



HAL
open science

Unraveling complex interplay between heat shock factor 1 and 2 splicing isoforms.

Sylvain Lecomte, Léa Reverdy, Catherine Le Quément, Florent Le Masson, Axelle Amon, Pascale Le Goff, Denis Michel, Elisabeth Christians, Yves Le Dréan

► To cite this version:

Sylvain Lecomte, Léa Reverdy, Catherine Le Quément, Florent Le Masson, Axelle Amon, et al.. Unraveling complex interplay between heat shock factor 1 and 2 splicing isoforms.. PLoS ONE, 2013, 8 (2), pp.e56085. 10.1371/journal.pone.0056085 . inserm-00814784

HAL Id: inserm-00814784

<https://inserm.hal.science/inserm-00814784v1>

Submitted on 17 Apr 2013

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Unraveling Complex Interplay between Heat Shock Factor 1 and 2 Splicing Isoforms

Sylvain Lecomte¹, Léa Reverdy¹, Catherine Le Quément¹, Florent Le Masson^{2^{aa}}, Axelle Amon³, Pascale Le Goff¹, Denis Michel¹, Elisabeth Christians^{2^{ab}}, Yves Le Dréan^{1*}

1 Transcription, Environment and Cancer group, Institut de Recherche sur la Santé, l'Environnement et le Travail, Inserm U1085, Université de Rennes 1, Rennes, France, **2** Centre de Biologie du Développement, UMR CNRS 5547, Université de Toulouse 3, Toulouse, France, **3** UMR CNRS 6251, Institut de Physique de Rennes, Université de Rennes 1, Rennes, France

Abstract

Chaperone synthesis in response to proteotoxic stress is dependent on a family of transcription factors named heat shock factors (HSFs). The two main factors in this family, HSF1 and HSF2, are co-expressed in numerous tissues where they can interact and form heterotrimers in response to proteasome inhibition. HSF1 and HSF2 exhibit two alternative splicing isoforms, called α and β , which contribute to additional complexity in HSF transcriptional regulation, but remain poorly examined in the literature. In this work, we studied the transcriptional activity of HSF1 and HSF2 splicing isoforms transfected into immortalized Mouse Embryonic Fibroblasts (iMEFs) deleted for both *Hsf1* and *Hsf2*, under normal conditions and after proteasome inhibition. We found that HSF1 α is significantly more active than the β isoform after exposure to the proteasome inhibitor MG132. Furthermore, we clearly established that, while HSF2 had no transcriptional activity by itself, short β isoform of HSF2 exerts a negative role on HSF1 β -dependent *transactivation*. To further assess the impact of HSF2 β inhibition on HSF1 activity, we developed a mathematical modelling approach which revealed that the balance between each HSF isoform in the cell regulated the strength of the transcriptional response. Moreover, we found that cellular stress such as proteasome inhibition could regulate the splicing of *Hsf2* mRNA. All together, our results suggest that relative amounts of each HSF1 and HSF2 isoforms quantitatively determine the cellular level of the proteotoxic stress response.

Citation: Lecomte S, Reverdy L, Le Quément C, Le Masson F, Amon A, et al. (2013) Unraveling Complex Interplay between Heat Shock Factor 1 and 2 Splicing Isoforms. PLoS ONE 8(2): e56085. doi:10.1371/journal.pone.0056085

Editor: Brendan D. Price, Dana-Farber/Harvard Cancer Institute, United States of America

Received: July 28, 2012; **Accepted:** January 4, 2013; **Published:** February 13, 2013

Copyright: © 2013 Lecomte et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This study was supported by National Research Agency (ANR), France, under Grant No. 10-CESA-017-01 (BioREF project). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: yves.le-drean@univ-rennes1.fr

^{aa} Current address: Developmental Biology Program, Memorial Sloan-Kettering Cancer Center, New York, New York, United States of America

^{ab} Current address: Division of Cardiology, Department of Internal Medicine, University of Utah Health Sciences Center, Salt Lake City, Utah, United States of America

Introduction

Proteasome is a major protein complex responsible for regulated degradation of intracellular proteins, and its activity is modified in many disorders. For example, a decrease in proteasome activity is associated with neurodegenerative diseases, whereas an increase catalytic activity is associated with cancers [1]. Thus, proteasome is a prime target in cancer therapy and bortezomib was the first proteasome inhibitor authorized as anti-tumor agent in humans. In a previous work, we have shown that proteasome subunit expression was regulated by heat shock factors [2]. Heat shock factor 1 (HSF1) and heat shock factor 2 (HSF2) belong to the family of transcription factors, which are essential for the expression of heat shock proteins (Hsps) in response to protein insults. HSF1 is the main factor responsible for Hsp induction, which is abolished in HSF1 deficient cells or organism, and cannot be rescued by HSF2 alone [3]. Among proteotoxic stress, proteasome inhibition, but not heat shock, activates HSF2 [4]. Interestingly, it was shown that after treatment with MG132, a classical proteasome inhibitor, HSF1 and HSF2 can form heterotrimers and bind to DNA [5]. The exact role of such

heterotrimers is not yet fully understood, but it was proposed that HSF2 could act as a modulator of HSF1 activity [6–7].

Moreover, both HSF1 and HSF2 exist under two different isoforms produced by alternative splicing. This increases the diversity of potential HSF homo and heterotrimers and adds more complexity to HSF regulation. Regarding HSF1, only two publications refer to the existence of two splicing isoforms (Hsf1 α and Hsf1 β) in mice [8–9]. Hsf1 α results from a splicing process maintaining the insertion of the exon 11 (66 bp), and thus producing a longer protein in comparison to the β isoform. This exon is flanked by two introns presenting a consensus-splicing site. The 22 amino acids encoded by exon 11 are located in the C-terminal domain, adjacent to the hydrophobic region C (HR-C) that is important to maintain HSF1 under an inactive form in absence of stress signal. This additional region could form a leucine-zipper pattern possibly involved in the temperature activation of HSF1 α [9]. Furthermore, it was shown that the relative quantity of both isoforms was regulated in a tissue specific manner in mice, but surprisingly, their transcriptional activities have never been studied so far. Like HSF1, the long and short HSF2 isoforms are named HSF2 α and HSF2 β , respectively [10].

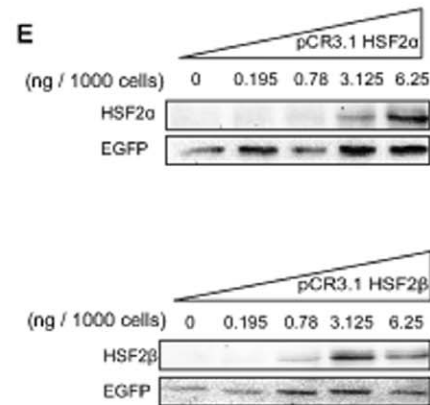
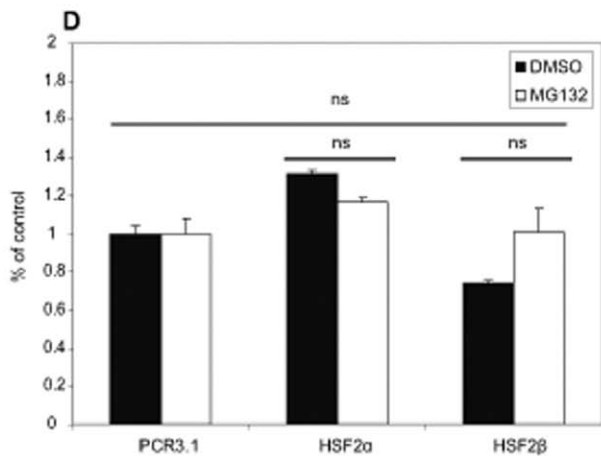
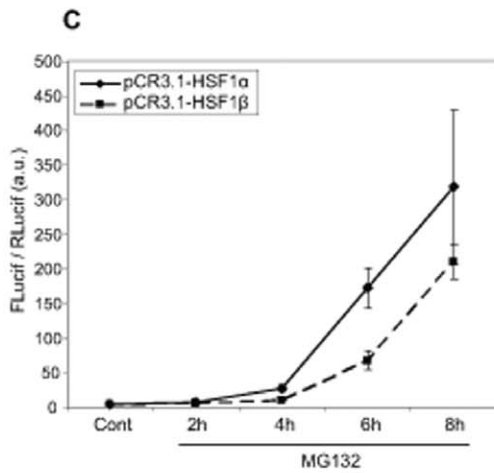
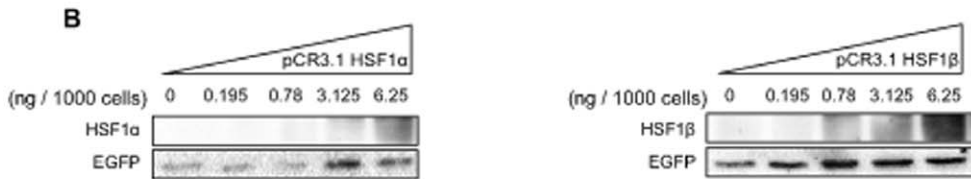
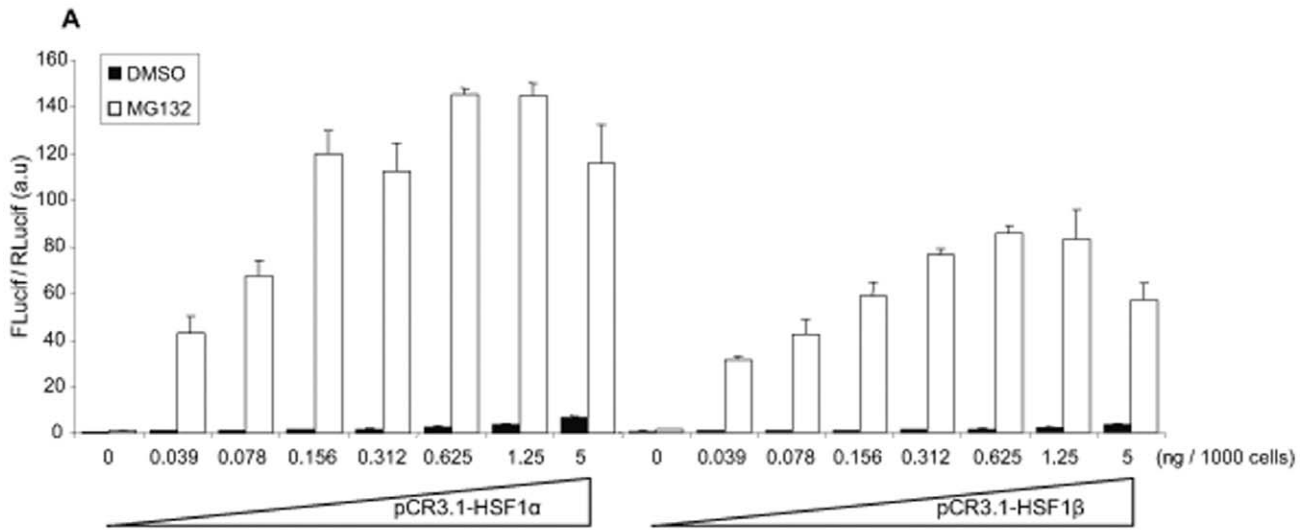


Figure 1. Transcriptional activity of HSF1 and HSF2 isoforms. (A) *Hsf1.2*^{-/-} iMEFs were co-transfected with increasing quantity of pCR3.1-HSF1 α (left), or pCR3.1-HSF1 β (right), in addition to pHSE_{2x}-TATA-Luc used as a reporter gene. DNA quantities were adjusted with empty pCR3.1. Transfection efficiency was assessed using the pTK-Rluc reporter gene. Cells were treated with MG132 at 2.5 μ M (white) or with DMSO (black) as control, for 8 h. Results correspond to the ratio between firefly luciferase (FLucif) and renilla luciferase (RLucif) activities. The data are from a representative experiment including three independent replicates (mean \pm SD). (B) Representative Western-blot showing the expression of HSF1 isoforms after transfection. *Hsf1.2*^{-/-} iMEFs were co-transfected with increasing quantity of pCR3.1-HSF1 α (left) or pCR3.1-HSF1 β (right) and pEGFP as control for transfection efficiency. Cells were treated with MG132 at 2.5 μ M and immunoblots for HSF1 and GFP were performed. (C) *Hsf1.2*^{-/-} iMEFs were co-transfected with 12.5 ng of pCR3.1-HSF1 α (full line), or pCR3.1-HSF1 β (dotted line), with two reporter genes described previously. Cells were treated with MG132 at 2.5 μ M or with DMSO, for 2 h, 4 h, 6 h or 8 h. Results correspond to the ratio between firefly luciferase (FLucif) and renilla luciferase (RLucif) activities and are the mean of three independent experiments \pm SD. (D) *Hsf1.2*^{-/-} iMEFs were co-transfected with 12.5 ng of pCR3.1-HSF2 α , or pCR3.1-HSF2 β , with the reporter genes as described in (A). Cells were treated with MG132 at 2.5 μ M (black), or with DMSO (white), for 8 h. Results are expressed in percentage of empty vector and represent the mean of three independent experiments \pm SD (Student's t test, ns: no significant). (E) Representative Western-blot showing the expression of HSF2 isoforms after transfection. *Hsf1.2*^{-/-} iMEFs were co-transfected with increasing quantity of pCR3.1-HSF2 α (high panel) or pCR3.1-HSF2 β (low panel) and pEGFP as control for efficiency. Cells were treated with MG132 at 2.5 μ M and immunoblots for HSF2 and GFP were performed.
doi:10.1371/journal.pone.0056085.g001

HSF2 α contains also an alternative exon 11, which is similarly flanked by two introns containing consensus-splicing sites. Hsf2 exon 11 encodes an 18 amino acids region located after HR-C and partially overlapping the activation domain called AD-1. The relative quantity of HSF2 isoforms is tissue specific [8,10]. Activities of HSF2 isoforms have been better documented in studies mainly conducted in K562 erythroleukemia cells, since HSF2 activity is particularly efficient in these cells. Overexpression of HSF2 β resulted in a weaker Hsp70 induction compared to HSF2 α [11]. This confirmed previous work showing that HSF2 α is a more potent transcriptional activator than the HSF2 β isoform [10]. However, as K562 erythroleukemia cells contain a mixture of both HSFs in their two existing isoforms, it is difficult to distinguish the actual role of each isoform in the context of heterimers.

Since the consequences of proteasome inhibition on HSF1 and HSF2 isoform expression and activity have not yet been fully characterized, we took advantage of existing cells which are deficient for both HSF1 and HSF2. Using defined transfections, we sought to determine the distinct role of each HSF1 and HSF2 isoforms independently of endogenous HSFs. Here, we show that HSF1 α and HSF1 β exert different transcriptional activity and we provide evidence for a specific repressor role of HSF2 β . Finally, our data and analyses establish that the relative quantity of each HSF2 isoforms is regulated in a stress-dependent manner.

Materials and Methods

Animals, Embryos and Cell Culture

Mixed genetic background wild type, *hsf1*^{-/-} and *Hsf2*^{-/-} mice, previously provided by Dr IJ Benjamin (University of Utah, Salt Lake City) and described elsewhere [12], were used in those experiments. For embryos collection, females were superovulated and mated with males of corresponding genotypes as described in [13]. Plug was considered as day 0.5 after fertilization (0.5 days post coitum (dpc)) and embryos were collected at the 2-cell stage (1.5 dpc) to be cultured to the blastocyst stage as previously described [13]. Blastocysts were obtained on 3–3.5 dpc and used to perform real time RT-PCR.

Immortalized Mouse Embryonic Fibroblasts wild type (WT) or deleted for both *Hsf1* and *Hsf2* (*Hsf1.2*^{-/-}) were obtained by intercrossing single knockout mice [12,14]. The primary cells were then immortalized using SV40 large T antigen by Dr Valérie Mezger (UMR CNRS 7216, Paris, France). Cells were cultivated as previously described [2].

Plasmid Constructs

pHSE_{2x}-TATA-Firefly luciferase was obtained by cloning two synthetic HSEs (5'-GAAgC TTCaGAAgC TTCaGAAgC TTC-3')

followed by a TATA box into the pGL3 vector (Promega) at *KpnI* and *SacI* sites. The plasmid pTK-Renilla luciferase (gift from Dr Island, INSERM U991, Rennes, France) was used as control to normalize transfection efficiency. All Hsf isoforms were cloned in pCR3.1 expression vector (Invitrogen). For Hsf1 α , the coding sequence was amplified by RT-PCR from total RNA of iMEFs WT, using primers flanking the open reading frame and providing restriction site for *EcoRI* and *NotI*. Then, PCR fragment was cloned to pCR3.1 and the cloned isoform was verified by sequencing. The Hsf1 β and Hsf2 β cDNAs (in pGEM1-HSF1 β and pGEM-HSF2 β vector, respectively) were a gift from Dr Richard Morimoto (Northwestern University, Evanston IL, USA) and they were subcloned in pCR3.1 at *EcoRI* site. The p β actin-HSF2 α vector was a gift of Dr Kevin Sarge (University of Kentucky, Lexington KY, USA) and the cDNA was also subcloned in pCR3.1 at *EcoRI* and *NotI* sites.

Transfection and Luciferase Assay

All transfections were performed using JetPEI (Polyplus Transfection) according to the manufacturer's instructions. Cells were harvested 24 h after transfection and 8 h of treatment with MG132 at 2.5 μ M, or with an equivalent volume of dimethyl sulfoxide (DMSO) as control. For heat shock experiment, cells were maintained at 45°C during 20 min and then transferred at 37°C during 6 h for recovery. Firefly luciferase and Renilla luciferase activities were determined with Dual luciferase reporter assay (Promega).

RT-PCR Analysis

One million of WT iMEFs were plated on a 10 cm dish and treated with MG132 at 1 μ M during 2 h, 4 h, 6 h, 8 h or 10 h. After 10 h of MG132 treatment, cells were allowed to recover for 1 h, 6 h or 10 h, and then harvested. Total RNAs were extracted with TRIzol reagent (Invitrogen) and 5 μ g of RNAs were retro-transcribed using M-MLV RT (Invitrogen). To discriminate Hsf2 α and Hsf2 β , primers flanking alternative exon (*forward*: CATGTCTAGTGTGCTGTCCAGC and *reverse*: 5'-GAGCT-CATCGACTTCTATGG-3') were used in RT-PCR experiment. RT-PCR of the housekeeping gene Glyceraldehyde-3-phosphate dehydrogenase (*Gapdh*) (*forward*: 5'-TGAAGCAGGCATCT-GAGGG-3' and *reverse*: 5'-CGAAGGTGGAAGAGTGGGAG-3') was used as control for the quality of extraction and retro-transcription. Densitometric analysis was made with Quantity One software (Bio-Rad).

RT-real time PCR experiments were performed on blastocysts as described in [15]. Experiments to determine the relative abundance of Hsf2 isoforms used primers to detect the total population of Hsf2 transcripts (*Hsf2 forward*: 5'-AGGGGAGTA-

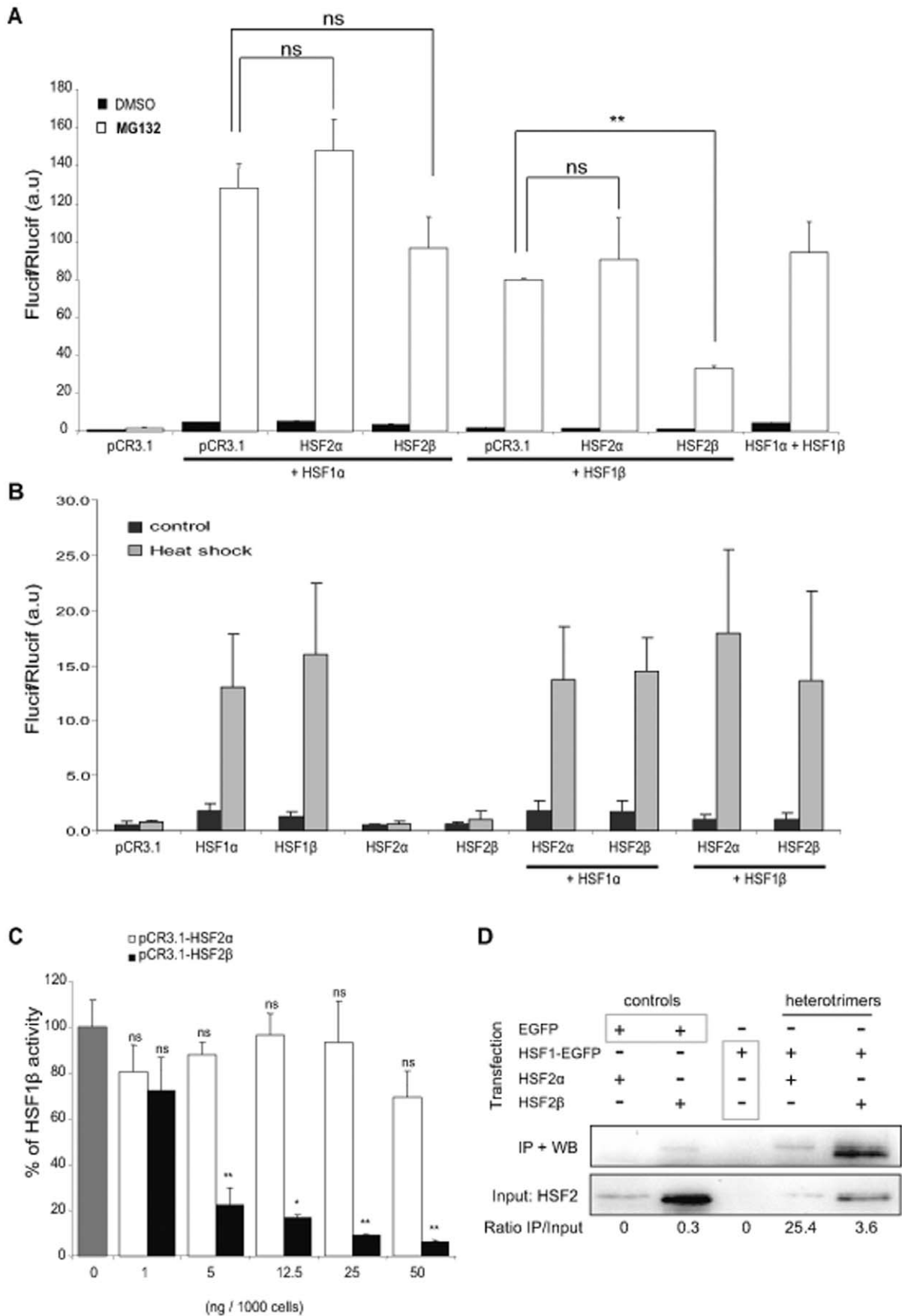


Figure 2. HSF2 β interacts with HSF1 β and inhibits its transcriptional activity. (A) *Hsf1.2*^{-/-} iMEFs were co-transfected with 12.5 ng of pCR3.1-HSF1 α or pCR3.1-HSF1 β in combination with 12.5 ng of pCR3.1-HSF2 α , or pCR3.1-HSF2 β . Transcriptional activity was followed with pHSE_{2x}-TATA-Luc, and pTK-Rluc was used as control for transfection efficiency. Cells were treated with MG132 at 2.5 μ M (white), or with DMSO as control (black). Results correspond to the ratio between firefly luciferase (FLucif) and renilla luciferase (RLucif) activities and are a representative experiment made with three independent replicates (Student's t test: HSF1 α or β alone compared to others conditions. **p<0.01, ns: no significant). (B) Cells were transfected with the indicated expression vectors and the reporter genes, as described in (A). Cells were submitted to heat shock at 45°C for 20 min and put to recovery at 37°C for 6 h (grey). Untreated cells served as control (black). Results are the mean \pm SD of 9 to 12 independent transfections. (C) *Hsf1.2*^{-/-} iMEFs were co-transfected with pCR3.1-HSF1 β (gray) and with an increase quantity of pCR3.1-HSF2 α (white), or pCR3.1-HSF2 β (black). Then, cells were treated with MG132 at 2.5 μ M for 8 h. Results are expressed in percentage of HSF1 β activity and are the mean of three independent experiments \pm SD (Student's t test: HSF1 β alone compare to others conditions. *p<0.05, **p<0.01, ns: no significant). (D) Representative co-immunoprecipitation experiments. *Hsf1.2*^{-/-} iMEFs were co-transfected with pEGFP or pHSF1 β -EGFP, in combination with pCR3.1-HSF2 α or pCR3.1-HSF2 β . Cells were treated with MG132 at 1 μ M for 8 h and nuclear protein extracts were submitted to EGFP immunoprecipitation, followed by immunoblotting for HSF2. The value of the IP/Input ratio is indicated below the panel.

doi:10.1371/journal.pone.0056085.g002

CAACTGCATCG-3' and reverse: 5'-CAGGCGGACAAGCT-TACTC-3') [16] and primers designed to amplify only Hsf2 α isoform (forward: 5'-AGTTCTGTGCAGATGAATCCCACAG-3' and reverse: 5'-GCAGATGCAGATTCCCATCC-3'). Experiments performed to measure Hsp70.1 transcripts used the following primers: Hsp70.1 forward: 5'-TTGTCCATGT-TAAGGTTTTGTGGTATA-3', Hsp70.1 reverse: 5'-GTTTTTTTTCATTAGTTTGTAGTGATGCAA-3'. The experiments were performed at least in duplicate with one or two independent groups of embryos included in each experiment (n=20 blastocysts). Results were normalized using 18S RNA amplification.

Protein Extracts and Immunoblot Analysis

To evaluate protein expression after transfection, 200 000 *Hsf1.2*^{-/-} iMEFs per well were seeded in a 6-well plate. Cells were transfected as described above and were harvested after 24 h of transient expression and 8 h of treatment with MG132 at 2.5 μ M. To analyze the expression of endogenous HSF2, 1 million of WT cells were plated in a 10 cm dish and were treated the next day with MG132 at 1 μ M, or with DMSO for 10 h. Whole cell extracts were prepared with NP-40 lysis buffer (50 mM Tris-HCl pH 8, 150 mM NaCl, 10 mM EDTA, 1% NP-40, 0.2% sarkosyl). Proteins were separated on an 8.5% or a 12% polyacrylamide gel to discriminate HSF2 isoforms and transferred to a nitrocellulose membrane (Amersham Bioscience). Anti-HSF1 (4B4) antibody (ab44819), anti-HSF2 (3E2) antibody (sc-13517) and Anti-GFP antibody (a11122) were purchased from Abcam, Santa Cruz Biotechnology and Molecular Probes, respectively.

Four hours of migration at 40 mA were necessary to separate HSF2 α and HSF2 β . Ponceau staining was performed to assess equal loading in place of classical actin control, which was excluded from the gel during the long electrophoresis.

Co-immunoprecipitation

Hsf1.2^{-/-} iMEFs were transfected with pHSF1 β -EGFP and/or with pCR3.1-HSF2 α , or with pCR3.1-HSF2 β . As control, cells were co-transfected with pEGFP and pCR3.1-HSF2 α , or pCR3.1-HSF2 β . Cells were treated with MG132 at 1 μ M during 10 h. Anti-GFP antibody (A11122, Molecular Probes) was used for immunoprecipitation, carried out with Nuclear Complex Co-immunoprecipitation kit (Active Motif), according to manufacturer's instructions.

Results

Transcriptional Activities of HSF1 and HSF2 Splicing Isoforms

To characterize the *transactivation* abilities of HSF1 isoforms, we first co-transfected increasing doses of expression vector coding

either HSF1 α or HSF1 β , in *Hsf1.2*^{-/-} iMEFs, with pHSE_{2x}-TATA-Firefly luciferase as a reporter gene. Then, cells were treated with MG132 at 2.5 μ M for 8 h. As presented in Figure 1A, low quantities of HSF1 α expression vector provided high level of *transactivation* and HSF1 α exhibited 1.4 times higher plateau values than HSF1 β . These data suggested that the α isoform was more efficient than the β one to activate HSE-dependent transcription under proteasome inhibition. Nevertheless, increasing quantities of transfected HSF1 α vector was accompanied by a more pronounced basal activity (around 10 fold for 5 ng of transfected vector/1000 cells), indicating that the α isoform presents a weak stress-independent activity. To verify that the difference observed in *transactivation* abilities was not due to a defect in the level of expression of the transcription factors, immunoblots were performed with extracts from *Hsf1.2*^{-/-} iMEFs that had been co-transfected with an increased quantity of pCR3.1-HSF1 α or pCR3.1-HSF1 β , in addition to pEGFP used as a transfection efficiency control (Figure 1B). All cells were treated with MG132 at 2.5 μ M for 8 h. HSF1 α protein (left panel) became detectable when 3.125 ng of vector were transfected per 1000 cells, whereas HSF1 β was detected after transfection of a lower amount of vector (0.78 ng/1000 cells), suggesting that the β isoform was more stable than the α isoform, but less effective from a transcriptional point of view. Moreover, both HSF1 isoforms required approximately 80 and 20 fold higher quantity of transfected vector to be detected by immunoblot in comparison to the amount needed to obtain a visible effect by luciferase assay. These results suggest that a very low HSF1 intracellular concentration is sufficient to activate transcription.

To study the kinetic of *transactivation* of each isoform, we co-transfected 12.5 ng of expression vector in *Hsf1.2*^{-/-} iMEFs. Then, cells were treated with MG132 at 2.5 μ M for 2 h, 4 h, 6 h or 8 h. Stress-dependent induction of transcription was detectable after 4 h of treatment (Figure 1C), showing that both isoforms exhibited similar responses in terms of activation time course.

Finally, HSF2 α or HSF2 β expression vector were co-transfected with pHSE_{2x}-TATA-Firefly luciferase in *Hsf1.2*^{-/-} iMEFs. Neither HSF2 α nor HSF2 β induced the transcription of the HSE reporter gene after treatment with MG132 at 2.5 μ M (Figure 1D). Two other reporter genes, containing either HSP70 or p35 gene promoters, were used in similar experiments and gave comparable results since both HSF2 isoforms were unable to significantly trigger Hsp70 or p35 *transactivation* (data not shown). These data suggest that by itself, HSF2 is a very poor transcription factor, whatever the splicing isoform considered. As previously tested for HSF1, *Hsf1.2*^{-/-} iMEFs were co-transfected with increasing quantity of pCR3.1-HSF2 α or pCR3.1-HSF2 β , in addition to pEGFP used as a transfection efficiency control, to assess the level of expression of the different vectors (Figure 1E). Like for HSF1 isoforms, a minimum of 3.125 ng transfected vector

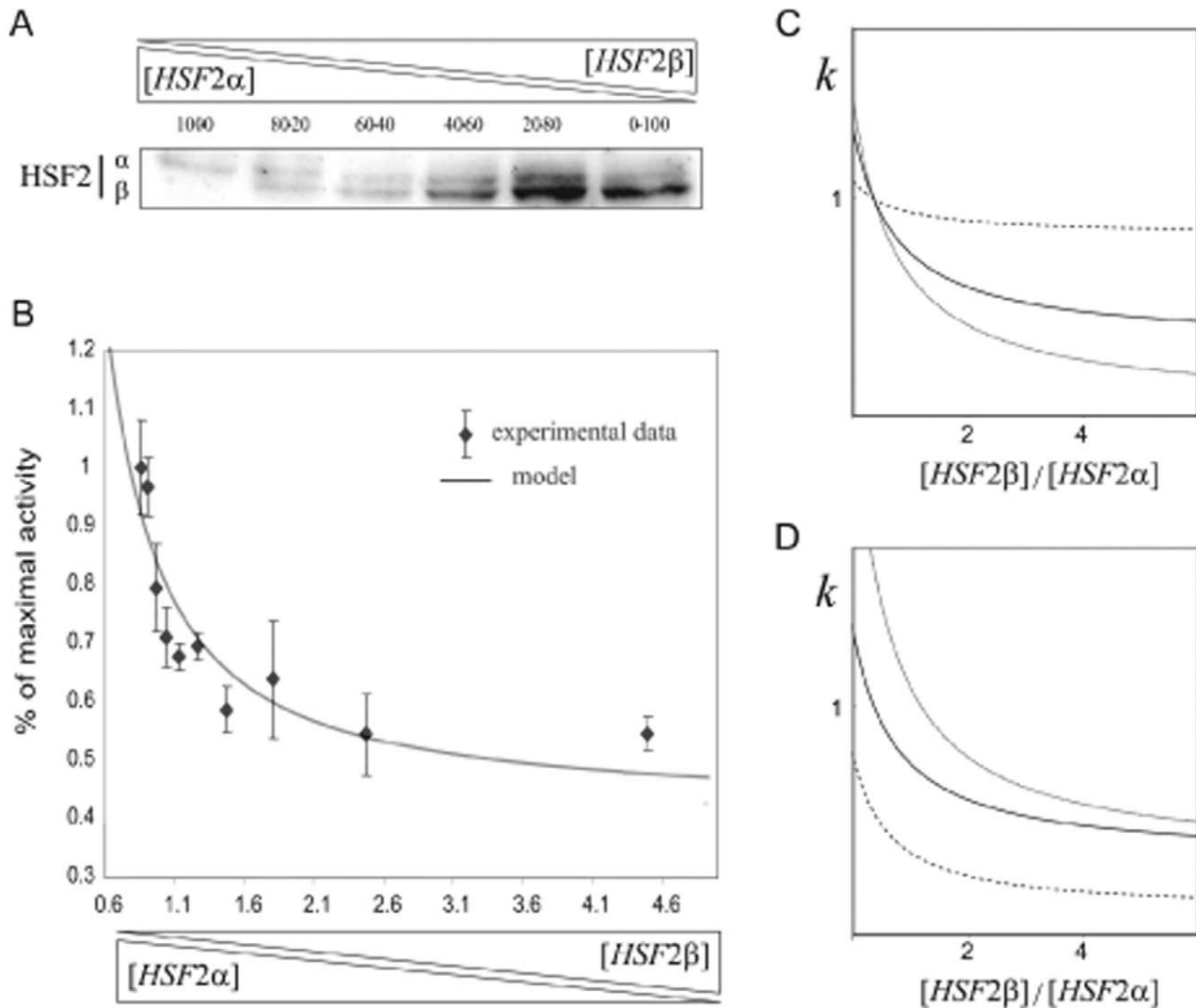


Figure 3. Ratio between HSF2 α and HSF2 β controls HSF1 β transcriptional activity. (A) Representative Western-blot showing the expression of HSF2 isoforms after transfection. *Hsf1.2*^{-/-} iMEFs were co-transfected with pCR3.1-HSF1 β and pCR3.1-HSF2 α/β to obtain the expression of equivalent amounts of HSF1 and HSF2, and increasing concentration of HSF2 β relatively to HSF2 α . Cells were treated with 2.5 μ M MG132 for 8 h. Protein extracts were loaded on 12% polyacrylamide gel and submitted to a long migration to separate efficiently HSF2 isoforms. HSF2 was revealed by immunoblotting. (B) Transcriptional activity induced, with fixed concentrations of HSF1 and total HSF2, but varying combinations of HSF2 β /HSF2 α . *Hsf1.2*^{-/-} iMEFs were co-transfected with 12.5 ng of pCR3.1-HSF1 β and with the same quantity of pCR3.1-HSF2 α/β , as previously described. Diamonds correspond to the experimental data obtained in independent triplicates, and expressed as percentage of maximal activity. Solid line drawn to equation 2 with random multimerization and considering that HSF1 is active only in absence of HSF2 β in the trimer. (C) HSE-driven transcriptional activities expected from Eq. 2, with a constant and identical amounts of HSF1 and total HSF2, and when increasing the ratio HSF2 β /HSF2 α . The transcriptional strength of a trimer is assumed to be proportional to the number of HSF1 monomers present, and the unit of transcriptional strength ($k=1$) corresponds to that of an HSF1 monomer. The strength of trimerization is considered as either (i) identical between hetero- and homodimers (plain line), (ii) 10 fold higher for homodimers (dashed line), or (iii) 10 fold higher for heterodimers (dotted line). (D) HSE-driven transcriptional activities drawn to Eq. 2, with constant and equivalent amounts of HSF1 and HSF2, capable to either randomly homo- or heterotrimerize. Plain line: as for panel C, the transcriptional strength of a trimer is proportional to the number of HSF1 monomers included in the trimer and the strength of a HSF1 monomer is set to 1 ($k_3=3 k_1$ and $k_2=2 k_1$ and $k_1=1$). Dashed line: Transcriptional strength independent on whether the trimer contains 1, 2 or 3 HSF1 monomers ($k_1=k_2=k_3=1$). Dotted line: same rule with $k_1=k_2=k_3=3$. doi:10.1371/journal.pone.0056085.g003

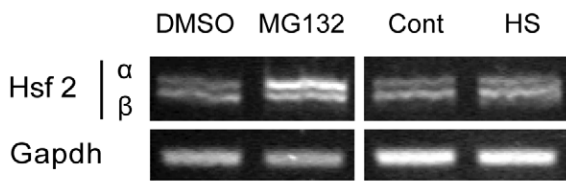
per 1000 cells was necessary to detect HSF2 α whereas only 0.78 ng per 1000 cells was required for HSF2 β , suggesting that this latter isoform is more stable than HSF2 α .

HSF2 β Forms Heterotrimers with HSF1 β and Inhibits its Transcriptional Activity

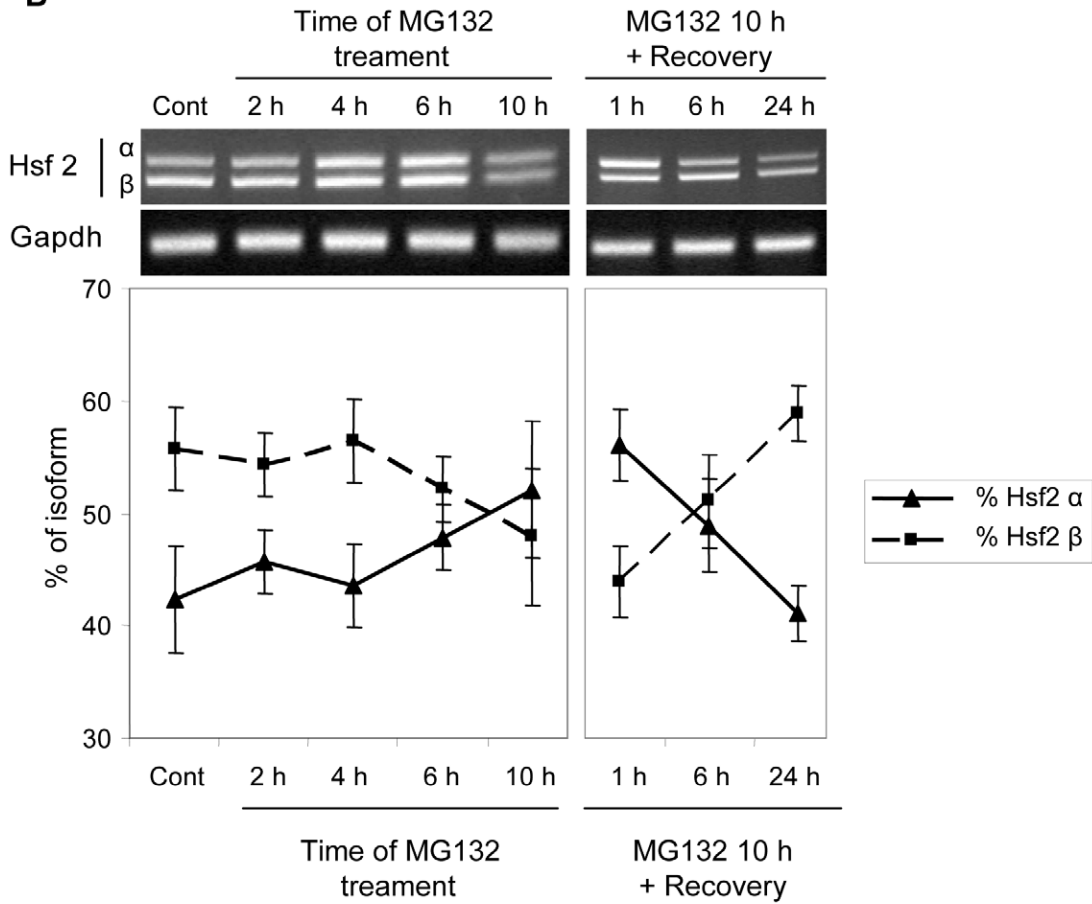
Previous work from our team had shown that proteasome inhibition was associated with the formation of HSF1/HSF2

heterotrimers [5]. To determine how the different splicing isoforms could impact heterotrimer activity, we co-transfected different combinations of expression vectors with reporter genes in *Hsf1.2*^{-/-} iMEFs (Figure 2A). HSF1 α transcriptional activity was not statistically different in absence or presence of HSF2 isoforms. Likewise, HSF1 β transcriptional activity was not altered when co-transfected with HSF2 α . However, it was strongly and significantly decreased (3 fold) after co-transfection with HSF2 β . The activity of

A



B



C

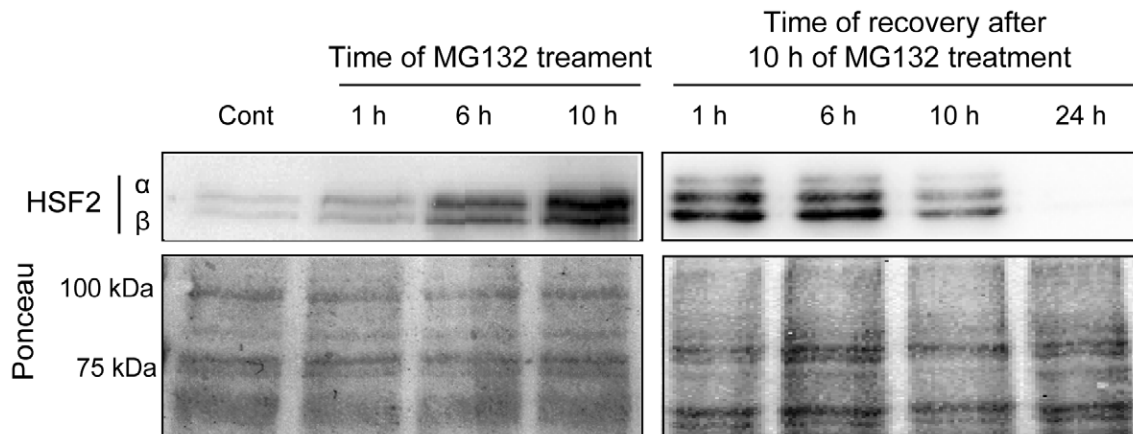


Figure 4. Proteasome inhibition modifies the relative quantity of Hsf2 α and Hsf2 β at the mRNA level. (A) WT iMEFs were treated with MG132 at 1 μ M or with DMSO for 10 h, or were heat shocked 20 min at 45°C and allowed to recover during 6 h. The relative quantity of Hsf2 α and Hsf2 β transcripts was assessed by RT-PCR using primers flanking the alternative exon. Gapdh RT-PCR served as control for efficient retrotranscription and amplification. (B) WT iMEFs were treated during 2 h, 4 h, 6 h or 10 h with MG132 at 1 μ M or with DMSO before being harvested. In addition, WT iMEFs exposed to MG132 during 10 h were allowed to recover during 1 h, 6 h or 24 h before collection. Relative quantity of Hsf2 α and Hsf2 β was assessed by RT-PCR, as described above. Quantity one software (Bio Rad) was used to assess band intensity. Results were expressed in mean of percentage of each isoform \pm SD from four independent experiments. (C) WT iMEFs treated as described in (B) were harvested for protein analysis. Protein extracts were loaded on 12% polyacrylamide gel and submitted to a long migration to separate efficiently HSF2 isoforms. HSF2 was revealed by immunoblotting. Ponceau staining was used to verify the equal loading. doi:10.1371/journal.pone.0056085.g004

the heterotrimer was also assessed after heat shock (Figure 2B). When HSF1 isoforms were transfected alone, the same transactivation level was observed whatever the isoform used. As observed after MG132 treatment, HSF2 α and HSF2 β did not show any significant transcriptional activity under thermal stress. Co-transfection of HSF2 with HSF1 did not affect HSF1 activity. Especially, the combination of HSF1 β with HSF2 β did not decrease the trimer activity after heat shock, in contrast to MG132 treatment.

To verify that the inhibition observed in MG132 experiment was due to HSF2 β itself and not to a side effect related to difference in transfection efficiency, we performed a dose effect experiment (Figure 2C). *Hsf1.2*^{-/-} iMEFs were co-transfected with a constant quantity of pCR3.1-HSF1 β and increasing doses of pCR3.1-HSF2 α or pCR3.1-HSF2 β . HSF1 β transactivation remained similar whatever the amount of co-transfected HSF2 α . On the contrary, HSF2 β transfection provoked a sharp decrease of transactivation with low dose (5 ng/1000 cells), confirming the inhibitory role of HSF2 β .

The ability of each HSF2 isoform to associate with HSF1 β in response to MG132 treatment was determined by co-immunoprecipitation experiment (Figure 2D). *Hsf1.2*^{-/-} iMEFs were co-transfected with expression vector coding either HSF1 β -EGFP or only EGFP, in association with HSF2 isoforms. Immunoprecipitation was performed with antibodies against EGFP and blots were probed with HSF2 antibodies. Cells transfected with HSF1 β -EGFP, but not HSF2, were used as control and showed no signal after immunoprecipitation, demonstrating that the HSF2 anti-

bodies do not cross-react with HSF1 β (lane 3). Interestingly, we found that both HSF2 α and β isoforms interact with HSF1 β (lane 4 and 5) indicating the formation of heterotrimers. Moreover, HSF2 co-immunoprecipitation required the presence of HSF1, as demonstrated by its absence in the EGFP control (lane 1 and 2).

Influence of the HSF2 β /HSF2 α Ratio on HSF1 Transcriptional Activity

To determine how the HSF2 β /HSF2 α ratio impacts the transcriptional activity of HSF1 β , *Hsf1.2*^{-/-} iMEFs were transfected with HSF1 β and the same amount of total HSF2, including variable HSF2 β /HSF2 α ratios (Figure 3). HSF2 proteins were first quantified by immunoblotting (Figure 3A) and densitometric analyses. Due to the differences in protein stability of HSF2 isoforms, we calculated from three densitometric analyses a corrective coefficient, linking transfected vector ratio and protein expression. Then, the same transfection conditions were used to measure the transcriptional activity with a luciferase reporter vector (Figure 3B). HSE-driven transcriptional activity diminished when increasing the HSF2 β /HSF2 α protein ratio. The capacity of HSF2 and HSF1 to heterotrimerize has been evidenced [5,17] but the stoichiometry of the different HSFs in the trimers remains unknown. Moreover, the kinetic and thermodynamic parameters of HSF trimerization, as well as the precise mode of stepwise HSF assembly, ordered or random, have to be characterized. Hence, in absence of defined data, minimal rules were selected, as listed below. It is considered that HSF2 α has no influence on the transcriptional activity of the HSF1 protomers included in a

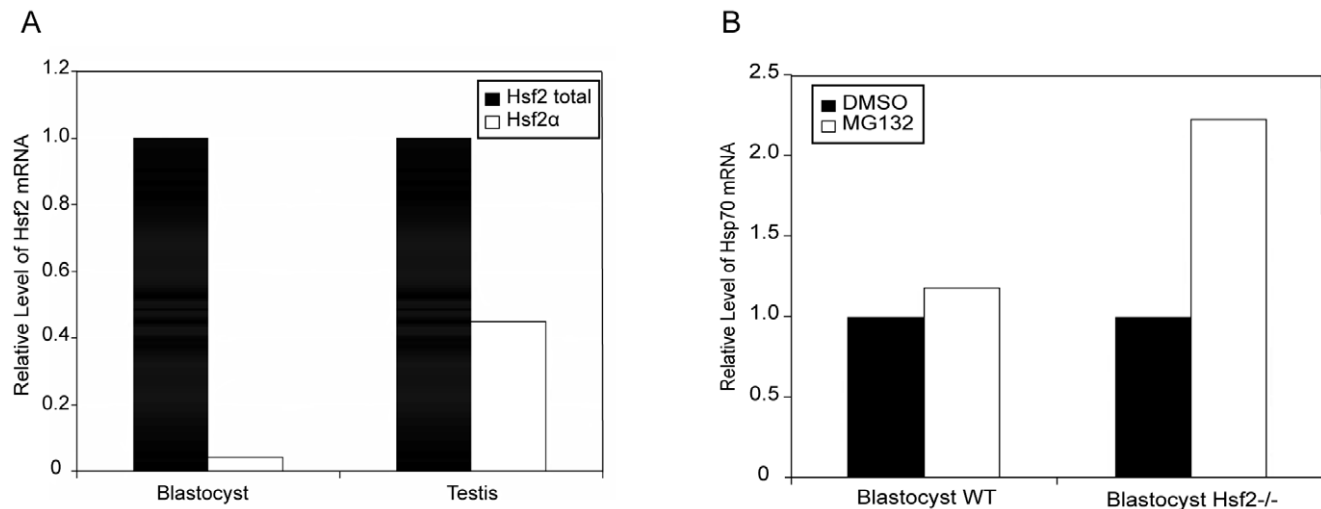


Figure 5. Hsf2 deletion in blastocyst is characterized by a higher Hsp70 induction. (A) Relative quantity of total Hsf2 and Hsf2 α transcripts was assessed in blastocyst and testis using RT-real time PCR. Results are expressed as relative expression of Hsf2 α compared to total Hsf2 and correspond to the mean of two independent experiments. (B) Hsp70 induction after 4 h of MG132 treatment was assessed by RT-real time PCR in WT and *Hsf2*^{-/-} blastocysts. Results are expressed as fold induction of Hsp70 expression after MG132 treatment, normalized to DMSO for each sample, and correspond to the mean of two independent experiments. doi:10.1371/journal.pone.0056085.g005

and HSF2 β were barely detectable, because HSF2 is a labile protein, constitutively expressed and degraded. During MG132 treatment, HSF2 was highly stabilized which adds an additional level of regulation, in addition to the translation. Differences between isoforms remained difficult to detect because of the lack of sensitivity of the immunoblot method and the absence of isoform specific antibodies.

Low Hsf2 α /Hsf2 β Ratio in Blastocyst is Associated with a Weak Response to Proteasome Inhibition

HSF2 exhibits a developmentally regulated DNA binding activity at the blastocyst stage (E 3.5 d) [19], which remains unexplored so far. This prompted us to examine the relative abundance of HSF2 isoforms, and the blastocyst response to proteasome inhibition. Due to the limited amount of starting material, we designed primers enabling real time RT-PCR to amplify either the total Hsf2 cDNA, or specifically the α isoform. For comparison, the same strategy was applied to testis total extract (Figure 5A). In blastocysts, Hsf2 α represented around 4% of total Hsf2 transcripts (implying that the β isoform would count for 96%), while in testis, the proportion of Hsf2 α rose up to 46% of Hsf2 total (Figure 5A). When WT blastocysts were treated with MG132 for 4 h, Hsp70 expression assessed by real time RT-PCR (Figure 5B) was only slightly induced (around 1.2 fold), compared to non-treated ones. On the contrary, in Hsf2 $^{-/-}$ blastocysts, Hsp70 was 2.2 fold induced in MG132 treated versus non-treated embryos. In accordance with our previous results, and speculating that in WT blastocysts where HSF2 β was the dominant isoform, the response to proteasome inhibition was blunted. Conversely, in Hsf2 $^{-/-}$ blastocyst where there was no HSF2 β protein, the Hsp70 response could be elicited.

Discussion

Both HSF1 and HSF2 exist as two main splicing isoforms, but very few studies have focused on their respective transcriptional activity. It is uneasy to address this question in cells expressing endogenous HSF1 and 2, because of the coexistence of various HSF complexes, either homo- or hetero-trimers, using potentially both existing isoforms. Therefore, we took advantage of iMEFs genetically depleted for both factors, which provide an invaluable tool to dissect the precise role of each splicing isoforms of HSF1 and HSF2.

We found that HSF1 α and HSF1 β possessed distinct transcriptional activities, HSF1 α being a more potent activator under proteasome inhibition. When HSF1 isoforms were discovered, the authors hypothesized that HSF1 α should be activated at higher temperatures than HSF1 β , because exon 11 encodes a putative leucine zipper increasing the length of the HR-C domain, known to stabilize HSF1 in the monomeric non-DNA binding form [9]. In our study, we showed that HSF1 α had the same kinetic of transcription activation than HSF1 β , suggesting that these two isoforms do not differ by their sensitivity to stress intensity but by their transcriptional efficiency. Moreover, this difference in transcriptional competency depends on the type of stress experienced by the cells, since we found that the two isoforms have the same transcriptional activities after heat shock treatment.

Regarding HSF2, previous works in yeast [20] and in K562 erythroleukemia cells [21–22] reported a modest, but detectable transcriptional activity for HSF2. It was even suggested that the short HSF2 β isoform is a less potent activator than the long HSF2 α isoform [10]. Our study revealed that both HSF2 isoforms are transcriptionally inefficient in our cellular model. Taken together, these results suggest that HSF2 transcriptional activity

depends upon the cellular context and the presence of indispensable co-factors, or particular stimuli. The absence of HSF2 transcriptional activity also indicates that its main role is not to directly recruit the transcriptional apparatus, but rather to play a pioneer role in chromatin preparation. This hypothesis is in agreement with the fact that HSF2 can prevent compaction of HSF target genes during mitosis [23]. Our study confirmed the role of HSF2 isoforms in the modulation of HSF1 activity [11,17,24]. While HSF2 α does not affect significantly HSF1 transcriptional activity, HSF2 β clearly reduces HSF1 activity. Moreover, this repression is specific to the association of HSF1 β with HSF2 β , under MG132 treatment. Indeed, it was shown that HSF2 is not activated after heat shock treatment, but rather after inhibition of the ubiquitin-proteasome pathway [4]. Therefore, HSF1-HSF2 heterotrimers can be assembled only after proteasome inhibition, and not after heat shock [5], which might explain why HSF2 β inhibitory effect cannot be found after heat shock. According to our results, the presence or absence of a short isoform in the heterotrimer appears to be critical for this modulation of HSF activity. We propose that the leucine zipper conserved in the long isoform could create a novel interaction domain. Thus, this domain could stabilize the activation domain in an optimal conformation, while in the short isoforms, the activation domains could be in a more relaxed conformation, resulting in a less efficient co-factors binding, especially in the context of an HSF1 β -HSF2 β heterotrimer.

The absence of specific antibody for each isoform, and the technical limitations to accurately determine the ratio of isoforms in a trimer, remain a real concern to further decipher HSF2 mechanism in HSF1 inhibition. In an attempt to alternatively address this question, we have developed a mathematical modelling approach. Our model is different from those previously published on the dynamic of the eukaryotic heat shock response [25–27]. In contrast with previous heat shock response models, we did not focus on the various steps involved in the kinetics of Hsp gene activation and its feedback regulation. We rather concentrated on the active HSF trimer that binds to the promoter. We analyzed the impact of the presence of HSF2 α and/or HSF2 β isoforms on the steady state of HSF1 activity. We postulated that the presence of several HSF isoforms induces the coexistence of different types of heterotrimers. These complexes do not equally transactivate, and they are expected to compete to bind to target promoters. The model we propose satisfies the experimental data obtained by transfection. Based on this model, the strength of the response depends on the proportion of each isoform in the cell. Accordingly, the combination of HSF isoforms synthesized in each cell determines the level of response. This implies that all the cells do not have the same capabilities to respond to proteotoxic stress, and that expression of these HSF isoforms is crucial for the modulation of chaperone expression.

Expression of HSF2 isoforms is tissue-specific. For example HSF2 α is the main isoform in testis, while HSF2 β is dominant in brain [10]. In addition to this tissue-specific regulation of HSF isoforms, we have shown that the differential splicing of Hsf1 and Hsf2 messenger is also dependent on the type of stress. The switch between the two isoforms is specific to proteasome inhibition, as clearly shown by our RT-PCR experiments.

Taken together our data demonstrate how HSF1 and HSF2 splicing isoforms contribute a new level of complexity to HSF regulation. Our results raise additional questions: how efficient is the proteotoxic response in tissues where HSF1 β and HSF2 β are dominant? Which evolutionary mechanisms established such complex regulation of the relative quantity of HSF isoforms? It might be critical for stressed cells to possess various ways to

modulate HSF1 activity, to achieve better adaptation, and thus survival. Heat shock is an acute stress with massive protein alteration, which requires a rapid and high HSP expression. In that case, HSF1 is found in homotrimeric complex, while HSF2 become insoluble and is found in the perinuclear fraction [28]. On the other hand, proteasome inhibition induces a proteotoxic stress, which can be considered as a chronic stress with the requirement for longer-term adaptation. This should trigger a more limited response, since it could be deleterious for cells or tissues to express excessive amount of HSPs. Proteasome inhibition activates both HSF1 and HSF2 [29], leading to the formation of a diverse population of HSF heterotrimers, with variable transcriptional efficiency [5]. This mechanism could be the best option to control proteotoxic response in cells exposed to in case of chronic stress.

References

- Paul S (2008) Dysfunction of the ubiquitin-proteasome system in multiple disease conditions: therapeutic approaches. *Bioessays* 30: 1172–1184.
- Lecomte S, Desmots F, Le Masson F, Le Goff P, Michel D, et al. (2010) Roles of heat shock factor 1 and 2 in response to proteasome inhibition: consequence on p53 stability. *Oncogene* 29: 4216–4224.
- Pirkkala L, Alastalo TP, Zuo X, Benjamin IJ, Sistonen L (2000) Disruption of heat shock factor 1 reveals an essential role in the ubiquitin proteolytic pathway. *Mol Cell Biol* 20: 2670–2675.
- Mathew A, Mathur SK, Morimoto RI (1998) Heat shock response and protein degradation: regulation of HSF2 by the ubiquitin-proteasome pathway. *Mol Cell Biol* 18: 5091–5098.
- Loison F, Debure L, Nizard P, Le Goff P, Michel D, et al. (2006) Up-regulation of the clusterin gene after proteotoxic stress: implication of HSF1-HSF2 heterocomplexes. *Biochem J* 395: 223–231.
- Ostling P, Björk JK, Roos-Mattjus P, Mezger V, Sistonen L (2007) Heat shock factor 2 (HSF2) contributes to inducible expression of hsp genes through interplay with HSF1. *J Biol Chem* 282: 7077–7086.
- Åkerfelt M, Morimoto RI, Sistonen L (2010) Heat shock factors: integrators of cell stress, development and lifespan. *Nat Rev Mol Cell Biol* 11: 545–554.
- Fiorenza MT, Farkas T, Dissing M, Kolding D, Zimarino V (1995) Complex expression of murine heat shock transcription factors. *Nucleic Acids Res* 23: 467–474.
- Goodson ML, Sarge KD (1995) Regulated expression of heat shock factor 1 isoforms with distinct leucine zipper arrays via tissue-dependent alternative splicing. *Biochem Biophys Res Commun* 211: 943–949.
- Goodson ML, Park-Sarge OK, Sarge KD (1995) Tissue-dependent expression of heat shock factor 2 isoforms with distinct transcriptional activities. *Mol Cell Biol* 15: 5288–5293.
- Leppä S, Pirkkala L, Saarento H, Sarge KD, Sistonen L (1997) Overexpression of HSF2- β inhibits hemin-induced heat shock gene expression and erythroid differentiation in K562 cells. *J Biol Chem* 272: 15293–15298.
- McMillan DR, Christians E, Forster M, Xiao X, Connell P, et al. (2002) Heat shock transcription factor 2 is not essential for embryonic development, fertility, or adult cognitive and psychomotor function in mice. *Mol Cell Biol* 22: 8005–8014.
- Le Masson F, Christians E (2011) HSFs and regulation of Hsp70.1 (Hspa1b) in oocytes and preimplantation embryos: new insights brought by transgenic and knockout mouse models. *Cell Stress Chaperones* 16: 275–285.
- McMillan DR, Xiao X, Shao L, Graves K, Benjamin IJ (1998) Targeted disruption of heat shock transcription factor 1 abolishes thermotolerance and protection against heat-inducible apoptosis. *J Biol Chem* 273: 7523–7528.
- Metchat A, Åkerfelt M, Bierkamp C, Delsinne V, Sistonen L, et al. (2009) Mammalian heat shock factor 1 is essential for oocyte meiosis and directly regulate Hsp90alpha expression. *J Biol Chem* 284: 9521–9528.
- Le Masson F, Razak Z, Kaigo M, Audouard C, Charry C, et al. (2011) Identification of heat shock factor 1 molecular and cellular target during embryonic and adult female meiosis. *Mol Cell Biol* 31: 3410–3423.
- Sandqvist A, Björk JK, Åkerfelt M, Chitikova Z, Grichine A, et al. (2009) Heterotrimerization of heat-shock factors 1 and 2 provides a transcriptional switch in response to distinct stimuli. *Mol Cell Biol* 20: 1340–1347.
- Michel D (2010) How transcription factors can adjust the gene expression floodgates. *Prog Biophys Mol Biol* 102: 16–37.
- Mezger V, Rallu M, Morimoto RI, Morange M, Renard JP (1994) Heat shock factor 2-like activity in mouse blastocysts. *Dev Biol* 166: 819–822.
- Liu XD, Liu PC, Santoro N, Thiele DJ (1997) Conservation of a stress response: human heat shock transcription factors functionally substitute for yeast HSF. *EMBO J* 16: 6466–6477.
- Sistonen L, Sarge KD, Phillips B, Abravaya K, Morimoto RI (1992) Activation of heat shock factor 2 during hemin-induced differentiation of human erythroleukemia cells. *Mol Cell Biol* 12: 4104–4111.
- Yoshima T, Yura T, Yanagi H (1998) Function of the C-terminal transactivation domain of human heat shock factor 2 is modulated by the adjacent negative regulatory segment. *Nucleic Acids Res* 26: 2580–2585.
- Xing H, Wilkerson DC, Mayhew CN, Lubert EJ, Skaggs HS, et al. (2005) Mechanism of hsp70i gene bookmarking. *Science* 307: 421–423.
- He H, Soncin F, Grammatikakis N, Li Y, Sigano A, et al. (2003) Elevated expression of heat shock factor (HSF) 2A stimulates HSF1-induced transcription during stress. *J Biol Chem* 278: 65465–65475.
- Rieger T, Morimoto RI, Hatzimanikatis V (2005) Mathematical modeling of eukaryotic heat shock response: dynamics of the hsp70 promoter. *Biophys J* 88: 1646–1658.
- Szymańska Z, Zyllicz M (2009) Mathematical modeling of heat shock protein synthesis in response to temperature change. *J Theor Biol* 259: 562–569.
- Petre I, Mizera A, Hyder C, Meinander A, Mikhailov A, et al. (2011) A simple mass-action model for the eukaryotic heat shock response and its mathematical validation. *Nat Comput* 10: 595–612.
- Mathew A, Mathur SK, Jolly C, Fox SG, Kim S, et al. (2001) Stress-specific activation and repression of heat shock factors 1 and 2. *Mol Cell Biol* 21: 7163–7171.
- Kawazoe Y, Nakai A, Tanabe M, Nagata K (1998) Proteasome inhibition leads to the activation of all members of the heat-shock-factor family. *Eur J Biochem* 255: 356–362.

Acknowledgments

We thank Frédéric Percevault for his assistance in cell culture and Christophe Audouard for his contribution to the experiments on blastocysts. We particularly thank Catherine Martin for her technical assistance in heat shock experiments. We thank Dr Yann Le Page for his careful and critical reading.

Author Contributions

Conceived and designed the experiments: YLD SL EC. Performed the experiments: SL LR CLQ FLM PLG. Analyzed the data: YLD SL AA DM EC. Wrote the paper: SL YLD EC.