



HAL
open science

$\gamma\delta$ T cell receptor ligands and modes of antigen recognition.

Eric Champagne

► **To cite this version:**

Eric Champagne. $\gamma\delta$ T cell receptor ligands and modes of antigen recognition.: Antigen recognition by gamma delta T cells. *Archivum Immunologiae et Therapiae Experimentalis*, 2011, 59 (2), pp.117-37. 10.1007/s00005-011-0118-1 . inserm-00568107

HAL Id: inserm-00568107

<https://inserm.hal.science/inserm-00568107>

Submitted on 22 Feb 2011

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

$\gamma\delta$ T cell receptor ligands and modes of antigen recognition

Eric Champagne *

CPTP, Centre de physiopathologie de Toulouse Purpan INSERM : U1043 , CNRS: UMR5282 , Université Paul Sabatier - Toulouse III , Hôpital Purpan Place du Docteur Baylac BP 3028 31024 Toulouse Cedex 3, FR

* Correspondence should be addressed to: Eric Champagne <eric.champagne@inserm.fr >

Abstract

T lymphocytes expressing the $\gamma\delta$ -type of T cell receptors for antigens contribute to all aspects of immune responses, including defenses against viruses, bacteria, parasites and tumors, allergy and autoimmunity. Multiple subsets have been individualized in humans as well as in mice and they appear to recognize in a TCR-dependent manner antigens as diverse as small non-peptidic molecules, soluble or membrane-anchored polypeptides and molecules related to MHC antigens on cell surfaces, implying diverse modes of antigen recognition. We review here the $\gamma\delta$ TCR ligands which have been identified along the years and their characteristics, with emphasis on a few systems which have been extensively studied such as human $\gamma\delta$ T cells responding to phosphoantigens or murine $\gamma\delta$ T cells activated by allogeneic MHC antigens. We discuss a speculative model of antigen recognition involving simultaneous TCR recognition of MHC-like and non-MHC ligands which could fit with most available data and shares many similarities with the classical model of MHC-restricted antigen recognition for peptides or lipids by T cells subsets with $\alpha\beta$ -type TCRs.

MESH Keywords Animals ; Antigens ; metabolism ; Humans ; Immunity, Innate ; Ligands ; Models, Immunological ; Receptors, Antigen, T-Cell, gamma-delta ; metabolism ; Signal Transduction ; T-Lymphocytes ; immunology

Author Keywords Gamma delta lymphocytes ; T cell receptor ; Innate immunity ; Antigen recognition ; Major Histocompatibility complex

Introduction

On the basis of the T cell antigen receptor (TCR) chains which they express and use to recognize antigens, human and murine T lymphocytes comprise two subtypes: T cells using an $\alpha\beta$ TCR heterodimer ($\alpha\beta$ T cells) and those using a $\gamma\delta$ heterodimer ($\gamma\delta$ T cells), which associate to a common CD3 signal transduction module (Hayday 2000 ; Kabelitz and Wesch 2003 ; Pennington et al. 2005). The majority of $\alpha\beta$ T cells in blood and secondary lymphoid organs express clonotypic TCR chains with a strong diversity originating from usage of a large number of V α and V β genes, as well as extensive junctional diversity allowing the recognition of a large array of antigenic peptides in complex with polymorphic presenting MHC class I or class II molecules. Besides these "classical" T cells, NKT cells are present in small numbers in blood, liver and spleen and recognize non-polymorphic CD1d, an MHC-class I-like antigen (MHC-Ib) which associates with lipids. Among these, a subset of invariant NKT cells (iNKT) use a precise combination of V α and V β genes and carry very limited junctional diversity, having in particular no N-region insertions in the third complementarity-determining region (CDR3) of their TCR. They may recognize a limited set of antigenic endogenous lipids. Other NKT cells use a limited set of V genes and an extensive junctional diversity to recognize exogenous bacterial lipids presented by CD1d (Godfrey et al. 2004 ; Godfrey et al. 2010). Mucosal-associated invariant T cells (MAIT) are abundant in blood and are also found in epithelial tissues. They are in fact semi-invariant as although they also use a defined combination of V α /J α and use preferentially a limited set of V β genes, these can combine with multiple V β genes carrying some N-region junctional diversity (Treiner et al. 2005). This limited diversity is probably used to recognize uncharacterized antigens from pathogens in combination with MR1, another MHC-Ib antigen (Le Bourhis et al. 2010). All these $\alpha\beta$ T cell populations are present in both humans and mice and their biology appears closely homologous.

In contrast and although they are also present in both species, there is accumulating evidence that human and murine $\gamma\delta$ T cells have substantially diverged. Subpopulations of $\gamma\delta$ T cells are recognized in both systems. In the murine system, most of these are characterized by their predominant localization in defined epithelial tissues and have characteristics of NKT or MAIT cells by their expression of preferential V γ and V δ TCR region genes. V γ 5V δ 1 cells and V γ 6V δ 1 cells colonize the skin (DETC) and the reproductive tract epithelium (r-IELs), respectively, early during the fetal life and do not express junctional diversity. Other $\gamma\delta$ populations appear later and use particular V γ genes in correlation with their homing to preferential epithelial tissues which continues after birth (intestinal, lung and other mucosal $\gamma\delta$ IELs). These V γ genes are used in combination with multiple V δ genes and the diversity of their repertoire is further increased by an extensive junctional diversity (Allison and Raulet 1990). In humans, and especially in adults, one $\gamma\delta$ subset expressing V γ 9 and V δ 2 TCR regions starts to expand early after birth and is particularly abundant in adult blood and peripheral lymphoid tissues, although it is also present in epithelia (usually ~70% of $\gamma\delta$ cells). These V γ 9V δ 2 cells, also called V γ 2V δ 2, carry extensive junctional diversity in CDR3 regions but share a similar antigenic reactivity although they may display diverse effector functions (Nedellec et al. 2010). A specific tissue distribution for other $\gamma\delta$ cell populations in humans as a function of V gene expression is not as clear as in mice. Although $\gamma\delta$ T cells expressing V δ 1 regions predominantly reside in epithelial tissues (and in large proportion in the intestine), they are diverse and may express different V γ regions (Holtmeier 2003).

It thus appears that there are for $\gamma\delta$ cells as well as for $\alpha\beta$ cells multiple populations which may differ significantly in their biological properties. It seems likely that invariant DETC in the mouse recognize a tissue-specific and non-polymorphic endogenous ligand to perform homeostatic functions in the skin (Jameson and Havran 2007). Other populations may have completely different roles from the recognition of endogenous stress signals in epithelia to the recognition of pathogen-derived antigens so that a single antigen recognition mechanism may not apply to all $\gamma\delta$ cell populations (Born et al. 2010). In mice as well as in humans, the total $\gamma\delta$ TCR repertoire is characterized by a limited number of available V γ and V δ genes, and preferential usage of a few V segments in preferential V γ /V δ combinations. In contrast, there is a potential for extensive junctional diversity in particular in CDR3 δ which results from usage of multiple D δ gene segments in tandem, so that most of the TCR diversity is confined to CDR3 regions (Hayday 2000). This potential diversity is not used by the murine invariant $\gamma\delta$ populations and how it is used by other populations is still not really understood. The limited V region repertoire probably reflects the specialization of $\gamma\delta$ cells for the recognition of a limited number of conserved ligands expressed in selected tissues during stress or infection.

Activation of $\gamma\delta$ cells may be achieved through multiple non-TCR activating receptors, among which TLRs (Martin et al. 2009; Mokuno et al. 2000; Pietschmann et al. 2009), CD16, CD226 (Gertner-Dardenne et al. 2009; Toutirais et al. 2009), NKRs (Das et al. 2001; Fisch et al. 2000; Halary et al. 1999; Lafarge et al. 2005), CD28 (Sperling et al. 1993) and NKG2D. These receptors are thought to be co-stimulators rather than providers of a primary stimulus (Nedellec et al. 2010). Focusing on TCR activation, the list of antigens which have been found to activate $\gamma\delta$ T cells in a TCR-dependent manner include polypeptides, non-peptidic molecules and an increasing list of MHC-like structures so that no unifying concept about $\gamma\delta$ T cell reactivity has yet emerged. The necessity for antigen presentation in the case of $\gamma\delta$ T cells has been questioned as some soluble antigens can activate $\gamma\delta$ cells in the absence of antigen presenting cells (Born and O'Brien 2009). The understanding of $\gamma\delta$ cell biology is further complicated by the wide expression of NK receptors for MHC and MHC-like molecules (NKR) by murine and human $\gamma\delta$ T cells and these may act in conjunction with the TCR to modulate its activity.

We will review here the different putative $\gamma\delta$ TCR ligands that have been found recently and in the past and discuss the data which support their role in TCR ligation. Through a more detailed description of a few murine and human models which have been extensively studied, we will discuss how the recognition of many diverse structures may be integrated in a common model of antigen recognition involving simultaneously MHC-like and non-MHC-like ligands and sharing many features of classical antigen restriction. For V γ / δ gene nomenclature used here, refer to (Kabelitz and Wesch 2003).

$\gamma\delta$ TCR ligands belonging to the MHC family

Besides few exceptions, $\gamma\delta$ T cells do not recognize antigens restricted by MHC complex class I and class II molecules as $\alpha\beta$ T cells do. Spits *et al.* reported isolation of cytotoxic human $\gamma\delta$ T cell clones which proliferated against JY EBV-B-cells and were inhibited by anti-MHC-I antibody and anti- β 2m. The fine specificity of these clones was however not reported (Spits et al. 1989). MHC-Ia and Ib molecules interact with NKRs and modulate antigen recognition on many $\gamma\delta$ T cell populations. This has led to the concept that these T cells recognize antigens while sensing the context of their expression on target cells (Bonneville and Fournie 2005). Nevertheless, although $\gamma\delta$ cells recognizing MHC-like molecules were initially thought to be exceptions, the list of MHC-Ib molecules which seem to interact directly with $\gamma\delta$ TCRs has recently grown and the concept that MHC molecules are essentially targets for NKRs deserves to be revisited.

CD1

CD1 is a non polymorphic MHC-Ib protein associated with β 2-microglobulin (β 2m). It is expressed on immature dendritic cells, can present phospholipids and comprises four isoforms in humans (CD1a, b, c, d) and one in mice (CD1d) (Vincent et al. 2003). Human $\gamma\delta$ T cell clones activated by the recognition of CD1c have been isolated from peripheral blood (Faure et al. 1990; Leslie et al. 2002; Spada et al. 2000). In the study by Faure *et al.*, CD1c reactivity was found in one $\gamma\delta$ T cell clone out of 43 obtained from peripheral blood. In the study by Spada *et al.*, these were raised by repeated stimulation with dendritic cells and mycobacterial extracts, although these extracts were not required for reactivation. Due to the ability of CD1c to present lipids, it was assumed that these clones were recognizing endogenous lipids bound to CD1c. $\gamma\delta$ TCR transfection in TCR-deficient J.RT3-T3.5 cells together with the usage of CD1c-transfected target HeLa or C1R cells have confirmed the specific recognition of CD1c in a TCR-dependent manner. All four clones analyzed expressed V δ 1 associated with a different V γ chain (V γ 1.2, V γ 1.3, V γ 1.4, V γ 2) with no recognizable homology in the CDR3 γ or δ regions.

Human T cell clones recognizing the lipid antigen phosphatidyl-ethanolamine have been isolated from normal blood and were enriched in the blood and nasal mucosa of patients allergic to pollen lipids. These were all found to be strictly antigen-specific and restricted by CD1d (Russano et al. 2006; Russano et al. 2007) when tested on HeLa cells transfected with various CD1 isoforms. Some expressed V δ 2 whereas the majority expressed V δ 1. Most of them were also CD4⁺ as opposed to most human $\gamma\delta$ cells. Although their reactivity appears very similar to that of NKT cells, the involvement of the TCR in this reactivity has not been formally proven. In the same line, Cui *et al.* (Cui et al. 2005; Cui et al. 2009) have reported that human $\gamma\delta$ T cells from peripheral blood could be activated by

lipid-A, a component of LPS, loaded on monocyte-derived dendritic cells or CD1-transfected C1R cells. The response appeared to involve specifically CD1b or CD1c (not CD1a or CD1d), and could be blocked by anti-TCR $\gamma\delta$, anti-lipid-A and anti-CD1b/c antibodies. Nevertheless, the direct involvement of the TCR in lipid-A recognition is not ascertained and lipid-A/CD1-mediated stimulation may represent a co-stimulus for $\gamma\delta$ T cells since anti-TLR-4 antibodies also inhibited $\gamma\delta$ cell reactivity. In addition, V δ 2⁺ as well as V δ 1⁺ cells seemed to be responding and anti-TCR antibodies used for $\gamma\delta$ -cell preparation may have had a role in activation.

MHC class-I-related chain A/B (MICA/B)

MICA/B are polymorphic β 2m-linked MHC-Ib molecules constitutively expressed in intestinal epithelial cells and many tumors. They are inducible by stress on multiple cell types. Both are ligands for the activating receptor NKG2D expressed on NK cells, some cytolytic CD8⁺ T cells and subsets of $\gamma\delta$ cells (Champsaur and Lanier 2010). Groh *et al.* have obtained MICA/MICB-reactive $\gamma\delta$ T cell clones from human intestinal mucosae and found that these reacted specifically against MICA-transfected C1R targets but not against C1R transfected with another NKG2D ligand (ULBP1). All 5 clones described expressed V δ 1 associated with different V γ chains (V γ 1.3, 1.4, 1.5 or 1.8), again with no obvious homology in CDR3 γ/δ besides a common usage of J δ 1, suggesting that CDR3 regions do not contribute significantly to MIC reactivity (Groh *et al.* 1998). The usage of MICA tetramers indicated that these specifically stained MICA-reactive clones (Wu *et al.* 2002). Strikingly, some V δ 1/V γ 1.4 clones were non-reactive to MICA and did not stain with MICA tetramers, suggesting either a role for CDR3 or the involvement of co-receptors or co-activators besides the TCR. As $\gamma\delta$ T cells frequently express the NKG2D receptor which also binds to MIC proteins, this may have been involved. TCR-dependence of this recognition was however analyzed using TCR transfer in HPB-ALL T cells which lack NKG2D expression. Although these cells could not be functionally tested for antigen reactivity, MICA tetramers specifically stained cells transfected with reactive TCRs. Strikingly again, although the contribution of gamma chains did not appear previously, transfection of mixed combinations of V γ and V δ chains from responder clones produced TCRs which did not stain with the tetramers.

MICA reactivity was also described for $\gamma\delta$ clones infiltrating ovarian carcinomas. These clones were expanded using recombinant α 1 α 2 MICA protein constructs and expressed V δ 1 associated with V γ 2, V γ 3 or V γ 4 (Zhao *et al.* 2006). V δ 1⁺ iIEs also react against MICA/B from multiple primate species despite considerable variation or alterations in the structure of α -helical regions of these α 1 α 2 domains (Steinle *et al.* 1998). Soluble recombinant single-chain TCR constructs corresponding to these TCRs as well as mixed combinations of V γ and V δ chains were subsequently tested for binding on HeLa cells which expressed MICA. Authors show a strong dependence on V δ 1 chains for soluble TCR binding to HeLa cells with little dependence on the associated γ -chain. In both studies, TCR-MICA interactions could be inhibited by antibodies recognizing the α 1 or α 2 domains of MICA.

UL16-binding protein family (ULBP/RAET1)

This family of MHC-Ib antigens comprises six known functional members in humans (Eagle *et al.* 2009; Eagle and Trowsdale 2007; Samarakoon *et al.* 2009). They are non-polymorphic, do not associate with β 2m and mice also express genes homologous to the human ones. Members of this family of NKG2D ligands are frequently expressed in tumors, virally-infected cells and selective normal tissues, and are inducible by retinoic acid and cellular stress. They can be linked to the plasma membrane through a trans-membrane domain or a GPI-anchor. Unlike MIC proteins, they lack the α 3 MHC-I domain (Champsaur and Lanier 2010). Their expression is modulated following CMV infection through binding to UL16 viral protein and through miRNAs (Stern-Ginossar *et al.* 2007; Stern-Ginossar and Mandelboim 2009). In normal tissues, ULBP4 expression is restricted to skin and small intestine (Bacon *et al.* 2004; Chalupny *et al.* 2003; Conejo-Garcia *et al.* 2003; Eagle *et al.* 2006; Radosavljevic *et al.* 2002). $\gamma\delta$ T cells infiltrating colonic and ovarian tumors could be stimulated and amplified with immobilized recombinant ULBP4, and the majority of these expressed V δ 2 although V δ 1 cells were also detected in cultures (Kong *et al.* 2009). The pairing of V δ 2 with V γ 9 in these tumor infiltrating lymphocytes (TIL) was not documented. Sequencing of V δ 2 chains revealed considerable diversity in CDR3 regions although most were associated with J δ 1. Using a V γ 9V δ 2 TCR-Fc recombinant soluble construct carrying the V δ 2 TIL sequence and a synthetic V γ 9-J γ 1.2 (also called JP) γ -chain, the authors have shown that this could stain specifically EL4 cells transfected with ULBP4. It also stained Daudi lymphoma cells which are classical targets for V γ 9V δ 2 cells, although it remains to be shown that this is due to ligation of ULBP4 on these cells. Reactivity of V γ 9V δ 2 cells against EL4-ULBP4 could be inhibited by both anti-TCR and anti-NKG2D antibodies, indicating that both receptors were involved in ULBP4 recognition.

ULBP1 has been shown to be determinant in the recognition of hematopoietic tumors by V γ 9V δ 2 lymphocytes expanded from peripheral blood. Nevertheless, in Lanca's study (Lanca *et al.* 2010), this appeared to be mediated essentially through ULBP1 interaction with NKG2D rather than with the TCR and was not hampered by anti-TCR antibodies. Similarly, although ULBP3 expression on B-CLL is determinant for their recognition by some V δ 1 cells, there is no evidence in favor or against an involvement of the TCR in their recognition of ULBP3 (Poggi *et al.* 2004).

HLA-E and Qa-1

Barkonyi *et al.* reported that V γ 9V δ 2 cells expanded from peripheral blood recognized preferentially and made conjugates with choriocarcinoma targets (JAR) transfected with HLA-G/E whereas V δ 1 cells did not. There is no evidence for an involvement of the TCR in HLA-E/G recognition (Barkonyi *et al.* 2002). Nevertheless, in the mouse, the activation of some $\gamma\delta$ cells were reported to be dependent on the recognition of Qa-1, the murine counterpart of HLA-E. These included a V γ 2/V δ 6.1 hybridoma (DGT3) recognizing Glu-Tyr polymer + Qa-1b (Vidovic and Dembic 1991 ; Vidovic *et al.* 1989), and, more recently, a population of CD8⁺ iIEL expressing V γ 4 and expanding in the course of salmonella infections (Davies *et al.* 2004). Although Qa-1 can activate CD94/NKG2D family of receptors on $\gamma\delta$ T cells, these receptors were found absent on this particular $\gamma\delta$ subset. The presence of these cells in the intestine is strongly dependent on TAP and Qa-1. Although not a definitive proof, this strong dependency on Qa-1 for development suggests an involvement of the TCR in Qa-1 recognition. Nevertheless, since Qa-1 expression is dependent on endogenous peptides for expression and may be altered by TAP deficiency, TAP sensitivity of V γ 4 T cell development does not imply that they recognize peptides in a Qa-1-restricted manner (Kambayashi *et al.* 2004).

T10/T22

Since the initial description that some murine $\gamma\delta$ T cells were stimulated by MHC antigens encoded in the T region of the MHC (Bluestone *et al.* 1988 ; Bonneville *et al.* 1989), T cell clones recognizing the two closely homologous MHC-Ib antigens T10 and T22 have been isolated and extensively studied (reviewed in (Chien and Konigshofer 2007 ; Meyer *et al.* 2010)). These molecules are not ubiquitously expressed and cannot present peptides due to an alteration of their α 1/ α 2 domains similar to that of MICA (Wingren *et al.* 2000). KN6 and G8 $\gamma\delta$ T cell clones can be directly activated by immobilized T10 or T22 molecules in complexes with β 2m, independently of peptides. T22 tetramers can stain 0.2 to 2% of splenic $\gamma\delta$ T cells in normal mice and most are V γ 1⁺ or V γ 4⁺ (Crowley *et al.* 2000). They also stain few V γ 7⁺ cells in the intestine. Although T22-specific clones may also use different V γ regions, it was found that there is a strict requirement for the expression of a CDR3 δ region bearing the mostly D δ 2-encoded motif (S)EGYEL, whereas surrounding N, P and J δ -encoded diversity had little influence on antigen recognition. The crystallographic structure of the G8 TCR in complex with T22 has been obtained and confirmed the involvement of the CDR3 δ loop in the contact with T22. Strikingly however, it was found that G8 binding to T22 differed considerably from that of $\alpha\beta$ TCRs with peptide-MHC complexes, in that the two complexes were bound at an angle and CDR loops other than CDR3 δ did not seem to contribute significantly to this binding (Adams *et al.* 2005). Thus, the G8 TCR does not recognize its target MHC antigen as do $\alpha\beta$ TCRs which must align with the MHC molecule and use all six CDR regions to make contacts with the MHC-peptide complex.

This unexpected interaction suggests that the G8 TCR might interact simultaneously with molecules surrounding the T22 complex on stimulatory cells, an interaction which may involve other CDR regions and could influence fine specificity. The T22 molecule would thus be only part of the physiological TCR ligand. T22 tetramers can stain efficiently cells expressing the relevant TCRs and accordingly T10 or T22 recombinant complexes can stimulate G8 cells in experimental settings (Chien and Konigshofer 2007). This may be explained by an inherent high affinity of the (S)EGYEL CDR3 δ motif for T10/T22, multivalence of the tetramers and the high amounts of antigen used experimentally for stimulation. This interaction may however not be sufficient in more physiological conditions. This hypothesis would lead to reconsider the interpretation of findings relating to G8-TCR T cell development in mice lacking T10/T22 and will be discussed further below.

MHC-class II molecules

A human $\gamma\delta$ T cell clone (V γ 9V δ 2) with dual specificity has been reported to recognize mycobacterial antigens in a non-MHC restricted fashion and a tetanus toxoid (TT) peptide in the context of an MHC-II molecule (DRw53) (Holoshitz *et al.* 1989). Kozbor *et al.* reported the isolation of TT-specific $\gamma\delta$ T cell clones from a TT-hyper-immune individual. These responded specifically to TT in the presence of autologous APCs, were CD8⁺, could be inhibited by anti-DR antibodies and appeared DR4-restricted (Kozbor *et al.* 1989). The frequency of such apparent MHC restriction to protein antigens is unknown. Moreover, although the reactivity was strikingly similar to that of MHC-II restricted $\alpha\beta$ T cells, evidence that TT was recognized in the form of classical peptide-MHC complexes was not reported. MHC-II-alloreactive murine $\gamma\delta$ clones have been obtained (Matis *et al.* 1989). LKB5 (specific for IE alleles) and LKD1 (specific for IA^d) have been isolated and LKB5 has been extensively studied (see (Chien *et al.* 1996 ; Chien and Konigshofer 2007) for a complete review on this system). Both clones were found to bind MHC independently of peptide and peptide-loading mechanisms and could be stimulated by purified peptide-stabilized MHC-II complexes as well as MHC-II-transfected CHO cells. Moreover, they share identical V γ and V δ sequences except in CDR3 regions (Matis *et al.* 1989 ; Meyer *et al.* 2010). The scanning of IE mutations which affected TCR recognition have revealed that the $\gamma\delta$ TCR epitopes did not involve amino acid locations usually involved in MHC-II recognition by $\alpha\beta$ TCRs but instead a functional polymorphic epitope near β 67/70. Recognition was also affected by a residue at one end of the peptide groove (α 79) possibly affecting the glycosylation state of the molecule on the α 82 residue (Schild *et al.* 1994). From these data, the interaction between LKB5 TCR and IE must differ fundamentally from that of $\alpha\beta$ TCR and IE. Nevertheless IE^k +peptide tetramer complexes could stain relevant IE^k-restricted $\alpha\beta$ T cell clones but did not stain LKB5, possibly because of a particularly low affinity of the

TCR for the MHC ligand (Chien and Konigshofer 2007). We suggest the possibility that the LKB5 TCR co-recognizes an unknown membrane ligand associated with IE outside of the peptide binding groove resulting in a low affinity for IE alone.

$\gamma\delta$ TCR ligands non related to MHC

Many of these putative antigens have been reviewed recently (Born and O'Brien 2009 ; Konigshofer and Chien 2006). For convenience, non-MHC ligands for V γ 9V δ 2 cells are described in separate sections.

Herpes simplex virus 1-glycoprotein I (HSV-gI)

HSV-gI (US7 gene product) is thought to be involved in viral assembly and intracellular routing in neurones. In HSV-infected mice, Bluestone and coworkers have found that $\gamma\delta$ T cells bearing V γ 1.2 and V δ 8 expand in infected mice and react in vitro specifically to gI (Johnson et al. 1992). T cell stimulation could be obtained with purified protein in the absence of APCs, did not require glycosylation and was conformation-dependent (Sciammas and Bluestone 1998). gI could also be recognized on the surface of gI-transfected fibroblasts, and cells expressing gI mutants incapable to reach the cell surface were not recognized. Recognition did not require MHC-I, MHC-II or TAP, excluding conventional recognition of antigenic peptides (Sciammas et al. 1994). gI was thus recognized either as a conformed soluble or membrane-anchored protein and specialized APCs were not required. HSV-reactive $\gamma\delta$ cells have also been found in humans (Bukowski et al. 1994). These belonged to the V γ 9V δ 2 subset and probably recognized an endogenous, virus-induced ligand instead of a viral protein as they were equally reactive with T cell blasts infected with HSV and the unrelated vaccinia virus.

Lipids

The possible involvement of lipids in conjunction with CD1 isoforms in the stimulation of $\gamma\delta$ cells has been mentioned above in the CD1 section. $\gamma\delta$ T cells responsive to glycerophospholipids in an unusual manner have also been described (Born et al. 2003). Indeed, some murine $\gamma\delta$ T cell hybridomas derived from non-immunized mice were found to respond in a TCR-dependent manner to cardiolipin and, although more weakly, to the related phospholipids phosphatidyl glycerol, phosphatidic acid and cytidine diphosphate-phosphatidyl glycerol, whereas phosphatidyl inositol, phosphatidyl-serine and phosphatidyl choline were not stimulatory. In addition to particular head groups, the acyl chains were also important for antigenic activity as deacylated lipids did not stimulate. Responsive $\gamma\delta$ cells all expressed murine V γ 1 and two characterized V δ chains belonged to the V δ 6 family. Stimulation did not require APCs. TCR transfer experiments have confirmed the strict TCR dependence of lipid recognition. However, these hybridomas as well as the TCR transfectants were characterized by a strong, also TCR-dependent, self reactivity (lymphokine secretion in absence of added lipid) as well as by their dependence on a serum component which could be replaced by β 2-glycoprotein-1 (β 2-GP1). This protein (also called apolipoprotein H) can be a component of circulating lipoproteins and was independently shown to bind to cell surfaces, in particular apoptotic cells, and to undergo conformation and orientation changes upon lipid binding (Wang et al. 2002 ; Wang et al. 2000). As pathogenic human anti-phospholipid autoantibodies have been shown to recognize cardiolipin- β 2-GP1 complexes, it is quite possible that the murine hybridomas recognize similar complexes through their TCR. Nevertheless, one cannot exclude that another component present on the T cell or hybridoma surface is co-recognized and is responsible for the self-reactivity of their TCR.

Heat shock proteins (HSP)

In humans as well as in mice, $\gamma\delta$ T cells expand in response to mycobacterial infections. Earlier works have shown that murine $\gamma\delta$ T cells could be stimulated with HSP65 from mycobacterial extracts and similarly with recombinant proteins including the close relative endogenous mammalian HSP60 (Kabelitz et al. 1990 ; Kobayashi et al. 1994). Reactive murine cells expressed V γ 1 in association with several V δ 2 genes, predominantly V δ 6, and represented 10–20% of unselected splenic or lymph node $\gamma\delta$ T cells (Kobayashi et al. 1994 ; O'Brien et al. 1992). As for lipid-reactive $\gamma\delta$ cells, HSP-reactive T cell clones and hybridomas were found to have significant autoreactivity. Autoreactivity as well as HSP60 reactivity could be conferred to non-reactive cells by relevant $\gamma\delta$ TCR transfer experiments. A minimal 7aa FGLQLEL conserved peptide (HSP60^{181–190}) was also found efficient with an essential contributions of amino acid positions 181 and 183 (Fu et al. 1994). APCs and antigen processing were not required for HSP recognition and recognition was not restricted by MHC-I or MHC-II classical antigens. This establishes a direct involvement of the TCR in HSP60 reactivity. A cognate interaction of the TCR with the stimulating peptide is not ascertained and HSP peptides could possibly alter the expression or the conformation of another TCR ligand expressed on the surface of hybridomas and responsible for their inherent autoreactivity.

Endogenous HSP70 family proteins were also found to stimulate murine V δ 8 cells (Kishi et al. 2001). In humans, V γ 9V δ 2 T cells which react against mycobacterial antigens were initially found to be stimulated by HSP60 homologs (Haregewoin et al. 1989) and an endogenous protein of this family was found to be expressed on the surface of Daudi lymphoma cells which can activate cells expressing a V γ 9V δ 2 TCR after TCR transfer (Bukowski et al. 1998 ; Fisch et al. 1990 ; Kaur et al. 1993). Although anti-HSP antibodies could stain cells which were not V γ 9V δ 2 targets, they could however block recognition of V γ 9V δ 2-targeted tumors (Selin et al. 1992). Since these earlier studies, potent non-peptidic phosphoantigens rather than HSPs have been found to be the essential stimulatory components of mycobacterial extracts for human V γ 9V δ 2 cells and no TCR transfer experiment have demonstrated that human $\gamma\delta$ TCRs have a direct role

in HSP recognition. Expansion of V γ 9V δ 2 cells have been reported using HSP60 as well as HSP70 proteins (Chauhan et al. 2007 ; Wadia et al. 2005 ; Zhang et al. 2005).

Although they are essentially cytosolic, secreted through exosomes (Gupta and Knowlton 2007 ; Lancaster and Febbraio 2005) and/or transported to the cell surface by mechanisms which are not well defined, HSPs were found expressed on normal, transformed or apoptotic cells and their surface expression can be up-regulated by various stimuli, in particular TLR stimulation, bacterial and parasitic infections (Hirsh et al. 2006 ; Hirsh and Junger 2008). Endogenous or exogenous HSPs and HSP peptides can bind to cell surfaces via multiple receptors including CD91, may provide a potent stimulus for some $\gamma\delta$ T cell subpopulations and this is not be limited to HSP60 family (Binder et al. 2004 ; Habich et al. 2002 ; MacAry et al. 2004). Zhang *et al.* have reported the stimulation of V γ 9V δ 2 cells by EBV-transformed LCL expressing HSP70 and this was abrogated by silencing HSP70 expression in target cells (Zhang et al. 2005). HSP70 expression in lung neutrophils may also be involved in their killing by $\gamma\delta$ cells in a murine model of sepsis (Hirsh et al. 2006). In many of these systems, there is no substantial evidence that HSP recognition involves the TCR. HSP70 was found to bind to TLR2 and TLR4 (Asea 2008). More generally, HSPs may signal through TLRs, directly or indirectly through binding peptides or hydrophobic ligands, including LPS (Tsan and Gao 2009 ; Zhao et al. 2007). Nevertheless, HSPs could also be non-specific ligands for the TCR or target antigens and affect specific TCR-antigen cognate interaction.

Insulin peptide

Insulin provided in aerosol has been reported to induce $\gamma\delta$ cells with regulatory activity in autoimmune diabetic NOD mice (Harrison et al. 1996). In addition to insulin peptide-specific $\alpha\beta$ T cells, $\gamma\delta$ T cell hybridomas responding to a diabetogenic insulin B:9-23 epitope could be derived from NOD mice with or without peptide immunization. Peptide-reactive $\gamma\delta$ hybridomas were diverse in terms of V γ and V δ gene usage. Unlike $\alpha\beta$ -T cell hybridomas which responded to the same epitope, reactive $\gamma\delta$ hybridomas did not require APCs and peptide mutations altered differently the response of $\alpha\beta$ and $\gamma\delta$ hybridomas. Simultaneous transfer of the γ and δ chains from a responsive hybridoma to $\alpha\beta$ TCR-deficient cells could confer peptide reactivity although unlike the original hybridoma their response was amplified by provision of APCs. Finally, B:9-23-specific $\gamma\delta$ cells responded to the processed peptide but not to native insulin (Zhang et al. 2010).

Glu-Tyr polymer

The biological relevance of Glu-Tyr polymer is not clear although it may mimic natural proteins from diverse pathogens. A reassessment of the reactivity of Glu-Tyr polymer-reactive $\gamma\delta$ cells revealed a similar frequency of these cells in β 2m-deficient mice, questioning the role of Qa-1 in polymer recognition (Cady et al. 2000). Unlike the DGT3 clone (see above HLE/Qa-1 section), new $\gamma\delta$ hybridomas recognized Glu-Tyr polymer in the absence of APCs and did not require Qa-1. They expressed mainly V γ 1 in association with various possible V δ genes. In one case, reactivity could be transferred through γ and δ TCR chain expression, did not require APCs and was maximal with the Glu⁵⁰Tyr⁵⁰ polypeptide.

6-kDa early secreted antigenic target protein (ESAT-6)

ESAT-6 from *M. tuberculosis* is one member of 23 related proteins secreted by the pathogen which constitute determinant virulence factors and are thought to play a role in pathogen escape from phagolysosomes. This family of proteins carries potent immunogenic epitopes in particular for CD4⁺ $\alpha\beta$ T lymphocytes. ESAT-6 binds to and can disrupt phospholipid membranes, an interaction which can induce conformational changes in the protein (Meher et al. 2006). The protein can be post-transcriptionally modified by cleavage or acetylation and makes heterodimers with another family member, culture filtrate component-10 (CFP10) (Brodin et al. 2004 ; Renshaw et al. 2005). A number of reports link $\gamma\delta$ T cell responses to the recognition of ESAT-6 following stimulation with mycobacterial extracts. When screening PPD components for stimulatory activity on bovine T cells, Rhodes *et al.* found that purified ESAT-6 could recapitulate the effects of PPD stimulation on WC1 (+) bovine $\gamma\delta$ cells (Welsh et al. 2002). Reindeer $\gamma\delta$ cells proliferate in response to ESAT-6 (Waters et al. 2006). Bovine $\gamma\delta$ cells were also found to respond, although more weakly, to the pyrophosphate antigens (Welsh et al. 2002) and thus some of the stimulatory activity of ESAT-6 on $\gamma\delta$ cells could be due to co-purified phosphoantigens (see below), although in the latter study recombinant ESAT-6 was used. Li and Wu have observed a similar activation of human peripheral blood V γ 9V δ 2 T cells with ESAT-6 which increased their CD69 and CD25 expression, produced IFN γ , underwent cell division, and stimulations were increased by CD28 co-stimulation (Li and Wu 2008). Although V γ and V δ expression was not determined, the responding cells are assumed to belong to the V γ 9V δ 2 subset which is responsive to mycobacterial extracts. Nevertheless, Casetti *et al.* could not confirm the responsiveness of human V δ 1 or V δ 2 subsets to ESAT-6 and the co-immunization of cynomolgus monkeys with an ESAT-6 fusion protein and a phosphoantigen did not significantly modulate the *in vivo* phosphoantigen response (Casetti et al. 2008 ; Cendron et al. 2007). Discrepancies between studies may be due to differences in ESAT-6 antigen preparations which may contain uncharacterized components, including phosphoantigens, or on the conformation of the protein. The properties of ESAT-6 as a $\gamma\delta$ TCR antigen have thus to be ascertained notably because TLR-2 can also be expressed on $\gamma\delta$ T cells (Martin et al. 2009 ; Mokuno et al. 2000 ; Pietschmann et al. 2009), and because ESAT-6 was found to bind TLR-2 (Pathak et al. 2007).

V γ 9V δ 2 T cell antigens

Human V γ 9V δ 2 T cells have been the focus of recent reviews (Bonneville and Fournie 2005 ; Nedellec et al. 2010). Their response to antigens does not require prior immunization and may result in a dramatic polyclonal amplification, leading to the concept that they recognize frequent pathogen- or tumor-associated determinants, and participate to the innate line of immune defense (reviewed in (Kabelitz and Wesch 2003)). They are activated by multiple tumor cells types *in vitro* , infiltrate human tumors *in vivo* and are thought to play a role in tumor immunity (Tanaka 2006). They also proliferate in the course of bacterial and parasitic infections and mediate protective responses against these pathogens (Morita et al. 1999). It is clear that the TCR is only one of the receptors which can promote their activation in these different contexts and that TLRs, CD16, NKG2D and NKRs, either of the lectin or immunoglobulin super-families tightly determine or regulate their activity. Nevertheless, multiple proteins and non-peptidic antigens have been described as potential TCR stimuli and the diversity of these ligands excludes a common mechanism of action. Some of them may however contribute in different ways to the mechanism of TCR-mediated antigen recognition. Highly specific stimulators comprise the non peptidic substances phosphoantigens, aminobisphosphonates, alkylamines. Candidate proteins involved in their activation include apolipoprotein A-I, F1-ATPase, ULBP proteins and the above mentioned HSPs and ESAT-6.

Phosphoantigens and related non-peptidic compounds

Phosphoantigens were first isolated as phosphatase-sensitive and protease-resistant stimulatory components of mycobacterial extracts (Constant et al. 1994 ; Pfeffer et al. 1990 ; Schoel et al. 1994 ; Tanaka et al. 1995). The main antigen characterized is hydroxymethyl butenyl pyrophosphate (HMBPP, also called HDMAPP), a metabolite produced in many bacteria and apicomplexan parasites through the methyl-erythritol phosphate biochemical pathway (MEP, also called DOXP pathway) leading to isopentenyl pyrophosphate (IPP), a precursor for isoprenoid and steroid synthesis (Eberl et al. 2003 ; Hintz et al. 2001 ; Jomaa et al. 1999 ; Poupot and Fournie 2004). Together with HMBPP, nucleotide derivatives have been isolated from bacterial extracts and can also activate specifically V γ 9V δ 2 T cells (Constant et al. 1994 ; Poquet et al. 1996a ; Poquet et al. 1996b). The direct role of phosphoantigens in T cell stimulation is confirmed by the fact that multiple synthetic molecules with stimulatory activity have been produced, and this has confirmed the general requirement for a pyrophosphate or triphosphate nucleotide moiety linked through the terminal phosphate to a short alkyl chain, usually of five carbons (Espinosa et al. 2001 ; Morita et al. 2001 ; Zhang et al. 2006). Few molecules qualified as phosphoantigens do not follow this structure, in particular monoethyl phosphate, 2,3-diphosphoglyceric acid, β -D-ribosyl phosphate, xylose-1-phosphate and glycerol-3 phosphoric acid which require high micro or millimolar concentrations for stimulation (Burk et al. 1997 ; Burk et al. 1995 ; Morita et al. 1999 ; Tanaka et al. 1994). As the the ability of pyrophosphate antigens has been linked to the ability of the phosphodiester bond to be cleavable (Belmant et al. 1999 ; Belmant et al. 2000), it should be ascertained that a similar mechanism accounts for the stimulatory activity of all these phosphorylated compounds. Replacement of the phosphoester bond linking the alkyl chain to its proximal phosphate by a phosphatase-resistant phosphonate bond however leads to compounds with increased activity (Zgani et al. 2004). Several phosphoantigens carry one unsaturation on the alkyl chain and their stereochemical conformation was found determinant for activity (Boedec et al. 2008). Although active at concentrations far above that of the bacterial HMBPP (μ M versus nM range), IPP and its isomer DMAPP are phosphoantigens universally produced by bacterial and eukaryotic cells through the mevalonate pathway of isoprenoid synthesis (Poupot and Fournie 2004). Experimental manipulation of the mevalonate pathway in human tumor cells affects their stimulatory activity. Daudi Burkitt lymphoma cells are natural activators and targets for cytotoxic V γ 9V δ 2 and their stimulatory activity is abrogated by statins such as lovastatin and mevastatin, which block hydroxymethylglutaryl-CoA reductase (HMGR), an enzyme of the mevalonate pathway upstream of IPP/DMAPP synthesis (Gober et al. 2003 ; Thompson et al. 2002). Conversely, non-stimulatory cells become stimulant following accumulation of these compounds resulting from inhibiting farnesyl pyrophosphate synthase (FPP), the enzyme which consumes IPP for downstream isoprenoid synthesis. *E. coli* , a bacterium which cannot make HDMAPP because it does not carry the MEP pathway, can induce HMGR activation through dephosphorylation and alters the mevalonate metabolism in infected macrophages resulting in their stimulatory activity for V γ 9V δ 2 cells (Kistowska et al. 2008). T cell reactivity against pyrophosphate antigens can be transferred by V γ 9V δ 2 TCR expression in TCR-deficient cells and this confers reactivity to soluble antigens as well as against stimulatory tumors, demonstrating a direct involvement of the TCR in these responses (Bukowski et al. 1995).

Alkylamines (AA) and aminobisphosphonates (ABP) are two other classes of compounds which were found to have stimulatory activity for V γ 9V δ 2 cells. AAs are small aminated alkyl molecules from plants and microorganisms (Bukowski et al. 1999 ; Kamath et al. 2003). ABPs such as pamidronate and zoledronate are synthetic drugs used for treating bone resorption. Although their structure is reminiscent of that of phosphoantigens, they have different requirements for activity (Thompson et al. 2010). Whereas phosphoantigens of the pyrophosphate group can directly activate $\gamma\delta$ T cells in the absence of APCs, alkylamines and aminobisphosphonates have an absolute requirement for APCs of human or primate origin, although many non-professional APCs are efficient for presentation (Green et al. 2004 ; Kato et al. 2003 ; Miyagawa et al. 2001b). It is now clear that ABPs act indirectly as inhibitors of FPP thus inducing phosphoantigen accumulation in APCs which subsequently become stimulatory (Li et al. 2009 ; Rogers 2003). A similar property has been ascribed to AAs (Thompson et al. 2006). In addition to IPP, ABP stimulation of cells induces accumulation of triphosphoric acid 1-adenosin-5-yl ester 3- (3-methylbut-3-enyl) ester, an ATP derivative of isopentenyl (ApppI) (Monkkonen et al. 2006 ; Monkkonen et al. 2007 ; Rogers et al. 1996 ; Rogers et al. 1994). This product can be detected in Daudi cells as well as in cells treated with pamidronate (Monkkonen et al. 2007 ; Vantourout et al. 2009). Strikingly, we could not stimulate directly $\gamma\delta$ T cells with synthetic ApppI which required to be

pulsed on APCs. Its activity was not abrogated by statins and thus it did not appear to act as a modulator of the mevalonate pathway (Vantourout et al. 2009). It is not clear whether other nucleotidic phosphoantigens have a similar requirement for APCs, as their activity was generally measured in the presence of APCs (Morita et al. 2001).

Although pyrophosphate antigens can directly activate purified V γ 9V δ 2 cells, early studies uncovered a minimal requirement for autologous T cell-T cell contact (Lang et al. 1995; Morita et al. 1995). Although this might reveal the involvement of co-stimulatory or adhesion molecules, there is accumulating evidence that phosphoantigens are displayed on cellular membranes for T cell recognition. V γ 9V δ 2 reactivity with tumor surfaces supports this hypothesis. Sarikonda *et al.* could crosslink photoreactive phosphoantigens on cell surfaces and these were stimulatory in the absence of MHC-I, MHC-II or CD1 antigens (Sarikonda et al. 2008). In another study, Wei *et al.* have used macaque V γ 9V δ 2 TCR tetramer to reveal specific ligands on HMBPP-pulsed cells of monocytes, T or B cells of human or primate origin as well as mycobacteria-infected human monocytes. These tetramers did not bind mouse, rat or pig cells. In the latter case, recognition was abrogated by trypsin, suggesting that the phosphoantigen was recognized in a protein context (Wei et al. 2008).

Apolipoprotein A-I and F₁-ATPase

As Daudi Burkitt lymphoma cells are killed almost constantly by V γ 9V δ 2 cells, we have produced anti-Daudi monoclonal antibodies and selected one which did not react with non-stimulatory Raji Burkitt lymphoma cells and had modulatory activity on Daudi cell killing by V γ 9V δ 2 cells. This antibody, M5, could decrease cytotoxicity by 30% and was found to recognize apolipoprotein A-I (ApoA-I) adsorbed on the surface of Daudi cells (Scotet et al. 2005). Strikingly, positive M5 antibody staining of hematopoietic tumors correlated with low MHC-I expression (unpublished data). ApoA-I addition in killing assays performed in the absence of serum revealed that although the cytolytic activity of several $\gamma\delta$ T cell clones was increased by apoA-I, this was not always required. A possible effect of MHC-I interaction with inhibitory NKRs could have accounted for the observed correlation between M5 positivity (low MHC-I) and sensitivity to $\gamma\delta$ killing. Nevertheless, purified apoA-I but not apoA-II, was found to bind specifically to TCR V γ 9V δ 2 tetramers by surface plasmon resonance and was thus a possible TCR ligand. Subsequently, known apoA-I receptors were screened for their presence on V γ 9V δ 2-targeted tumor lines. The ecto-F₁-ATPase is one such receptor expressed on hepatocytes (Martinez et al. 2003). As for M5 staining, the detection of the α and or β chains of the mitochondrial ATP synthase/F₁-ATPase on tumor lines correlated, although not perfectly, with sensitivity to V γ 9V δ 2 killing. A purified F₁-ATPase bovine complex could also bind specifically to V γ 9V δ 2 TCR tetramers in surface plasmon resonance and ELISA assays, and this binding was increased by apoA-I, suggesting that apoA-I is an optional ligand which can stabilize F₁-ATPase-TCR interaction. When immobilized on polystyrene beads, the F₁-ATPase complex could be used to stimulate T cells from peripheral blood and induced detectable lymphokine production in the V γ 9V δ 2 subset (Scotet et al. 2005).

Ecto-F₁-ATPase (also called cell surface ATP synthase) is thought to be similar to the ATP synthase located in the inner membrane of mitochondria. This complex uses the energy of the electrochemical proton gradient generated by the respiratory chain between the intermembrane space and the mitochondrial matrix to catalyze ATP synthesis from ADP and inorganic phosphate while translocating protons across the inner mitochondrial membrane. In the absence of a proper proton gradient, the complex can hydrolyze ATP into ADP, a process which is normally regulated in mitochondria by the soluble polypeptide IF1 (inhibitor of F₁). The mitochondrial complex comprises an extramembrane F₁ domain ($\alpha\beta\beta_3$) which binds nucleotides and performs ATP synthesis/hydrolysis, and an intramembrane domain which functions as a rotor (c12 ring) and is the proton channel. The two domains are connected by a peripheral stalk of subunits (a , e , f , g , $A6L$, b , $F6$, d and OSCP) and a mobile central stalk ($\gamma\delta\epsilon$) which is attached to the rotor and produces conformational changes in the F₁ domain during rotation of the c-ring associated with proton translocation (Devenish et al. 2008). The α and β chains can be detected on cell surfaces of many cell types which orientates the complex with the catalytic F₁-domain on the outside (Champagne et al. 2006). F₁ and other components of this complex have been detected on rat cell membranes (b , g and OSCP) (Mangiullo et al. 2008), on osteosarcoma cells (d and OSCP) (Yonally and Capaldi 2006), or in membrane lipid raft or caveolae-enriched extracts (Bae et al. 2004; Bini et al. 2003; Kim et al. 2004; Sprenger et al. 2004; von Haller et al. 2001; Yu et al. 2005). F₁-ATPase-expressing cells include endothelial cells, hepatocytes, neurones, adipocytes (reviewed in. (Champagne et al. 2006)). This probably underscores the real tissue expression as detection of the complex on cell surfaces has proven difficult on tissue sections in particular due to the sensitivity of epitopes to fixation procedures. The strong mitochondrial expression hampers detection of the membrane complex by fluorescence techniques. F₁ epitopes are also removed by trypsinization of cells (unpublished observations). There is increasing evidence that the membrane complex is enzymatically active although there are discrepancies on the reaction it performs. ATP synthesis (Burrell et al. 2005; Kim et al. 2004; Mangiullo et al. 2008; Moser et al. 2001) as well as ATP hydrolysis (Fabre et al. 2006; Mangiullo et al. 2008; Martinez et al. 2003; Radojkovic et al. 2009; Yegutkin et al. 2001) have been reported and may depend on the extracellular environment of the cell as well as on its metabolic status. The metabolic activity of ecto-F₁-ATPase has been shown to modulate lipoprotein internalization by hepatocytes, to regulate endothelial and keratinocyte proliferation and apoptosis, and to regulate intestinal lipid absorption. In most instances, these functions were uncovered through evidence that this complex acts as a receptor for specific regulatory polypeptides: apoA-I, enterostatin, angiostatin, EMAPII and the regulatory subunits IF1 and F6 which can be found as circulating serum proteins (Champagne et al. 2006). How F₁-ATPase reaches the cell surface is unknown and presumably involves an intracellular vesicular pathway. A possible involvement of F₁-ATPase in intracellular vesicular trafficking is further supported by its association with phagosomes (Garin et al. 2001; Kovarova et

al. 2002 ; Li et al. 2003). The α -subunit may be glycosylated on the surface of murine cells suggesting that the complex may transit through the endoplasmic reticulum or Golgi apparatus (Schmidt et al. 2008).

Strikingly we could immunoprecipitate F_1 -ATPase components from the membrane extracts of cells where it was not detected by immunofluorescence, such as Raji or Jurkat cells. We assumed that F_1 -ATPase was indeed expressed on Raji and many other tumor lines but was not detected due to a masking of antibody epitopes by surrounding membrane components. MHC-I antigens are in large part involved in this interaction as stimulation of their expression leads to lower detection of F_1 -ATPase, whereas their disruption through β 2m removal by acid treatment leads to higher detection. MHC-I and F_1 -ATPase can be co-precipitated and colocalize partially in membrane patch-like domains on HepG2 cells (Vantourout et al. 2008). These findings indicate a close association of MHC-I antigens and F_1 -ATPase on the cell membrane. ApoA-I binding (detected with the M5 antibody) as well as the binding of most anti- F_1 -ATPase antibodies is likely to occur on an area of the F_1 surface which is alternatively occupied by an interaction with MHC-I, presumably close to the plasma membrane.

Possible role of F_1 -ATPase in phosphoantigen presentation

The presence of F_1 -ATPase in intracellular non-mitochondrial compartments and the modulator properties of chloroquine on phosphoantigen presentation (Rojas et al. 2002) are compatible with an involvement of this complex in some form of phosphoantigen antigen processing and presentation analogous to the mechanisms used for peptides and lipids. More investigations are required to determine if this is truly the case. Nevertheless, F_1 -ATPase is probably one of the determinant factors involved in phosphoantigen recognition by V γ 9V δ 2 cells. Although IPP does not induce a calcium flux in cells which do not make contacts with other cells, contact with beads coated with purified bovine F_1 -ATPase in the presence of IPP induces a weak and delayed calcium signal in individual cells indicating that F_1 -ATPase partly replaces the required interaction with autologous cell membranes. In accordance with this, the ability of contacting cells to induce a calcium flux in response to IPP correlated with their expression of F_1 -ATPase. In addition, cells which did not promote calcium flux were equally unable to promote V γ 9V δ 2 T cell responses after treatment with aminobisphosphonates and this correlated with undetectable F_1 -ATPase on their surface. These data are in favor of a strict requirement for F_1 -ATPase in phosphoantigen responses (Mookerjee-Basu et al. 2010).

Pichler *et al.* have shown that some $\alpha\beta$ T cells specific for hapten drugs in an MHC-II restricted fashion did not require stable binding of the drug to the MHC molecule although a TCR-dependent recognition of both the drug and the MHC were required. Accordingly, sulfametoxazole (SMX) cannot be pulsed on APCs but, when in solution, SMX can activate drug-specific T cells provided fixed APC with the relevant MHC are available for contact (Pichler et al. 2006 ; Schnyder et al. 1997 ; Zanni et al. 1998). A similar situation seems to occur with IPP: although some pyrophosphate antigens can be pulsed on APCs, this is usually regarded as poorly efficient, in particular when weak antigens are concerned, and the response to pyrophosphates requires contact with APCs, partially replaceable by beads carrying F_1 -ATPase (Mookerjee-Basu et al. 2010).

Purified F_1 -ATPases of multiple origins can bind multiple nucleotide analogs on three catalytic sites and additional non-catalytic sites of unknown function. It is thus not surprising that bovine F_1 -ATPase immobilized on polystyrene beads stably binds ApppI. Such complexes were found to induce TCR aggregation on V γ 9V δ 2 cells although F_1 -ATPase alone did not. We have observed however that they were unable to promote lymphokine secretion and expression of cytolytic activity marker CD107 unless a nucleotide pyrophosphatase (NPP) cleaving ApppI into IPP + AMP was added in the assay (Mookerjee-Basu et al. 2010).

There are conflicting reports on the ability of pyrophosphate antigens to induce TCR modulation: in the study by Lafont et al. (Lafont et al. 2001), $\gamma\delta$ T cell stimulation with IPP was unable to induce TCR down-modulation, whereas Sireci *et al.* (Sireci et al. 2001) found modulation with weak agonists including IPP but not with a stronger agonist BrHPP. Pyrophosphate antigens are probably too small to crosslink TCRs and it seems thus likely that TCR cross-linking and internalization results from the recognition of a putative presentation molecule (O'Brien et al. 2007). The efficiency of cross-linking and modulation may require a physical link between the phosphoantigen and the presentation molecule and this link may depend on chemistry of the phosphoantigen and experimental procedures. As opposed to the recognition of soluble phosphoantigens, the recognition of antigenic molecules on tumors, on APB-treated or on ApppI-treated cells invariably induces TCR aggregation and down-modulation. In these cases, antigen-loaded cells can be washed and remain stimulatory, suggesting that these situations lead to efficient antigen presentation through an endogenous pathway (APB, tumors), or through an exogenous route (ApppI and possibly phosphate antigens). ApppI can be captured by APCs (tumor cells or dendritic cells) and is efficiently presented to $\gamma\delta$ cell. In our hands this process was more efficient for ApppI than for IPP on a molar concentration basis. Although it is not clear whether this is due to better stability, more efficient capture by the cell or to a better efficiency of the presentation mechanism, this suggests that nucleotide derivatives could be a form of storage, transport and/or presentation for phosphoantigens produced intracellularly. Once ApppI was loaded on APCs, the stimulatory activity was similar to that of APB-treated cells or stimulatory tumors since it was resistant to extensive washing of stimulatory cells and did not require exogenous pyrophosphatase activity as opposed to ApppI loaded on purified F_1 -ATPase.

A definitive proof that F_1 -ATPase is a presentation molecule for phosphoantigens will require demonstration of its physical interaction with endogenously produced antigens and demonstration of an association of both components on the cell surface in physiological conditions. Another degree of complexity in the mechanism of phosphoantigen recognition is suspected from the observation that hydrolysis of ApppI is required for full T cell activation by F_1 -ATPase/ApppI complexes, although this hydrolysis is not necessary for recognition and TCR aggregation. This may indicate that ApppI/ F_1 are not physiological ligands. In support of this possibility, V γ 9V δ 2 TCR tetramers from macaques (Wei et al. 2008) can stain surface ligands on cells loaded with HMBPP, suggesting that HMBPP itself is efficiently presented. Additionally, cells which are stimulatory either naturally or following pulsing with ABP or ApppI do not require exogenous NPP activity. Nevertheless, an alternative possibility is that HMBPP, like IPP, can be converted into a nucleotide derivative by cells and presented as such in complex with the presentation molecule for recognition. In support, IPP does not make stable stimulatory complexes with F_1 -ATPase in vitro. A requirement for such complexes for initial recognition does not preclude the necessity of a hydrolysis step for full TCR activation, as is observed with ApppI/ F_1 complexes. The known hydrolytic activity of F_1 -ATPase cannot perform this hydrolysis as it would release isopentenyl monophosphate which is not a stimulus for V γ 9V δ 2 cells. Although it is not excluded that the F_1 -ATPase complex has unknown activities, such hydrolysis could be performed by one of the multiple ecto-nucleotide pyrophosphatases present on APC surfaces (Bollen et al. 2000; Goding et al. 2003; Yegutkin 2008).

Finally, it is possible that F_1 -ATPase is one among several possible presentation molecules for phosphoantigens. Spencer *et al.* have found that the T cells which were responsive to mycobacterial antigens presented by mycobacteria-infected macrophages, presumably through an endogenous route, were only a subset of the V γ 9V δ 2 which were expanded with pyrophosphate antigens loaded on dendritic cells, suggesting an involvement of different antigen presentation molecules or processing pathways (Spencer et al. 2008).

V γ 9V δ 2 TCR epitopes involved in ligand recognition

$\gamma\delta$ cells responding to mycobacteria and purified phosphate antigens all express V γ 9 and V δ 2 and TCR transfer experiments with mixed TCR chains from responder and non-responder $\gamma\delta$ clones have demonstrated the contribution of both V γ 9 and V δ 2 chains. Although responder clones carry an extreme diversity in the CDR3 γ and δ regions, most of them express J γ 1.2 and mutational analysis have demonstrated the tight contribution of two lysine residues encoded by J γ 1.2 (K¹⁰⁸-K¹⁰⁹). TCR transfer experiment with TCRs mutated in other CDR regions highlighted the contribution of a frequent N-region neutral or hydrophobic residue in position 97 of the δ chain (5' end of CDR3 δ), and an arginine residue in CDR2 δ (R51) (Bukowski et al. 1998; Miyagawa et al. 2001a; Morita et al. 2001; Yamashita et al. 2003). The three lysine residues and R⁵¹ are thought to provide an anchor for negatively charged phosphates in the antigen and the existence of a possible binding pocket for phosphoantigens involving these residues was substantiated by the crystal structure of a V γ 9V δ 2 TCR (Allison et al. 2001; Morita et al. 2001). Recently, a more extensive analysis of mutations which influence reactivity to phosphoantigens revealed that the area covered by critical mutations was much larger than the pocket initially defined, involved all six CDR regions and was largely germ-line encoded. Since this area exceeds the size of pyrophosphate antigens, Wang *et al.* conclude that these new critical residues likely contact a presentation molecule whereas pyrophosphate recognition mainly involves CDR3 $\gamma\delta$ (Wang et al. 2010).

Xu *et al.* have shown that a peptide derived from the CDR3 δ sequence of a V δ 2⁺ lymphocyte infiltrating an ovarian tumor, either alone or grafted onto an IgG, could be used to detect a surface ligand on epithelial tumors as well as in protein extracts from selected tissues (Xu et al. 2007). This CDR3 δ peptide displayed selectivity for certain tumors. Probing of a random peptide library with this peptide allowed the identification of sequences homologous to possible ligands, two of which HSP60 and MSH-2 were also detected with monoclonal antibodies on the cell surface of epithelial tumors (Chen et al. 2008). The implication of these candidate proteins as TCR targets has to be further explored, as well as the relationship between V δ 2⁺ ovarian and colon carcinoma infiltrating cells and the V γ 9V δ 2 T cell population which reacts to phosphoantigens.

Other possible protein ligands for $\gamma\delta$ TCRs

Recently, the *Skint1* gene was identified by a genetic approach as a member of a multigene family which determines the presence of V γ 5/V δ 1 DETCs in the skin of mice (Boyden et al. 2008). There is multiple evidence that DETCs recognize an endogenous ligand expressed in murine skin upon infections or stress (Jameson and Havran 2007). The *Skint1* gene encodes a protein of the immunoglobulin superfamily. Its expression is confined to murine thymus and epidermis and thus it could possibly be a TCR ligand determining thymic differentiation and tissue homing of DETC cells. Evidence for surface expression of the protein on murine cells which do not overexpress the gene artificially is still lacking but surface expression could be driven by contact with pathogens as suggested for H-2M3 or MR1 which are ligands for $\alpha\beta$ TCRs (Gulden et al. 1996; Le Bourhis et al. 2010; Lenz et al. 1996). Nevertheless, Skint may not be directly involved in TCR-mediated antigen recognition and an alternative possibility is that it is somehow involved in TCR ligand expression. Skint proteins share significant homology with proteins of the butyrophilin (BTN) family which are expressed in humans as well as in mice. In humans, the butyrophilin family comprises seven members some of which are expressed on the cell surface and have a wide tissue distribution (Rhodes et al. 2001). BTN products have also been shown to affect TCR-dependent stimulation of CD4 and CD8 $\alpha\beta$ T cells (Smith et al. 2010) but it is not known whether this is also true human $\gamma\delta$ T cells.

Non-V δ 2 $\gamma\delta$ T cells expand in the course of CMV infection or reactivation in transplanted patients (Dechanet et al. 1999 ; Pitard et al. 2008). CMV-specific $\gamma\delta$ T cell clones cross-react with determinants expressed on tumoral epithelial cells suggesting recognition of an endogenous ligand induced by CMV or tumoral processes (Halary et al. 2005). Similar expansions are observed in the course of CMV infection *in utero* (Vermijlen et al. 2010). In this latter study, $\gamma\delta$ T cells bearing a public TCR consisting of V δ 1 paired with V γ 8 were strikingly enriched in infected newborns. These TCRs were mostly germline-encoded as they had no P/N nucleotide additions in CDR3 and carried very similar CDR3 γ and δ sequences. This may be explained by the immaturity of the immune system. This population of cells is undetectable in uninfected newborns and is a unique example of $\gamma\delta$ T cell which can mount a vigorous oligoclonal response to a viral infection.

For completeness, one should mention the streptococcal superantigens SEA and SEB which have been shown to activate human V δ 1 (SEB) and V δ 2 (SEA) T cells, respectively, and do so by cross-linking the TCR γ -chain and MHC-II molecules from APCs in much the same way they bridge selected V β regions with MHC-II molecules to activate $\alpha\beta$ -T cells (Fikri et al. 2001 ; Loh et al. 1994 ; Maeurer et al. 1995 ; Ramesh et al. 1995). Superantigens represent an alternative way to activate $\gamma\delta$ as well as $\alpha\beta$ T cells through bypassing their fine antigen specificity (Rust and Koning 1993).

MHC-dependent antigen recognition and the “docking partner hypothesis”

Based on the analysis of CDR3 length distributions, Chien and collaborators have shown that the antigen-binding region of $\gamma\delta$ TCRs more closely resembles that of immunoglobulins than that of $\alpha\beta$ TCRs (Chien and Konigshofer 2007 ; Rock et al. 1994). The discovery of non-peptidic antigens and peptidic ones which did not require classical processing comforted the hypothesis that $\gamma\delta$ TCRs can recognize soluble antigens. Nevertheless, in most instances if not all, antigen recognition is potentiated by APCs and multiple evidence that antigens are preferentially recognized on cell surfaces have already been presented. In most instances, antigen recognition by $\gamma\delta$ TCRs appears to be dependent on germ line-encoded TCR elements. Born et al. thus proposed that $\gamma\delta$ TCRs have evolved to recognize conserved antigenic structures and that the particularly high potential variability of CDR sequences serves to provide TCR adaptability to diverse cell surface molecules which constitute the context in which these antigens are expressed.

From this review, it appears that candidate ligands for $\gamma\delta$ cells fall in three categories: MHC-like molecules, unrelated polypeptides and, in the case of V γ 9V δ 2 cells, small non-peptidic molecules. T cell activation by such diverse structures cannot involve similar TCR-ligand interactions. Although the example of superantigens (and also mitogens) shows that TCR activation can be achieved through multiple alternative interactions with MHC molecules, data in the V γ 9V δ 2 system do not fit well with this situation. The specificity of V γ 9V δ 2 cells for targeted tumors has been shown to be determined through the recognition of phosphoantigens produced endogenously, F₁-ATPase expressed at the cell surface, and MHC-Ib molecules. As described above, there is molecular evidence for TCR interaction with each of these structures although the TCR interaction with phosphoantigens still relies on indirect data. The involvement of molecules of the MHC family is ambiguous as they also participate to T cell activation through NKRs. The case of MICA recognition by human V δ 1 cells is striking as this antigen can bind the NKD2D receptor as well as the TCR. A similar situation is observed with V δ 2 TCRs which possibly bind ULBP4. To the diverse antigenic context proposed by Born et al, we thus suggest the variant hypothesis that antigens appear in a relatively defined context and our view is that above observations converge to a preferential involvement of MHC-related molecules recognized simultaneously with antigenic ligand.

The model of peptide-MHC recognition by $\alpha\beta$ T cells indicates that MHC molecules perform different essential functions: a) they provide an anchor for antigens on cell surfaces, the antigen binding cleft; b) they insure that these antigens are efficiently targeted by TCRs by providing a recognizable framework for recognition. This is usually called self-MHC restriction and is achieved through thymic positive selection; c) they are involved in specific presentation routes for exogenous or endogenous antigens which ensure that the antigens are recognized on defined APCs and initiate an appropriate immune response. This model is valid for $\alpha\beta$ TCRs which recognize polymorphic MHC-I or MHC-II molecules and those recognizing non-polymorphic CD1 together with lipids. How does it apply to $\gamma\delta$ -TCR antigen recognition? There is no evidence that homologous binding clefts in MHC-related $\gamma\delta$ TCR ligands provide anchors for antigens since they either do not have a proper binding cleft (ULBPs, MICA/B, T10/T22) or are recognized independently of peptides (IE). In the case of $\gamma\delta$ cells recognizing CD1, the role of lipids may similarly have no role in CD1-dependent recognition by $\gamma\delta$ TCRs, although they may be required for proper surface expression of complexes. Available data fit with a general model based on a simultaneous recognition of MHC molecules and ligands which are tightly associated outside the peptide groove. The complex would be recognized by the TCR through CDR regions although other framework TCR regions may also be involved. This model is supported by the following observations.

Based on available crystallographic data, the G8 TCR in complex with T22 uses almost exclusively CDR3 δ to contact the MHC ligand, with a minor contribution of CDR3 γ , none from other CDRs and with some docking flexibility. The difference in G8 TCR docking on T10 is extreme as compared with peptide/MHC restricted TCRs. Differences in docking on MHC ligand, as compared to $\alpha\beta$ -TCRs, is also reported for the TCRs of NKT cells on CD1, revealing that homologous MHC molecules can be ligated by different TCRs without

using conserved docking sites and alignment (Borg et al. 2007 ; Gadola et al. 2006 ; Godfrey et al. 2010 ; Kjer-Nielsen et al. 2006 ; Zajonc et al. 2008). The crystallographic analysis of unbound V γ 9V δ 2 TCR also uncovered a considerable difference between $\alpha\beta$ and $\gamma\delta$ TCRs architecture, that is an acute V γ /C γ interdomain angle of 42° observed in the G115 TCR (Allison et al. 2001). Although it is not yet reported that other $\gamma\delta$ TCRs have a similar tilted V domain, these structural features suggest that docking of $\gamma\delta$ TCRs on their ligands obeys to constraints which are different from those of other TCRs. CDR3 δ peptides can be used to detect ligands on the surface of target cells in an apparently specific manner supporting a relative autonomy for CDR3 δ in ligand binding. Finally, as already mentioned, CDR3 δ length distribution is uniquely long and is not constrained as other CDR regions in $\alpha\beta$ and $\gamma\delta$ TCRs (Chien and Konigshofer 2007 ; Rock et al. 1994). As CDR3 δ makes the essential contribution to the binding to T10, it seems plausible that other CDR regions are available for docking on tightly associated partners.

Some of the $\gamma\delta$ TCR ligands are membrane-anchored proteins which may represent such docking partners: the viral HSV-gI protein for V δ 8 murine TCRs and the F $_1$ -ATPase for human V δ 2 cells. Whereas association with MHC-like proteins is not documented for gI, F $_1$ -ATPase appears to associate with classical MHC-I molecules. We find plausible that it can also interact with ULBP proteins which would then be co-recognized with F $_1$ -ATPase. The presence or absence of an α 3 domain in the MHC-Ib molecule would then be determinant for defining the docking area on the [MHC/docking partner] complex, so that classical MHC-I antigens might not allow proper docking as opposed to ULBPs. In the case of F $_1$ -ATPase which is a self ubiquitous ligand, antigenicity could be conferred by associated phosphoantigens, whereas HSV-gI would be antigenic by itself. An inherent and somewhat autonomous affinity of different regions of the $\gamma\delta$ TCRs for MHC-like proteins and their partners would explain the physical associations observed in vitro between soluble TCRs and ULBP, MICA, T10/T22 on one side and F $_1$ -ATPase or gI on the other.

There are multiple lines of evidence that thymic positive selection occurs for murine V γ 5/V δ 1 DETC and related epidermic T cells (Lewis et al. 2006 ; Mallick-Wood et al. 1998 ; Xiong et al. 2004) although the selecting ligand is not yet identified. Mice which are deficient for MHC-II or MHC-I have normal numbers of $\gamma\delta$ cells indicating that classical MHC expression is not generally required for their development (Bigby et al. 1993 ; Correa et al. 1992). Nevertheless, in the case of murine $\gamma\delta$ T cells reacting against the β 2m-associated MHC-Ib T10/T22, it is intriguing that thymic antigen expression does not determine thymic selection of cells carrying relevant TCRs and G8 TCR-transgenic mice bred on β 2m 0 background have increased numbers of fully mature antigen-reactive $\gamma\delta$ cells (Schweighoffer and Fowlkes 1996). The recent work by Jensen *et al.* demonstrates that thymic expression on T10/T22 influences a functional switch in lymphokine secretion ability of the mature $\gamma\delta$ progeny and the straightforward conclusion was that these cells could have a unique thymic selection program and do not need positive selection (Jensen and Chien 2009). Thymic MHC-I/T10 expression appears to affect the phenotype of T10 tetramer-positive mature T cells, leading to higher expression of CD122 and lower HSA expression, indicative that they have encountered antigen in the thymus whereas cells developing in the absence of T10 are naive (Jensen and Chien 2009 ; Shibata et al. 2008). An alternative explanation would be that a binding partner of T10/T22 promotes thymic selection whereas T10/T22 are dispensable or replaceable by another MHC-like, β 2m-independent ligand for positive selection whereas T10/T22 would only influence TCR binding affinity.

The recognition of a combination of MHC-Ib molecules and a docking partner implies that each partner could contribute to the antigenicity of the complex. The T10/T22 system suggests at first sight that antigenicity is conferred by the polymorphic T10. This may however not be the case: although the original T10 b -reactive clone G8 does not react with T10 d , G8 transgenic mice carrying the T10 d haplotype have reduced numbers of $\gamma\delta$ thymocytes and Schweighoffer and Fowlkes provide evidence that in these mice the transgenic thymocytes undergo thymic deletion and have thus made cognate interactions with T10 d (Schweighoffer and Fowlkes 1996). Although we cannot exclude that T10 d induces anergy in mature cells, an alternative possibility is that T10 d cannot pair with the selecting partner of T10 b , altering thymic selection. The thymic partner of T10 b would possibly be replaced by another which induces thymic deletion of G8 cells. The non-reactivity of G8 cells towards T10 d would imply that this alternative partner is not expressed by APCs used for specificity assays. The existence of multiple tissue-specific binding partners would also explain why $\gamma\delta$ cells use different V genes for T22 recognition within different tissues while retaining the common CDR3 δ WSEGYEL motif (Shin et al. 2005). If this is the case, the antigenicity would be conferred by a combination of T10 with its partner rather than T10 itself and the MHC would confer only part of the $\gamma\delta$ -TCR recognizable framework.

Soluble molecules identified as possible $\gamma\delta$ TCR ligands may play different roles in antigen recognition. Phosphoantigens could confer antigenicity to F $_1$ -ATPase which could provide the anchoring function for the antigen. Similarly, phospholipids such as cardiolipin could confer antigenicity to some unknown conserved ligand. Hydrophobic proteins such as HSPs and apolipoproteins (apoA-I, apoH) could participate in TCR-ligand complexes by their ability to bind TCRs, MHC or partners through less specific interactions, and could facilitate or stabilize interactions. Their role may be more significant in the case of low affinity TCR-ligand interactions.

The possible recognition of NKG2D ligands by both the TCR and NKG2D raises the question of a possible co-ligand function for NKG2D. However, structural data on NKG2D/ligand complexes do not leave much room for a possible simultaneous interaction of both receptors on the same molecule (Radaev and Sun 2003). Together with other NKRs, NKG2D might rather signal in separate complexes

and allow $\gamma\delta$ cells to sense the cellular context of the antigenic stimulus. This would be essential in the case of endogenous and ubiquitous antigens such as IPP/ApppI in tumor cells. These putative autoantigens are however weak stimulators of V γ 9/V δ 2 cells and probably become significant only when they are strongly over-expressed or when the inhibitory signals by NKRs are decreased. They may as well provide a basal TCR signal licensing the $\gamma\delta$ T cell for a response to other receptors activated by stress or pathogen-associated stimuli.

Conclusion

The hypothetical [MHC + docking partner] recognition hypothesis is not much different from classical MHC-restriction for peptide-MHC recognition, and differs only by the nature of the ligand, the mode of ligand binding to the MHC, and the mode of docking of the TCR on the combined ligand. If many of the possible MHC-Ib molecules possibly involved are non-polymorphic, MICA proteins display high polymorphism concentrated on the outer edge of the residual cleft as well as in the α 3 domain which may influence interactions with surrounding membrane proteins and putative ligands. Together with the emerging view that $\gamma\delta$ T cells recognize antigens on cell surfaces, the points discussed here seem to indicate that antigen recognition by $\gamma\delta$ T cells shares most of the characteristics of peptide recognition by $\alpha\beta$ TCRs. Nevertheless it is likely that most of the antigenic ligands for $\gamma\delta$ T cells have not been fully identified and if a unifying model for antigen recognition by $\gamma\delta$ cells is proposed here, experimental evidence are still scarce and our view of antigen diversity is still unclear.

Acknowledgements:

This work was supported by the French Association pour la Recherche sur le Cancer.

List of abbreviations

AA : alkylamin
ABP : aminobisphosphonate
apoA-I : apolipoprotein A-I
ApppI : triphosphoric acid 1-adenosin-5'-yl ester 3- (3-methylbut-3-enyl) ester
 β 2m : β 2-microglobulin
BTN : butyrophilin family
BrHPP : bromohydrin pyrophosphate
CDR : complementarity determining region of the TCR
DMAPP : dimethylallyl pyrophosphate
EMAP II : endothelial monocyte-activating polypeptide II
ESAT-6 : early secreted antigenic target protein 6
HMBPP : (E)-1-hydroxy-2-methylbut-2-enyl 4-diphosphate
HMGR : 3-hydroxy-3-methyl-glutaryl-CoA reductase
HSP : heat shock protein
HSV : herpes simplex virus
IEL : intraepithelial lymphocytes
IF1 : inhibitor of F₁-ATPase
F₁-ATPase : F₁ adenosine triphosphatase
FPP : farnesyl pyrophosphate synthase
IPP : isopentenyl pyrophosphate
MAIT : Mucosal-associated invariant T cells
MEP : 2-C-methyl-D-erythritol-4-phosphate
MICA/B : MHC class-I-related chain A/B
NKR : Natural killer receptor
PPD : partially purified tuberculin derivative
SEA/B : staphylococcal enterotoxin A/B
SMX : sulfamethoxazole
TAP : transporter associated with antigen processing
TIL : tumor infiltrating lymphocyte
TT : tetanus toxoid
ULBP/RAET1 : unique long 16 (UL16)-binding protein/retinoic acid early transcript 1

References:

- Adams EJ , Chien YH , Garcia KC . 2005 ; Structure of a gammadelta T cell receptor in complex with the nonclassical MHC T22 . Science . 308 : 227 - 231
- Allison JP , Raulet DH . 1990 ; The immunobiology of gamma delta+ T cells . Seminars in immunology . 2 : 59 - 65
- Allison TJ , Winter CC , Fournie JJ . 2001 ; Structure of a human gammadelta T-cell antigen receptor . Nature . 411 : 820 - 824
- Asea A . 2008 ; Heat shock proteins and toll-like receptors . Handbook of experimental pharmacology . 111 - 127

- Bacon L, Eagle RA, Meyer M. 2004; Two human ULBP/RAET1 molecules with transmembrane regions are ligands for NKG2D. *J Immunol*. 173: 1078 - 1084
- Bae TJ, Kim MS, Kim JW. 2004; Lipid raft proteome reveals ATP synthase complex in the cell surface. *Proteomics*. 4: 3536 - 3548
- Barakonyi A, Kovacs KT, Miko E. 2002; Recognition of nonclassical HLA class I antigens by gamma delta T cells during pregnancy. *J Immunol*. 168: 2683 - 2688
- Belmont C, Espinosa E, Halary F. 1999; Conventional and non-conventional recognition of non-peptide antigens by T lymphocytes. *Comptes rendus de l'Academie des sciences*. 322: 919 - 924
- Belmont C, Espinosa E, Halary F. 2000; A chemical basis for selective recognition of nonpeptide antigens by human delta T cells. *Faseb J*. 14: 1669 - 1670
- Bigby M, Markowitz JS, Bleicher PA. 1993; Most gamma delta T cells develop normally in the absence of MHC class II molecules. *J Immunol*. 151: 4465 - 4475
- Binder RJ, Vatner R, Srivastava P. 2004; The heat-shock protein receptors: some answers and more questions. *Tissue antigens*. 64: 442 - 451
- Bini L, Pacini S, Liberatori S. 2003; Extensive temporally regulated reorganization of the lipid raft proteome following T-cell antigen receptor triggering. *The Biochemical journal*. 369: 301 - 309
- Bluestone JA, Cron RQ, Cotterman M. 1988; Structure and specificity of T cell receptor gamma/delta on major histocompatibility complex antigen-specific CD3+, CD4-, CD8- T lymphocytes. *J Exp Med*. 168: 1899 - 1916
- Boedec A, Sicard H, Dessolin J. 2008; Synthesis and biological activity of phosphonate analogues and geometric isomers of the highly potent phosphoantigen (E)-1-hydroxy-2-methylbut-2-enyl 4-diphosphate. *J Med Chem*. 51: 1747 - 1754
- Bollen M, Gijssbers R, Ceulemans H. 2000; Nucleotide pyrophosphatases/phosphodiesterases on the move. *Critical reviews in biochemistry and molecular biology*. 35: 393 - 432
- Bonneville M, Fournie JJ. 2005; Sensing cell stress and transformation through Vgamma9Vdelta2 T cell-mediated recognition of the isoprenoid pathway metabolites. *Microbes and infection/Institut Pasteur*. 7: 503 - 509
- Bonneville M, Ito K, Krecko EG. 1989; Recognition of a self major histocompatibility complex TL region product by gamma delta T-cell receptors. *Proc Natl Acad Sci USA*. 86: 5928 - 5932
- Borg NA, Wun KS, Kjer-Nielsen L. 2007; CD1d-lipid-antigen recognition by the semi-invariant NKT T-cell receptor. *Nature*. 448: 44 - 49
- Born WK, O'Brien RL. 2009; Antigen-restricted gammadelta T-cell receptors?. *Archivum immunologiae et therapiae experimentalis*. 57: 129 - 135
- Born WK, Vollmer M, Reardon C. 2003; Hybridomas expressing gammadelta T-cell receptors respond to cardiolipin and beta2-glycoprotein I (apolipoprotein H). *Scand J Immunol*. 58: 374 - 381
- Born WK, Yin Z, Hahn YS. 2010; Analysis of gammadelta T cell functions in the mouse. *J Immunol*. 184: 4055 - 4061
- Boyden LM, Lewis JM, Barbee SD. 2008; Skint1, the prototype of a newly identified immunoglobulin superfamily gene cluster, positively selects epidermal gammadelta T cells. *Nat Genet*. 40: 656 - 662
- Brodin P, Rosenkrands I, Andersen P. 2004; ESAT-6 proteins: protective antigens and virulence factors?. *Trends in microbiology*. 12: 500 - 508
- Bukowski JF, Morita CT, Band H. 1998; Crucial role of TCR gamma chain junctional region in prenyl pyrophosphate antigen recognition by gamma delta T cells. *J Immunol*. 161: 286 - 293
- Bukowski JF, Morita CT, Brenner MB. 1994; Recognition and destruction of virus-infected cells by human gamma delta CTL. *J Immunol*. 153: 5133 - 5140
- Bukowski JF, Morita CT, Brenner MB. 1999; Human gamma delta T cells recognize alkylamines derived from microbes, edible plants, and tea: implications for innate immunity. *Immunity*. 11: 57 - 65
- Bukowski JF, Morita CT, Tanaka Y. 1995; V gamma 2V delta 2 TCR-dependent recognition of non-peptide antigens and Daudi cells analyzed by TCR gene transfer. *J Immunol*. 154: 998 - 1006
- Burk MR, Carena I, Donda A. 1997; Functional inactivation in the whole population of human V gamma 9/V delta 2 T lymphocytes induced by a nonpeptidic antagonist. *J Exp Med*. 185: 91 - 97
- Burk MR, Mori L, De Libero G. 1995; Human V gamma 9-V delta 2 cells are stimulated in a cross-reactive fashion by a variety of phosphorylated metabolites. *Eur J Immunol*. 25: 2052 - 2058
- Burrell HE, Wlodarski B, Foster BJ. 2005; Human keratinocytes release ATP and utilize three mechanisms for nucleotide interconversion at the cell surface. *J Biol Chem*. 280: 29667 - 29676
- Cady CT, Lahn M, Vollmer M. 2000; Response of murine gamma delta T cells to the synthetic polypeptide poly-Glu50Tyr50. *J Immunol*. 165: 1790 - 1798
- Casetti R, Martino A, Sacchi A. 2008; Do human gammadelta T cells respond to M tuberculosis protein antigens?. *Blood*. 112: 4776 - 4777 author reply 4777
- Cendron D, Ingoure S, Martino A. 2007; A tuberculosis vaccine based on phosphoantigens and fusion proteins induces distinct gammadelta and alphabeta T cell responses in primates. *Eur J Immunol*. 37: 549 - 565
- Chalupny NJ, Sutherland CL, Lawrence WA. 2003; ULBP4 is a novel ligand for human NKG2D. *Biochem Biophys Res Commun*. 305: 129 - 135
- Champagne E, Martinez LO, Collet X. 2006; Ecto-F1Fo ATP synthase/F1 ATPase: metabolic and immunological functions. *Curr Opin Lipidol*. 17: 279 - 284
- Champsaur M, Lanier LL. 2010; Effect of NKG2D ligand expression on host immune responses. *Immunol Rev*. 235: 267 - 285
- Chauhan SK, Singh M, Nityanand S. 2007; Reactivity of gamma/delta T cells to human 60-kd heat-shock protein and their cytotoxicity to aortic endothelial cells in Takayasu arteritis. *Arthritis Rheum*. 56: 2798 - 2802
- Chen H, He X, Wang Z. 2008; Identification of human T cell receptor gammadelta-recognized epitopes/proteins via CDR3delta peptide-based immunobiochemical strategy. *J Biol Chem*. 283: 12528 - 12537
- Chien YH, Jores R, Crowley MP. 1996; Recognition by gamma/delta T cells. *Annu Rev Immunol*. 14: 511 - 532
- Chien YH, Konigshofer Y. 2007; Antigen recognition by gammadelta T cells. *Immunol Rev*. 215: 46 - 58
- Conejo-Garcia JR, Benencia F, Courreges MC. 2003; Letal, A tumor-associated NKG2D immunoreceptor ligand, induces activation and expansion of effector immune cells. *Cancer biology & therapy*. 2: 446 - 451
- Constant P, Davodeau F, Peyrat MA. 1994; Stimulation of human gamma delta T cells by nonpeptidic mycobacterial ligands. *Science*. 264: 267 - 270
- Correa I, Bix M, Liao NS. 1992; Most gamma delta T cells develop normally in beta 2-microglobulin-deficient mice. *Proc Natl Acad Sci U S A*. 89: 653 - 657
- Crowley MP, Fahrner AM, Baumgarth N. 2000; A population of murine gammadelta T cells that recognize an inducible MHC class Ib molecule. *Science*. 287: 314 - 316
- Cui Y, Cui L, He W. 2005; Unraveling the mystery of gammadelta T cell recognizing lipid A. *Cell Mol Immunol*. 2: 359 - 364
- Cui Y, Kang L, Cui L. 2009; Human gammadelta T cell recognition of lipid A is predominately presented by CD1b or CD1c on dendritic cells. *Biology direct*. 4: 47 -
- Das H, Groh V, Kuijl C. 2001; MICA engagement by human Vgamma2Vdelta2 T cells enhances their antigen-dependent effector function. *Immunity*. 15: 83 - 93
- Davies A, Lopez-Briones S, Ong H. 2004; Infection-induced expansion of a MHC Class Ib-dependent intestinal intraepithelial gammadelta T cell subset. *J Immunol*. 172: 6828 - 6837
- Dechanet J, Merville P, Berge F. 1999; Major expansion of gammadelta T lymphocytes following cytomegalovirus infection in kidney allograft recipients. *J Infect Dis*. 179: 1 - 8
- Devenish RJ, Prescott M, Rodgers AJ. 2008; The structure and function of mitochondrial F1F0-ATP synthases. *International review of cell and molecular biology*. 267: 1 - 58
- Eagle RA, Traherne JA, Ashiru O. 2006; Regulation of NKG2D ligand gene expression. *Human immunology*. 67: 159 - 169
- Eagle RA, Traherne JA, Hair JR. 2009; ULBP6/RAET1L is an additional human NKG2D ligand. *Eur J Immunol*. 39: 3207 - 3216
- Eagle RA, Trowsdale J. 2007; Promiscuity and the single receptor: NKG2D. *Nature reviews*. 7: 737 - 744
- Eberl M, Hintz M, Reichenberg A. 2003; Microbial isoprenoid biosynthesis and human gammadelta T cell activation. *FEBS Lett*. 544: 4 - 10
- Esmon CT. 2004; Structure and functions of the endothelial cell protein C receptor. *Critical care medicine*. 32: S298 - 301
- Espinosa E, Belmont C, Pont F. 2001; Chemical synthesis and biological activity of bromohydrin pyrophosphate, a potent stimulator of human gamma delta T cells. *J Biol Chem*. 276: 18337 - 18344

- Fabre AC , Vantourout P , Champagne E . 2006 ; Cell surface adenylate kinase activity regulates the F(1)-ATPase/P2Y (13)-mediated HDL endocytosis pathway on human hepatocytes . *Cell Mol Life Sci* . 63 : 2829 - 2837
- Faure F , Jitsukawa S , Miossec C . 1990 ; CD1c as a target recognition structure for human T lymphocytes: analysis with peripheral blood gamma/delta cells . *Eur J Immunol* . 20 : 703 - 706
- Fikri Y , Denis O , Pastoret P . 2001 ; Purified bovine WC1+ gamma delta T lymphocytes are activated by staphylococcal enterotoxins and toxic shock syndrome toxin-1 superantigens: proliferation response, TCR V gamma profile and cytokines expression . *Immunol Lett* . 77 : 87 - 95
- Fisch P , Malkovsky M , Kovats S . 1990 ; Recognition by human V gamma 9/V delta 2 T cells of a GroEL homolog on Daudi Burkitt's lymphoma cells . *Science* . 250 : 1269 - 1273
- Fisch P , Moris A , Rammensee HG . 2000 ; Inhibitory MHC class I receptors on gamma delta T cells in tumor immunity and autoimmunity . *Immunol Today* . 21 : 187 - 191
- Fu YX , Vollmer M , Kalataradi H . 1994 ; Structural requirements for peptides that stimulate a subset of gamma delta T cells . *J Immunol* . 152 : 1578 - 1588
- Gadola SD , Koch M , Marles-Wright J . 2006 ; Structure and binding kinetics of three different human CD1d-alpha-galactosylceramide-specific T cell receptors . *J Exp Med* . 203 : 699 - 710
- Garin J , Diez R , Kieffer S . 2001 ; The phagosome proteome: insight into phagosome functions . *J Cell Biol* . 152 : 165 - 180
- Gertner-Dardenne J , Bonnafous C , Bezombes C . 2009 ; Bromohydrin pyrophosphate enhances antibody-dependent cell-mediated cytotoxicity induced by therapeutic antibodies . *Blood* . 113 : 4875 - 4884
- Gober HJ , Kistowska M , Angman L . 2003 ; Human T cell receptor gammadelta cells recognize endogenous mevalonate metabolites in tumor cells . *J Exp Med* . 197 : 163 - 168
- Godfrey DI , MacDonald HR , Kronenberg M . 2004 ; NKT cells: what's in a name? . *Nature reviews* . 4 : 231 - 237
- Godfrey DI , Pellicci DG , Patel O . 2010 ; Antigen recognition by CD1d-restricted NKT T cell receptors . *Seminars in immunology* . 22 : 61 - 67
- Goding JW , Grobden B , Slegers H . 2003 ; Physiological and pathophysiological functions of the ecto-nucleotide pyrophosphatase/phosphodiesterase family . *Biochimica et biophysica acta* . 1638 : 1 - 19
- Green AE , Lissina A , Hutchinson SL . 2004 ; Recognition of nonpeptide antigens by human V gamma 9V delta 2 T cells requires contact with cells of human origin . *Clin Exp Immunol* . 136 : 472 - 482
- Groh V , Steinle A , Bauer S . 1998 ; Recognition of stress-induced MHC molecules by intestinal epithelial gammadelta T cells . *Science* . 279 : 1737 - 1740
- Gulden PH , Fischer P 3rd , Sherman NE . 1996 ; A *Listeria monocytogenes* pentapeptide is presented to cytolytic T lymphocytes by the H2-M3 MHC class Ib molecule . *Immunity* . 5 : 73 - 79
- Gupta S , Knowlton AA . 2007 ; HSP60 trafficking in adult cardiac myocytes: role of the exosomal pathway . *American journal of physiology* . 292 : H3052 - 3056
- Habich C , Baumgart K , Kolb H . 2002 ; The receptor for heat shock protein 60 on macrophages is saturable, specific, and distinct from receptors for other heat shock proteins . *J Immunol* . 168 : 569 - 576
- Halary F , Fournie JJ , Bonneville M . 1999 ; Activation and control of self-reactive gammadelta T cells . *Microbes and infection/Institut Pasteur* . 1 : 247 - 253
- Halary F , Pitard V , Dlubek D . 2005 ; Shared reactivity of V(delta)2(neg) {gamma}{delta} T cells against cytomegalovirus-infected cells and tumor intestinal epithelial cells . *J Exp Med* . 201 : 1567 - 1578
- Haregewoin A , Soman G , Hom RC . 1989 ; Human gamma delta+ T cells respond to mycobacterial heat-shock protein . *Nature* . 340 : 309 - 312
- Harrison LC , Dempsey-Collier M , Kramer DR . 1996 ; Aerosol insulin induces regulatory CD8 gamma delta T cells that prevent murine insulin-dependent diabetes . *J Exp Med* . 184 : 2167 - 2174
- Hayday AC . 2000 ; [gamma][delta] cells: a right time and a right place for a conserved third way of protection . *Annu Rev Immunol* . 18 : 975 - 1026
- Hintz M , Reichenberg A , Altincicek B . 2001 ; Identification of (E)-4-hydroxy-3-methylbut-2-enyl pyrophosphate as a major activator for human gammadelta T cells in *Escherichia coli* . *FEBS Lett* . 509 : 317 - 322
- Hirsh MI , Hashiguchi N , Chen Y . 2006 ; Surface expression of HSP72 by LPS-stimulated neutrophils facilitates gammadelta T cell-mediated killing . *Eur J Immunol* . 36 : 712 - 721
- Hirsh MI , Junger WG . 2008 ; Roles of heat shock proteins and gamma delta T cells in inflammation . *American journal of respiratory cell and molecular biology* . 39 : 509 - 513
- Holoshitz J , Koning F , Coligan JE . 1989 ; Isolation of CD4- CD8- mycobacteria-reactive T lymphocyte clones from rheumatoid arthritis synovial fluid . *Nature* . 339 : 226 - 229
- Holtmeier W . 2003 ; Compartmentalization gamma/delta T cells and their putative role in mucosal immunity . *Crit Rev Immunol* . 23 : 473 - 488
- Jameson J , Havran WL . 2007 ; Skin gammadelta T-cell functions in homeostasis and wound healing . *Immunol Rev* . 215 : 114 - 122
- Jensen KD , Chien YH . 2009 ; Thymic maturation determines gammadelta T cell function, but not their antigen specificities . *Curr Opin Immunol* . 21 : 140 - 145
- Johnson RM , Lancki DW , Sperling AI . 1992 ; A murine CD4-, CD8- T cell receptor-gamma delta T lymphocyte clone specific for herpes simplex virus glycoprotein I . *J Immunol* . 148 : 983 - 988
- Jomaa H , Wiesner J , Sanderbrand S . 1999 ; Inhibitors of the nonmevalonate pathway of isoprenoid biosynthesis as antimalarial drugs . *Science* . 285 : 1573 - 1576
- Kabelitz D , Bender A , Schondelmaier S . 1990 ; A large fraction of human peripheral blood gamma/delta + T cells is activated by *Mycobacterium tuberculosis* but not by its 65-kD heat shock protein . *J Exp Med* . 171 : 667 - 679
- Kabelitz D , Wesch D . 2003 ; Features and functions of gamma delta T lymphocytes: focus on chemokines and their receptors . *Crit Rev Immunol* . 23 : 339 - 370
- Kamath AB , Wang L , Das H . 2003 ; Antigens in tea-beverage prime human Vgamma 2Vdelta 2 T cells in vitro and in vivo for memory and nonmemory antibacterial cytokine responses . *Proc Natl Acad Sci U S A* . 100 : 6009 - 6014
- Kambayashi T , Kraft-Leavy JR , Dauner JG . 2004 ; The nonclassical MHC class I molecule Qa-1 forms unstable peptide complexes . *J Immunol* . 172 : 1661 - 1669
- Kato Y , Tanaka Y , Tanaka H . 2003 ; Requirement of species-specific interactions for the activation of human gamma delta T cells by pamidronate . *J Immunol* . 170 : 3608 - 3613
- Kaur I , Voss SD , Gupta RS . 1993 ; Human peripheral gamma delta T cells recognize hsp60 molecules on Daudi Burkitt's lymphoma cells . *J Immunol* . 150 : 2046 - 2055
- Kim BW , Choo HJ , Lee JW . 2004 ; Extracellular ATP is generated by ATP synthase complex in adipocyte lipid rafts . *Exp Mol Med* . 36 : 476 - 485
- Kishi A , Ichinohe T , Hirai I . 2001 ; The cell surface-expressed HSC70-like molecule preferentially reacts with the rat T-cell receptor Vdelta6 family . *Immunogenetics* . 53 : 401 - 409
- Kistowska M , Rossy E , Sansano S . 2008 ; Dysregulation of the host mevalonate pathway during early bacterial infection activates human TCR gammadelta cells . *Eur J Immunol* . 38 : 2200 - 2209
- Kjer-Nielsen L , Borg NA , Pellicci DG . 2006 ; A structural basis for selection and cross-species reactivity of the semi-invariant NKT cell receptor in CD1d/glycolipid recognition . *J Exp Med* . 203 : 661 - 673
- Kobayashi N , Matsuzaki G , Yoshikai Y . 1994 ; V delta 5+ T cells of BALB/c mice recognize the murine heat shock protein 60 target cell specificity . *Immunology* . 81 : 240 - 246
- Kong Y , Cao W , Xi X . 2009 ; The NKG2D ligand ULBP4 binds to TCR{gamma}9/{delta}2 and induces cytotoxicity to tumor cells through both TCR{gamma}{delta} and NKG2D . *Blood* . 214 : 310 - 317
- Konigshofer Y , Chien YH . 2006 ; Gammadelta T cells - innate immune lymphocytes? . *Curr Opin Immunol* . 18 : 527 - 533
- Kovarova H , Halada P , Man P . 2002 ; Proteome study of *Francisella tularensis* live vaccine strain-containing phagosome in Bcg/Nramp1 congenic macrophages: resistant allele contributes to permissive environment and susceptibility to infection . *Proteomics* . 2 : 85 - 93
- Kozbor D , Trinchieri G , Monos DS . 1989 ; Human TCR-gamma+/delta+, CD8+ T lymphocytes recognize tetanus toxoid in an MHC-restricted fashion . *J Exp Med* . 169 : 1847 - 1851

- Lafarge X , Pitard V , Ravet S . 2005 ; Expression of MHC class I receptors confers functional intraclonal heterogeneity to a reactive expansion of gammadelta T cells . *Eur J Immunol* . 35 : 1896 - 1905
- Lafont V , Liautard J , Sable-Teychene M . 2001 ; Isopentenyl pyrophosphate, a mycobacterial non-peptidic antigen, triggers delayed and highly sustained signaling in human gamma delta T lymphocytes without inducing down-modulation of T cell antigen receptor . *J Biol Chem* . 276 : 15961 - 15967
- Lanca T , Correia DV , Moita CF . 2010 ; The MHC class Ib protein ULBP1 is a nonredundant determinant of leukemia/lymphoma susceptibility to gammadelta T-cell cytotoxicity . *Blood* . 115 : 2407 - 2411
- Lancaster GI , Febbraio MA . 2005 ; Exosome-dependent trafficking of HSP70: a novel secretory pathway for cellular stress proteins . *J Biol Chem* . 280 : 23349 - 23355
- Lang F , Peyrat MA , Constant P . 1995 ; Early activation of human V gamma 9V delta 2 T cell broad cytotoxicity and TNF production by nonpeptidic mycobacterial ligands . *J Immunol* . 154 : 5986 - 5994
- Le Bourhis L , Martin E , Peguillet I . 2010 ; Antimicrobial activity of mucosal-associated invariant T cells . *Nature immunology* . 11 : 701 - 708
- Lenz LL , Dere B , Bevan MJ . 1996 ; Identification of an H2-M3-restricted Listeria epitope: implications for antigen presentation by M3 . *Immunity* . 5 : 63 - 72
- Leslie DS , Vincent MS , Spada FM . 2002 ; CD1-mediated gamma/delta T cell maturation of dendritic cells . *J Exp Med* . 196 : 1575 - 1584
- Lewis JM , Girardi M , Roberts SJ . 2006 ; Selection of the cutaneous intraepithelial gammadelta+ T cell repertoire by a thymic stromal determinant . *Nature immunology* . 7 : 843 - 850
- Li J , Herold MJ , Kimmel B . 2009 ; Reduced expression of the mevalonate pathway enzyme farnesyl pyrophosphate synthase unveils recognition of tumor cells by Vgamma9Vdelta2 T cells . *J Immunol* . 182 : 8118 - 8124
- Li L , Wu CY . 2008 ; CD4+ CD25+ Treg cells inhibit human memory gammadelta T cells to produce IFN-gamma in response to M tuberculosis antigen ESAT-6 . *Blood* . 111 : 5629 - 5636
- Li N , Mak A , Richards DP . 2003 ; Monocyte lipid rafts contain proteins implicated in vesicular trafficking and phagosome formation . *Proteomics* . 3 : 536 - 548
- Loh EY , Wang M , Bartkowiak J . 1994 ; Gene transfer studies of T cell receptor-gamma delta recognition. Specificity for staphylococcal enterotoxin A is conveyed by V gamma 9 alone . *J Immunol* . 152 : 3324 - 3332
- MacAry PA , Javid B , Floto RA . 2004 ; HSP70 peptide binding mutants separate antigen delivery from dendritic cell stimulation . *Immunity* . 20 : 95 - 106
- Maeurer M , Zitvogel L , Elder E . 1995 ; Human intestinal V delta 1+ T cells obtained from patients with colon cancer respond exclusively to SEB but not to SEA . *Nat Immun* . 14 : 188 - 197
- Mallick-Wood CA , Lewis JM , Richie LI . 1998 ; Conservation of T cell receptor conformation in epidermal gammadelta cells with disrupted primary Vgamma gene usage . *Science* . 279 : 1729 - 1733
- Mangiullo R , Gnoni A , Leone A . 2008 ; Structural and functional characterization of F(o)F(1)-ATP synthase on the extracellular surface of rat hepatocytes . *Biochimica et biophysica acta* . 1777 : 1326 - 1335
- Martin B , Hirota K , Cua DJ . 2009 ; Interleukin-17-producing gammadelta T cells selectively expand in response to pathogen products and environmental signals . *Immunity* . 31 : 321 - 330
- Martinez LO , Jacquet S , Esteve JP . 2003 ; Ectopic beta-chain of ATP synthase is an apolipoprotein A-I receptor in hepatic HDL endocytosis . *Nature* . 421 : 75 - 79
- Matis LA , Fry AM , Cron RQ . 1989 ; Structure and specificity of a class II MHC alloreactive gamma delta T cell receptor heterodimer . *Science* . 245 : 746 - 749
- Meher AK , Bal NC , Chary KV . 2006 ; Mycobacterium tuberculosis H37Rv ESAT-6-CFP-10 complex formation confers thermodynamic and biochemical stability . *The FEBS journal* . 273 : 1445 - 1462
- Meyer C , Zeng X , Chien YH . 2010 ; Ligand recognition during thymic development and gammadelta T cell function specification . *Seminars in immunology* . 22 : 207 - 213
- Miyagawa F , Tanaka Y , Yamashita S . 2001a ; Essential contribution of germline-encoded lysine residues in Jgamma1.2 segment to the recognition of nonpeptide antigens by human gammadelta T cells . *J Immunol* . 167 : 6773 - 6779
- Miyagawa F , Tanaka Y , Yamashita S . 2001b ; Essential requirement of antigen presentation by monocyte lineage cells for the activation of primary human gamma delta T cells by aminobisphosphonate antigen . *J Immunol* . 166 : 5508 - 5514
- Mokuno Y , Matsuguchi T , Takano M . 2000 ; Expression of toll-like receptor 2 on gamma delta T cells bearing invariant V gamma 6/V delta 1 induced by Escherichia coli infection in mice . *J Immunol* . 165 : 931 - 940
- Monkkonen H , Auriola S , Lehenkari P . 2006 ; A new endogenous ATP analog (ApppI) inhibits the mitochondrial adenine nucleotide translocase (ANT) and is responsible for the apoptosis induced by nitrogen-containing bisphosphonates . *Br J Pharmacol* . 147 : 437 - 445
- Monkkonen H , Ottewill PD , Kuokkanen J . 2007 ; Zoledronic acid-induced IPP/ApppI production in vivo . *Life Sci* . 81 : 1066 - 1070
- Mookerjee-Basu J , Vantourout P , Martinez LO . 2010 ; F1-adenosine triphosphatase displays properties characteristic of an antigen presentation molecule for Vgamma9Vdelta2 T cells . *J Immunol* . 184 : 6920 - 6928
- Morita CT , Beckman EM , Bukowski JF . 1995 ; Direct presentation of nonpeptide prenyl pyrophosphate antigens to human gamma delta T cells . *Immunity* . 3 : 495 - 507
- Morita CT , Lee HK , Leslie DS . 1999 ; Recognition of nonpeptide prenyl pyrophosphate antigens by human gammadelta T cells . *Microbes and infection/Institut Pasteur* . 1 : 175 - 186
- Morita CT , Lee HK , Wang H . 2001 ; Structural features of nonpeptide prenyl pyrophosphates that determine their antigenicity for human gamma delta T cells . *J Immunol* . 167 : 36 - 41
- Moser TL , Kenan DJ , Ashley TA . 2001 ; Endothelial cell surface F1-F0 ATP synthase is active in ATP synthesis and is inhibited by angiostatin . *Proc Natl Acad Sci U S A* . 98 : 6656 - 6661
- Nedellec S , Bonneville M , Scotet E . 2010 ; Human Vgamma9Vdelta2 T cells: from signals to functions . *Seminars in immunology* . 22 : 199 - 206
- O'Brien RL , Fu YX , Cranfill R . 1992 ; Heat shock protein Hsp60-reactive gamma delta cells: a large, diversified T-lymphocyte subset with highly focused specificity . *Proc Natl Acad Sci U S A* . 89 : 4348 - 4352
- O'Brien RL , Roark CL , Jin N . 2007 ; gammadelta T-cell receptors: functional correlations . *Immunol Rev* . 215 : 77 - 88
- Pathak SK , Basu S , Basu KK . 2007 ; Direct extracellular interaction between the early secreted antigen ESAT-6 of Mycobacterium tuberculosis and TLR2 inhibits TLR signaling in macrophages . *Nature immunology* . 8 : 610 - 618
- Pennington DJ , Vermijlen D , Wise EL . 2005 ; The integration of conventional and unconventional T cells that characterizes cell-mediated responses . *Adv Immunol* . 87 : 27 - 59
- Pfeffer K , Schoel B , Gulle H . 1990 ; Primary responses of human T cells to mycobacteria: a frequent set of gamma/delta T cells are stimulated by protease-resistant ligands . *Eur J Immunol* . 20 : 1175 - 1179
- Pichler WJ , Beeler A , Keller M . 2006 ; Pharmacological interaction of drugs with immune receptors: the p-i concept . *Allergol Int* . 55 : 17 - 25
- Pietschmann K , Beetz S , Welte S . 2009 ; Toll-like receptor expression and function in subsets of human gammadelta T lymphocytes . *Scand J Immunol* . 70 : 245 - 255
- Pitard V , Roumanes D , Lafarge X . 2008 ; Long-term expansion of effector/memory Vdelta2-gammadelta T cells is a specific blood signature of CMV infection . *Blood* . 112 : 1317 - 1324
- Poggi A , Venturino C , Catellani S . 2004 ; Vdelta1 T lymphocytes from B-CLL patients recognize ULBP3 expressed on leukemic B cells and up-regulated by trans-retinoic acid . *Cancer research* . 64 : 9172 - 9179
- Poquet Y , Constant P , Halary F . 1996a ; A novel nucleotide-containing antigen for human blood gamma delta T lymphocytes . *Eur J Immunol* . 26 : 2344 - 2349
- Poquet Y , Constant P , Peyrat MA . 1996b ; High-pH anion-exchange chromatographic analysis of phosphorylated compounds: application to isolation and characterization of nonpeptide mycobacterial antigens . *Anal Biochem* . 243 : 119 - 126
- Poupot M , Fournie JJ . 2004 ; Non-peptide antigens activating human Vgamma9/Vdelta2 T lymphocytes . *Immunol Lett* . 95 : 129 - 138
- Radaev S , Sun PD . 2003 ; Structure and function of natural killer cell surface receptors . *Annual review of biophysics and biomolecular structure* . 32 : 93 - 114
- Radojkovic C , Genoux A , Pons V . 2009 ; Stimulation of cell surface F1-ATPase activity by apolipoprotein A-I inhibits endothelial cell apoptosis and promotes proliferation . *Arteriosclerosis, thrombosis, and vascular biology* . 29 : 1125 - 1130

- Radosavljevic M, Cuillerier B, Wilson MJ . 2002 ; A cluster of ten novel MHC class I related genes on human chromosome 6q24.2–q25.3 . *Genomics* . 79 : 114 - 123
- Ramesh N, Horner A, Ahern D . 1995 ; Bacterial superantigens induce the proliferation of resting gamma/delta receptor bearing T cells . *Immunol Invest* . 24 : 713 - 724
- Renshaw PS, Lightbody KL, Veverka V . 2005 ; Structure and function of the complex formed by the tuberculosis virulence factors CFP-10 and ESAT-6 . *The EMBO journal* . 24 : 2491 - 2498
- Rhodes DA, Stammers M, Malcherek G . 2001 ; The cluster of BTN genes in the extended major histocompatibility complex . *Genomics* . 71 : 351 - 362
- Rock EP, Sibbald PR, Davis MM . 1994 ; CDR3 length in antigen-specific immune receptors . *J Exp Med* . 179 : 323 - 328
- Rogers MJ . 2003 ; New insights into the molecular mechanisms of action of bisphosphonates . *Curr Pharm Des* . 9 : 2643 - 2658
- Rogers MJ, Brown RJ, Hodkin V . 1996 ; Bisphosphonates are incorporated into adenine nucleotides by human aminoacyl-tRNA synthetase enzymes . *Biochem Biophys Res Commun* . 224 : 863 - 869
- Rogers MJ, Ji X, Russell RG . 1994 ; Incorporation of bisphosphonates into adenine nucleotides by amoebae of the cellular slime mould *Dictyostelium discoideum* . *The Biochemical journal* . 303 : (Pt 1) 303 - 311
- Rojas RE, Torres M, Fournie JJ . 2002 ; Phosphoantigen presentation by macrophages to mycobacterium tuberculosis--reactive Vgamma9Vdelta2+ T cells: modulation by chloroquine . *Infect Immun* . 70 : 4019 - 4027
- Russano AM, Agea E, Corazzi L . 2006 ; Recognition of pollen-derived phosphatidylethanolamine by human CD1d-restricted gammadelta T cells . *J Allergy Clin Immunol* . 117 : 1178 - 1184
- Russano AM, Bassotti G, Agea E . 2007 ; CD1-Restricted Recognition of Exogenous and Self-Lipid Antigens by Duodenal {gamma}{delta}+ T Lymphocytes . *J Immunol* . 178 : 3620 - 3626
- Rust CJ, Koning F . 1993 ; Gamma delta T cell reactivity towards bacterial superantigens . *Seminars in immunology* . 5 : 41 - 46
- Samarakoon A, Chu H, Malarkannan S . 2009 ; Murine NKG2D ligands: "double, double toil and trouble" . *Mol Immunol* . 46 : 1011 - 1019
- Sarikonda G, Wang H, Puan KJ . 2008 ; Photoaffinity Antigens for Human {gamma}{delta} T Cells . *J Immunol* . 181 : 7738 - 7750
- Schild H, Mavaddat N, Litzengerger C . 1994 ; The nature of major histocompatibility complex recognition by gamma delta T cells . *Cell* . 76 : 29 - 37
- Schmidt C, Lepsverdze E, Chi SL . 2008 ; Amyloid precursor protein and amyloid beta-peptide bind to ATP synthase and regulate its activity at the surface of neural cells . *Molecular psychiatry* . 13 : 953 - 969
- Schnyder B, Mauri-Hellweg D, Zanni M . 1997 ; Direct, MHC-dependent presentation of the drug sulfamethoxazole to human alphabeta T cell clones . *J Clin Invest* . 100 : 136 - 141
- Schoel B, Sprenger S, Kaufmann SH . 1994 ; Phosphate is essential for stimulation of V gamma 9V delta 2 T lymphocytes by mycobacterial low molecular weight ligand . *Eur J Immunol* . 24 : 1886 - 1892
- Schweighoffer E, Fowlkes BJ . 1996 ; Positive selection is not required for thymic maturation of transgenic gamma delta T cells . *J Exp Med* . 183 : 2033 - 2041
- Sciammas R, Bluestone JA . 1998 ; HSV-1 glycoprotein I-reactive TCR gamma delta cells directly recognize the peptide backbone in a conformationally dependent manner . *J Immunol* . 161 : 5187 - 5192
- Sciammas R, Johnson RM, Sperling AI . 1994 ; Unique antigen recognition by a herpesvirus-specific TCR-gamma delta cell . *J Immunol* . 152 : 5392 - 5397
- Scotet E, Martinez LO, Grant E . 2005 ; Tumor Recognition following Vgamma9Vdelta2 T Cell Receptor Interactions with a Surface F1-ATPase-Related Structure and Apolipoprotein A-I . *Immunity* . 22 : 71 - 80
- Selin LK, Stewart S, Shen C . 1992 ; Reactivity of gamma delta T cells induced by the tumour cell line RPMI 8226: functional heterogeneity of clonal populations and role of GroEL heat shock proteins . *Scand J Immunol* . 36 : 107 - 117
- Shibata K, Yamada H, Nakamura R . 2008 ; Identification of CD25+ gamma delta T cells as fetal thymus-derived naturally occurring IL-17 producers . *J Immunol* . 181 : 5940 - 5947
- Shin S, El-Diwanry R, Schaffert S . 2005 ; Antigen recognition determinants of gammadelta T cell receptors . *Science* . 308 : 252 - 255
- Sireci G, Espinosa E, Di Sano C . 2001 ; Differential activation of human gammadelta cells by nonpeptide phosphoantigens . *Eur J Immunol* . 31 : 1628 - 1635
- Smith IA, Knezevic BR, Ammann JU . 2010 ; BTN1A1, the mammary gland butyrophilin, and BTN2A2 are both inhibitors of T cell activation . *J Immunol* . 184 : 3514 - 3525
- Spada FM, Grant EP, Peters PJ . 2000 ; Self-recognition of CD1 by gamma/delta T cells: implications for innate immunity . *J Exp Med* . 191 : 937 - 948
- Spencer CT, Abate G, Blazevic A . 2008 ; Only a subset of phosphoantigen-responsive gamma9delta2 T cells mediate protective tuberculosis immunity . *J Immunol* . 181 : 4471 - 4484
- Sperling AI, Linsley PS, Barrett TA . 1993 ; CD28-mediated costimulation is necessary for the activation of T cell receptor-gamma delta+ T lymphocytes . *J Immunol* . 151 : 6043 - 6050
- Spits H, Paliard X, De Vries JE . 1989 ; Antigen-specific, but not natural killer, activity of T cell receptor-gamma delta cytotoxic T lymphocyte clones involves secretion of N alpha-benzyloxycarbonyl-L-lysine thiobenzyl ester serine esterase and influx of Ca2+ ions . *J Immunol* . 143 : 1506 - 1511
- Sprenger RR, Speijer D, Back JW . 2004 ; Comparative proteomics of human endothelial cell caveolae and rafts using two-dimensional gel electrophoresis and mass spectrometry . *Electrophoresis* . 25 : 156 - 172
- Steinle A, Groh V, Spies T . 1998 ; Diversification, expression, and gamma delta T cell recognition of evolutionarily distant members of the MIC family of major histocompatibility complex class I-related molecules . *Proc Natl Acad Sci U S A* . 95 : 12510 - 12515
- Stern-Ginossar N, Elefant N, Zimmermann A . 2007 ; Host immune system gene targeting by a viral miRNA . *Science* . 317 : 376 - 381
- Stern-Ginossar N, Mandelboim O . 2009 ; An integrated view of the regulation of NKG2D ligands . *Immunology* . 128 : 1 - 6
- Tanaka Y . 2006 ; Human gamma delta T cells and tumor immunotherapy . *J Clin Exp Hematop* . 46 : 11 - 23
- Tanaka Y, Morita CT, Nieves E . 1995 ; Natural and synthetic non-peptide antigens recognized by human gamma delta T cells . *Nature* . 375 : 155 - 158
- Tanaka Y, Sano S, Nieves E . 1994 ; Nonpeptide ligands for human gamma delta T cells . *Proc Natl Acad Sci U S A* . 91 : 8175 - 8179
- Thompson K, Dunford JE, Ebetino FH . 2002 ; Identification of a bisphosphonate that inhibits isopentenyl diphosphate isomerase and farnesyl diphosphate synthase . *Biochem Biophys Res Commun* . 290 : 869 - 873
- Thompson K, Roelofs AJ, Jauhainen M . 2010 ; Activation of gammadelta T Cells by Bisphosphonates . *Advances in experimental medicine and biology* . 658 : 11 - 20
- Thompson K, Rojas-Navea J, Rogers MJ . 2006 ; Alkylamines cause Vgamma9Vdelta2 T-cell activation and proliferation by inhibiting the mevalonate pathway . *Blood* . 107 : 651 - 654
- Toutirais O, Cabillie F, Le Fric G . 2009 ; DNAX accessory molecule-1 (CD226) promotes human hepatocellular carcinoma cell lysis by Vgamma9Vdelta2 T cells . *Eur J Immunol* . 39 : 1361 - 1368
- Treiner E, Duban L, Moura IC . 2005 ; Mucosal-associated invariant T (MAIT) cells: an evolutionarily conserved T cell subset . *Microbes and infection/Institut Pasteur* . 7 : 552 - 559
- Tsan MF, Gao B . 2009 ; Heat shock proteins and immune system . *Journal of leukocyte biology* . 85 : 905 - 910
- Vantourout P, Martinez LO, Fabre A . 2008 ; Ecto-F1-ATPase and MHC-class I close association on cell membranes . *Mol Immunol* . 45 : 485 - 492
- Vantourout P, Mookerjee-Basu J, Rolland C . 2009 ; Specific requirements for Vgamma9Vdelta2 T cell stimulation by a natural adenylylated phosphoantigen . *J Immunol* . 183 : 3848 - 3857
- Vermijlen D, Brouwer M, Donner C . 2010 ; Human cytomegalovirus elicits fetal gammadelta T cell responses in utero . *J Exp Med* . 207 : 807 - 821
- Vidovic D, Dembic Z . 1991 ; Qa-1 restricted gamma delta T cells can help B cells . *Curr Top Microbiol Immunol* . 173 : 239 - 244
- Vidovic D, Roglic M, McKune K . 1989 ; Qa-1 restricted recognition of foreign antigen by a gamma delta T-cell hybridoma . *Nature* . 340 : 646 - 650
- Vincent MS, Gumperz JE, Brenner MB . 2003 ; Understanding the function of CD1-restricted T cells . *Nature immunology* . 4 : 517 - 523
- von Haller PD, Donohoe S, Goodlett DR . 2001 ; Mass spectrometric characterization of proteins extracted from Jurkat T cell detergent-resistant membrane domains . *Proteomics* . 1 : 1010 - 1021

- Wadia P, Atre N, Pradhan T. 2005 ; Heat shock protein induced TCR gammadelta gene rearrangements in patients with oral cancer . *Oral Oncol* . 41 : 175 - 182
- Wang F, Xia XF, Sui SF. 2002 ; Human apolipoprotein H may have various orientations when attached to lipid layer . *Biophysical journal* . 83 : 985 - 993
- Wang H, Fang Z, Morita CT. 2010 ; Vgamma2Vdelta2 T Cell Receptor recognition of prenyl pyrophosphates is dependent on all CDRs . *J Immunol* . 184 : 6209 - 6222
- Wang SX, Sun YT, Sui SF. 2000 ; Membrane-induced conformational change in human apolipoprotein H . *The Biochemical journal* . 348 : (Pt 1) 103 - 106
- Waters WR, Palmer MV, Thacker TC. 2006 ; Antigen-specific proliferation and activation of peripheral blood mononuclear cells from *Mycobacterium bovis*-infected reindeer . *Veterinary immunology and immunopathology* . 111 : 263 - 277
- Wei H, Huang D, Lai X. 2008 ; Definition of APC presentation of phosphoantigen (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate to Vgamma2Vdelta2 TCR . *J Immunol* . 181 : 4798 - 4806
- Welsh MD, Kennedy HE, Smyth AJ. 2002 ; Responses of bovine WC1(+) gammadelta T cells to protein and nonprotein antigens of *Mycobacterium bovis* . *Infect Immun* . 70 : 6114 - 6120
- Wingren C, Crowley MP, Degano M. 2000 ; Crystal structure of a gammadelta T cell receptor ligand T22: a truncated MHC-like fold . *Science* . 287 : 310 - 314
- Wu J, Groh V, Spies T. 2002 ; T cell antigen receptor engagement and specificity in the recognition of stress-inducible MHC class I-related chains by human epithelial gamma delta T cells . *J Immunol* . 169 : 1236 - 1240
- Xiong N, Kang C, Raulat DH. 2004 ; Positive selection of dendritic epidermal gammadelta T cell precursors in the fetal thymus determines expression of skin-homing receptors . *Immunity* . 21 : 121 - 131
- Xu C, Zhang H, Hu H. 2007 ; gammadelta T cells recognize tumor cells via CDR3delta region . *Mol Immunol* . 44 : 302 - 310
- Yamashita S, Tanaka Y, Harazaki M. 2003 ; Recognition mechanism of non-peptide antigens by human {gamma}{delta} T cells . *Int Immunol* . 15 : 1301 - 1307
- Yegutkin GG. 2008 ; Nucleotide- and nucleoside-converting ectoenzymes: Important modulators of purinergic signalling cascade . *Biochimica et biophysica acta* . 1783 : 673 - 694
- Yegutkin GG, Hentinen T, Jalkanen S. 2001 ; Extracellular ATP formation on vascular endothelial cells is mediated by ecto-nucleotide kinase activities via phosphotransfer reactions . *Faseb J* . 15 : 251 - 260
- Yonally SK, Capaldi RA. 2006 ; The F(1)F(0) ATP synthase and mitochondrial respiratory chain complexes are present on the plasma membrane of an osteosarcoma cell line: An immunocytochemical study . *Mitochondrion* . 6 : 305 - 314
- Yu C, Alterman M, Dobrowsky RT. 2005 ; Ceramide displaces cholesterol from lipid rafts and decreases the association of the cholesterol binding protein caveolin-1 . *J Lipid Res* . 46 : 1678 - 1691
- Zajonc DM, Savage PB, Bendelac A. 2008 ; Crystal structures of mouse CD1d-iGb3 complex and its cognate Valpha14 T cell receptor suggest a model for dual recognition of foreign and self glycolipids . *Journal of molecular biology* . 377 : 1104 - 1116
- Zanni MP, von Greyerz S, Schnyder B. 1998 ; HLA-restricted, processing- and metabolism-independent pathway of drug recognition by human alpha beta T lymphocytes . *J Clin Invest* . 102 : 1591 - 1598
- Zgani I, Menut C, Seman M. 2004 ; Synthesis of prenyl pyrophosphonates as new potent phosphoantigens inducing selective activation of human Vgamma9Vdelta2 T lymphocytes . *J Med Chem* . 47 : 4600 - 4612
- Zhang H, Hu H, Jiang X. 2005 ; Membrane HSP70: the molecule triggering gammadelta T cells in the early stage of tumorigenesis . *Immunol Invest* . 34 : 453 - 468
- Zhang L, Jin N, Nakayama M. 2010 ; Gamma delta T cell receptors confer autonomous responsiveness to the insulin-peptide B:9-23 . *Journal of autoimmunity* .
- Zhang Y, Song Y, Yin F. 2006 ; Structural studies of Vgamma2Vdelta2 T cell phosphoantigens . *Chem Biol* . 13 : 985 - 992
- Zhao J, Huang J, Chen H. 2006 ; Vdelta1 T cell receptor binds specifically to MHC I chain related A: molecular and biochemical evidences . *Biochem Biophys Res Commun* . 339 : 232 - 240
- Zhao Y, Yokota K, Ayada K. 2007 ; *Helicobacter pylori* heat-shock protein 60 induces interleukin-8 via a Toll-like receptor (TLR)2 and mitogen-activated protein (MAP) kinase pathway in human monocytes . *Journal of medical microbiology* . 56 : 154 - 164