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Estelle Marion, Sara Eyangoh, Edouard Yeramian, Julien Doannio, Jordi Landier, et al.. Seasonal and regional dynamics of *M. ulcerans* transmission in environmental context: deciphering the role of water bugs as hosts and vectors.. PLoS Neglected Tropical Diseases, 2010, 4 (7), pp.e731. 10.1371/journal.pntd.0000731 . inserm-00691396

HAL Id: inserm-00691396

<https://inserm.hal.science/inserm-00691396v1>

Submitted on 26 Apr 2012

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Seasonal and Regional Dynamics of *M. ulcerans* Transmission in Environmental Context: Deciphering the Role of Water Bugs as Hosts and Vectors

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Abstract

Background: Buruli ulcer, the third mycobacterial disease after tuberculosis and leprosy, is caused by the environmental mycobacterium *M. ulcerans*. Various modes of transmission have been suspected for this disease, with no general consensus acceptance for any of them up to now. Since laboratory models demonstrated the ability of water bugs to transmit *M. ulcerans*, a particular attention is focused on the transmission of the bacilli by water bugs as hosts and vectors. However, it is only through detailed knowledge of the biodiversity and ecology of water bugs that the importance of this mode of transmission can be fully assessed. It is the objective of the work here to decipher the role of water bugs in *M. ulcerans* ecology and transmission, based on large-scale field studies.

Methodology/Principal Findings: The distribution of *M. ulcerans*-hosting water bugs was monitored on previously unprecedented time and space scales: a total of 7,407 water bugs, belonging to large number of different families, were collected over one year, in Buruli ulcer endemic and non endemic areas in central Cameroon. This study demonstrated the presence of *M. ulcerans* in insect saliva. In addition, the field results provided a full picture of the ecology of transmission in terms of biodiversity and detailed specification of seasonal and regional dynamics, with large temporal heterogeneity in the insect tissue colonization rate and detection of *M. ulcerans* only in water bug tissues collected in Buruli ulcer endemic areas.

Conclusion/Significance: The large-scale detection of bacilli in saliva of biting water bugs gives enhanced weight to their role in *M. ulcerans* transmission. On practical grounds, beyond the ecological interest, the results concerning seasonal and regional dynamics can provide an efficient tool in the hands of sanitary authorities to monitor environmental risks associated with Buruli ulcer.

Citation: Marion E, Eyangoh S, Yeramian E, Doannio J, Landier J, et al. (2010) Seasonal and Regional Dynamics of *M. ulcerans* Transmission in Environmental Context: Deciphering the Role of Water Bugs as Hosts and Vectors. PLoS Negl Trop Dis 4(7): e731. doi:10.1371/journal.pntd.0000731

Editor: Mathieu Picardeau, Institut Pasteur, France

Received: October 12, 2009; **Accepted:** May 12, 2010; **Published:** July 6, 2010

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Funding: This work was supported by the Fondation Francaise Raoul Follereau, by the Institut National de la Sante et de la Recherche Medicale (INSERM), by European community (FEDER 10250), by Region Pays de Loire, by Pasteur Network (PTR 212), and by Fondation Pierre Ledoux-Jeunesse Internationale. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

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Introduction

Mycobacterium ulcerans is the causative agent of Buruli ulcer. This devastating necrotic human skin disease is the third most common mycobacterial disease, after tuberculosis and leprosy. The detection rates of Buruli ulcer were found to increase gradually and steadily. The range of the disease extends from 10°N to 10°S latitude in Africa and spans 16 endemic countries, 10 potential endemic countries and 20 non endemic countries. The majority of cases are localized in Africa, with cases also reported in Asia, Australia and South America. In Africa, Buruli ulcer occurs

mainly in poor rural communities [1–5]. As a consequence, very often treatment is sought and prescribed too late. Treatment of later stages requires extensive surgery at major hospitals, involving prolonged and very expensive stays, with 25% of those who experienced Buruli ulcer -in particular children- becoming permanently disabled [6–7]. More recently, it was possible to cure the disease at an early stage with antibiotics, without the need for surgery. The administration of a combination of rifampin and an aminoglycoside for four to eight weeks led to the healing of early lesions, without radical surgery. This antibiotics-based treatment is now the recommended standard regimen in areas

Author Summary

Buruli ulcer, caused by *Mycobacterium ulcerans*, is a devastating skin disease. Most cases of Buruli ulcer occur in poor rural communities. As a result, treatment is frequently sought too late and about 25% of those infected—particularly children—become permanently disabled. Outbreaks of Buruli ulcer have always been associated with swampy areas. However, the route(s) of bacillus transmission is (are) still unclear. This *Mycobacterium* species resides in water where it colonizes many ecological niches such as aquatic plants, herbivorous animals and predatory/carnivorous insects. For several years the role of water bugs as hosts and vectors of *M. ulcerans* was suspected and was demonstrated under laboratory conditions. The aim of this work was to further assess the role of water bugs as hosts and vectors of *M. ulcerans* in environmental context. This work identifies several water bug families as hosts of *M. ulcerans* in Buruli ulcer endemic area. The detection of bacilli in saliva of human biting insects provides further evidence for their role in *M. ulcerans* transmission. Interestingly, three of these insects are good flyers, and as such could participate in *M. ulcerans* dissemination.

where information about Buruli ulcer was made accessible, resulting in early diagnosis of the disease [8–9].

In 1998, the World Health Organization launched the Global Buruli Ulcer Initiative to intensify surveillance, disease control, treatment and also to promote understanding of the ecology and the mode of transmission of *M. ulcerans*.

There is at present no clear understanding of the exact mode(s) of transmission of *M. ulcerans*. Populations affected by Buruli ulcer are those living close to humid and swampy zones. Indeed the foci of the disease are associated with the creation or the extension of swampy areas, such as construction of dams or lakes for the development of agriculture (irrigation) [2–3,10–17].

Over the last few decades, different mechanisms have been proposed for the transmission of *M. ulcerans* from aquatic environments to human skin, ranging from aerosol contamination (an hypothesis invoked in Australia but never confirmed) [12] to insect-dependent transmission. The role of insects is indeed suggested by various studies over the past ten years. Recently, *M. ulcerans* DNA was detected in 0.04% of mosquito populations [18–20]. One possible transmission route is through the subversion of *Aedes* and *Anopheles* aquatic larvae (inhabiting *M. ulcerans*-loaded water) as hosts, with the human blood-feeding adults delivering *M. ulcerans* in the skin. However at present this hypothesis has not been confirmed experimentally, with no detection of *M. ulcerans* in the saliva or salivary glands. As the blood-feeding adults emerging from aquatic larvae cannot account for *M. ulcerans* transmission in any general way, different investigators have proposed that biting water bugs could act as vectors for *M. ulcerans*. In 1999, Portaels was first to raise this hypothesis, and also to isolate *M. ulcerans* from water bug tissues [21–22]. Our pioneering experimental, laboratory-based, studies on mice have given weight to this hypothesis. These studies showed that *M. ulcerans* was able to colonize the salivary glands of water bugs, which could then transmit *M. ulcerans* to mice through biting [23–28]. Other field investigations allowed detecting *M. ulcerans* DNA in the water bugs captured in Buruli ulcer endemic areas [21,28–30]. On the other hand, a field study conducted in Ghana [30] did not support the role of biting Hemiptera or other invertebrates as possible *M. ulcerans* hosts/reservoirs or vectors. This study rather pointed out the need for

further research to better understand *M. ulcerans* transmission [31]. In this context, our first study on the ability of water bugs to transmit bacterium through biting was confirmed recently in an invertebrate model [32].

Water bugs are familiar insects in aquatic habitats throughout the world. They are present in all Buruli ulcer endemic areas. They belong to the order of Hemiptera, containing several families. There are basically two kinds of water bugs: the semi-aquatic bugs, which live upon the water surface and the true water bugs, which live beneath the water surface. These invertebrates live in a wide variety of natural habitats, lakes and rivers ranging from small to large and also small ponds. Most water bugs can be characterized as predatory feeders, preying on aquatic invertebrates (insect larvae, snails, etc.) [33]. Water bugs can also feed on small vertebrates such as fishes and amphibians. In addition, a water bug family was reported to feed on plant material [33]. Most water bug species are able to fly, flying mainly at night when attracted by light, a feature that could account for *M. ulcerans* dissemination in the environment as suggested by Portaels and Meyers [34]. In this direction, *M. ulcerans* DNA was detected recently in water bugs collected out of their aquatic environment, thus demonstrating their flying capacity (Marsollier, personal communication).

In tropical areas, water bug biodiversity and biology are poorly documented, making it difficult (i) to define their role as *M. ulcerans* hosts and vectors and (ii) to characterize the relations between *M. ulcerans* and these aquatic insects.

In this context, the present investigations had three objectives:

- 1- To establish an inventory of water bug genera and species in central Cameroon, comparing Buruli ulcer endemic and non endemic areas.
- 2- To investigate the water bug population dynamics throughout the seasons.
- 3- To monitor the presence of *M. ulcerans* DNA i) not only in bodies of water bugs from the inventoried family but also ii) in the saliva collected from these water bugs during the four seasonal periods. Of note, when the *M. ulcerans* DNA positive saliva was inoculated in the mouse tail skin, lesions displaying features of Buruli ulcer did develop.

Materials and Methods

Sites of study

Regular sampling of aquatic insects was performed between October 2007 and July 2008, in two areas of the Centre Province of Cameroon: a Buruli ulcer endemic area in the Nyong River basin (Akonolinga, 3.77334N 12.24135E) and a Buruli ulcer non endemic area (Mbalmayo, 3.51552N 11.50085E) situated 100 km downstream (Figure 1). The two areas, along the Nyong River, were selected on the basis of their accessibility all year long (i.e. including in rainy season) and of the availability of relevant epidemiological studies. Prevalence of Buruli ulcer endemic site (Akonolinga district) is estimated at 0.47% [16,35–36] and no case of Buruli ulcer has been reported at this day in Mbalmayo. Both populations are primarily involved in fishing, which is supplemented by riverbank agriculture in the endemic site. The population densities are 21 h/km² and 40 h/km² in the endemic and non-endemic sites, respectively. Six and two large sampling collections were carried out respectively in the endemic site (October, November, December, January, April and July) and in the non-endemic site (April and July). Water bodies were sampled from the main water sources: domestic washing, bathing, fishing

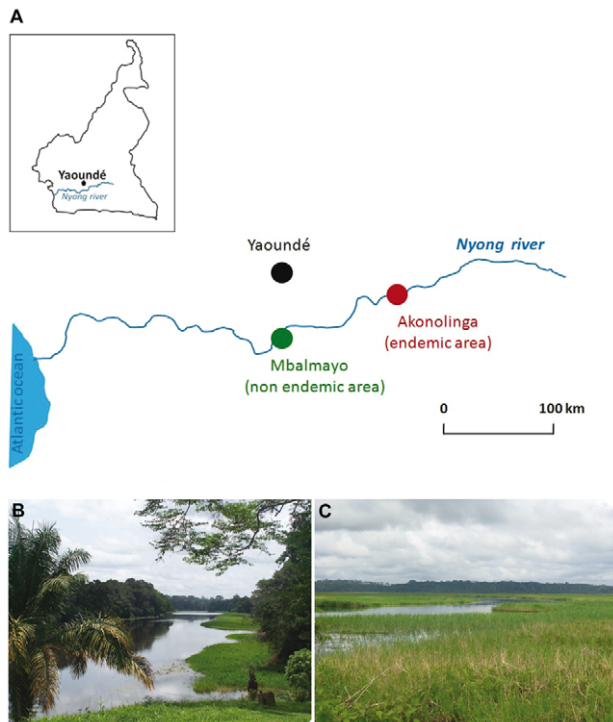


Figure 1. Locations of sampling sites along the Nyong River (Cameroon). (A) The samples of aquatic insects were collected on the Nyong river in an endemic area (Akonolinga) and in a non endemic area (Mbalmayo) for Buruli ulcer. In the non endemic site, the bank of the Nyong (B) harbours a dense evergreen forest contrary to the endemic sites with an open vegetation landscape (C) caused by intense agricultural activities.

doi:10.1371/journal.pntd.0000731.g001

and recreational sources. It is also noticeable that the sites under study corresponded to meeting points for the population, to cross the Nyong River in dugout canoes.

Aquatic insect capture, sampling and pooling

Sampling was conducted between 8:00 am and 12:00 noon for all sites, with the same sampling methods. As aquatic bugs are associated in general with aquatic plants, the exploration was restricted to this ecological niche, along the bank of the Nyong River. In order to minimize escape of insects, a canoe was used to access the capture site. The insects were captured with a square-net (32×32 cm and 1 mm in mesh size) from the surface to a depth of 1 meter, and over a distance of 1 meter. Each month, 5 samplings were performed on each of 3 consecutive mornings. A given sample corresponds to the mix of all insects collected after 10 sweeps. All insects were preserved in 70% ethanol for laboratory identification. The adults as well as nymph insects were numbered individually for taxa identification. To detect *M. ulcerans* DNA, the insects were sorted into pooled groups, including fewer than 10 specimens from the same family.

Water bug family identification

The water bugs were classified in phylum Arthropoda, class Insecta, order Hemiptera and suborder Heteroptera. The main criteria for Heteroptera identification were as follows :

- piercing-sucking mouthparts, with a segmented rostrum arising from the front of the head

- two pairs of wings in adults: partly membranous forewings - hemelytra- and fully membranous hind wings

The classification of the collected samples into families was performed based on the application of the heteroptera family determination criteria to each specimen [37].

Insect saliva collection

Additional Belostomatidae (*Appasus sp.*) were selectively gathered by sweep sampling. They were transported to the laboratory in plastic containers with an air pump (Pafex 3 aerator, Pafex) in fresh water. It should be noted that these collected insects were not included in the counting of water bugs to study variation of water bug density and detection of *M. ulcerans* in water bug tissues. The saliva was collected by a method similar to that used for mosquito saliva [38–39], with appropriate modifications. As a difference to the case of mosquito salivation, there was no need to remove legs and wings in order to sedate the insect and to inoculate a solution in the thorax region. The body of the water bug was grasped with metallic blunt pincers and its rostrum was placed into a conventional plastic pipette tip containing 10 µl of sterile water. With such manipulation white saliva fluid could be observed after 2 minutes time. From each individual saliva sample, 5 µl were used for quantitative PCR and 5 µl kept for Ziehl-Neelsen staining and mouse tail inoculation experiments in PCR positive cases (Figure S1).

DNA extraction from water bug tissues and saliva

Pooled insect bodies were ground and homogenized in 50 mM NaOH solution. Tissue homogenates were heated at 95°C for 20 min. The samples were neutralized by 100 mM Tris-HCl, pH 8.0. DNA from homogenized insect tissues was purified using Power Soil DNA isolation kit (MO Bio lab, Carlsbad, CA), according to manufacturer's recommendations. 5 µl of each individual saliva sample was resuspended in 50 µl of 50 mM NaOH, heated and neutralized as described above. No purification was needed because no PCR inhibitor was present after DNA extraction from saliva. To eliminate DNA traces after each extraction, homogenizers were decontaminated overnight in 1 M NaOH and rinsed in distilled water before sterilization at 130°C for 20 min.

Detection of *M. ulcerans* DNA by quantitative PCR

Oligonucleotide primer and TaqMan probe sequences were selected from the GenBank IS2404 sequence [40] and the ketoreductase B (KR) domain of the mycolactone polyketide synthase (mks) gene (Table 1) from the plasmid pMUM001 [40–43]. PCR mixtures contained 5 µl of template DNA, 0.3 µM

Table 1. Primers and probes used to detect *M. ulcerans* DNA sequences by Taq Man real-time PCR.

Primer or Probe Name	Sequence (5' to 3')
IS2404 forward primer	ATTGGTGGCCGATCGAGTTG
IS2404 reverse primer	TCGCTTTGGCCGCTAAA
IS2404 probe	FAM-CACCACGCAGCATTCTTGCCGT-TAMRA
KR-B forward primer	TCACGGCCTGCGATATCA
KR-B reverse primer	TTGTGTGGGCACTGAATTGAC
KR-B probe	FAM-ACCCCGAAGCACTGGCCGC-TAMRA

doi:10.1371/journal.pntd.0000731.t001

concentration of each primer, 0.25 μ M concentration of the probe, and IQSupermix (Bio-Rad Lab) in a total volume of 25 μ l. Amplification and detection were performed with Thermocycler (MX3000P, Stratagene) using the following program: 1 cycle of 50°C for 2 min, 1 cycle of 95°C for 15 min, 40 cycles of 95°C for 15 s and 60°C for 1 min. DNA extracts were tested at least as duplicates, and negative controls were included in each assay. Quantitative readout assays were set up, based on external standard curve with *M. ulcerans* (strain 1G897) DNA serially diluted over 8 logs. Samples were considered positive only if both IS2404 sequence and the gene sequence encoding the ketoreductase B domain (KR) of the mycolactone polyketide synthase were detected, with threshold cycle (Ct) values strictly <35 cycles.

Quality control and quality assurance of clinical and environmental diagnostic PCR

The university laboratory is enrolled in quality control of clinical specimens, along with three partners: Angers University Hospital, Pasteur Centre of Yaoundé (Cameroon), and Institut Pasteur of Bangui (Central African Republic). The quality assurance program of the laboratory has been also involved in the analysis of environmental samples, under the coordination of the WHO Collaborating Centre for *Mycobacterium ulcerans* (Victorian Infectious Diseases Reference Laboratory in Melbourne).

Detection of acid-fast bacilli in saliva

Positive PCR saliva of *Appasus sp.* were pooled according to the sampling month and diluted in PBS (final volume 160 μ l). To detect the acid-fast bacilli, smears of suspensions (10 μ l of saliva suspension pool) were stained by the Ziehl-Neelsen procedure and examined using an oil immersion lens (100 \times) in an Olympus binocular microscope (model CH30 Olympus) (Figure S1).

Inoculation of saliva collected from captured *Appasus* into the mouse tail skin

Six week old female BALB/c mice (Charles River France, <http://www.criver.com/ico>) were maintained under conventional conditions in the animal house facility of the Centre Hospitalier Universitaire, Angers, France (Agreement A 49 007 002), adhering to the institution's guidelines for animal husbandry. From each saliva pool, 3 mice were inoculated subcutaneously with 50 μ l of suspension, using 26-gauge needles. The first lesions appeared after the fourth month of inoculation, with inflammatory lesions of the tails of three mice (two mice were inoculated with positive

PCR saliva from the April pool, and one mouse from the July pool). Two weeks later, a small oedema was observed and mice were sacrificed. Tissue specimens from mice were weighed, minced with disposable scalpels in a Petri dish and ground with a Potter-Elvehjem homogeniser, size 22, (Kimble/Kontes, Vineland, NJ), in 0.15 M NaCl to obtain a tenfold dilution. Smears of suspensions (10 μ l) were stained by the Ziehl Neelsen procedure. DNA was extracted from this material and purified to detect and quantify *M. ulcerans* by quantitative PCR, as described above. Tissue suspensions were decontaminated using an equal volume of *N*-acetyl-L-cysteine sodium hydroxide (2%), as previously described [44], and 0.2 ml of each suspension was inoculated onto two Löwenstein-Jensen slants and incubated at 30°C (Figure S1).

Statistical analysis

The number of water bugs collected per sampling was modelled using a negative binomial regression, with a random intercept to allow for within-day correlations of samples collected the same day. In this model, the effect of "month of collection" was studied as a fixed effect by introducing dummy variables associated with months of collection. Pearson Chi-square tests were used to compare proportions, and in particular the proportion of insects belonging to a given family (e.g., Belostomatidae, Notonectidae, ...) by month of the study, the proportion of insect pools positive for the presence of *M. ulcerans* DNA by month of the study, the proportion of insect pools positive for the presence of *M. ulcerans* DNA by insect family, and the proportion of saliva samples of *Appasus sp.* (Belostomatidae) positive for *M. ulcerans* DNA by month of the study.

Results

Taxonomic composition of water bug community

The inventory of water bug families in Centre Province of Cameroon was undertaken in Buruli ulcer endemic and non endemic areas, along the Nyong River. Among 7,407 collected specimens, seven aquatic Heteroptera families (Four true aquatic bugs and three semi-aquatic bugs) present in both areas were identified: Belostomatidae, Notonectidae, Nepidae, Corixidae, Gerridae, Mesoveliidae and Hydrometridae (Table 2 and Figure 2). The two most diversified families in our study were the families Notonectidae and Belostomatidae. Seven undetermined morphotypes were present in the Notonectidae family, two belonging to Anisopinae subfamily and five to Notonectinae

Table 2. Identification of aquatic heteroptera collected in endemic and non-endemic areas for Buruli ulcer.

Category	Family	Sub family	Genus	Humans-biting	Endemic site	Non endemic site
True water bugs	Belostomatidae	Belostomatinae	<i>Appasus</i>	Yes	+	+
	Belostomatidae	Lethocerinae	<i>Lethocerus</i>	Yes	+	+
	Notonectidae	Anisopinae	ND*	Yes	+	+
	Notonectidae	Notonectinae	ND	Yes	+	+
	Nepidae	Ranatrinae	ND	Yes	+	+
	Corixidae	Micronectinae	ND	No	+	+
Semi-aquatic bugs	Gerridae	Gerrinae	ND	?	+	+
	Mesoveliidae		ND	No	+	+
	Hydrometridae		ND	No	+	+

*No determination key available.

doi:10.1371/journal.pntd.0000731.t002

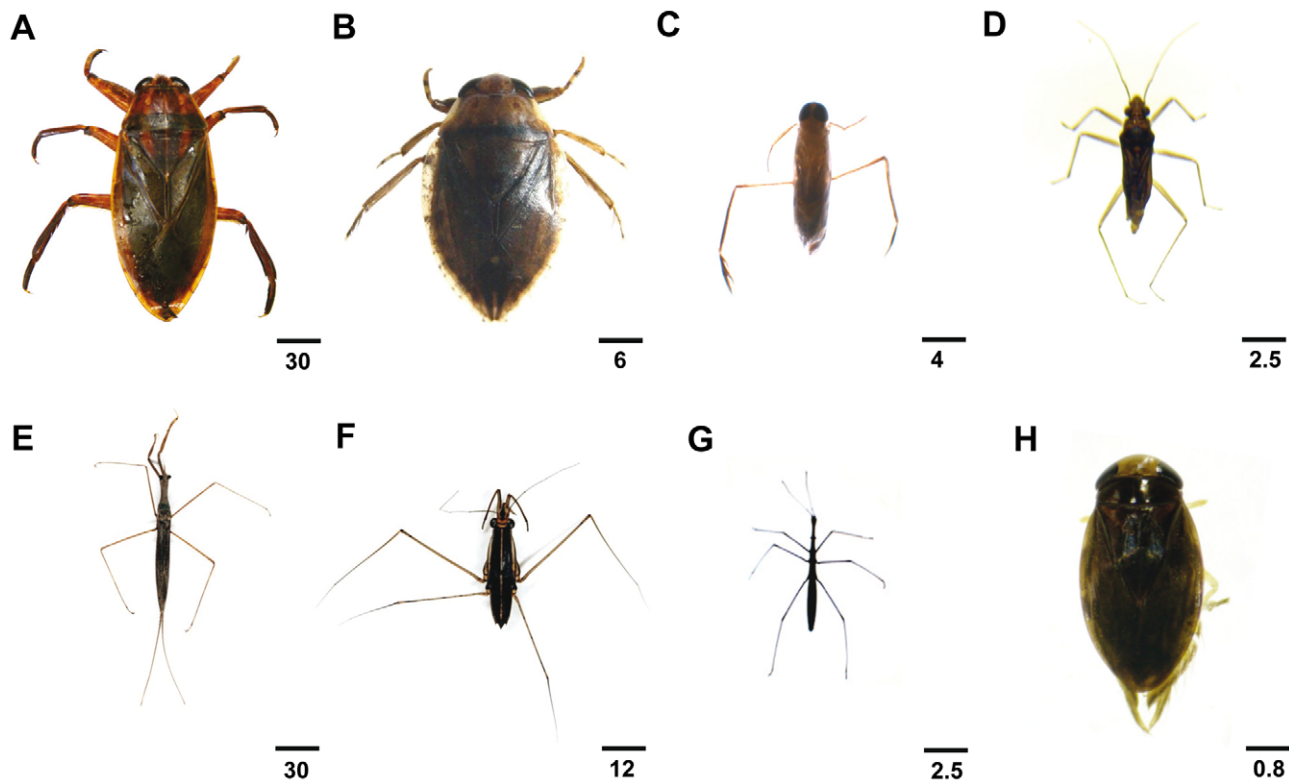


Figure 2. Specimens of aquatic bugs (Hemiptera) collected during the study (dorsal habitus view). Two species of Belostomatidae family: (A) *Appasus sp.*, a biting and flying water bug, (B) *Lethocerus sp.*, the biggest water bug, (C) Notonectinae, biting and flying water bug, (D) Mesoveliidae, a semi-aquatic bug, (E) Ranatrinae, family of Nepidae, a biting water bug, (F) Gerridae, a semi-aquatic bug, (G) Hydrometridae, a semi-aquatic bug, and (H) Micronectinae, family of Corixidae, a fluid feeder water bug. Scale bar: (A) (E) 30 mm, (B) 6 mm, (C) 4 mm, (D) (G) 2.5 mm, (F) 12 mm, (H) 0.8 mm.

doi:10.1371/journal.pntd.0000731.g002

subfamily. *Appasus sp.*, *Lethocerus* and another unidentified morphotype were present in the Belostomatidae family. Only one subfamily was identified in Nepidae and Corixidae families, respectively Micronectinae and Ranatrinae. All families identified in this study can be characterized as carnivorous predatory fluid-feeders, with the exception of the Corixidae family (plant feeders). Three families (Belostomatidae, Notonectidae, Nepidae), among the six carnivorous ones, are able to bite humans and to fly.

Population dynamics of water bugs in Buruli ulcer endemic area

Cameroon has a tropical climate which varies from equatorial in the South to Sahelian in the North. The equatorial South, where the Buruli ulcer endemic area is located, has two wet seasons and two dry seasons. One wet season occurs between March and June and the main wet season occurs between August and November. One dry season occurs between June and August and the main dry season occurs between December and March. The population dynamics of water bugs was investigated in an endemic area for Buruli Ulcer (district of Akonolinga). In order to get comparable results, insects were captured at periodic intervals by the same operator (with standardized sampling methods), in the same water body each time.

Large fluctuations of water bug density were observed among the samples (Figure 3A). The highest number of collected insects per sampling was recorded in January, during the long dry season (median = 369), whereas in other months, the median number of captured insects per sampling varied between 25 and 94 specimens

($p < 0.001$, according to the negative binomial regression model estimating the count of insects per sampling).

With respect to water bug families, the following variations in the sample composition were observed: out of seven families, four families (Belostomatidae, Notonectidae, Gerridae, Nepidae) were collected throughout the study period (Figure 3B), whereas the three other families (Corixidae, Mesoveliidae and Hydrometridae) were found only in January and/or April (Figure 3B). The most abundant family was Notonectidae (67% of total collected water bugs). The proportions for the other families were: 14.2% (Belostomatidae), 10.5% (Gerridae), 5.6% (Corixidae), 1% (Hydrometridae) and 0.1% (Mesoveliidae). The relative abundance of families fluctuated over the year. For example, Belostomatidae and Notonectidae represented respectively 59.8% and 34.3% of total insects in October, and 10.1% and 88.4% in November (Figure 3B). Moreover it was observed that in January Corixidae reached the highest abundance (10.2% out of total water bugs), whereas in January and April the highest water bug diversity was noticed (Figure 3B). All these data suggest that the long dry season corresponds to the period during which highest water bug diversity and abundance occur.

M. ulcerans DNA detection in insect tissues from Buruli ulcer endemic site

Detection of *M. ulcerans* in samples collected in Buruli ulcer endemic and non endemic sites was performed by PCR targeting the IS2404 insertion and the KR domain which encodes a polyketide synthase.

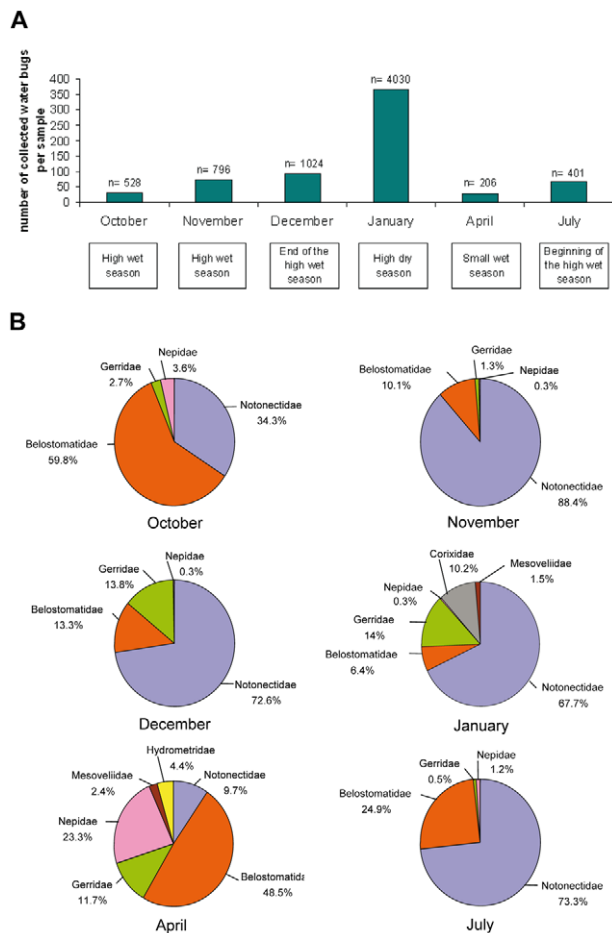


Figure 3. Variation in water bug abundance and taxa distribution in collected samples from an endemic area for Buruli ulcer. (A) Variation in water bug abundance per month, n corresponding to total collected water bugs. (B) Family heteroptera composition and distribution from October to July. doi:10.1371/journal.pntd.0000731.g003

From 3647 water bugs collected in the endemic area, 68 pools out of 616 (11%) were positive for both markers, IS2404 and Ktoreductase (Figure 4A). In addition *M. ulcerans* DNA was detected in five out of seven analyzed insect families. The rate of colonization in these pools was around 10%, except for the Corixidae family (Micronectinae) captured only in January, for which the rate reached 43.7% ($p = 0.008$, Pearson Chi-square test) (Figure 4B). Of note, this result was confirmed for individual Corixidae specimens ($n = 72$). However, given the very low number of collected water bugs from Mesoveliidae and Hydrometridae families (33 and 9 specimens, respectively), it is difficult to draw clear conclusions, in this case, about their possible subversion as hosts for *M. ulcerans*.

The rate of insect colonization by *M. ulcerans* fluctuated between 1.4% and 33.9% according to the sampling period, with a peak in July (33.9%) ($p = 0.008$, Pearson Chi-square test) (Figure 4A). Moreover, in the present study, no correlation was established between abundance of water bugs and rate of colonization of water bugs by *M. ulcerans*. All families -notably Belostomatidae, Notonectinae and Gerridae- displayed very similar fluctuations in colonisation rates by *M. ulcerans* (Figure 4C), with the exception of Nepidae (for which the sample size was limited).

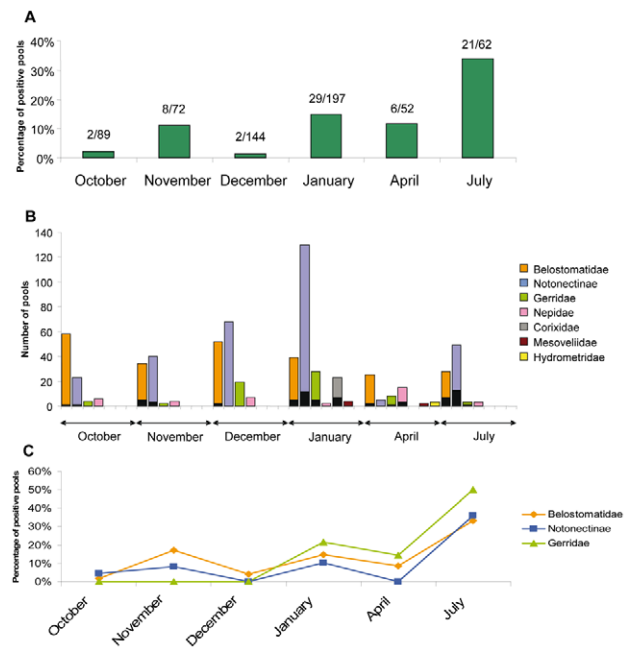


Figure 4. Detection of *M. ulcerans* DNA in insect tissues. The analysis was performed on 616 insect pools corresponding to 3647 individual specimens. (A) Percentage of positive pools over the period of study, (n) corresponding to the number of positive pools out of total pools. (B) Number of pools and number of positive pools (black part of bars represent the number of positive PCR pools) following water bug families and period of collection. (C) Monthly trends in *M. ulcerans* DNA positivity rate by family. doi:10.1371/journal.pntd.0000731.g004

Detection of *M. ulcerans* DNA in insect tissues from Buruli ulcer non endemic site

From 422 water bugs caught in a Buruli ulcer non endemic area, no pool out of 80 was found positive (Table 3). Significantly, in these same April and July periods, 11.5% and 33.9%, respectively, pools were found positive in the endemic site situated 100 kms away.

Detection of *M. ulcerans* in the saliva collected from *Appasus sp.* sampled in Buruli ulcer endemic area

Only living Belostomatidae insects of the genus *Appasus sp.* (Figure 5A and B) were allowed to salivate, for technical reasons. The saliva of these insects was first monitored for the presence of *M. ulcerans* DNA (Figure 5A and B). The individual saliva samples analysed by IS2404 and KR PCR were found positive in 51/293 of cases (17.4%), with a peak in July. Similar patterns were observed with homogenate tissue of *Appasus sp.* (Figure 5C). Interestingly, few acid-fast bacilli were observed in saliva samples of three individual positive pools (November, April and July). Their viability was evaluated by inoculation of PCR positive saliva into the tails of 21 Balb/c mice. Using quantitative PCR, quantity of inoculated bacilli was determined to range between 1×10^2 and 5×10^3 bacilli per ml. Four months after the subcutaneous injection, three mice displayed lesions typical of *M. ulcerans*, in which acid-fast bacilli were detected. Quantity of bacilli was estimated by quantitative PCR to range between 6×10^4 and 3×10^5 bacilli per ml of grounded tissue. This result suggests growth of *Mycobacterium ulcerans* in mouse tail. It can be noticed, however, that conventional methods failed to isolate the bacilli by culture from mouse tissues presenting clinical lesions.

Table 3. Detection of *M. ulcerans* DNA in aquatic heteroptera tissues collected in Buruli ulcer endemic and non-endemic areas.

	Endemic site			Non endemic site			
	Family	No. of water bugs	Pools		No. of water bugs	Pools	
No.			Positive (%)	No.		Positive (%)	
	Belostomatidae	100	23	2 (8.7)	21	5	0
	Notonectidae	20	5	0	17	3	0
	Nepidae	48	12	3 (25)	95	19	0
April	Corixidae	0	0	-	0	0	-
	Gerridae	24	7	1 (14.3)	0	0	-
	Mesoveliidae	5	2	0	0	0	-
	Hydrometridae	9	3	0	0	0	-
	Belostomatidae	100	21	7 (33.3)	14	5	0
	Notonectidae	294	36	13 (36.1)	32	9	0
	Nepidae	5	3	0	0	0	-
July	Corixidae	0	0	-	44	4	0
	Gerridae	2	2	1 (50)	24	5	0
	Mesoveliidae	0	0	-	6	3	0
	Hydrometridae	0	0	-	10	3	0
Total		607	114	27 (16.8)	422	80	0

doi:10.1371/journal.pntd.0000731.t003

Discussion

The mode of transmission of *M. ulcerans* to humans remains unclear, with several different mechanisms proposed in the last few decades. Within the WHO, a consensus therefore emerged to investigate the processes underlying i) the transmission of *M.*

ulcerans to humans and ii) the dissemination of *M. ulcerans* in the environment.

Objectives and rationale of the study

The aim of this study was to assess the role of water bugs as hosts and vectors of *M. ulcerans*, in the complete environmental context. To this end, we carried out an extensive field study on unprecedented temporal and spatial scales, monitoring the distribution of water bugs harboring *M. ulcerans* and the dissemination of *M. ulcerans* in the environment. We assessed water bug diversity and determined the frequency of insect tissue colonization by *M. ulcerans* in the various seasons, in an area in which Buruli ulcer is endemic (Akonolinga, Cameroon). For the purposes of comparison, the study also covered an area in which Buruli ulcer is not endemic. In short, the specificities of the study are: (i) focusing entirely on water bugs; (ii) the collection of samples from the same water body in all four seasons and (iii) large-scale sample collection in Buruli ulcer endemic and non endemic areas, with subsequent analysis of the captured water bug specimens (7407 specimens collected, 696 pools analyzed).

Inventory of water bugs and fluctuations in their density

We document here, for the first time, fluctuations in the density of water bug families in an area in which Buruli ulcer is endemic, over the course of a year. The highest density of water bugs is recorded in January, during the long dry season. Variations of water bug density described here are in agreement with those in another study in a tropical area (Costa Rica) [45]. The causes of these fluctuations remain unclear, but several possible factors have been identified, including abrupt changes in environmental conditions or prey density [33,45–46].

Seven water bug families were identified in Buruli ulcer endemic and non endemic areas, including many unknown species. This latter point is not surprising, as determination keys for water bug species are not yet available for West Africa. One key finding of our study was the striking difference between the

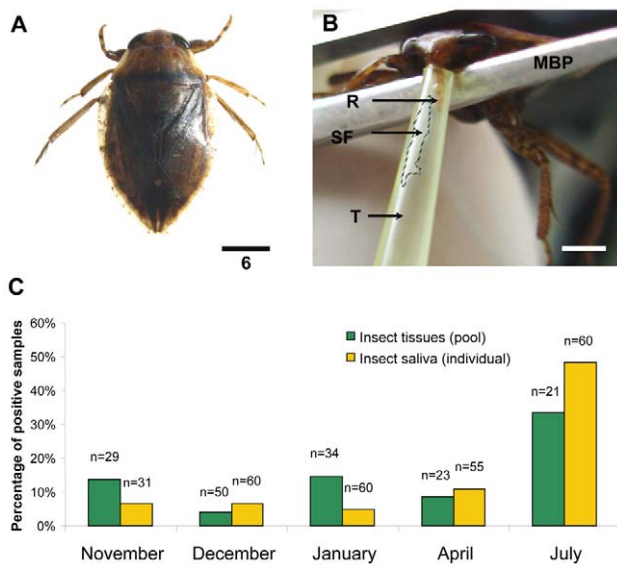


Figure 5. Detection of *M. ulcerans* DNA in tissues and in saliva of *Appasus* sp. (Belostomatidae). (A) Detection of *M. ulcerans* DNA in saliva was performed on *Appasus* sp. (Scale: 6 mm). (B) Water bug was grasped with metallic blunt pincers (MBP). The saliva was collected by introduction of rostrum (R) in a tip (T). The white saliva fluid (SF) (dotted line) could be observed. (C) Detection of *M. ulcerans* DNA in saliva was performed on saliva samples and on insect tissues over the period of study.

doi:10.1371/journal.pntd.0000731.g005

endemic and non endemic areas in terms of the density of water bugs, with the density in the endemic area more than 10 times higher than in the non endemic area. It can be noticed that this observation is not in accordance with results in another published study [31]. However, the conclusions of this previous study were drawn based on data for a significantly smaller number of specimens (only 200 water bugs; 2% of the invertebrates collected [31]).

Water bugs as hosts for *M. ulcerans*

M. ulcerans DNA was detected in five of the seven families of water bugs in the endemic area. The mean rate of colonization was about 10%. However, large fluctuations were observed in the rate of insect colonization by *M. ulcerans* (1.4 to 33.9%). As observed for other hosts of microorganisms [47–48], there was no correlation between water bug density and rates of water bug colonization by *M. ulcerans*. We detected no *M. ulcerans* DNA in insects from the non endemic area, despite the high rates of colonization reported for the nearby endemic area.

One key difference between the endemic and non endemic areas concerned human activity. Despite their close physical proximity, the non endemic area was largely unaffected by human activity, whereas, in the endemic area, the bank of the Nyong River had been deforested, for agricultural and fishing activities (Figure 1B and C). Interventions disrupting the environment have been identified, in several studies, as factors potentially favoring the establishment of *M. ulcerans* in remodeled environments [13–15,49].

The observed fluctuations in colonization rate may be accounted for by changes in the level of water in the Nyong River, which falls markedly in the dry season. Our results are supported by the findings of an epidemiological study performed in 1993 [50], in which low water levels in the dry season were found to favor the transmission of *M. ulcerans* to humans. Several factors may be involved in this phenomenon, including greater access to nutrients (resulting in an increase in *M. ulcerans* density) and the abundance of aquatic vegetation favoring *M. ulcerans* biofilm formation [26,30,51] (increasing the level of contact between *M. ulcerans* and water bugs).

The water bugs from the family Corixidae were the only phytophagous insects inventoried here. These water bugs were detected only in January, when they were present in high abundance, and displayed the highest rate of colonization by *M. ulcerans* (43.7%). The high frequency of colonization by *M. ulcerans* in Corixidae supports the hypothesis that aquatic plants may be the primary reservoir of *M. ulcerans*, as previously suggested [26]. This specific family may be involved in spreading the bacillus to other trophic levels, as they are eaten by other water bugs, aquatic invertebrates and vertebrates. These observations suggest that *M. ulcerans* may colonize different levels within the trophic chain (aquatic plants, invertebrates and vertebrates), as already considered [21,30,52–53]. In addition, three water bug families are known to be good flyers [33,45,54–57] and were identified as *M. ulcerans* hosts in our study. These water bugs may therefore be involved in disseminating *M. ulcerans* in the environment, as previously proposed by Portaels and Meyers [34].

Water bugs as vectors for *M. ulcerans*

We showed that water bug saliva could harbor bacilli. *M. ulcerans* may therefore be present in the saliva under natural conditions, with the bacilli colonizing the salivary glands of the insect. This could provide a route for *M. ulcerans* transmission in natural conditions, in accordance with previous experimental demonstrations in laboratory conditions [24,28]. It should be

noted that the bacilli present in the saliva of *Appasus sp.* induced an *M. ulcerans*-containing lesion following the inoculation of mouse tail. However, it was not possible to isolate these bacilli by conventional culture methods. The lack of appropriate culture media and decontamination procedures adapted to the isolation of *M. ulcerans* (from the environment) thus remain a major handicap, hindering investigations of the ecology and mode of transmission of this mycobacterium.

Highlights, difficulties and perspectives

The various results presented above provide further evidence that water bugs are hosts and vectors of *M. ulcerans*, and provide insight into the environmental context underlying transmission. However, no definitive conclusion can yet be drawn concerning the precise importance of this route of transmission. The presence, in human sera, of antibodies binding water bug salivary gland extracts may be accounted for by the exposure of humans to water bug bites [27]. Indeed, reports of the exposure of humans to blood-feeding arthropods, which are known to act as hosts and vectors for parasitic microorganisms, are becoming increasingly frequent [38–39].

To gain a complete picture of the transmission route it would have been desirable to explore the relationship between the incidence of the disease in humans and the rate of colonization of water bugs by *M. ulcerans*, over a one-year period. Such exploration was however hampered by the amount of accessible information, with several important parameters not available: (i) incubation time between exposure to *M. ulcerans* and the appearance of the first clinical lesions (currently estimated at between a few weeks and several months); (ii) the slow progression of clinical lesions, resulting in patients being diagnosed at different stages of the disease (rarely at early stages) and (iii) the small number of Buruli ulcer patients diagnosed with early lesions between October 2007 and July 2008 in Akonolinga (84 Buruli ulcer patients, including 15 with early lesions).

In conclusion, this study sheds light on the natural history of *M. ulcerans* within its ecosystem. The observed fluctuations in insect colonization rates suggest that there may be a particularly favorable period for the development of *M. ulcerans* in natural conditions, and a favorable period for the transmission of *M. ulcerans* to humans (as previously suggested [50,58] and observed for *Plasmodium sp.* [47,59–60]). It is also noticeable that the results here can be put to advantage for practical applications, with surveillance and prevention purposes [27,61]. More precisely, our work suggests that the detection of *M. ulcerans* in water bug saliva could be used as an environmental indicator of the risk of *M. ulcerans* transmission to humans. It would then be possible to set up environmental surveillance (detection of *M. ulcerans* DNA in water bug tissue and saliva) in non endemic areas close to Buruli ulcer endemic areas. Health messages concerning environmental risk factors could be specifically targeted at populations newly exposed to the risk of *M. ulcerans* infection, as is already the case for protective factors (wearing long clothing during farming activities and use of bed nets) [16].

Supporting Information

Figure S1 Main steps followed to detect *M. ulcerans* in water bug saliva.

Found at: doi:10.1371/journal.pntd.0000731.s001 (0.10 MB TIF)

Acknowledgments

We thank all the staff working for Médecin Sans Frontière in Akonolinga, and Francis Décheron and his team (Service des Expéditions, Institut

Pasteur Paris) for their appraised care of either reagents or biological materials.

We are grateful to Caroline Lavender (WHO Collaborating Centre for *Mycobacterium ulcerans* at the Victorian Infectious Diseases Reference Laboratory in Melbourne) for the organization and the quality control done on the quantitative PCR.

Special thanks to Geneviève Milon for critical reading of the manuscript.

We thank Doctor François Sihom, Chief Health District of Akonolinga, who provided the data of Buruli ulcer patients.

References

- World Health Organization (2004) Buruli ulcer. *Mycobacterium ulcerans* infection. WER 79: 145–149.
- Asiedu K, Sherpier R, Raviglione MC (2000) Buruli Ulcer *Mycobacterium ulcerans* infection. W.H.O. Global Buruli Ulcer initiative. Report 2000 World Health Organisation Geneva Switzerland.
- Johnson PD, Stinear T, Small PL, Pluschke G, Merritt RW, et al. (2005) Buruli ulcer (*M. ulcerans* infection): new insights, new hope for disease control. *PLoS Med* 2: e108. doi: 10.1371/journal.pmed.0020108.
- Johnson PD, Veitch MG, Leslie DE, Flood PE, Hayman JA (1996) The emergence of *Mycobacterium ulcerans* infection near Melbourne. *Med J Aust* 164: 76–78.
- Johnson RC, Makoutode M, Sopoh GE, Elsen P, Gbovi J, et al. (2005) Buruli ulcer distribution in Benin. *Emerg Infect Dis* 11: 500–501.
- Asiedu K, Etuaful S (1998) Socioeconomic implications of Buruli ulcer in Ghana: a three-year review. *Am J Trop Med Hyg* 59: 1015–1022.
- Stienstra Y, van Roest MH, van Wezel MJ, Wiersma IC, Hospers IC, et al. (2005) Factors associated with functional limitations and subsequent employment or schooling in Buruli ulcer patients. *Trop Med Int Health* 10: 1251–1257.
- Chauty A, Ardant MF, Adeye A, Euverte H, Guedenon A, et al. (2007) Promising clinical efficacy of streptomycin-rifampin combination for treatment of Buruli ulcer (*Mycobacterium ulcerans* disease). *Antimicrob Agents Chemother* 51: 4029–4035.
- Etuaful S, Carbonnelle B, Grosset J, Lucas S, Horsfield C, et al. (2005) Efficacy of the combination rifampin-streptomycin in preventing growth of *Mycobacterium ulcerans* in early lesions of Buruli ulcer in humans. *Antimicrob Agents Chemother* 49: 3182–3186.
- Uganda Buruli Group (1971) Epidemiology of *Mycobacterium ulcerans* infection (Buruli ulcer) at Kinyara, Uganda. *Trans R Soc Trop Med Hyg* 65: 763–775.
- Roberts B, Hirst R (1997) Immunomagnetic separation and PCR for detection of *Mycobacterium ulcerans*. *J Clin Microbiol* 35: 2709–2711.
- Ross BC, Johnson PD, Oppedisano F, Marino L, Sievers A, et al. (1997) Detection of *Mycobacterium ulcerans* in environmental samples during an outbreak of ulcerative disease. *Appl Environ Microbiol* 63: 4135–4138.
- Brou T, Broutin H, Elguero E, Assé H, Guegan JF (2008) Landscape Diversity Related to Buruli Ulcer Disease in Côte d'Ivoire. *PLoS Negl Trop Dis* 2: e271. doi: 10.1371/journal.pntd.0000271.
- Marston BJ, Diallo MO, Horsburgh CR, Jr., Diomande I, Saki MZ, et al. (1995) Emergence of Buruli ulcer disease in the Daloa region of Côte d'Ivoire. *Am J Trop Med Hyg* 52: 219–224.
- Oluwasanmi JO, Solanke TF, Olurin EO, Itayemi SO, Alabi GO, et al. (1976) *Mycobacterium ulcerans* (Buruli) skin ulceration in Nigeria. *Am J Trop Med Hyg* 25: 122–128.
- Pouillot R, Matias G, Wondje CM, Portaels F, Valin N, et al. (2007) Risk factors for Buruli ulcer: a case control study in Cameroon. *PLoS Negl Trop Dis* 1: e101. doi: 10.1371/journal.pntd.0000101.
- Raghunathan PL, Whitney EA, Asamo K, Stienstra Y, Taylor TH, Jr., et al. (2005) Risk factors for Buruli ulcer disease (*Mycobacterium ulcerans* Infection): results from a case-control study in Ghana. *Clin Infect Dis* 40: 1445–1453.
- Johnson PD, Aзуolas J, Lavender CJ, Wishart E, Stinear TP, et al. (2007) *Mycobacterium ulcerans* in mosquitoes captured during outbreak of Buruli ulcer, southeastern Australia. *Emerg Infect Dis* 13: 1653–1660.
- Johnson PD, Lavender CJ (2009) Correlation between Buruli ulcer and vector-borne notifiable diseases, Victoria, Australia. *Emerg Infect Dis* 15: 614–615.
- Quek TY, Athan E, Henry MJ, Pasco JA, Redden-Hoare J, et al. (2007) Risk factors for *Mycobacterium ulcerans* infection, southeastern Australia. *Emerg Infect Dis* 13: 1661–1666.
- Portaels F, Elsen P, Guimaraes-Peres A, Fonteyne PA, Meyers WM (1999) Insects in the transmission of *Mycobacterium ulcerans* infection. *Lancet* 353: 986.
- Portaels F, Meyers WM, Ablordey A, Castro AG, Chemlal K, et al. (2008) First Cultivation and Characterization of *Mycobacterium ulcerans* from the Environment. *PLoS Negl Trop Dis* 2: e178. doi: 10.1371/journal.pntd.0000178.
- Marsollier L, Andre JP, Frigui W, Reyssat G, Milon G, et al. (2007) Early trafficking events of *Mycobacterium ulcerans* within *Nauoris cimicoides*. *Cell Microbiol* 9: 347–355.
- Marsollier L, Aubry J, Coutanceau E, Andre JP, Small PL, et al. (2005) Colonization of the salivary glands of *Nauoris cimicoides* by *Mycobacterium ulcerans* requires host plasmacytes and a macrolide toxin, mycolactone. *Cell Microbiol* 7: 935–943.
- Marsollier L, Aubry J, Milon G, Brodin P (2007) Aquatic insects and transmission of *Mycobacterium ulcerans*. *Med Sci (Paris)* 23: 572–575.
- Marsollier L, Brodin P, Jackson M, Kordulakova J, Tafelmeyer P, et al. (2007) Impact of *Mycobacterium ulcerans* biofilm on transmissibility to ecological niches and Buruli ulcer pathogenesis. *PLoS Pathog* 3: e62. doi: 10.1371/journal.ppat.0030062.
- Marsollier L, Deniaux E, Brodin P, Marot A, Wondje CM, et al. (2007) Protection against *Mycobacterium ulcerans* lesion development by exposure to aquatic insect saliva. *PLoS Med* 4: e64. doi: 10.1371/journal.pmed.0040064.
- Marsollier L, Robert R, Aubry J, Saint Andre JP, Kouakou H, et al. (2002) Aquatic insects as a vector for *Mycobacterium ulcerans*. *Appl Environ Microbiol* 68: 4623–4628.
- Marsollier L, Severin T, Aubry J, Merritt RW, Saint Andre JP, et al. (2004) Aquatic snails, passive hosts of *Mycobacterium ulcerans*. *Appl Environ Microbiol* 70: 6296–6298.
- Williamson HR, Benbow ME, Nguyen KD, Beachboard DC, Kimbirauskas RK, et al. (2008) Distribution of *Mycobacterium ulcerans* in Buruli Ulcer Endemic and Non-Endemic Aquatic Sites in Ghana. *PLoS Negl Trop Dis* 2: e205. doi: 10.1371/journal.pntd.0000205.
- Benbow ME, Williamson H, Kimbirauskas R, McIntosh MD, Kolar R, et al. (2008) Aquatic invertebrates as unlikely vectors of Buruli ulcer disease. *Emerg Infect Dis* 14: 1247–1254.
- Mosi L, Williamson H, Wallace JR, Merritt RW, Small PL (2008) Persistent association of *Mycobacterium ulcerans* with the West African predaceous insect, Belostomatidae. *Appl Environ Microbiol* 74(22): 7036–7042.
- Andersen NM, Weir TA, eds. Australian Water Bugs. Their biology and identification Apollo books. 344 p.
- Portaels F, Meyers WM (2007) Buruli ulcer. In: R FW, Hay JH, Naafs B, eds. Imported skin diseases Elsevier. pp 117–129.
- Noeske J, Kuaban C, Rondini S, Sorlin P, Ciaffi L, et al. (2004) Buruli ulcer disease in Cameroon rediscovered. *Am J Trop Med Hyg* 70: 520–526.
- Porten K, Sailor K, Comte E, Njikap A, Sobry A, et al. (2009) Prevalence of Buruli ulcer in Akonolinga health district, Cameroon: results of a cross sectional survey. *PLoS Negl Trop Dis* 3: e466. doi: 10.1371/journal.pntd.0000466.
- Dethier M (1981) Introduction à la morphologie, la biologie et la classification des Hétéropères. *Bulletin Romand d'Entomologie* 1: 11–16.
- Orlandi-Pradines E, Almeras L, Denis de Senneville L, Barbe S, Remoue F, et al. (2007) Antibody response against saliva antigens of *Anopheles gambiae* and *Aedes aegypti* in travellers in tropical Africa. *Microbes Infect* 9: 1454–1462.
- Remoue F, Cisse B, Ba F, Sokhna C, Herve JP, et al. (2006) Evaluation of the antibody response to *Anopheles* salivary antigens as a potential marker of risk of malaria. *Trans R Soc Trop Med Hyg* 100: 363–370.
- Rondini S, Mensah-Quainoo E, Troll H, Bodmer T, Pluschke G (2003) Development and application of real-time PCR assay for quantification of *Mycobacterium ulcerans* DNA. *J Clin Microbiol* 41: 4231–4237.
- Stinear TP, Mve-Obiang A, Small PL, Frigui W, Pryor MJ, et al. (2004) Giant plasmid-encoded polyketide synthases produce the macrolide toxin of *Mycobacterium ulcerans*. *Proc Natl Acad Sci U S A* 101: 1345–1349.
- Stinear TP, Pryor MJ, Porter JL, Cole ST (2005) Functional analysis and annotation of the virulence plasmid pMUM001 from *Mycobacterium ulcerans*. *Microbiology* 151: 683–692.
- Fyfe JA, Lavender CJ, Johnson PD, Globan M, Sievers A, et al. (2007) Development and application of two multiplex real-time PCR assays for the detection of *Mycobacterium ulcerans* in clinical and environmental samples. *Appl Environ Microbiol* 73: 4733–4740.
- Kubica GP, Dye WE, Cohn ML, Middlebrook G (1963) Sputum digestion and decontamination with N-acetyl-L-cysteine-sodium hydroxide for culture of mycobacteria. *Am Rev Respir Dis* 87: 775–779.
- Stout JR (1982) Effects of harsh environment on the life history patterns of two species of tropical aquatic hemiptera (Famili : Naucoridae). *Ecology* 63: 75–83.
- Kopp K, Wachlewski M, Eterovick PC (2006) Environmental complexity reduces tadpole predation by water bugs. *Can J Zool* 84: 136–140.
- Bigoga JD, Manga L, Titanji VP, Coetzee M, Leke RG (2007) Malaria vectors and transmission dynamics in coastal south-western Cameroon. *Malar J* 6: 5.
- Wanji S, Tanke T, Atanga SN, Ajonina C, Nicholas T, et al. (2003) Anopheles species of the mount Cameroon region: biting habits, feeding behaviour and entomological inoculation rates. *Trop Med Int Health* 8: 643–649.
- Veitch MG, Johnson PD, Flood PE, Leslie DE, Street AC, et al. (1997) A large localized outbreak of *Mycobacterium ulcerans* infection on a temperate southern Australian island. *Epidemiol Infect* 119: 313–318.
- Daric H, Le Guyadec T, Touze JE (1993) Epidemiological and clinical aspects of Buruli ulcer in Ivory Coast. 124 recent cases. *Bull Soc Pathol Exot* 86: 272–276.

The authors acknowledge many insightful comments and suggestions by four anonymous referees.

Author Contributions

Conceived and designed the experiments: EM SE LM. Performed the experiments: EM VC YK PL. Analyzed the data: EM SE JD JL JA AF JC AM ME CD LM. Contributed reagents/materials/analysis tools: LM. Wrote the paper: EM SE EY JA AF CR ME CD JP SA LM.

51. Marsollier L, Stinear T, Aubry J, Saint Andre JP, Robert R, et al. (2004) Aquatic plants stimulate the growth of and biofilm formation by *Mycobacterium ulcerans* in axenic culture and harbor these bacteria in the environment. *Appl Environ Microbiol* 70: 1097–1103.
52. Eddyani M, Ofori-Adjei D, Teugels G, De Weirtd D, Boakye D, et al. (2004) Potential role for fish in transmission of *Mycobacterium ulcerans* disease (Buruli ulcer): an environmental study. *Appl Environ Microbiol* 70: 5679–5681.
53. Kotowski R, Martin A, Ablordey A, Chemlal K, Fonteyne PA, et al. (2004) One-tube cell lysis and DNA extraction procedure for PCR-based detection of *Mycobacterium ulcerans* in aquatic insects, molluscs and fish. *J Med Microbiol* 53: 927–933.
54. Bowden J (1964) The relation of activity of two species of belostomatidae to rainfall and moonlight in Ghana. *Journal of Entomology Society of South of Africa* 26: 293–301.
55. Lytle DA (1999) Use of rainfall cues by *Abedus herberti* (hemiptera: Belostomatidae): A mechanism for avoiding flash floods. *J Insect Behav* 12: 1–12.
56. Mukai Y, Ishii M (2007) Habitat utilization by giant water bug, *Appasus* (= *Diplonychus*) major (Hemiptera: Belostomatidae), in traditional rice paddy water system in northern Osaka, central Japan. *Appl Entomol Zool* 42: 595–605.
57. Robertson IAD (1976) Records of insects taken at light traps in Tanzania. Distribution and seasonal change in catches of Belostomatidae (Hemiptera: Heteroptera) in relation to rainfall. London: Centre for Overseas Pest Research.
58. Portaels F (1989) Epidemiology of ulcers due to *Mycobacterium ulcerans*. *Ann Soc Belg Med Trop* 69: 91–103.
59. Fontenille D, Lochouarn L, Diatta M, Sokhna C, Dia I, et al. (1997) Four years' entomological study of the transmission of seasonal malaria in Senegal and the bionomics of *Anopheles gambiae* and *A. arabiensis*. *Trans R Soc Trop Med Hyg* 91: 647–652.
60. Boudin C, Robert V, Verhave JP, Carnevale P, Ambroise-Thomas P (1991) *Plasmodium falciparum* and *P. malariae* epidemiology in a West African village. *Bull World Health Organ* 69: 199–205.
61. Silva MT, Portaels F, Pedrosa J (2007) Aquatic insects and *Mycobacterium ulcerans*: an association relevant to Buruli ulcer control? *PLoS Med* 4: e63. doi: 10.1371/journal.pmed.0040063.