methods* 247 (2001) 163-
42 74.
- 43 [70] G. Raposo, W. Stoorvogel. Extracellular vesicles: exosomes, microvesicles, and
44 friends, *J Cell Biol* 200 (2013) 373-83.
- 45 [71] D. S. Choi, D. K. Kim, Y. K. Kim, Y. S. Gho. Proteomics, transcriptomics and
46 lipidomics of exosomes and ectosomes, *Proteomics* (2013).
- 47 [72] A. Shabbir, D. Zisa, H. Lin, M. Mastro, G. Roloff, G. Suzuki, T. Lee. Activation of
48 host tissue trophic factors through JAK-STAT3 signaling: a mechanism of
49 mesenchymal stem cell-mediated cardiac repair, *Am J Physiol Heart Circ Physiol* 299
50 (2010) H1428-38.

- 1 [73] A. Shabbir, D. Zisa, G. Suzuki, T. Lee. Heart failure therapy mediated by the trophic
2 activities of bone marrow mesenchymal stem cells: a noninvasive therapeutic regimen,
3 *Am J Physiol Heart Circ Physiol* 296 (2009) H1888-97.
- 4 [74] L. Timmers, S. K. Lim, F. Arslan, J. S. Armstrong, I. E. Hofer, P. A. Doevendans, J.
5 J. Piek, R. M. El Oakley, A. Choo, C. N. Lee, G. Pasterkamp, D. P. de Kleijn.
6 Reduction of myocardial infarct size by human mesenchymal stem cell conditioned
7 medium, *Stem Cell Res* 1 (2007) 129-37.
- 8 [75] L. Timmers, S. K. Lim, I. E. Hofer, F. Arslan, R. C. Lai, A. A. van Oorschot, M. J.
9 Goumans, C. Strijder, S. K. Sze, A. Choo, J. J. Piek, P. A. Doevendans, G.
10 Pasterkamp, D. P. de Kleijn. Human mesenchymal stem cell-conditioned medium
11 improves cardiac function following myocardial infarction, *Stem Cell Res* 6 (2011)
12 206-14.
- 13 [76] H. C. Zhang, X. B. Liu, S. Huang, X. Y. Bi, H. X. Wang, L. X. Xie, Y. Q. Wang, X.
14 F. Cao, J. Lv, F. J. Xiao, Y. Yang, Z. K. Guo. Microvesicles derived from human
15 umbilical cord mesenchymal stem cells stimulated by hypoxia promote angiogenesis
16 both in vitro and in vivo, *Stem Cells Dev* 21 (2012) 3289-97.
- 17 [77] L. A. Reis, F. T. Borges, M. J. Simoes, A. A. Borges, R. Sinigaglia-Coimbra, N.
18 Schor. Bone marrow-derived mesenchymal stem cells repaired but did not prevent
19 gentamicin-induced acute kidney injury through paracrine effects in rats, *PLoS One* 7
20 (2012) e44092.
- 21 [78] S. Gatti, S. Bruno, M. C. Deregibus, A. Sordi, V. Cantaluppi, C. Tetta, G. Camussi.
22 Microvesicles derived from human adult mesenchymal stem cells protect against
23 ischaemia-reperfusion-induced acute and chronic kidney injury, *Nephrol Dial*
24 *Transplant* 26 (2011) 1474-83.
- 25 [79] J. He, Y. Wang, S. Sun, M. Yu, C. Wang, X. Pei, B. Zhu, J. Wu, W. Zhao. Bone
26 marrow stem cells-derived microvesicles protect against renal injury in the mouse
27 remnant kidney model, *Nephrology (Carlton)* 17 (2012) 493-500.
- 28 [80] A. Mokarizadeh, N. Delirezh, A. Morshedi, G. Mosayebi, A. A. Farshid, K. Mardani.
29 Microvesicles derived from mesenchymal stem cells: potent organelles for induction
30 of tolerogenic signaling, *Immunol Lett* 147 (2012) 47-54.
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1 **Legends to figures**

2

3 **Graphical abstract.** Role of trophic factors released by mesenchymal stem cells (MSC) in
4 the treatment of rheumatic diseases: possible mechanisms.

5

6 **Fig. 1.** Role of various paracrine factors released by mesenchymal stem cells. Secreted factors
7 may exert different functions on cells via the release of different types of molecules
8 depending on the microenvironment. Angiopoietin (Ang), Basic fibroblast growth factor
9 (bFGF), brain derived neurotrophic factor (BDNF), chemokine ligand (CCL), chemokine (C-
10 X-C motif) ligand (CXCL), erythropoietin (EPO), glial cell line-neurotrophic factor (GDNF),
11 granulocyte macrophage-colony stimulating factor (GM-CSF), heme oxygenase (HO),
12 hepatocyte growth factor (HGF), human leucocyte antigen (HLA), indoleamine 2,3-
13 dioxygenase (IDO), insulin growth factor (IGF), interleukin (IL), keratinocyte growth factor
14 (KGF), leukemia inhibitory factor (LIF), human cathelicidin (LL37), monocyte
15 chemoattractant protein (MCP), metalloproteinase (MMP), nerve growth factor (NGF), nitric
16 oxide (NO), platelet derived growth factor (PDGF), prostaglandin (PGE), placental growth
17 factor (PIGF), stem cell factor (SCF), stromal cell-derived factor (SDF), tissue inhibitor of
18 metalloproteinase (TIMP), transforming growth factor (TGF), thrombopoietin (TPO), TNF- α -
19 stimulated gene/protein (TSG), vascular endothelial growth factor (VEGF)

20

21 **Fig. 2.** Schematic representation of mesenchymal stem cell (MSC)-based therapy after intra-
22 articular injection in the treatment of rheumatic diseases. Through the release of trophic
23 factors, MSCs may act to reduce fibrosis, apoptosis and/or hypertrophic phenotypes
24 associated with cartilage degeneration and osteophyte formation during osteoarthritis. They
25 may control synovial inflammation occurring during rheumatoid arthritis and/or induce the
26 proliferation of chondrocytes to stimulate regeneration of endogenous cartilage.