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The Small Chill: mild hypothermia for cardioprotection?

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1 <u>1- Introduction</u>

Therapeutic whole body hypothermia has been considered for centuries.¹ For 2 3 example, the Russian approach to resuscitation of a patient in cardiac arrest in 1803 4 consisted of covering him with snow and then hoping for the resumption of a spontaneous circulation.¹ A century and a half later, the beneficial effect of therapeutic hypothermia 5 during cardiac surgery was proven in canine models.^{2, 3} Outside of the operating room, 6 7 hypothermia has been demonstrated to protect the brain following cardiac arrest in both animal models and humans.^{4, 5} The American Heart Association and the European 8 9 Resuscitation Council both recommend the use of hypothermia in comatose patients resuscitated from cardiac arrest to improve the subsequent neurological recovery.^{6,7} 10

11 Beside hypothermia's beneficial effect during cardiac surgery or following cardiac 12 arrest, it has also been clearly demonstrated that even very mild hypothermia can greatly 13 increase the heart's tolerance to myocardial ischemia resulting in decreases of infarct size in 14 animal models of coronary artery occlusion. Mild hypothermia (body temperature down to 15 32 °C) has the advantage that the heart continues to pump normally and no extracorporeal 16 support is needed. However, despite the encouraging results in animals, clinical trials of 17 mild hypothermia in patients being revascularized for acute myocardial infarction have yielded surprisingly disappointing effects on infarct size.⁸⁻¹⁰ One goal of the present review 18 19 is to consider why past trials have failed and propose what might be done to make cooling-20 induced cardioprotection more effective. We will review the literature to see if an optimal 21 target temperature and cooling method can be recommended. Finally, the mechanism of 22 mild cooling's protection will be revisited. While deep hypothermia stops the heart and 23 clearly preserves ATP during ischemia, cooling to 34°C has only a modest effect on ATP depletion during ischemia¹¹ but it is as protective to the rabbit heart as is ischemic 24 preconditioning. ¹² That has caused some to suspect that the mechanism of the 25

cardioprotective effect of mild hypothermia might be more complex than energy
 preservation alone.¹³

28

29 **<u>2. Physiological effects of mild hypothermia</u>**

Hypothermia has distinct physiological effects. Hypothermia can be classified as 30 31 mild (32-35°C), moderate (28-32°C), severe (20-28°C) or profound (<20°C). Mild 32 hypothermia is the only one to be used for a whole-body therapeutic purpose as lower 33 temperatures are increasingly life-threatening. Below 28°C, the heart spontaneously 34 fibrillates in most mammalian species. Conversely, mild whole-body hypothermia is 35 remarkably well tolerated by both animals and humans. The physiological effects of mild 36 hypothermia include a decrease in heart rate and a subsequent fall in cardiac output (-7% for each $^{\circ}C$),^{3, 14} while stroke volume and mean arterial pressure remain unchanged. Cooling of 37 38 the skin provokes an increase in systemic vascular resistance. With respect to regional hemodynamics, cerebral blood flow and intracranial pressure decrease,^{3, 14} whereas renal 39 blood flow and subsequent diuresis are increased.¹⁵ Mild hypothermia also attenuates the 40 cerebral metabolic rate (-6 to -7% for each $^{\circ}$ C).¹⁶ Intestinal motility is also depressed by 41 42 mild hypothermia. Blood pH changes by +0.016 unit for each decrease of 1°C, and serum potassium decreases as a result of its enhanced cellular uptake.¹⁷ Hypothermia-induced 43 44 hypokalemia should be corrected with caution during hypothermia since raising serum K⁺ 45 usually leads to hyperkalemia during subsequent rewarming. Other adverse effects include increased infection risk, a mild coagulopathy¹⁷ and hyperglycemia caused by decreased 46 plasma insulin levels.¹⁸ 47

48

49 **<u>3. The infarct limiting property of cooling: experimental** *in vivo* **investigations**</u>

50 3.1. Proof of concept of the myocardial infarct-limiting property of hypothermia

Heart temperature is known to be a major determinant of infarct size in animal models of acute myocardial ischemia.¹⁹⁻²¹ As an example, infarct size resulting from 30 min of regional myocardial ischemia in *in situ* rabbit hearts decreases by 8% of the risk zone for each °C decrement.²¹ The infarct-limiting property of mild hypothermia has been confirmed in several species including dogs,¹⁹ rabbits,^{12, 21-25} pigs^{20, 26-28} and rats.²⁹

56 *3.2.* What is the optimal target temperature?

Importantly, there is no threshold temperature for the cardioprotective effect of 57 hypothermia.¹⁹⁻²¹ In fact, warming the heart above normothermia extends infarct size by the 58 same degree as cooling reduces it.²¹ For example, Hamamoto et al.³⁰ demonstrated in sheep 59 60 submitted to 60 min of coronary artery occlusion that infarct size was progressively reduced 61 when cardiac temperature during ischemia and reperfusion was lowered by 1°C decrements from 39.5 to 35.5°C. As shown in Table I, temperature decrement during ischemia within 62 63 the mild hypothermic range has demonstrated a powerful anti-infarct effect in many studies. Figure 1 illustrates cooling to only 35°C at the onset of ischemia elicits ~70% decrease in 64 infarct size following 30 min of regional ischemia in the open-chest rabbit.¹² And protection 65 66 is still realized if the onset of cooling is delayed for 10 min. Cooling the heart to 32°C at the onset of ischemia essentially prevented infarction, and a protective effect was still evident 67 when cooling was initiated as long as 20 min after the onset of ischemia and, therefore, 10 68 min before reperfusion.¹² Furthermore, even when the ischemic period was extended to 60-69 70 120 min and cooling to 30-32°C was initiated 20-30 min after the onset of ischemia, the 71 infarction process appeared to be halted when cooling was started and significant myocardial salvage was realized.^{12, 22} And the heart continued to beat strongly and support 72 the rabbit's circulation. 73

74 *3.3.* What is the window of protection?

Cooling the heart during the ischemic period reduces infarct size. Very early studies
 from the Corday laboratory^{31, 32} clearly showed that if the dog heart were retrogradely

77 perfused with cooled arterial blood through the coronary sinus beginning 30 min after 78 coronary occlusion and persisting for the remaining 2¹/₂-3 hr of the ischemic period, infarct 79 size was decreased by 65-90%. Table 1 reveals that the degree of protection afforded by 80 cooling is not only determined by the depth of cooling but also by its duration during the ischemic period. Miki et al.¹² demonstrated, for example, that the magnitude of infarct size 81 82 reduction decreased when the onset of hypothermia was delayed from the beginning to the 20th min of a 30-min period of ischemia in rabbits. As illustrated in Figure 1, cooling during 83 84 just the last 10 minutes of ischemia still afforded significant cardioprotection when the target 85 temperature was 32°C but not when it was 35°C. Figure 2 merges the results of several 86 studies investigating the effect of hypothermia to 32°C initiated at different times during a 87 30-min period of ischemia in rabbits, again illustrating that cardioprotection is attenuated 88 when the onset of cooling is delayed. This raises a major logistical problem in the clinical 89 setting since these patients with acute myocardial infarction present with ischemia already in 90 progress. To significantly shorten the normothermic ischemic time cooling would have to 91 be accomplished very soon after arrival in the hospital and efforts to effect early cooling 92 must not come at the expense of increasing the time to reperfusion. That schedule has been 93 difficult to implement.

94 Cooling the body at the time of reperfusion or beyond seems to be protective in the central nervous system.^{3, 33} That has led investigators to ask whether mild hypothermia 95 96 might be cardioprotective when instituted only at the onset of the reperfusion phase, i.e., a 97 postconditioning maneuver. Cooling to 10° C at the onset of reoxygenation and glucose 98 resupply of chick cardiomyocytes subjected to simulated ischemia and reperfusion improved 99 cell viability, and initiation of cooling towards the end of the simulated ischemia further diminished cell necrosis.³⁴ These observations strongly suggested that cooling during the 100 101 peri-reperfusion period might be successful as well. Table 2 shows the results of several intact animal studies that have investigated this possibility.^{25, 28, 35-39} Unfortunately, all but 102

one³⁷ failed to see any effect on infarct size and in the one positive study cooling still 103 104 included that last 5 min of a 30 min ischemic period. Either the much lower temperature or 105 the non-mammalian cell type was likely responsible for the protection in the chick cardiomyocyte study.³⁴ Of course, cooling a patient to 10°C is not feasible because that 106 107 would stop the heart. However, it was found that hypothermia during reperfusion prevents 108 microvascular damage and thereby offers some protection against the no-reflow phenomenon.^{24, 35} One might, therefore, speculate that hypothermia at reperfusion protects 109 against vascular alteration and subsequent microvascular obstruction³⁵ but cannot protect the 110 111 cardiomyocytes. This could mean that one should maintain cooling during the first hours of 112 reperfusion even if cooling had been instituted early in the ischemic period. Surprisingly, 113 the question of whether prolonging hypothermia after reperfusion measurably adds to the 114 anti-infarct effect has yet to be addressed. It should be noted that most patients spend 115 approximately 90 minutes in hospital prior to revascularization because of delays associated with admitting, diagnosis, and preparation for intervention.⁴⁰ Whole-body cooling during 116 117 that period could produce significant reduction of the resulting infarct.

118 *3.4. How can myocardium be cooled?*

119 As emphasized earlier, the sooner hypothermia is achieved following the onset of 120 ischemia, the more cardioprotective it would be. Experimentally, some studies have been 121 performed using topical epicardial cooling to quickly lower cardiac temperature, but this 122 method would be difficult to implement in patients. In patients with acute myocardial 123 infarction, the critical parameters determining benefit would be the time when cooling could 124 be started and also the rate at which cooling could be achieved. As explained earlier, the 125 benefit would be expected to be small or absent if the target temperature is only reached far 126 into the reperfusion phase. Obviously, the least invasive strategy for implementing 127 therapeutic hypothermia in patients is external cooling. This can be done using ice packs or 128 with specific medical devices designed to promote heat exchange by a more efficient

contact between the skin and the cooling medium.^{2, 41, 42} Unfortunately the cooling rate 129 afforded by these strategies is rather low, averaging only 1-2°C/h in humans.⁴³ That is 130 131 because the cutaneous microcirculation tends to constrict when cold in an attempt to thermally insulate the body from the environment. Small laboratory animals cool faster with 132 133 skin cooling than humans since their ratio of body mass to surface area is much smaller than 134 in man. The time required to cool a rabbit weighing 2.0-3.0 kg to 32°C is still ~45 min, and this does not significantly limit infarct size after 60 min of ischemia.⁴⁴ While it is unlikely 135 136 that patients with acute myocardial infarction would experience any anti-infarct benefit from 137 conventional external cooling, this strategy is reported to be beneficial following cardiac 138 arrest when even delayed hypothermia improves overall survival and neurological recovery after resuscitation.^{2, 41, 42} 139

140 Other strategies have been proposed to induce therapeutic hypothermia. Some 141 examples include endovascular cooling with intravenous thermodes, infusion of cold 142 intravenous fluids, gastric lavage with cold fluid through a nasogastric catheter and even urinary bladder lavage.⁴² Most of those techniques have been investigated in the clinics for 143 their neuroprotective abilities.⁴² The one that has been investigated for a cardioprotective 144 therapy is endovascular cooling.^{26, 45, 46} In human-sized pigs, this strategy afforded 145 promising results when cooling was initiated early during ischemia.^{26, 45} A clinical trial in 146 147 patients with ST-segment elevation myocardial infarction (STEMI) clearly demonstrated the feasibility of that strategy,⁴⁶ but disappointingly did not show a significant cardioprotective 148 benefit,⁸ probably because of an insufficient cooling rate that resulted in normothermia 149 during most of the ischemic period.⁴⁷ 150

151 *3.5. Ultra-fast cooling*

Accordingly other strategies have been proposed that elicit a much faster rate of cooling which should increase the degree of cardioprotection in the clinical setting. Examples include extracorporeal blood cooling¹² or total liquid ventilation with

temperature-controlled perfluorocarbons.^{25, 48} These techniques can decrease cardiac 155 temperature to 32°C within 3-5 min in rabbits. A pericardial perfusion circuit has also been 156 proposed.⁴⁹ Unfortunately all these strategies would be challenging to implement in patients 157 presenting with myocardial infarction since they are very invasive and would significantly 158 159 delay reperfusion by angioplasty or thrombus extraction. A promising technique could be 160 peritoneal lavage, which, although still invasive, should be easier to institute than any of the 161 above-mentioned methods. As seen in patients with malignant hyperthermia it can cool a patient very quickly.⁵⁰ Whether any of these ultrafast cooling strategies would be beneficial 162 163 in humans with STEMI remains to be investigated.

164

165 <u>4- Mechanism of cooling-induced cardioprotection</u>

166 The mechanism of hypothermia-induced cardioprotection has mostly been 167 investigated in experimental models receiving cold cardioplegia. The temperature of the heart is reduced to very low levels which, among other things, arrests it.^{13, 51} However, 168 169 mechanisms are likely to be quite different for mild hypothermia of an *in vivo* beating heart 170 and an arrested heart exposed to cold cardioplegia. In beating hearts, one could, for example, 171 argue that the cardioprotection might be related to the bradycardia elicited during ischemia. 172 However, this is unlikely because the relationship between infarct size and temperature was unchanged when normothermic heart rate was restored by pacing in hypothermic rabbits.^{21,} 173 ⁵² It is therefore commonly assumed that hypothermia protects the heart, at least in part, 174 175 through reduced cardiac metabolism. This assumption has been amply supported by studies performed during cold cardioplegia ($<20^{\circ}$ C).^{51, 53-60} Most enzymes have a Q₁₀ of about 2, 176 which means the reaction rate doubles for every 10°C increase in temperature. Reducing the 177 178 cardiac temperature by 20°C should therefore decrease metabolism by a factor of 4. Mild hypothermia (>30°C) also decreases the rate of high energy phosphate^{11, 61, 62} and glucose⁶³ 179 utilization as well as lactate accumulation⁶³ but to a lesser extent than deep hypothermia. 180

Deep hypothermia may also alter ion exchange because it inhibits Na^+/Ca^{2+} and 181 Na^+/K^+ sarcoplasmic exchangers, although it paradoxically activates the Na^+/H^+ 182 exchanger.⁶⁴ Interestingly, hibernating hypothermic frogs increase their resistance to 183 hypoxia through a decreased demand for ATP by reduced Na⁺/K⁺-ATPase activity.⁶⁵ An 184 NMR study in isolated newborn rabbit hearts further confirmed that deep cooling (12°C) 185 186 with cold crystalloid cardioplegia limited acidosis and calcium and sodium cellular overload during ischemia/reperfusion.¹³ Deep hypothermia (17°C) also limited mitochondrial calcium 187 overload in Langendorff guinea pig hearts undergoing ischemia.⁶⁶ While deep hypothermia 188 189 tends to increase baseline reactive oxygen species formation during normoxia, it limits the burst following ischemia-reperfusion.⁶⁶ Using electron spin resonance spectroscopy in 190 191 isolated reperfused rat heart investigators have observed reduced free-radical generation at reperfusion following ischemia at 4°C.⁶⁷ 192

193 Deep hypothermia reduces several modulators of the mitochondrial permeability 194 transition pore (mPTP) *i.e.*, ATP depletion, calcium overload, and generation of reactive 195 oxygen species. Mild hypothermia (32°C) inhibited calcium-induced mPTP opening in 196 ventricular samples from rabbit hearts subjected to ischemia alone or to ischemia followed by 10 min of reperfusion.⁴⁸ Suppression of mPTP formation at reperfusion is thought to be 197 the mode of action of ischemic preconditioning⁶⁸ and ischemic postconditioning.⁶⁹ Thus it 198 199 is reasonable to speculate that hypothermia acts, at least in part, through inhibition of MPTP 200 formation. However, the manner of modulation of mPTP opening is probably different than 201 that afforded by pre- and postconditioning since the latter strategies are believed to trigger 202 signal transduction pathways that determine the fate of the previously ischemic myocardium during the first minutes of reperfusion,⁶⁹⁻⁷³ while hypothermia seems to exert its protection 203 during ischemia.⁴⁸ Opening of mPTP at reperfusion only occurs if the heart has been injured 204 205 by a period of prolonged ischemia. The nature of that injury is poorly understood, but Honda and colleagues⁷⁴ referred to it as "priming". Mild hypothermia may act to lessen that
ischemic injury.

208 The protection from mild hypothermia is proportional to the decrease in temperature 209 which argues against any off/on type of mechanism. A direct effect of mild hypothermia on 210 enzyme kinetics seems unlikely to be the protective mechanism since most enzymes are not so temperature-dependent. Hearts function quite well over the entire range of mild 211 212 hypothermia (32-38°C). An intriguing possibility is that mild hypothermia might somehow activate cardioprotective signal transduction pathways. Ning et al.⁷⁵ demonstrated that cold 213 214 cardioplegia (30°C) preserves mitochondrial protein gene expression during hypoxia, including genes coding for HSP70, ANT₁, and β -F₁-ATPase.⁷⁶ They observed that neither 215 216 ATP levels nor anaerobic metabolism are linked to mRNA expression of these latter proteins.⁷⁷ Ning et al.⁷⁷ also showed that moderate hypothermia (30°C) promotes expression 217 218 of proteins involved in cell survival, while it inhibits induction of p53 protein. It is, 219 therefore, reasonable to hypothesize that mild hypothermia triggers its protection through 220 thermal sensors. This hypothesis is supported by studies from Halestrap's group which 221 reported that a short period of perfusion at 26°C in isolated rat hearts induced a 222 preconditioning-like protection which increased protein kinase C-E translocation to the 223 particulate fraction (an index of its activation) and phosphorylated AMP-activated protein kinase (AMPK).⁷⁸ In one recent study in chick cardiomyocytes undergoing simulated 224 ischemia, cooling to 25°C just before reoxygenation protected the cells.³⁴ More importantly 225 226 that protection could be blocked by either a PKC or a nitric oxide synthase (NOS) blocker 227 indicating signal transduction. Protection in the latter study, however, seemed to mimic that 228 of postconditioning rather than that of mild hypothermia since the critical time for cooling 229 was during reoxygenation rather than simulated ischemia, and PKC and NOS are well 230 known components of the signal transduction pathways of pre- and postconditioning. Also

cooling was much more severe. All chemical reactions are at some point temperaturedependent, but the temperature coefficients (Q_{10}) for the various mammalian enzymes vary widely. The possibility that one particular enzyme might be very sensitive to temperature and could serve as a sensor to implement the cardioprotective effect of mild hypothermia through cell signaling is an attractive hypothesis, but of course that enzyme could be very difficult to find.

237 Hypothermia seems to protect against injury during ischemia, while ischemic 238 preconditioning protects against injury during reperfusion. Because of the different mechanisms it is possible to add the two together to produce additive protection.¹² It is 239 currently possible to pharmacologically postcondition a heart^{79, 80} and such an intervention is 240 241 thought to protect by a mechanism identical to that of preconditioning. Thus, it should be 242 possible to combine mild hypothermia during ischemia with a postconditioning agent at 243 reperfusion. The primary impediment to using mild hypothermia clinically has been the 244 logistics of implementing it quickly after the onset of ischemia. However, if signaling 245 pathways turn out to be responsible for mild hypothermia's protection, then pharmacological 246 activation of those pathways would likely be simpler to implement and such an agent could 247 possibly even be given by EMS personnel.

248

249 <u>5- Cooling and cardioprotection: STEMI clinical data</u>

250 Soon after the turn of the 21st century the promising experimental results regarding

251 the cardioprotective effect of cooling inspired several clinical trials testing mild

252 hypothermia's feasibility and safety in STEMI patients.^{46, 81, 82} As shown in Table 3, two

253 large-scale clinical trials (COOL-MI⁸ and ICE-IT¹⁰) were conducted using endovascular

254 cooling. Both studies assessed infarct size using single photon emission computed

tomography (SPECT). The results of these studies have not yet been published in peer-

256 reviewed journals, but they have been summarized in several reviews.^{41, 83, 84}

257 In the COOL-MI study (*COOLing as an adjunctive therapy to percutaneous* 258 intervention in patients with acute Myocardial Infarction), 392 STEMI patients were enrolled within 6 hours following the onset of symptoms.⁸ Patients were assigned to either 259 260 treatment with a percutaneous coronary intervention alone or to cooling with an 261 endovascular cooling device (Reprieve Temperature Therapy System, Radiant Medical, Redwood City, CA, USA) prior to revascularization. The percutaneous coronary 262 263 intervention was accomplished a mean of 18 min after cooling was instituted, with a mean 264 temperature of 35.0°C at the moment of reperfusion. As shown in Table 3, cooling did not 265 provide an overall significant reduction in infarct size, except in a sub-group of patients with 266 anterior myocardial infarction that were cooled to less than 35°C at the time of revascularization.⁸ The overall negative result of that study might have been biased since the 267 268 average door-to-balloon time in cooled patients was 18 minutes longer than in control 269 patients (110 vs 92 min).

270 In the ICE-IT study (Intravascular Cooling Adjunctive to Primary Coronary 271 Intervention), the design was similar to COOL-MI with the inclusion of 228 patients 272 randomized to either normothermic revascularization or to revascularization with mild 273 hypothermia using another endovascular device (Innercool by Celsius Control System, San Diego, CA, USA).¹⁰ Hypothermia again did not provide a significant benefit regarding 274 275 infarct size in the whole population (Table 3). A trend for a benefit was observed in the sub-276 group with anterior infarction with a body temperature of less than 35°C at the time of 277 reperfusion. Interestingly, a subanalysis of the 6 sites with the best protocol compliance out 278 of the 22 participating sites also demonstrated a significant reduction in infarct size with 279 cooling compared to conventional revascularization (-44%). These data suggest, as 280 emphasized above, that the overall negative result of the COOL-MI and ICE-IT trials was 281 related to a delay in institution of hypothermia and/or to an insufficient cooling rate which 282 resulted in little shortening of the normothermic ischemic period. A further study (COOL-

MI II) was accordingly recently conducted with an earlier, deeper and faster cooling protocol, with cooling started in the emergency room rather than in the catheterization laboratory. The results of this last trial are not yet available.

It has conversely been suggested that hyperthermia may worsen the clinical outcome in patients with myocardial infarction.⁸⁵ In experimental conditions, Chien et al.²¹ found not only that hypothermia could diminish infarct size but that hyperthermia could increase it. One would, therefore, speculate that therapeutic hypothermia would also be indicated to reverse the adverse effects of hyperthermia. A general relationship between clinical outcome and body temperature may well exist.

292 *Cardiac Resuscitation*

293 Another setting in which the protective effect of cooling would be relevant is cardiopulmonary resuscitation. Cooling after the heart is restarted can indeed protect the 294 central nervous system, as has been previously shown,⁵but it may also protect the heart since 295 myocardial ischemia is a common cause of cardiac arrest.^{41, 84} Theoretically, the cardiac 296 297 benefit would even be greater for those patients than for "typical" STEMI as the time before 298 revascularization is prolonged by the resuscitation time. There is controversy as to whether resuscitated patients should be immediately submitted to a coronary intervention^{41, 84} since 299 300 the use of hypothermia is basically recommended for comatose survivors after out-ofhospital cardiac arrest with ventricular fibrillation.^{6,7} Importantly, the combination of 301 302 cooling and percutaneous intervention is at least feasible and should be safe in those patients.41,84 303

304

305 <u>6. Conclusion</u>

In conclusion, cooling the myocardium with whole body mild hypothermia is a very
potent cardioprotective maneuver, at least in the experimental setting. The benefit depends
upon the rapidity with which cooling is instituted and by how much it shortens the

309	normothermic ischemic time. To afford a clinical benefit, a cooling strategy should
310	accordingly be designed to achieve the target temperature well before the time of
311	revascularization. The depth of cooling is also important. Temperatures in the range of 32-
312	35°C are considered safe. Yet these temperatures exert a powerful anti-infarct effect in
313	animal studies. Finally, more studies of mild hypothermia are needed to determine its actual
314	mechanism, as compared to deep hypothermia that acts through energy preservation.

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323

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Legend of Figures

Figure 1

Infarct sizes (expressed as % of the area at risk) in different groups of anesthetized rabbits submitted to 30 min of coronary artery occlusion and 3 h of reperfusion. In the different groups, rabbits were either subjected to a normothermic protocol (Control group) or to extracorporeal blood cooling to 35°C or 32°C from the onset (0') or from the 10th or the 20th minute of ischemia (0', 10' and 20' ischemia). Open circles represent the individual infarct size of each animal and closed circles represent the mean±SEM of each group. *Data adapted from Miki et al.* ¹² * p<0.05 vs Control.

Figure 2

Infarct sizes (expressed as % of the area at risk) obtained from several studies in anesthetized rabbits subjected to 30 min of coronary artery occlusion and cooled to 32°C starting at different times after initiation of ischemia. Closed circles represent the mean±SEM of the cooled groups for each study.

Numbers next to the data points are reference citations.









Table 1: Summary of *in vivo* experimental studies investigating the infarct-limiting effect of myocardial cooling during ischemia, i.e., with cooling initiated before ischemia or at least 5 min before the end of coronary artery occlusion

Species	Ref.	Cooling procedure	Duration of CAO (min) / CAR (h)	Target heart temperature	Time of cooling*	IS with Cooling vs Control groups (% decrease)
Rabbit	36	Topical epicardial cooling	30 min / 3 h	~33°C	$10 \min \text{CAO} \rightarrow 15$ $\min \text{CAR}$	23±4 vs 44±4 (-48%)
Rabbit	22	Topical epicardial cooling	120 min / 3 h	~30°C	$\begin{array}{c} 30 \text{ min CAO} \rightarrow 15 \\ \text{min CAR} \end{array}$	59±3 vs 72±3 (-18%)
Rabbit	49	Closed pericardioperfusion circuit	30 min / 3 h	~34°C	$-30 \rightarrow 25 \min \text{CAO}$	18±3 vs 35±6 (-49%)
Rabbit	24	Topical epicardial cooling	30 min / 3 h	~32°C	$\begin{array}{c} 20 \text{ min CAO} \rightarrow 120 \\ \text{min CAR} \end{array}$	27±4 vs 51±5 (-47%)
					$0 \rightarrow 30 \min \text{CAO}$	11±3 vs 37±3 (-70%)
		Blood cooling through	20	~35°C	$10 \rightarrow 30 \min \text{CAO}$	18±3 vs 37±3 (-51%)
D-11:4	12				$20 \rightarrow 30 \min \text{CAO}$	34±2 vs 37±3 (NS)
Kabbit		heat exchanger	30 min / 3 n		$0 \rightarrow 30 \min \text{CAO}$	4±1 vs 37±3 (-89%)
				~32°C	$10 \rightarrow 30 \min \text{CAO}$	8±1 vs 37±3 (-78%)
Dabbit					$20 \rightarrow 30 \min \text{CAO}$	23±2 vs 37±3 (-38%)
Rabbit	25	Total liquid ventilation	30 min / 3 h	~32°C	$0 \rightarrow 30 \min \text{CAO}$	4±1 vs 38±1 (-89%)
D 111	48		20 : (72)	2200	$5 \rightarrow 30 \min \text{CAO}$	4±1 vs 39±2% (-90%)
Rabbit		Total liquid ventilation	30 min / 72 h	~32°C	$15 \rightarrow 30 \min \text{CAO}$	11±5 vs 39±2% (-72%)
					Before CAO→ 180 min CAR	30±5 vs 59±1% (-48%)
Rabbit	37	Surface cooling (water blankets)	30 min / 3 h	~37.0°C	$\begin{array}{c} 0 \text{ min CAO} \rightarrow 180 \\ \text{min CAR} \end{array}$	33±5 vs 59±1% (-43%)
					$15 \min \text{CAO} \rightarrow 180$ $\min \text{CAR}$	42±1 vs 59±1% (-28%)
Rabbit	52	Topical epicardial cooling	30 min / 3 h	~35°C	-20 min before CAO → 15 min CAR	16±3 vs 46±4 (-65%)
		Surface cooling			$\begin{array}{cccccccccccccccccccccccccccccccccccc$	78±10 vs 82±7 (NS)
Rabbit Rabbit Rabbit Pig	44	Total liquid ventilation	60 min / 4 h	~32°C	$5 \rightarrow 30 \min \text{CAO}$	45±18 vs 82±7 (-45%)
		Total liquid ventilation				58±5 vs 82±7 (-29%)
Pig	26	Endovascular cooling	60 min / 3 h	~34°C	$20 \min \text{CAO} \\ \rightarrow 15 \min \text{CAR}$	9±6 vs 45±8 (-80%)
Pig	27	Topical epicardial cooling	40 min / 3 h	~29°C	$0 \rightarrow 40 \min \text{CAO}$	25±2 vs 62±5 (-60%)
Pig	28	Intracoronary cold saline infusion	60 min / 3 h	~33°C	$\begin{array}{l} 15 \text{ min CAO} \\ \rightarrow 15 \text{ min CAR} \end{array}$	9±2 vs 36±4 (-75%)
Pig	35	I.V. cold saline + endovascular cooling	40 min / ~4.3h	~33°C	$\begin{array}{c} 25 \text{ min CAO} \\ \rightarrow 25 \text{ min CAR} \end{array}$	46±8 vs 75±5 (-39%)
	30		60 min / 3 h	~38.5°C	0 min CA0	63±2 vs 72±3 (-12%)
Sheen		Surface cooling (ice bags)		~37.5°C	\rightarrow 180 min CAR	49±1 vs 72±3 (-31%)
Sneep				~36.5°C	Control Group at	39±1 vs 72±3 (-46%)
				~35.5°C	39.5°C	22±2 vs 72±3 (-70%)
Dog	32	Hypothermic retroperfusion of autologous blood	210 min / 3 h	~28-30°C	$30 \rightarrow 210 \min \text{CAO}$	6±3 vs 24±7† (-75%)

CAO, coronary artery occlusion; CAR, coronary artery reperfusion; IS, infarct size (expressed as % of area at risk);

ref, reference number

*represents the time of the application of the cooling strategy and not the actual time at which the target temperature was reached. In several studies a delay was inevitable between the onset of the cooling protocol and the time of achievement of the target temperature (e.g., with low rate cooling strategy such as surface cooling). † the Control value corresponds to infarct sizes observed with normothermic retroperfusion. **Table II:** Summary of *in vivo* experimental studies investigating the infarct-limiting effect of myocardial cooling during the reperfusion phase, i.e., with cooling started only 5 min before reperfusion or later yielding normothermic ischemia and hypothermic reperfusion

Species	Ref.	Cooling procedure	Duration of CAO (min) / CAR (h)	Target heart temperature	Time of cooling*	IS with Cooling vs Control groups (% decrease)
Rabbit	36	Topical epicardial cooling	30 min / 3 h	~33°C	$25 \text{ min CAO} \rightarrow$ 15 min CAR	43±4 vs 44±4 (NS)
Rabbit	25	Total liquid ventilation	30 min / 3 h	~32°C	$\begin{array}{c} 25 \text{ min CAO} \rightarrow \\ 30 \text{ min CAR} \end{array}$	35±4 vs 38±1 (NS)
Rabbit	37	Surface cooling (water blankets)	30 min / 3 h	~37.0°C -	25 min CAO → 180 min CAR	44±2 vs 59±1% (-25%)
					30 min CAO → 180 min CAR	51±2 vs 59±1% (NS)
Pig	38	Regional blood cooling through heat exchanger	45 min / 3 h	~33°C	$\begin{array}{l} 43 \text{ min CAO} \rightarrow \\ 120 \text{ min CAR} \end{array}$	71±8 vs 68±1 (NS)
Pig	28	Intracoronary cold saline infusion	60 min / 3 h	~33°C	0 min CAR→ 30 min CAR	33±2 vs 45±5 (NS)
Pig	35	Intravenous infusion of cold saline + endovascular cooling	40 min / ~4.3h	~33°C	$\begin{array}{l} 0 \text{ min CAR} \rightarrow \\ 30 \text{ min CAR} \end{array}$	80±6 vs 75±5 (NS)

See Table 1 for legend.

Table III: Infarct size assessed by single photon emission computed tomography in patients presenting with ST-segment elevation myocardial infarction and treated by revascularization alone or in conjunction with endovascular cooling in the COOL-MI⁸ and the ICE-IT¹⁰ studies. Data were obtained from ref⁸³, ⁴¹ and ⁸⁴.

	Infarct size (% left ventricle)		р
	Cooled group	Normothermic group	<u>P</u> value
COOL-MI (n=392 patients)			
Overall population	13.8%	14.1%	0.86
Subgroup with anterior STEMI cooled to <35° at the time of revascularization	9.3%	18.2%	0.05
ICE-IT (n=228 patients)			
Overall population	10.2%	13.2%	0.14
Subgroup with anterior STEMI cooled to <35° at the time of revascularization	12.9%	22.7%	0.09

STEMI, ST-segment elevation myocardial infarction