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RESEARCH

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Twenty patients including 7 probands with autosomal dominant cutis laxa confirm clinical and molecular homogeneity

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Abstract

Background: Elastin gene mutations have been associated with a variety of phenotypes. Autosomal dominant cutis laxa (ADCL) is a rare disorder that presents with lax skin, typical facial characteristics, inguinal hernias, aortic root dilatation and pulmonary emphysema. In most patients, frameshift mutations are found in the 3' region of the elastin gene (exons 30-34) which result in a C-terminally extended protein, though exceptions have been reported.

Methods: We clinically and molecularly characterized the thus far largest cohort of ADCL patients, consisting of 19 patients from six families and one sporadic patient.

Results: Molecular analysis showed C-terminal frameshift mutations in exon 30, 32, and 34 of the elastin gene and identified a mutational hotspot in exon 32 (c.2262delA). This cohort confirms the previously reported clinical constellation of skin laxity (100%), inguinal hernias (51%), aortic root dilatation (55%) and emphysema (37%).

Conclusion: ADCL is a clinically and molecularly homogeneous disorder, but intra- and interfamilial variability in the severity of organ involvement needs to be taken into account. Regular cardiovascular and pulmonary evaluations are imperative in the clinical follow-up of these patients.

Keywords: Elastin, *ELN*, Autosomal dominant cutis laxa, Genotype, Phenotype

Background

Elastin gene (*ELN*) mutations have been associated with various skin, cardiovascular and pulmonary phenotypes. Microdeletions of the region on 7q11, encompassing the *ELN* gene, result in Williams-Beuren syndrome (WBS, OMIM #194050) [1]. This recognizable entity combines mental retardation with a happy demeanor, typical facial gestalt, a soft, slightly elastic and doughy skin, hypercalcaemia, and arterial stenoses, mainly of the supra-valvular aorta. A related disorder is the supra-valvular aortic stenosis (SVAS, OMIM #185500) phenotype. These patients present without the mental and dysmorphic

features encountered in the WBS, but do show arterial stenoses. SVAS is caused by missense and premature truncation mutations that are dispersed throughout the *ELN* gene [2].

In contrast, frameshift mutations at the 3' terminus that result in a C-terminally elongated and secreted elastin protein [3-8] have been identified in autosomal dominant cutis laxa (ADCL, OMIM #123700). This disorder comprises generalized skin redundancy, pulmonary emphysema and aortic root dilatation (ARD).

Different mechanisms underlying the perturbations subsequent to elastin mutations have been identified. In general, reduced elastin secretion and deposition, resulting in increased vascular smooth muscle cell proliferation and a higher number of smaller elastic lamellae, induces a phenotype dominated by arterial stenoses [9], while aberrant elastin protein production infers to elastic fiber

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fragmentation with manifest cutis laxa, arterial dilatation and/or emphysema [3].

To date, frameshift mutations in the 3'UTR of the elastin gene have been identified in 11 probands with ADCL, but it remains a matter of debate whether other molecular mechanisms could be involved. Indeed, two reports indicated a different molecular basis including a heterozygous mutation in exon 25 of the elastin gene [10], and a tandem duplication in the fibulin-5 gene [11], respectively. Moreover, in the absence of large patient cohorts, the ADCL phenotype is still insufficiently characterized and not always easy to discern from related forms of cutis laxa. Furthermore, the identification of an *ELN* variant in exon 34 (c.2318G>A; p.(G773D)) complicated genotype-phenotype correlations in the *ELN* gene. This variant was associated with pulmonary emphysema [12] and, in conjunction with a p.G202R alteration in the fibulin-5 protein, with acquired cutis laxa following a parasitic infection [13]. Finally, a case report mentions causal involvement of the *ELN* gene in a recessive form of cutis laxa due to a p.(P211S) alteration [14].

We report on six novel ADCL families, comprising 19 patients, and one sporadic patient, with a review of the literature. Our data favor homogeneity both at the clinical and molecular level.

Methods

Clinical data

Patients were evaluated by an experienced clinical geneticist or dermatologist. Written informed consent was obtained from each patient and family member before sample collection. Specific consents were obtained for publication of the clinical pictures. This study was approved by the medical ethical committee of Inserm (France) and of the Ghent University Hospital and the Declaration of Helsinki Principles were respected.

Mutation screening

Genomic DNA was extracted from peripheral blood leukocytes at Genethon's DNA Bank using standard procedures, followed by touchdown PCR amplification and sequencing of exons 28 through 34 of the *ELN* gene as previously described [3]. Sequences were compared to the wild-type sequence as submitted to Ensembl Accession number ENST00000358929. Nucleotides were numbered starting from the first base of the initiation codon (ATG) of the cDNA reference sequence. Amino acid residues are numbered from the first methionine residue of the reference sequence. Segregation was verified in all families and mutations were absent in a panel of 100 Caucasian controls.

Results

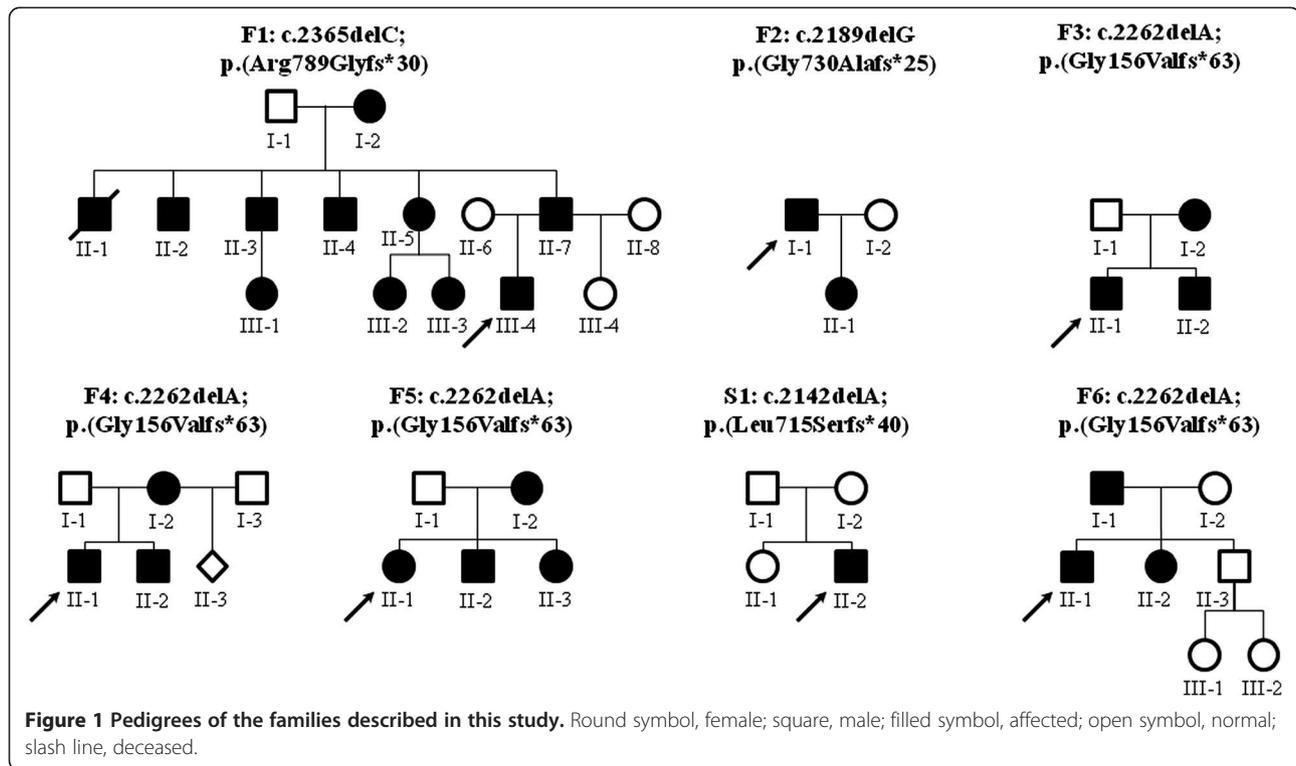
Clinical data

We present six families and one sporadic patient (S1:II-2) with ADCL. Pedigrees of the families are presented in Figure 1. Family 1 originates from the Netherlands, Family F2 from Switzerland, and all others from France. Twenty of the 27 patients were available for clinical evaluation (Table 1). Male to female ratio was 15/12. Curiously, in 20 out of 22 meioses the phenotype was inherited ($\chi^2=8.84$; p-value=0.003). Ages at last clinical evaluation ranged from 1 to 84 years, with 11 patients being over 21 years old.

All patients had areas of cutis laxa. A minority of patients (25%) showed skin redundancy only in the facial, neck, inguinal and/or axillary regions, while 75% of patients had extensive to generalized cutis laxa. The facial skin showed a spectrum ranging from a coarse face over an aged appearance to overt cutis laxa. Especially the lower half of the face and neck region were involved with sagging cheeks, a flabby skin around the chin and skin redundancy in the neck region. Blepharochalasis and ptosis of the upper eyelid were often present (Figure 2J). The skin of the limbs was relatively spared. Overall, the skin had a soft feeling to the touch with variable hyperextensibility (Figure 2L). Skin abnormalities tended to improve with age but were highly variable between and within families. Surgery both for aesthetic reasons [sagging breasts (F1:II-2, F1:III-1) or lax abdominal skin (F2:I-1 and F2:II-1)] and for functional reasons [ptosis (F1:II-2)] was disappointing with frequent relapse. Wound healing was reported normal, although F1:III-1 mentioned recurrent superficial erosions, wound infections and nail fragility.

Craniofacial dysmorphism was mild but recognizable and showed a long face, large pliant ears, and a long philtrum (Figure 2). A beaked nose was present in most patients (70%).

Inguinal hernias were frequent, appearing congenitally (F1:II-2, F1:II-3, F1:III-1, F2:II-1) or throughout life (F1:I-2 at age 80 and F1:III-4 at 18 years old) and often re-occurred after surgery (patient F1:II-2, F1:II-3, F1:II-3, F1:III-1). Patient F1:II-4 also had an umbilical hernia. Systemic involvement was noted in 15 out of 20 patients (75%). Chronic obstructive pulmonary disease was diagnosed in eight patients, seven of whom had emphysema and one had asthma. F1:II-2 and F1:II-1 were significant smokers and the latter died at age 55 due to end-stage respiratory failure with emphysematous lung changes. Vascular involvement included ARD in eight patients, and a dilatation of the aortic arch in one (patient F4:II-2). In addition, patient F3:II-2 had pulmonary artery dilatation and patient F6:II-1 had a global but stable increase of the diameters of the arteries, especially of the carotid arteries (diameter increase of 30%). Four



patients had aortic valve regurgitation (F1:II-4; F5:II-2, F5:II-3, F6:II-1), patient F1:II-3 had tricuspid valve stenosis, and patients F1:II-4 and F6:II-1 had mitral valve regurgitation. Other manifestations included gastro-oesophageal reflux (F1:II-7 and F1:III-4), urogenital prolapse (F1:II-2), and a hoarse voice (F2:I-1, F2:II-1, F3:II-1, and F3:II-2).

Hypomobility of spermatozooids required *in vitro* fertilization for one patient (F1:II-3). None of the patients reported ocular problems. Growth and psychomotor development were appropriate in all patients.

Genetic analysis

The *ELN* mutations identified in these families are reported in Table 2 and Figure 1. We found three novel mutations: p.2189delG; p.(Gly730Alafs*25) and c.2142delG; p.(Leu715Serfs*40) in exon 30; and c.2365delC; p.(Arg789Glyfs*30) in exon 34. One previously reported mutation (c.2262delA (p.Gly156Valfs*63)) in exon 32 was found in four families and may represent a mutational hotspot. All mutations are predicted to result in a C-terminal missense sequence with a read-through in the 3' UTR. Although exon 30 mutations induce a premature truncation in exon 32, (partial) splicing of this exon will result in a C-terminally elongated missense protein as previously reported for similar mutations [3,5].

Genotype-phenotype correlation

There are no obvious genotype-phenotype correlations when comparing patients with mutations in different

exons (Table 3). The recurrent c.2262delA mutation shows large interfamilial variability. Moreover, family 1, 3 and 5 also document high intrafamilial variability, both in skin features and internal organ involvement.

Although the risk for ARD was suggested to be smaller in patients with exon 32 mutations [3], we did identify vascular involvement in this patient subgroup. However, ARD seems more prominent in association with exon 30 mutations (Table 3), but further confirmation is needed.

Discussion

We present the largest patient cohort with ADCL thus far. The phenotype is homogeneous and comprises generalized skin involvement, inguinal and umbilical hernias, ARD and emphysema. The disorder shows full penetrance, but with variable expression. The aspect and topography of the skin lesions is highly variable but the skin changes are most pronounced on the axillar, inguinal, neck and lower facial regions. In congruence with earlier reports [5,8], skin manifestations improve with aging and the phenotype may be unrecognized in older patients. Although ADCL is often considered 'benign', systemic involvement, although variable, is frequently present. Both lung (present in 35% of patients) and aortic involvement (present in 57% of patients) have led to premature death [5,7]. Patient management therefore warrants close monitoring of pulmonary function and aortic root diameters. No current treatment guidelines are available for ADCL patients. Because earlier reports

Table 1 Clinical details of the patients available for clinical evaluation

Patient	Sex, Age (y)	Skin							Inguinal hernia Left (L)/Right (R)/Bilateral (B) (times surgery)	Cardiovascular	Pulmonary emphysema	Other
		Aspect	Topography									
			Trunk	Axillar region	Inguina region	Limbs	Neck	Face				
F1:I-1	F, 84	Redundant	+	+	+				L (1)	-	+	Bilateral ptosis, sagging breasts, pelvic prolapse
F1:II-2	M, 59	Smooth, sagging, loose		+	+				B (10).	-	+	Pliant ears
F1:II-3	M, 57	Smooth, redundant sagging	+	+					B (12)	TVS ARD	+	Nycturia, decreased fertility
F1:II-4	M, 53	Sagging loose, sagging, redundant		+	+	+	+	+	B (15)	ARD (85mm – surgery), AR, MR	+	Umbilical hernia Inguinal hernia surgery (13 times)
F1:II-5	F, 51	Sagging loose, sagging, redundant	+	+	+				-	-	-	
F1:II-7	M, 46	Loose, sagging	+						-	-	-	Gastroesophageal reflux disease, strabismus
F1:III-1	F, 23	Sagging, loose, redundant	+	+	+				B (2)	-	-	Sagging breasts
F1:III-4	M, 22	Sagging	+						R (2)	ARD	-	Gastroesophageal reflux disease, coarse face
F2:I-1	M, 48	Redundant		+	+				-	-	+	Hoarse voice, corrective skin surgery
F2:II-1	F, 21	Redundant		+	+				B (0)	MVP	+	Hoarse voice, corrective skin surgery (3 times)
F3:I-2	F, 31	Excessive, sagging	+	+	+	+	+	+	-	ARD	-	
F3:II-1	M, 7	Excessive, sagging							-	ARD	-	Hoarse voice
F3:II-2	M, 4	Excessive, sagging	+	+	+	+	+	+	-	Pulmonary artery dilatation	Asthma	Atopic dermatitis, hoarse voice
F4:II-1	M, 16	Redundant	+	+	+				-	ARD		Coarse face
F4:II-2	M, 14	-							-	Dilated aortic arch (27mm)		Coarse face
F5:II-1	F, 14	Excessive, sagging	+	+	+	+	+	+	-	-	-	Labiae major coalescence
F5:II-2	M, 10	Excessive, sagging	+	+	+	+	+	+	-	AR	-	Testis ectopia
F5:II-3	F, 7	Excessive, sagging	+	+	+	+	+	+	-	AR, Borderline ARD (23mm)	-	
S1:II-2	M, 1	Excessive, sagging,	+	+	+				-	ARD		
F6:II-1	M, 39	Redundant	+	+					-	AR, global increase of arterial diameter	-	

M, male; F, female; y, years; L, left; R, right; B, bilateral; AR, aortic valve regurgitation; ARD, aortic root dilatation; MVP, mitral valve prolapse; MR, mitral valve regurgitation; TVS, tricuspid valve stenosis. F1 to F6: familial cases, S1: sporadic case.



indicated increased TGF β signaling in the pathogenesis [3], thresholds for aortic surgery could be based on similar parameters as in elastinopathies/TGF β signalopathies associated with ARD (actual aortic diameter, rate of aortic growth, family history, the patient's estimation of the risk, and pregnancy wish) [15]. Moreover, this mechanism might reveal a role for treatment with losartan, an

angiotensin II type 1 receptor blocker with TGF β antagonistic effect. Surgery for skin manifestations and hernias is disappointing and should be discouraged if done purely for aesthetic reasons.

The phenotype can be distinguished from autosomal recessive types of cutis laxa (ARCL) based on facial gestalt, the absence of skeletal, ocular, and central nervous involvement (type 2 and 3 ARCL) [16,17], and the absence of severe early onset emphysema and gastrointestinal and genitourinary diverticula (type 1a/c ARCL [18]) or arterial involvement (type 1b related cutis laxa) [19] (Table 4). The small number of patients described to date does not allow drawing straight-forward genotype-phenotype correlations, although a tendency for a milder vascular phenotype exists in exon 32 mutations. Mutations in this exon might be partially rescued by enhanced splicing of this exon [3].

The clinical homogeneity of the phenotype corresponds to a striking molecular homogeneity with mutations found

Table 2 ELN mutations identified in this cohort

Patients	Origin	cDNA level	Protein level
Family 1	Netherlands	c.2365delC (exon 34)	p.(Arg789GlyfsX30)
Family 2	Switzerland	c.2189delG (exon 30)	p.(Gly730AlafsX25)
Family 3	France	c.2262delA (exon32)	p.(Gly156ValfsX63)
Family 4	France	c.2262delA (exon32)	p.(Gly156ValfsX63)
Family 5	France	c.2262delA (exon32)	p.(Gly156ValfsX63)
Sporadic case	France	c.2142delG (exon 30)	p.(Leu715Serfsx40)
Family 6	France	c.2262delA (exon32)	p.(Gly156ValfsX63)

Table 3 Genotype – phenotype correlations in ADCL

Missense	F/M individuals	Mean age	FH	IH	FG	ARD	BAV	VA	Emphys	transmission
Exon 30	5/7	21.9	50	64	100	82	10	70	40	100
Exon 32	6/6	17.1	90	17	100	50	0	25	0	95
Exon 33	2/2	21	33	50	100	50	25	50	100	100
Exon 34	3/5	49.5	100	75	100	38	0	25	50	91
Chr rearr	3/0	42	100	67	100	?	0	0	100	56
Total	19/20	27.3	63	50	100	57	5	38	35	83

Frequencies (%) of the clinical characteristics are given for mutations in the different exons. Chr rear, Chromosomal rearrangement as described by [7]. Mean age in years. F, female; M, male; FH, family history; IH, Inguinal hernia; FG, Facial gestalt; ARD, aortic root dilatation; BAV, bicuspid aortic valve; VA, other valve anomalies; Emphys, emphysema. References: [3-8].

in the 3' terminus of the *ELN* gene. All mutations reside in exons 30, 32, 33, and 34 and result in a missense and elongated sequence at the C-terminus of the elastin protein [3-8]. Mutation analysis should focus firstly on these exons, although splice site mutations elsewhere in the *ELN* gene could also result in a C-terminal extension of the protein. Frameshift mutations resulting in C-terminal protein extensions have been identified in a variety of other diseases resulting in mislocalization, altered secretion, increased or decreased stability, resistance to proteases, loss-of-function, gain-of-function, and novel functions of the affected protein [23-28]. Extended proteins may also result from impaired processing of propeptides. A well-known example within the field of connective tissue disorders is the Ehlers-Danlos syndrome type VII that results from mutations in the substrate collagen type 1 or in its type 1 procollagen N-proteinase (type VIIC). In both cases the N-terminus of the procollagen type 1 is not removed and the mutant collagen is deposited into the extracellular matrix with severe dermal and internal organ consequences [29,30]. In ADCL it was shown that the altered and extended C-terminus resulted in increased

endoplasmic reticulum stress (due to protein misfolding), increased coacervation and elastin globule formation leading to lower amounts of insoluble, mature elastin. Enhanced TGFβ signaling is probably secondary to altered elastic fiber anatomy [3].

Two reports indicated molecular heterogeneity in ADCL. In a single ADCL patient, a tandem duplication in the *FBLN5* gene was shown to result in secretion of mutant protein, suggesting a dominant negative effect [11]. However, mutant protein integration in the extracellular matrix has been shown in *FBLN5* related ARCL type 1 [31] and carriers of these mutations do not have a cutis laxa phenotype. A second report showed a mutation in exon 25 of the *ELN* gene which encodes a conserved cross-linking domain important for insoluble elastin formation [10]. This patient also presented with severe neurological involvement, which was not attributed to his cutis laxa syndrome. His father carried the same alteration, but did not exhibit any phenotype. Future confirmation of these two defects may shed further insights on the molecular heterogeneity of ADCL. Until then, ADCL can be considered as a phenotypically and genetically homogeneous disorder in which analysis

Table 4 Clinical characteristics of ADCL and type 1 recessive cutis laxa syndromes

	ADCL (<i>ELN</i>)	ARCL type 1a (<i>FBLN5</i>)	ARCL type 1c (<i>LTBP4</i>)	ARCL type 1b (<i>FBLN4</i>)
Dysmorphism	Long philtrum, beaked nose	Long philtrum, large ears, beaked nose	Long philtrum, large ears, beaked nose, sparse hair on temporal sides	
Skin	Loose redundant skin folds Coarse face	Loose redundant skin folds	Loose redundant skin folds	Hyperextensible skin Skin redundancy
Cardiovascular	ARD, valvular anomalies	SVAS, PPAS	PPAS, CPAS	ARD, multiple arterial aneurysms
Emphysema	+	+++	+++	+/-
Genitourinary diverticula	+	+++	++	?
Gastro-intestinal diverticula/tortuosity	-	+	+++	?
Inheritance	AD	AR	AR	AR

ARD, aortic root dilatation; SVAS, supra-aortic root stenosis; PPAS, peripheral pulmonary artery stenosis; CPAS, common pulmonary artery stenosis; AD, autosomal dominant; AR, autosomal recessive. References: [3-8,18-22].

of exon 30-34 of the *ELN* gene is indicated in first instance.

Conclusion

This cohort of patients further indicates that ADCL is a systemic disorder with a risk for major complications including aortic root dilatation and pulmonary emphysema, warranting regular follow-up. The molecular data favor a single mutational mechanism resulting in a stable C-terminal extension of the elastin protein.

Abbreviations

ADCL: Autosomal dominant Cutis Laxa; ARCL: Autosomal recessive cutis laxa; ARD: Aortic root dilatation; *ELN*: Elastin; SVAS: Supravalvular aortic stenosis; WBS: Williams-Beuren syndrome; TGF β : Transforming Growth Factor β .

Competing interests

The authors declare no competing interests.

Authors' contributions

SHR and BLC reviewed patients, designed the study and drafted the manuscript. EB, MK, ASP, VL and DB reviewed patients. MR and PJC performed and supervised the molecular analyses. JDB, FM, OMV and ADP contributed to clinical assessment, sample collection and drafting of the manuscript. CB reviewed patients, and contributed in the design of the study. All authors have read and approved the final manuscript.

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References

1. Ewart AK, Morris CA, Atkinson D, Jin W, Sternes K, Spallone P, Stock AD, Leppert M, Keating MT: **Hemizyosity at the elastin locus in a developmental disorder, Williams syndrome.** *Nat Genet* 1993, **5**:11–16.
2. Curran ME, Atkinson DL, Ewart AK, Morris CA, Leppert MF, Keating MT: **The elastin gene is disrupted by a translocation associated with supravalvular aortic stenosis.** *Cell* 1993, **73**:159–168.
3. Callewaert B, Renard M, Huchtagowder V, Albrecht B, Hauser I, Blair E, Dias C, Albino A, Wachi H, Sato F, et al: **New insights into the pathogenesis of**

- autosomal-dominant cutis laxa with report of five *ELN* mutations. *Hum Mutat* 2011, **32**:445–455.
4. Rodriguez-Revena L, Iranzo P, Badenas C, Puig S, Carrio A, Mila M: **A novel elastin gene mutation resulting in an autosomal dominant form of cutis laxa.** *Arch Dermatol* 2004, **140**:1135–1139.
5. Szabo Z, Crepeau MW, Mitchell AL, Stephan MJ, Puntel RA, Yin Loke K, Kirk RC, Urban Z: **Aortic aneurysmal disease and cutis laxa caused by defects in the elastin gene.** *J Med Genet* 2006, **43**:255–258.
6. Tassabehji M, Metcalfe K, Hurst J, Ashcroft GS, Kiely C, Wilmot C, Donnai D, Read AP, Jones CJ: **An elastin gene mutation producing abnormal tropoelastin and abnormal elastic fibres in a patient with autosomal dominant cutis laxa.** *Hum Mol Genet* 1998, **7**:1021–1028.
7. Urban Z, Gao J, Pope FM, Davis EC: **Autosomal dominant cutis laxa with severe lung disease: synthesis and matrix deposition of mutant tropoelastin.** *J Invest Dermatol* 2005, **124**:1193–1199.
8. Zhang MC, He L, Giro M, Yong SL, Tiller GE, Davidson JM: **Cutis laxa arising from frameshift mutations in exon 30 of the elastin gene (*ELN*).** *J Biol Chem* 1999, **274**:981–986.
9. Pober BR, Johnson M, Urban Z: **Mechanisms and treatment of cardiovascular disease in Williams-Beuren syndrome.** *J Clin Invest* 2008, **118**:1606–1615.
10. Graul-Neumann LM, Hauser I, Essayie M, Rauch A, Kraus C: **Highly variable cutis laxa resulting from a dominant splicing mutation of the elastin gene.** *Am J Med Genet A* 2008, **146A**:977–983.
11. Markova D, Zou Y, Ringpfeil F, Sasaki T, Kostka G, Timpl R, Uitto J, Chu ML: **Genetic heterogeneity of cutis laxa: a heterozygous tandem duplication within the fibulin-5 (*FBLN5*) gene.** *Am J Hum Genet* 2003, **72**:998–1004.
12. Kelleher CM, Silverman EK, Broekelmann T, Litorjua AA, Hernandez M, Sylvia JS, Stoler J, Reilly JJ, Chapman HA, Speizer FE, et al: **A functional mutation in the terminal exon of elastin in severe, early-onset chronic obstructive pulmonary disease.** *Am J Respir Cell Mol Biol* 2005, **33**:355–362.
13. Hu Q, Raymond JL, Pinel N, Zabot MT, Urban Z: **Inflammatory destruction of elastic fibers in acquired cutis laxa is associated with missense alleles in the elastin and fibulin-5 genes.** *J Invest Dermatol* 2006, **126**:283–290.
14. Megarbane H, Florence J, Oliver Sass J, Schwonbeck S, Foglio M, de Cid R, Cure S, Saker S, Megarbane A, Fischer J: **An autosomal-recessive form of cutis laxa is due to homozygous elastin mutations, and the phenotype may be modified by a heterozygous fibulin 5 polymorphism.** *J Invest Dermatol* 2009, **129**:1650–1655.
15. Loeys BL, Dietz HC, Braverman AC, Callewaert BL, De Backer J, Devereux RB, Hilhorst-Hofstee Y, Jondeau G, Faivre L, Milewicz DM, et al: **The revised Ghent nosology for the Marfan syndrome.** *J Med Genet* 2010, **47**:476–485.
16. Kornak U, Reynders E, Dimopoulou A, van Reeuwijk J, Fischer B, Rajab A, Budde B, Nurnberg P, Foulquier F, Lefeber D, et al: **Impaired glycosylation and cutis laxa caused by mutations in the vesicular H⁺-ATPase subunit ATP6VOA2.** *Nat Genet* 2008, **40**:32–34.
17. Reversade B, Escande-Beillard N, Dimopoulou A, Fischer B, Chng SC, Li Y, Shboul M, Tham PY, Kayserili H, Al-Gazali L, et al: **Mutations in *PYCR1* cause cutis laxa with progeroid features.** *Nat Genet* 2009, **41**:1016–1021.
18. Callewaert B, Su CT, Van Damme T, Vlummens P, Malfait F, Vanakker O, Schulz B, Mac Neal M, Davis EC, Lee JG, et al: **Comprehensive clinical and molecular analysis of 12 families with type 1 recessive cutis laxa.** *Hum Mutat* 2013, **34**:111–121.
19. Renard M, Holm T, Veith R, Callewaert BL, Ades LC, Baspinar O, Pickart A, Dasouki M, Hoyer J, Rauch A, et al: **Altered TGF β signaling and cardiovascular manifestations in patients with autosomal recessive cutis laxa type I caused by fibulin-4 deficiency.** *Eur J Hum Genet* 2010, **18**:895–901.
20. Huchtagowder V, Sausgruber N, Kim KH, Angle B, Marmorstein LY, Urban Z: **Fibulin-4: a novel gene for an autosomal recessive cutis laxa syndrome.** *Am J Hum Genet* 2006, **78**:1075–1080.
21. Loeys B, Van Maldergem L, Mortier G, Coucke P, Gerniers S, Naeyaert JM, De Paepe A: **Homozygosity for a missense mutation in fibulin-5 (*FBLN5*) results in a severe form of cutis laxa.** *Hum Mol Genet* 2002, **11**:2113–2118.
22. Urban Z, Huchtagowder V, Schurmann N, Todorovic V, Zilberberg L, Choi J, Sens C, Brown CW, Clark RD, Holland KE, et al: **Mutations in *LTBP4* cause a syndrome of impaired pulmonary, gastrointestinal, genitourinary, musculoskeletal, and dermal development.** *Am J Hum Genet* 2009, **85**:593–605.
23. Arlt G, Brooks DA, Isbrandt D, Hopwood JJ, Bielicki J, Bradford TM, Bindloss-Petherbridge CA, von Figura K, Peters C: **Juvenile form of**

- mucopolysaccharidosis VI (Maroteaux-Lamy syndrome). A C-terminal extension causes instability but increases catalytic efficiency of arylsulfatase B. *J Biol Chem* 1994, **269**:9638–9643.
24. Choi BY, Kim DH, Chung T, Chang M, Kim EH, Kim AR, Seok J, Chang SO, Bok J, Kim D, *et al*: Destabilization and Mislocalization of POU3F4 by C-Terminal Frameshift Truncation and Extension Mutation. *Hum Mutat* 2013, **34**:309–316.
 25. Dickey DM, Yoder AR, Potter LR: A familial mutation renders atrial natriuretic Peptide resistant to proteolytic degradation. *J Biol Chem* 2009, **284**:19196–19202.
 26. Komaru K, Ishida Y, Amaya Y, Goseki-Sone M, Orimo H, Oda K: Novel aggregate formation of a frame-shift mutant protein of tissue-nonspecific alkaline phosphatase is ascribed to three cysteine residues in the C-terminal extension. Retarded secretion and proteasomal degradation. *FEBS J* 2005, **272**:1704–1717.
 27. Soldovieri MV, Castaldo P, Iodice L, Miceli F, Barrese V, Bellini G, Miraglia del Giudice E, Pascotto A, Bonatti S, Annunziato L, Tagliatela M: Decreased subunit stability as a novel mechanism for potassium current impairment by a KCNQ2 C terminus mutation causing benign familial neonatal convulsions. *J Biol Chem* 2006, **281**:418–428.
 28. Bohm J, Heinritz W, Craig A, Vujic M, Ekman-Joelsson BM, Kohlhase J, Froster U: Functional analysis of the novel TBX5 c.1333delC mutation resulting in an extended TBX5 protein. *BMC Med Genet* 2008, **9**:88.
 29. Colige A, Sieron AL, Li SW, Schwarze U, Petty E, Wertenleki W, Wilcox W, Krakow D, Cohn DH, Reardon W, *et al*: Human Ehlers-Danlos syndrome type VII C and bovine dermatosparaxis are caused by mutations in the procollagen I N-proteinase gene. *Am J Hum Genet* 1999, **65**:308–317.
 30. Smith LT, Wertenleki W, Milstone LM, Petty EM, Seashore MR, Braverman IM, Jenkins TG, Byers PH: Human dermatosparaxis: a form of Ehlers-Danlos syndrome that results from failure to remove the amino-terminal propeptide of type I procollagen. *Am J Hum Genet* 1992, **51**:235–244.
 31. Hu Q, Loeys BL, Coucke PJ, De Paepe A, Mecham RP, Choi J, Davis EC, Urban Z: Fibulin-5 mutations: mechanisms of impaired elastic fiber formation in recessive cutis laxa. *Hum Mol Genet* 2006, **15**:3379–3386.

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