



HAL
open science

Intracellular Pharmacokinetics of Antiretroviral Drugs in HIV-Infected Patients, and their Correlation with Drug Action

Caroline Bazzoli, Vincent Jullien, Clotilde Le Tiec, Elisabeth Rey, France Mentré, Anne-Marie Taburet

► **To cite this version:**

Caroline Bazzoli, Vincent Jullien, Clotilde Le Tiec, Elisabeth Rey, France Mentré, et al.. Intracellular Pharmacokinetics of Antiretroviral Drugs in HIV-Infected Patients, and their Correlation with Drug Action. *Clinical Pharmacokinetics*, 2010, 49 (1), pp.17-45. 10.2165/11318110-000000000-00000 . inserm-00461125

HAL Id: inserm-00461125

<https://inserm.hal.science/inserm-00461125v1>

Submitted on 3 Mar 2010

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29

Intracellular pharmacokinetics of antiretroviral drugs in HIV-infected patients and correlation with drug action

Caroline Bazzoli¹, Vincent Jullien², Clotilde Le Tiec³, Elisabeth Rey²,
France Mentré¹, Anne-Marie Taburet³

¹ INSERM, U738, Paris, France; Université Paris Diderot, UFR de Médecine, Paris, France.

² Université Paris Descartes ; AP-HP ; Groupe Hospitalier Cochin – Saint-Vincent de Paul ; Service de Pharmacologie Clinique ; Inserm U663

³ Clinical Pharmacy Department, Hôpital Bicêtre, Assistance Publique hôpitaux de Paris (AP-HP), France.

Running title: Intracellular pharmacokinetics of antiretrovirals

27 pages of text

9547 words for text only

2 Figures (attached file)

4 Tables

1 **Acknowledgements**

2 The authors are no conflicts of interest that are directly relevant to the content of this
3 review. No sources of funding were used to assiste the preparation of this review.

4

5 **Correspondence**

6 Pr France Mentré

7 UMR 738 INSERM - Université Paris Diderot

8 16 rue Henri Huchard

9 75018 Paris, France

10 tel: 33 (0) 1 57 27 75 34

11 fax: 33 (0) 1 57 27 75 21

12 email : france.mentre@inserm.fr

13

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32

Table of Contents

ABSTRACT	5
<u>1 INTRODUCTION.....</u>	<u>7</u>
<u>2 CLINICAL PHARMACOKINETICS OF ARVS.....</u>	<u>8</u>
2.1 ENTRY INHIBITORS	8
2.1.1 CCR5 INHIBITORS	8
2.1.2 ENFUVIRTIDE.....	9
2.2 NUCLEOSIDE AND NUCLEOTIDE ANALOG INHIBITORS OF REVERSE TRANSCRIPTASE	9
2.3 NON NUCLEOSIDE ANALOG INHIBITORS OF REVERSE TRANSCRIPTASE.....	10
2.4 INTEGRASE INHIBITORS	10
2.5 PROTEASE INHIBITORS	10
<u>3 METHODOLOGICAL CONSIDERATIONS</u>	<u>11</u>
3.1 CELL COLLECTION	11
3.2 ESTIMATION OF CELL NUMBER	12
3.3 ANALYTICAL METHODS FOR INTRACELLULAR ASSAYS	12
3.3.1 NUCLEOSIDE ANALOG INHIBITORS OF REVERSE TRANSCRIPTASE	13
3.3.2 NON NUCLEOSIDE ANALOGS AND PROTEASE INHIBITORS	15
<u>4 MECHANISMS INFLUENCING INTRACELLULAR ACCUMULATION</u>	<u>16</u>
4.1 GENERAL PRINCIPLES	16
4.2 NUCLEOSIDE AND NUCLEOTIDE ANALOG INHIBITORS OF REVERSE TRANSCRIPTASE	17
4.3 NON NUCLEOSIDE ANALOG INHIBITORS OF REVERSE TRANSCRIPTASE.....	18
4.4 PROTEASE INHIBITORS	18
4.5 IMPORTANCE OF GENETIC POLYMORPHISM.....	19
<u>5 CLINICAL STUDIES WITH INTRACELLULAR CONCENTRATIONS</u>	<u>20</u>
5.1 RELATIONSHIP BETWEEN INTRACELLULAR AND PLASMA PHARMACOKINETICS	20
5.2 DRUG-DRUG INTERACTIONS AT THE INTRACELLULAR LEVEL	21
5.3 RELATIONSHIP BETWEEN INTRACELLULAR CONCENTRATIONS AND EFFICACY.....	26
5.4 RELATIONSHIP BETWEEN INTRACELLULAR CONCENTRATIONS AND TOXICITY	30
<u>6 CONCLUSION</u>	<u>33</u>

1 **Figure captions**

2

3 **Fig. 1.** Host-cell-mediated sequential enzymatic phosphorylation steps required for
4 activating the nucleotide- and nucleoside-analogue reverse-transcriptase inhibitors
5 (NRTIs) to the triphosphate moiety (Reproduced from Anderson et al.^[17]). ABV =
6 abacavir; AMPD = adenosine monophosphate deaminase; AMPK = adenosine
7 monophosphate kinase (adenylate kinase); APT = adenosine phosphotransferase;
8 CBV = carbovir; dCK = deoxycytidine kinase; dCMPK = deoxycytidine
9 monophosphate kinase; ddA = 2',3'-dideoxyadenosine; ddl, = didanosine; DP, =
10 diphosphate; d4T = stavudine; FTC = emtricitabine; gK = guanylate kinase; MP
11 monophosphate; PMPA = tenofovir (PMPA DP is a triphosphate analogue); TFV =
12 tenofovir; TP = triphosphate; ZDV = zidovudine; 3TC = lamivudine; 5'NDPK = 5'
13 nucleoside diphosphate kinase; 5'NT = 5' nucleotidase.

14

15 **Fig. 2.** Schematic representation of uptake and efflux transporters that may influence
16 intracellular concentrations of antiretroviral drugs in peripheral blood cells.
17 Transporters are named by gene and proteins (Adapted from Ford et al.^[53] and
18 updated^[55-57]). OCT = Organic Cation Transporters, hCNT = Concentrative
19 Nucleoside Transporter, ENT = Equilibrative Nucleoside Transporter, P-gp = P-
20 glycoprotein, MRP = Multidrug Resistance Protein, BCRP = Breast Cancer
21 Resistance protein.

22

23

24

25

26

27

1 **Abstract**

2 In patients infected by human immunodeficiency virus (HIV), the efficacy of highly
3 active antiretroviral therapy (HAART) through the blockade of different steps of this
4 retrovirus life-cycle is now well established. As HIV is a retrovirus which replicates
5 within the cells of the immune system, intracellular drug concentrations are important
6 to determine antiretroviral efficacy and toxicity. Indeed, nucleoside reverse
7 transcriptase inhibitors (NRTI), non nucleoside reverse transcriptase inhibitors
8 (NNRTI), newly available integrase inhibitors and protease inhibitors (PI) act on
9 intracellular targets. NRTIs are prodrugs that require intracellular anabolic
10 phosphorylation to be converted into their active form: the triphosphorylated drug
11 metabolite (NRTI-TP), half-life of which being longer than plasma half-life of the
12 parent compound for most. Activity of intracellular kinases, expression of uptake
13 transporters which may be dependent upon cell functionality or their activation state
14 may greatly influence intracellular concentrations of NRTI-TP. In contrast, NNRTIs as
15 well as PIs are not prodrugs and exert their activity by inhibiting directly enzyme
16 targets. All PIs, are substrates of CYP3A, which explains most of them display poor
17 pharmacokinetic properties with intensive pre-systemic first pass metabolism and
18 short elimination half-lives. There are evidences that intracellular concentration of PIs
19 depends on P-gp and/or other efflux transporters activity, which is modulated by
20 genetic polymorphism and co-administration of drugs with inhibiting or inducing
21 properties. Assaying adequately the intracellular concentrations of antiviral (ARVs)
22 drugs is still a major technical challenge, together with the isolation and the counting
23 of peripheral blood mononuclear cells (PBMCs). Furthermore, intracellular drug could
24 be bound to cell membranes or proteins; the amount of intracellular ARV available for
25 antiretroviral effectiveness is never measured which is a limitation of all published
26 studies. In this review, we summarized the results of thirty-one articles that provided
27 results of intracellular concentrations of ARVs in HIV-infected patients. Most studies
28 also measured plasma concentrations but few of them studied the relationship
29 between plasma and intracellular concentrations. For NRTIs, most studies could not
30 established significant relationship between plasma and triphosphate concentrations.
31 Only eight published studies reported an analysis of the relationships between
32 intracellular concentrations and virological or immunological efficacy of antiretroviral
33 drugs in HIV patients. In prospective studies well designed and with a reasonable

1 number of patients, a significant correlation between virological efficacy and
2 intracellular concentrations of NRTIs was found with no influence of plasma
3 concentration. For PIs, the only prospectively design trial on lopinavir found both the
4 influence of trough plasma and intracellular concentrations. ARVs are known to
5 produce important adverse effects through their interferences with cellular
6 endogenous processes. The relationship between intracellular concentrations of
7 ARVs and their related toxicity were investigated in only four articles. For zidovudine,
8 the relative strength of the association between haemoglobin decrease and plasma
9 zidovudine compared to intracellular zidovudine-triphosphate is still unknown.
10 Similarly, for efavirenz and neuropsychological disorder methodological differences
11 penalize the comparison between studies. In conclusion, intracellular concentrations
12 of ARVs play a major role in their efficacy and toxicity and are influenced by
13 numerous factors. However the number of published clinical studies in that area is
14 limited; most studies were small and not always adequately designed. In addition,
15 standardization of assays and PBMC counts are warranted. Larger and prospectively
16 designed clinical studies are needed to further investigate the links between
17 intracellular concentrations of ARVs and clinical endpoints.

18
19
20
21

1 Introduction

2 Human Immunodeficiency Virus (HIV) is a retrovirus, which replicates within the cells
3 of the immune system. The efficacy of highly active antiretroviral therapy (HAART), is
4 now well established, and has provided extraordinary benefits to many patients with
5 HIV infection.^[1] The morbidity and mortality related to HIV infection have dramatically
6 decreased in countries in which HAART has been available, turning HIV infection into
7 a chronic manageable disease.^[2] Life-long antiretroviral treatment seems necessary,
8 as viral replication and loss of CD4 cells resume when HAART is interrupted. HAART
9 regimens have shown some limitations, the major one being the failure to eradicate
10 HIV even after several years of therapy. One of the reasons is that despite potent
11 antiretroviral (ARV) treatment, compartments of replication-competent virus persist,
12 suggesting that ARVs do not reach all the infected cells: however, there are no data
13 to support this theoretical assumption. This article will focus on pharmacologic
14 principles that govern intracellular concentrations of antiretroviral drugs and on
15 clinical studies which aimed at assessing whether intracellular concentrations of
16 ARVs could be a useful parameter to predict efficacy or toxicity of antiretroviral drug
17 regimen.

18 The different steps of HIV replication are now well identified and understood. A
19 number of antiretroviral drugs are now available and are grouped in five
20 pharmacologic classes according to their mechanism of action. These drugs target
21 essential receptors or enzymes at different steps of the life cycle of the virus and will
22 block the production of infectious retroviral particles from the cell.^[3] However virus
23 eradication cannot be achieved with the available treatments because of the pool of
24 latently infected CD4 cells.^[4]

25 HAART is the standard of care to avoid selection of viral mutations. Selection of
26 drugs for treatment naïve patients and experienced patients take into account the risk
27 benefit ratio and the viral genotype. Current guidelines recommend in
28 treatment-naïve patients a combination of a ritonavir boosted PI or a NNRTI plus two
29 NRTIs and in treatment experienced patients a combination of at least two active
30 ARV drugs from different classes based on viral genotype.^[5-7]

31 Besides entry inhibitors which act on receptors located on cell surface, most ARV
32 drugs inhibit viral replication inside the cell, therefore intracellular concentration
33 should be a reliable parameter to consider when relating pharmacokinetics and

1 efficacy. Results from several *in vitro* studies also exist in this area. But, because of
2 the difficulties of extrapolating the results from *in vitro* to *in vivo* studies, in this paper,
3 we focus only on *in vivo* studies.

4 This article summarizes clinical trials where intracellular concentrations were
5 measured and related to plasma concentrations, virological efficacy or toxicity. Prior
6 to this presentation and to understand limitations of such studies, the following topics
7 are presented and discussed: clinical pharmacokinetics of ARVs, intracellular drug
8 assays and mechanisms influencing intracellular diffusion and accumulation.

9

10 **2 Clinical pharmacokinetics of ARVs**

11 Pharmacokinetic parameters of ARVs are summarized in table I. ^[6, 8-13]

12 **2.1 Entry inhibitors**

13 Entry inhibitors block the virus attachment on receptor of the cell surface. They have
14 an extracellular mode of action and therefore differ from other available classes of
15 antiretroviral agents. Two drugs of this class are available, enfuvirtide and maraviroc,
16 considering their mechanism of action, they are outside the scope of this review. To
17 have an exhaustive overview of ARVs, their pharmacological properties are briefly
18 summarized below.

19 **2.1.1 CCR5 inhibitors**

20 CCR5 or CXCR4 chemokine co-receptor antagonists were promising entry inhibitors.
21 Maraviroc is the first approved drug of this new class. Maraviroc inhibits CCR5
22 chemokine co-receptor preventing HIV binding to cell membrane. Pharmacokinetic
23 characteristics have been summarized elsewhere.^[14] In brief, maraviroc is a CYP3A
24 substrate and dosing differs according to combined drugs (150 mg bid with ritonavir
25 boosted PI, 600 mg bid when combined with drugs with enzyme inducing properties
26 such as efavirenz and 300 mg bid when combined with nucleoside analogs).
27 Maraviroc is a P-gp substrate, which limits intracellular concentrations.
28 Concentrations in cervico-vaginal fluid and vaginal tissue are higher than in plasma.

1 **2.1.2 Enfuvirtide**

2 Enfuvirtide (T20) is a HIV-1 fusion inhibitor, which prevents fusion of HIV-1 and host
3 cell membranes. It is a synthetic peptide (4492 Da), which is not bioavailable when
4 taken orally and is administered subcutaneously twice daily (90 mg bid) which is
5 obviously a limitation to its long-term use. Pharmacokinetic properties have been
6 previously reported.^[15]

7 **2.2 Nucleoside and nucleotide analog inhibitors of reverse**
8 **transcriptase**

9 Zidovudine (ZDV) is the oldest antiretroviral drug; since a number of nucleoside
10 analogs were developed (zalcitabine, didanosine (ddI), stavudine (d4T) lamivudine
11 (3TC), emtricitabine (FTC), abacavir (ABC)). Tenofovir (TFV) is a nucleotide analog
12 obtained after drug administration of tenofovir disoproxil (TDF), its ester prodrug.
13 Apricitabine is a new NRTI under development.

14 Although absolute bioavailability is unknown, bioavailability is supposed to be high for
15 most nucleoside analogs but ddI, which is degraded at acid pH, and TDF. None of
16 these drugs are highly protein bound. Elimination of parent compound occurs as
17 unchanged drug via the kidney or non-CYP drug metabolizing enzymes, therefore
18 potential for drug-drug interaction is low, although TDF was demonstrated to inhibit
19 ddI metabolism.^[16] Triphosphate (TP) metabolites are the active component of all
20 nucleoside analogs. They also inhibit to varying degrees human mitochondrial
21 polymerase γ . Phosphorylation steps occur within the cell and involved kinases,
22 which are listed in figure 1.^[17] Half-life of the active moiety is longer than plasma half-
23 life of the parent compound for all nucleoside analogs. Long half-lives of TP
24 metabolites favor once daily dosing for most of nucleoside analogs except ZDV and
25 d4T, which are administered on a twice-daily basis. TFV is a nucleotide analog for
26 which the active form is a diphosphate (DP). All NRTIs compete with endogenous
27 analogs and stop DNA elongation. Nucleosides such as ABC (carbovir (CBV)) and
28 TFV are much less apt to cause mitochondrial toxicity compared with d4T.^[18]

29

1 **2.3 Non nucleoside analog inhibitors of reverse transcriptase**

2 NNRTIs do not require phosphorylation to inhibit reverse transcriptase. Nevirapine
3 (NVP) and efavirenz (EFV) are the most commonly used. Delavirdine is available in
4 some countries, and etravirine is a new NNRTI recently approved in the European
5 Union and in the United States. NVP and EFV have long half-life after single dose
6 administration. They are metabolized through CYP3A and CYP2B6 and a genetic
7 polymorphism has been described which explains at least part of interindividual
8 variability of their total clearance. They both have enzyme inducing and autoinducing
9 properties, which explains drug-drug interactions and their non linear
10 pharmacokinetics.^[19]

12 **2.4 Integrase inhibitors**

13 Integrase inhibitors represent a new class. These drugs inhibit the integration of HIV-
14 DNA into the host genome. Raltegravir was approved in early 2008 and elvitegravir is
15 under development. Raltegravir is rapidly absorbed and plasma concentrations
16 decline with a terminal half-life of 7 to 12 h which supports a twice daily dosing.^[20]
17 Plasma protein binding is 83%. Biotransformation pathway involved UGT1A1
18 therefore drug-drug interactions are limited.^[21] ATV which inhibits UGT1A1, increases
19 raltegravir concentrations modestly.^[22] Inducers such as EFV, TPV or rifampin
20 decrease raltegravir concentrations although the clinical consequences are currently
21 unclear.^[21]

23 **2.5 Protease inhibitors**

24 Protease inhibitors (PI) prevent cleavage of viral precursor protein into the subunits
25 required to form new virions. Approved PIs include amprenavir (APV), fosamprenavir,
26 atazanavir (ATV), darunavir (DRV), indinavir (IDV), lopinavir/ritonavir (LPV/r),
27 nelfinavir (NFV), ritonavir (RTV), saquinavir (SQV), and tipranavir (TPV).
28 They all are substrate and inhibitor of CYP3A, which explains part of their poor
29 pharmacokinetic properties: pre-systemic first pass metabolism, variable plasma
30 concentrations and short half-life in the 7 to 15 h range. RTV, which is the most
31 potent CYP3A inhibitor, is combined to all PIs but NFV to improve their
32 pharmacokinetic properties, increase plasma exposure and /or decrease the

1 administered dose.^[23, 24] As basic organic chemicals they all are bound to plasma
2 proteins, α 1-glycoprotein acid and albumin. They differ for some pharmacokinetic
3 parameters, extent of first pass metabolism, extent of protein binding (IDV 60%, LPV
4 98-99%) and some of them such as APV have inducing properties which make drug-
5 drug interaction prevision very difficult.
6

7 **3 Methodological considerations**

8 All intracellular assays described to date do not discriminate between drug localised
9 in cell membrane or in cytoplasm, either bound to intracellular proteins or truly
10 unbound which should be the effective antiretroviral moiety. Measurement of total cell
11 concentrations is somehow of limited value.

12 **3.1 Cell collection**

13 Isolation of PBMCs is the first step before analyzing the intracellular concentrations
14 of either NRTIs, NNRTIs or/and PIs. PBMCs can be isolated either using
15 conventional Ficoll gradient centrifugation or using cell preparation tubes (CPTs)
16 (from Becton Dickinson). The two procedures were compared by Becher et al.^[25] on
17 phosphorylated anabolites of two NRTIs and were shown to give identical results.
18 However using CPTs was found to be easier, less time-consuming and therefore
19 quicker which in the case of d4T-TP was most important as the drug was shown
20 instable in the cell ring of the Ficoll gradient (40% loss within 40 min) and lead the
21 authors to collect the ring in less than 10 min. However before this isolation step the
22 stability of the phosphorylated anabolites, that of NNRTIs and PIs in blood should be
23 considered.

24 Regarding d4T-TP, its stability has been checked in blood before PBMCs isolation
25 and the authors recommend to perform the isolation within 6 h after sampling.^[26]
26 Similar results were obtained with 3TC-TP and ZDV-TP.^[27] It therefore seems that for
27 the phosphorylated anabolites the storage of blood samples in CPTs before isolation
28 could last 6 h, although it has not been thoroughly investigated for CBV-TP and TFV-
29 DP.

30 Other issues during cell processing are to avoid contamination by red cells which
31 may phosphorylate some nucleoside analogs^[28] and efflux of PIs and NNRTIs out of
32 cells. In contrast the NRTI-TPs are ion-trapped intracellularly. For NNRTIs and PIs

1 intracellular measurements, the collection of PBMCs has not been systematically
2 studied. The authors mentioned that samples should be immediately taken to
3 laboratory (within 5 min) and that all the procedures should be performed at 4°C to
4 inhibit enzymatic activity and to prevent active drug efflux, the time between blood
5 sampling and the cell isolation and extraction procedure should be less than 1 h.^[29, 30]
6

7 **3.2 Estimation of cell number**

8 Since the number of cells normalizes intracellular concentration, a critical step in the
9 processus of intracellular assay is the determination of the number of cells from
10 which the compounds were quantified.

11 In most studies when the information is indicated the cells were determined on a
12 small aliquot with a Coulter Counter, or using a Malassez cell and a microscope.
13 However this last procedure may suffer from insufficient accuracy and precision,
14 specifically when multiple sites are involved which explains that a biochemical test
15 was developed based on the relationship between DNA content and cells count by
16 Malassez cell.^[31] This test could be performed in the analytical laboratories where
17 there is no Coulter Counter available.

18 The concentration is therefore expressed as amounts per 10^6 cells and can be
19 converted in amount per volume on the approximation that the PBMC volume is
20 0.4 pL in order to compare intracellular and plasma concentrations.^[32] The accuracy
21 of this volume may be questionable as it varies according to the state of the cells
22 (quiescent or stimulated) or to the nature of the cells (cell volume of human
23 lymphoblast : 2.1 pL).^[33] This highlights the pitfalls of the conversion. However the
24 0.4 pL volume is mostly used.^[29, 34] This calculation step is critical for the comparison
25 of the results from different teams, and a standardized procedure should therefore be
26 chosen.

27 **3.3 Analytical methods for intracellular assays**

28 The approaches regarding the analysis of the intracellular drugs due to the difference
29 in their concentrations (low about fmol/ 10^6 cells for intracellular TP anabolites and
30 ng/ 3×10^6 cells i.e. about pmol/ 10^6 cells for PIs) are quite different.

31

1 3.3.1 Nucleoside analog inhibitors of reverse transcriptase

2 The major problem in measuring intracellular TP anabolites is the small amount
3 present in cells of patients and the presence of the endogenous intracellular
4 nucleotides able to interfere. Thus, selective and sensitive analytical methodologies
5 should be developed.

6 Rodriguez et al.,^[35] in 2000 reviewed the latest information regarding the intracellular
7 *in vivo* quantification of NRTI-TP. The authors described the first methods used and
8 pointed out all their drawbacks (lack of sensitivity, cumbersome assays, inability to
9 differentiate NRTI-TP from the endogenous nucleotides, lack of internal standard).
10 More recent approaches were based on the same first steps, i.e. separation of ZDV-
11 anabolites using anion-exchange cartridges, cleavage of the phosphate group using
12 acid phosphatase, addition of an internal standard after enzymatic digestion,
13 desalting and quantification by HPLC-MS/MS. Moreover the calibration curve was
14 prepared from ZDV-TP contrary to the previous procedures, which used the parent
15 compound. The limit of detection was 4.0 fmol/10⁶ cells. The authors applied the
16 same procedure to the simultaneous determination of ZDV-TP and 3TC-TP.^[36] The
17 limit of quantification was 0.1 pmol and 4.0 pmol for ZDV-TP and 3TC-TP,
18 respectively. Moore et al.,^[37] improved this procedure describing an analytical
19 method which allows to measure simultaneously intracellular 3TC-TP, d4T-TP and
20 ZDV-TP with HPLC-MS/MS. The limits of detection were 5, 25, 25 pg on column for
21 3TC-TP, d4T-TP, ZDV-TP respectively. Similar methods were applied by King et
22 al.,^[38] to measure TFV-DP and by Robbins et al.,^[39] to measure simultaneously ZDV-
23 TP, TFV-DP and 3TC-TP in PBMC i.e. isolation by anion exchange, addition of a
24 stable labeled isotope, dephosphorylation, desalination and detection by LC MS/MS.
25 The lower limit of quantification were 10 fmol/10⁶ cells for TFV-DP^[38] and 0.11
26 pmol/10⁶ cells, 2 fmol/10⁶ cells and 3.75 fmol/10⁶ cells for 3TC-TP, ZDV-TP and TFV-
27 DP respectively for a sample size of 10⁶ cells.^[39] Most of these indirect methods are
28 quite labor-intensive, involving multiple steps, which may restrict their use to
29 specialized laboratories.

30 New methodologies were described based on direct HPLC-MS/MS determination on
31 the cellular extracts without dephosphorylation. However these processes need the
32 use of ion pairing agents to circumvent the poor retention of the nucleotides which
33 most of them are incompatible with ionisation mass spectrometry. Pruvost et al.^[26]

1 described the direct determination of d4T-TP as well as that of the endogenous
2 competitor deoxythymidine triphosphate (dT-TP). Just before cell lysis, an internal
3 standard was added. The instrument was operated in the electrospray negative ion
4 mode under MS/MS conditions. The limit of quantification was 9.8 fmol/10⁶ cells i.e.
5 20 pg injected for d4T-TP. In this article the authors focus on the stability of the d4T-
6 TP at the different steps (in blood, in the cells ring, in dry cells at 4°C, after cell lysis
7 at 4°C, in the injection solvent at room temperature). This procedure is very simple to
8 perform as it does not need any extraction step. However due to the very high pH of
9 the mobile phase (ion pairing agent: 1,5-Dimethylhexylamine) the column was
10 changed every two weeks.^[40]

11 With slight modifications regarding the internal standard and the chromatographic
12 column, the same authors were able to measure simultaneously d4T-TP, 3TC-TP
13 and ddA-TP (active anabolite of ddi) with their corresponding natural nucleotides in
14 the same run.^[25] However regarding ZDV-TP, a massive and tailing peak was
15 observed near the retention time of ZDV-TP, which precludes the analysis of ZDV-TP
16 simultaneously with d4T-TP, 3TC-TP and ddA-TP. To overcome this problem Becher
17 et al.,^[41] developed a specific extraction of ZDV-TP using immunoaffinity and
18 detection of ZDV-TP using LC-MS/MS. More recently the same group improved the
19 specificity and obtained a slightly better sensitivity for 3TC-TP, CBV-TP and TFV-TP
20 using a positive electrospray ionization mode.^[42]

21 Although the direct methods should be faster and more precise King et al.,^[43] were
22 unsuccessful in reproducing these methodologies. In particular they pointed out the
23 difficulty in analyzing ZDV-TP due to the large amount of ATP and to the interference
24 with dGTP, these latter compounds having the same precursor ion and the same
25 product ion. This was evidenced by Compain et al.,^[44] who developed an improved
26 method to determine ZDV-TP. The authors chose a minor but specific fragment ion
27 and had to spike their sample with a constant amount of ZDV-TP to allow the signal
28 to emerge from background in order to increase the sensitivity.

29 HPLC-MS/MS is susceptible to matrix effects, i.e. co-eluting matrix components that
30 affect the ionization of the target analyte, resulting in ion suppression, or, in some
31 cases ion enhancement.^[45] For intracellular assay the main parameter to study is the
32 influence of the number of cells in the sample, as it cannot be fixed. The matrix effect
33 plus recovery was tested by Becher et al.,^[46] on d4T-TP and ddA-TP and the
34 influence of the cell number was evidenced. The use of an appropriate internal

1 standard controlled the influence of the matrix effect between 7 and 14×10^6 cells for
2 the simultaneous assay of d4T-TP and ddA-TP. However the use of stable isotope
3 analog as internal standard would be the best choice to control the influence of the
4 matrix effect.

5 Monitoring the very low intracellular concentrations of these active anabolites
6 remains an analytical challenge. All the methods described have their drawbacks.
7 However they are all based on sophisticated methods, which can be hardly
8 reproduced, so each laboratory favors the analytical procedure in which it is familiar.

9 Whatever the choice regarding the procedure, indirect or direct, it appears that the
10 quantification of the compounds using HPLC coupled to MS/MS is very specific and
11 may circumvent all the drawbacks due to the multiple natural nucleotides that are
12 found in the complex mixture of the intracellular medium that can interfere with the
13 determination of intracellular phosphorylated anabolites of NRTIs.

14

15 **3.3.2 Non nucleoside analogs and protease inhibitors**

16 The measurement of intracellular concentrations of NNRTIs and PIs could be
17 obtained using HPLC-UV detection as was it published for EFV^[34] and for 10 of them
18 (NVP, DLV, APV, IDV, metabolite of NFV (M8), RTV, LPV, EFV, SQV, NFV)^[47].

19 However most reported data were obtained using LC-MS/MS methods either for one
20 drug^[48] or for the simultaneous measurement of several of them. The method

21 involved automated solid-phase extraction^[48], liquid-liquid extraction ((APV, LPV,
22 SQV, EFV)^[49], NVP^[30], (LPV, RTV)^[50]) or single-step extraction ((NVP, DLV, APV,
23 IDV, M8, RNV, LPV, EFV, SQV, NFV)^[47], (IDV, APV, SQV, RTV, NFV, LPV, ATV,
24 EFV)^[29]). Few quantitative immunoassays were published for the intracellular

25 determination of LPV and ATV.^[51, 52] These methods imply the preparation of a
26 polyclonal antibody obtained with a synthetic antiretroviral drug derivative coupled to
27 hemocyanin or serum albumin as the immunogen and the chemical synthesis of an
28 enzyme tracer. Obviously these methods are not available in most laboratories,
29 which preclude their use as useful tools to study the intracellular concentrations of
30 NNRTIs and PIs.

31

1 **4 Mechanisms influencing intracellular accumulation**

2 **4.1 General principles**

3 As already stated, most ARVs acting on cell receptors need to enter the cell to bind
4 to antiretroviral targets, reverse transcriptase, integrase or protease. In general,
5 disposition from systemic circulation and capillary lumen to the extravascular
6 compartment occurs by diffusion or involves active transporters

7 Simple diffusion is generally the most common mechanisms for transmembranal
8 movement of xenobiotics in the body. The rate of diffusion is defined by Fick's law
9 and accordingly the small, lipophilic, unionised and unbound molecules readily
10 diffuse across the membrane. Difference in the pH gradient between plasma and
11 lymphocytes could explain ion trapping. As reported by Ford et al.^[53], the pH gradient
12 between plasma and lymphocytes is subject to change depending on the membrane
13 potential. Binding of drugs to plasma proteins may slow diffusion rate as only free
14 unbound drug will cross biological membranes. However basic drugs which have
15 higher affinity for cells or tissues proteins than for plasma proteins may leave very
16 rapidly the blood stream and protein binding is not a limiting factor; for such drugs,
17 volume of distribution is high, the amount of drug in plasma small compared to the
18 amount in tissues and cells and small changes in plasma protein binding will not
19 affect the amount in extra vascular compartments.^[54] Membrane transporters (efflux,
20 influx) are now recognized to play an important role in drug absorption and
21 disposition and to explain, at least in part, the broad interindividual variability in
22 intracellular concentrations of drugs. Figure 2 summarizes the different carrier
23 proteins determining intracellular concentrations within a typical immune cell.^[53, 55-57]

24 Efflux transporters which operate at the expense of adenosine triphosphate (ATP)
25 hydrolysis are members of ATP-binding cassette (ABC)-type transport proteins and
26 are now well studied. P-gp was first described for its ability to reduce intracellular
27 concentrations of anticancer compounds. Other multidrug resistance proteins (MDR)
28 have been isolated since. They are expressed in the apical membrane of many
29 barrier tissues such as the intestine, liver, kidney, blood-brain-barrier, placenta, testis
30 and in immune cells. Relevance for pharmacotherapy of expression of ABC drug
31 transporters in peripheral blood cell have been reviewed recently.^[57] The Breast
32 Cancer Resistance Protein (BCRP) was found to play a major role in nucleoside

1 efflux.^[58, 59] Although influx transporters are not as well studied, several proteins have
2 been identified for nucleosides transport (Solute Carrier, SLC); they differ by their
3 mechanism of action. Some are powered by electrochemical gradient (Concentrative
4 Nucleoside Transporter hCNT), others are Equilibrative Nucleoside Transporter
5 (ENT), Organic Cation Transporters (OCT) or Organic Anion transporters (OAT),
6 although this later was not found to be expressed in immune cells.^[53, 60]
7 Several factors may influence transporters expression within cell membrane, cell
8 subsets and functionality, activation state of cells and polymorphism of coding genes.
9 Polymorphism in the coding region of the transporter genes has been evidenced
10 which lead to produce functional changes in the encoded transporter protein and
11 result in variation in drug disposition and response, however the studies are
12 scanty.^[55] Several factors which could affect intracellular concentrations of ARVs
13 have to be taken into consideration: drug affinity and expression of the transporters
14 according to different cells or tissues, and many of these transporters are known to
15 be modulated by co-administrated ARVs.

16

17 **4.2 Nucleoside and nucleotide analog inhibitors of reverse** 18 **transcriptase**

19 Data on nucleoside analogs (purine or pyrimidine base coupled to a sugar) cellular
20 penetration are scarce. As they are more hydrophilic compounds, it was suggested
21 that they could be substrates of the endogenous nucleoside transporters.^[61, 62]
22 Although studies demonstrated that cerebral penetration occurs mainly by passive
23 diffusion and that the low concentration of nucleoside in brain is the consequence of
24 active efflux transporters^[63], expression of uptake transporters in lymphocytes could
25 favor high intracellular concentrations. It was evidenced that ZDV-TP and 3TC-TP
26 concentrations were effluxed by MRP4 and BCRP. TFV as a nucleotide has an
27 ionized phosphate group, which confers acidic properties. It was demonstrated that
28 TFV uptake in the kidney proximal tubule basolateral membrane is mediated via
29 OAT1 and cellular efflux into the urine via MRP2 and MRP4.^[64] TP concentrations
30 differ according to cell type most likely as a consequence of influx and efflux
31 transporters expression. In healthy volunteers, 3TC-TP concentrations were close in
32 PBMCs and purified CD4 cells, whereas ZDV-TP concentrations were lower in CD4
33 cells than in PBMCs.^[65] Concentrations of TFV-DP were compared in PBMC, lymph

1 node tissue and digestive lymphatic tissue and were higher in PBMCs than in other
2 tissues.^[66] These data strongly suggest that transporters localisation may differ
3 according to cell functionality. Kinase activity could also influence the intracellular
4 concentration of TP. *In vitro* experiments suggested this activity varies greatly and is
5 lower in resting cells than in activated PBMC.^[67, 68] This could have important
6 consequences as kinases activity will govern the intracellular level of both
7 endogenous triphosphates and NRTI-TP which compete at the level of HIV-reverse
8 transcriptase. All NRTIs have been demonstrated to be more effective in monocyte
9 derived macrophages which are important HIV1 reservoirs than in CD4+T
10 lymphocytes.^[69] These could also well explain the differences in NRTI-TP intracellular
11 concentrations according to different cell types and different activation state.^[28]
12

13 **4.3 Non nucleoside analog inhibitors of reverse transcriptase**

14 NNRTIs are weakly acidic and predominantly bind to albumin. Neither EFV nor NVP
15 were thought to be substrate of P-gp.^[70] In a limited number of patients, Almond et
16 al.,^[34] demonstrated a relationship between intracellular concentration of EFV and
17 % bound EFV in plasma. Such data are in contrast with those obtained with NVP by
18 the same team.^[30] They demonstrated that intracellular concentrations of NVP are far
19 below those measured in plasma. Intracellular concentration was negatively related
20 to P-gp expression, but not related to plasma % unbound NVP.^[30] To explain these
21 data, the authors suggest that NVP could induce P-gp or co-regulated efflux
22 transporter. Clearly, to understand all mechanisms, which are involved in intracellular
23 concentrations of NNRTIs, further studies are needed.
24

25 **4.4 Protease inhibitors**

26 The intracellular pharmacology of PIs has been carefully reviewed by Ford et al.^[53].

27 The PI physio-chemical properties are in favour of passive transfer:

- 28 - The transfer is in agreement with lipophilicity measured by the n-octanol to water
29 partition coefficient. Accumulation of PIs in lymphocytes reflects the rank order of
30 lipophilicity: the less lipophilic PI being IDV and the most lipophilic NFV.
- 31 - PIs are weak bases and are mostly unionized in a basic environment; intracellular
32 sequestration is dependant upon pH gradient between plasma and cells.

1 - Protein binding of PIs to α 1-acid glycoprotein ranged from 60% for IDV to 97-99%
2 for RTV, LPV, SQV and NFV. However protein binding per se is not a limiting factor
3 to intracellular diffusion as IDV, which is 60% bound, has lower intracellular
4 concentrations than other PIs more highly bound. Within the cell PIs are bound to
5 cell proteins and HIV proteases and therefore their relative affinity for each protein
6 may influence their dynamic equilibrium.^[53]
7 However active transport may play a role in the intracellular accumulation. It is now
8 well established that PIs are substrates of P-gp and others efflux transporters such
9 as MRP1^[71] or MRP2^[72]. P-gp is expressed in the gastro-intestinal tract and the liver
10 and act with CYP3A to reduce their bioavailability. RTV combined to most PIs as a
11 pharmacologic enhancer inhibits both CYP3A and P-gp and markedly increases the
12 bioavailability of PIs. Such transporters are expressed on lymphocytes and may
13 reduce cellular accumulation. Meaden et al.,^[73] found a relationship between
14 combined expression of P-gp and MRP1 on PBMCs of HIV-infected patients and
15 intracellular accumulation of SQV and RTV. In summary, there are evidences that
16 intracellular concentration of PIs depends on P-gp and/or other efflux transporters
17 activity, which is modulated by genetic polymorphism and coadministration of drugs
18 with inhibiting or inducing properties. How these transporters will control intracellular
19 concentrations of PIs need further studies.
20

21 **4.5 Importance of genetic polymorphism**

22 At evidence, the role of transporters and their genetic polymorphism in drug
23 disposition should be considered^[74] and reviews have summarized findings from
24 recent pharmacogenetics studies.^[55, 75]

25 La Porte et al.,^[76] studied the relationship between ABCB1 (MDR1) genetic
26 polymorphism, P-gp expression and SQV or SQV/r pharmacokinetics in 150 healthy
27 volunteers. No relationship was found between the C3435T, G2677T/A or C1236T
28 polymorphisms of the ABCB1 gene and the pharmacokinetics of SQV or the
29 expression and activity of P-gp in PBMCs. Seventy one HIV-infected children treated
30 with a NFV backbone antiretroviral drug regimen were evaluated for MDR1
31 polymorphism (MDR1 C3435T), NFV plasma concentrations, CD4 cell count and
32 HIV-RNA.^[77] Children with the C/T genotypes had higher 8h post dose NFV
33 concentrations and more rapid response to HAART. Unfortunately, intracellular

1 concentrations of PIs were not measured in these studies. In contrast, in 12 HIV-
2 infected patients, Ford et al.,^[78] could not evidenced higher intracellular
3 concentrations of NFV or its M8 metabolite and lymphocyte cell surface expression of
4 P-gp. In a cohort of 47 patients treated with PI boosted or not by RTV, Chaillou et
5 al.,^[79] demonstrated that intracellular concentration of RTV was related to
6 undetectable plasma HIV-RNA, which was not related to MDR1 gene expression.
7 Interestingly the importance of MRP4 carrier was evidenced by Anderson et al.,^[72],
8 as they demonstrated that patients carrying MRP4 T4131G had elevated 3TC-TP
9 concentrations and patients with MRP4 G3724A had a trend for elevated ZDV-TP.
10 They also found that IDV clearance was faster in patients expressing CYP3A5 and in
11 patients carrying the MRP2-24C/T variant^[72]; whether this latter may contribute to
12 lower intracellular concentrations remains to be established.
13 Recently, Kiser et al.,^[80] demonstrated that intracellular concentrations of TFV-DP
14 were higher, first with decrease in kidney glomerular filtration rate and consequently
15 total and renal clearance of TFV (p=0.04) and second in presence of the ABCC4
16 3463 A>G variant (p=0.04 after adjustment for race, treatment group and glomerular
17 filtration rate). The authors pointed out the limitation of this small sample size study
18 for genetic association and thus results should be confirmed in larger study. Whether
19 those exploratory data could be extrapolated to intracellular concentration of TFV-DP
20 within renal proximal tubule cells is presently unknown.
21 At evidence, the control of intracellular concentrations of ARVs is complex and
22 dependant on many factors and more work is needed in this area taking into account
23 the differences in cell biology.

24

25 **5 Clinical studies with intracellular concentrations**

26 ***5.1 Relationship between intracellular and plasma*** 27 ***pharmacokinetics***

28 Clinical studies performed in HIV-infected patients reporting intracellular
29 concentrations are summarized in table II.^[9-13, 30, 34, 72, 78, 79, 81-101] They are displayed
30 by antiretroviral class, then within an antiretroviral class they are listed by molecule
31 with respect to the date of Health Authority approval. For each molecule, the more
32 recent studies are presented first. All those studies were published after 2000, except

1 for ZDV for which intracellular concentrations have been studied since 1994. Most
2 studies reported both plasma and intracellular concentrations but only few of them
3 studied the relationship between them.

4 It can be seen from table II that plasma pharmacokinetic parameters are rather
5 similar across studies but some differences are observed for intracellular parameters.
6 For NRTIs, most studies did not establish significant relationship between plasma
7 and TP concentrations. In contrast for NNRTIs and PIs results are more conflicting,
8 some studies evidencing correlation, while others could not. These results support
9 the use of plasma concentration of NNRTIs or PIs but not NRTIs to monitor antiviral
10 efficacy.

11 Such results are not surprising knowing first that PBMCs collection, preparation and
12 quantification are not an easy task (see section 3) and second that many factors
13 influence intracellular drug penetration and among them genetic polymorphism of
14 influx and efflux carriers (see section 4). Moreover, these intracellular studies have
15 been carried out in relatively few patients and larger studies would be needed to
16 address consistently the relationship between plasma and intracellular
17 concentrations of ARVs.

18 It is also important to note that there are potential methodological problems when
19 studying relationship between concentrations observed at single time points, as it is
20 done in a number of studies. Indeed, plasma and intracellular half-life are very
21 different. It is more adequate to assess the relationship through pharmacokinetic
22 parameters such as AUC. Surprisingly, most studies reported concentrations at some
23 time points or PK parameters obtained by non-compartmental analysis. Population
24 approaches were never used to analyse intracellular concentrations and their link
25 with plasma concentrations, although this approach seems more appropriate as it
26 allows to analyze sparse measurements.

27 **5.2 Drug-drug interactions at the intracellular level**

28 On a theoretical point of view, changes in the intracellular concentration of
29 antiretroviral drugs can be secondary to modifications of (i) plasma concentration of
30 the drug and/or the prodrug, (ii) activity of the enzymes responsible for drug
31 anabolism/metabolism at the cellular level, (iii) activity of membrane transporters
32 involved in cellular uptake or efflux. Since the intracellular amount of the active drug

1 is responsible for treatment efficacy, interactions leading to changes in intracellular
2 concentrations are a relevant issue regarding the virologic outcome.

3 The clinical impact of these interactions was first evidenced by the poor efficacy of
4 therapies combining ZDV with d4T.^[102] Though the likely mechanism of this result,
5 competitive inhibition of d4T phosphorylation by ZDV, was assessed only by *in vitro*
6 experiments, this phenomenon highlighted the necessity to investigate the possible
7 alteration in the intracellular concentrations of ARVs due to drug association.^[103, 104]

8 Potential interactions involving NRTIs at the intracellular level were therefore
9 investigated in several studies. Hawkins et al.,^[82] evaluated whether the high rate of
10 virological failure observed in patients receiving a triple NRTIs combination including
11 TDF could be explained by modifications in the intracellular anabolism of these
12 compounds.

13 So, intracellular levels of TFV-DP, CBV-TP and 3TC-TP were measured in 15 HIV-
14 infected patients receiving a triple NRTI combination (TDF-ABC-3TC or TDF-ABC-
15 d4T), before and after replacement of TDF or ABC by a NNRTI or a PI. No
16 modification in the intracellular concentrations of the active anabolites of the
17 remaining NRTIs was observed, which suggested the lack of significant interaction
18 between the investigated drugs. Another recent study confirmed these results on 27
19 patients.^[105] Taken together, these results suggest the clinical failure that was
20 observed with the triple NRTI (ABC/TDF/3TC) regimen was not due to drug
21 interactions but was more likely the consequence of lack of intrinsic power.^[106] This
22 latter study also evidenced a significant 50% increase in the intracellular
23 concentration of TFV-DP when TFV was combined to LPV/r. However, this result
24 could simply be the intracellular reflection of the systemic interaction between these
25 two drugs.^[107] This study found no significant difference in the intracellular
26 concentrations of CBV-TP and 3TC-TP with respect to LPV/r use, despite a 46%
27 decrease in ABC plasma concentration in the LPV/r group. Last, nevirapine was also
28 found not to significantly modify the intracellular concentrations of TFV-TP, CBV-TP,
29 and 3TC-TP.

30 Hoggard et al.,^[108] investigated whether prior exposure to ZDV could subsequently
31 inhibit d4T phosphorylation. The rationale for this study came from the observation
32 that naïve patients receiving a d4T -3TC combination experienced a further one log₁₀
33 decrease in viral DNA compared to patients previously treated by ZDV.^[109] A
34 subsequent inhibition of d4T phosphorylation due to a down regulation of thymidine

1 kinase induced by ZDV was one of the hypotheses raised to explain this result.
2 However, the cellular concentration of d4T-TP measured in 7 ZDV-experienced
3 patients was not different to the concentration measured in 20 ZDV-naïve subjects.
4 Furthermore, the ability of PBMCs to phosphorylate d4T was not different between
5 ZDV-experienced and ZDV-naïve subjects.^[108] Similarly, no influence of prior
6 exposure to ZDV on ZDV phosphorylation was observed during a 12 months period
7 on 23 HIV-infected patients.^[83] It is therefore likely that the decrease in efficacy
8 observed in ZDV-experienced patients was due to the acquisition of resistance
9 mutations rather to a modification in intracellular metabolism.

10 By measuring the TP moieties of ZDV and 3TC in the PBMCs of 8 patients, Fletcher
11 et al.,^[84] found a strong correlation between the intracellular concentrations of ZDV-
12 TP and 3TC-TP. If this result suggested the existence of interplay among the cellular
13 anabolism and/or metabolism of these drugs, its precise mechanism and possible
14 consequences have still not been elucidated.

15 TDF is known to increase the plasma concentration of ddl, the most likely
16 mechanism for this interaction being the inhibition by TVF of the enzyme responsible
17 for the hydrolysis of guanosine and adenosine analogues, the purine nucleoside
18 phosphorylase.^[16] This interaction is clinically relevant since it is responsible for
19 adverse effects^[110-113] or treatment failure^[114] which may be secondary to didanosine
20 overexposure. Because of this, this association is currently not recommended for the
21 initiation of HAART, but is nevertheless not contraindicated for ulterior lines of
22 treatment.^[6] Pruvost et al.,^[85] investigated the possible consequences of this
23 systemic interaction on the intracellular concentrations of the active moieties.
24 Intracellular concentrations of ddA-TP and TFV-DP were compared between 14
25 patients receiving the ddl/TDF (250 mg/300 mg) combination and 16 patients
26 receiving ddl (400 mg) without TDF or 14 patients receiving TDF (300 mg) without
27 ddl. The measured concentrations were found to be comparable between the groups
28 which validated the strategy consisting in decreasing ddl dose from 400 to 250 mg
29 when it is combined with TDF.^[115]

30 Apricitabine, a novel deoxycytidine analog currently under investigation, shares its
31 initial phosphorylation pathway by deoxycytidine kinase with 3TC and FTC. The
32 potential interaction between apricitabine (600 mg bid) and 3TC (300 mg qd) was
33 evaluated in a crossover study performed on 21 healthy volunteers who received
34 sequentially each drug separately and the combination of both. No significant

1 modification in the plasma pharmacokinetics of 3TC, or in the cellular
2 pharmacokinetics of its active TP moiety was observed during the combination
3 compared to the monotherapy period. However, if co-administration with 3TC had no
4 influence on apricitabine plasma pharmacokinetics, cellular concentration of
5 apricitabine TP dropped by 85% during the same period.^[116] These findings strongly
6 suggested that apricitabine should not be co-administered with deoxycitidine
7 analogues.

8 Hydroxyurea is an antiproliferative drug that was shown to provide a further 0.7 log₁₀
9 reduction in plasma HIV RNA when combined with ddl compared to patients
10 receiving ddl alone.^[117] By measuring intracellular deoxyadenosine triphosphate (dA-
11 TP) in 69 HIV-infected subjects, it was evidenced that patients receiving the
12 hydroxyurea-ddl combination achieved significantly lower dA-TP concentrations than
13 patients under ddl or hydroxyurea monotherapy, whereas no modification in the
14 plasma pharmacokinetics of the two drugs was observed.^[118] If the precise
15 mechanism of this interaction is still unknown, the likely explanation for the
16 enhancement of ddl's efficacy is the decrease in the intracellular dA-TP/ddA-TP ratio,
17 which would facilitate the incorporation ddA-TP in the replicating viral DNA.

18 Similarly to hydroxyurea, mycophenolic acid, an immunosuppressive agent, is known
19 to decrease the intracellular concentration of an endogenous nucleotide, the
20 deoxyguanosine triphosphate (dG-TP), which could enhance the antiviral activity of
21 abacavir by decreasing the dG-TP/CBV-TP ratio.^[119] Since this ratio could not be
22 measured to date in patients receiving the mycophenolate mofetil–ABC combination,
23 this hypothesis still needs to be confirmed. Nevertheless, the lack of influence of
24 mycophenolic acid on 3TC phosphorylation was suggested by the similar intracellular
25 concentration of 3TC-TP observed in patients receiving 3TC with or without
26 mycophenolate mofetil.^[86]

27 Ribavirin is a nucleoside analogue used for the treatment of hepatitis C virus (HCV)
28 infection. Although its mechanism of action is still not fully understood, it involves at
29 least in part an intracellular transformation into a TP moiety.^[120] Thus, because
30 ribavirin is used in HIV/HCV coinfecting patients, its potential interactions with NRTIs
31 were investigated in several studies.

32 Rodriguez-Torres et al.,^[121] evaluated the combination of ribavirin with 3TC, d4T, or
33 ZDV in HIV/HCV coinfecting patients. Plasma concentrations of ZDV, d4T, 3TC and
34 intracellular concentrations of ZDV-TP, d4T-TP, 3TC-TP were measured in 31

1 patients receiving concomitant ribavirin and compared to the concentrations obtained
2 in 25 patients receiving a placebo instead of ribavirin. No significant difference in
3 plasma and cellular concentrations of the measured compounds was observed,
4 suggesting ribavirin does not modify the plasma pharmacokinetics and the
5 intracellular phosphorylation of ZDV, d4T, 3TC. The lack of interaction between
6 ribavirin and ZDV was confirmed in another study performed on 14 HIV-infected
7 subjects.^[122]

8 It is noteworthy these results are discrepant with *in vitro* data which evidenced an
9 inhibition of the phosphorylation of ZDV^[123, 124] and d4T^[104] by ribavirin. However it is
10 still unexplained whether these discrepancies are due to a poor ability of *in vitro*
11 models to predict *in vivo* phenomenon or to some methodological drawbacks in the
12 *ex-vivo* quantification of intracellular TP moieties.

13 Oppositely, ribavirin was found to potentiate *in vitro* the phosphorylation of ddl via the
14 inhibition of inosine 5-monophosphate dehydrogenase.^[125] However, despite its
15 potential virologic interest, this interaction is also characterized by a high risk of
16 mitochondrial toxicity^[126, 127], so the ribavirin-ddl association is not recommended.

17 Concerning PIs, the influence of ATV on the plasma and intracellular
18 pharmacokinetics of SQV and RTV was investigated in 9 HIV-infected patients who
19 received the SQR/RTV (1600/100 mg qd) combination with and without ATV (200 mg
20 qd). ATV was found to significantly increase both plasma and intracellular
21 concentrations of SQV by a similar factor of approximately 4, but had no effect on
22 RTV concentrations.^[128] Interestingly, the cellular half-life of SQV was unaffected by
23 ATV, which suggested the increase in the intracellular concentration was secondary
24 to the increase in plasma concentration rather to the inhibition of a cellular
25 transporter.

26 The possible modification of plasma and cellular concentrations of SQV by quercetin,
27 a bioflavonoid displaying inhibitory properties on CYP3A4 and P-gp, was investigated
28 on 10 healthy adults who received SQV alone (1200 mg bid) for 11 days followed by
29 the association SQV/quercetin (1200 mg bid/500 mg bid) during the next three
30 days.^[129] If no change was observed for SQV plasma concentration, its intracellular
31 concentration surprisingly decreased by almost 50% when combined with quercetin.
32 However, the important intra and intersubject variability of the intracellular
33 concentrations prevented to draw conclusions from this result.

1 The accumulation ratio, equal to the cellular concentration divided by the plasma
2 concentration, of some PIs was found to be modified by low doses of RTV in HIV-
3 infected subjects.^[79] APV and IDV accumulation ratios in presence of 100 mg or 400
4 mg of RTV were indeed increased three-fold and five-fold respectively. However,
5 conflicting results were obtained in another study which found that RTV did not
6 increase the accumulation ratio of SQV and IDV.^[130]
7 Despite these discrepancies, assessing the consequences of drug interactions at the
8 cellular level is of great concern in order to validate new combinations. Some recent
9 surprising results, like the possible decrease in the efficacy of HCV therapy due to
10 ABC^[131], evidenced the need for a better understanding of drug interactions.

12 **5.3 Relationship between intracellular concentrations and efficacy**

13 Only eight studies were published reporting an analysis of the relationships between
14 intracellular concentrations and virological or immunological efficacy of antiretroviral
15 drugs in HIV patients. Five studied intracellular NRTIs and three PIs. These articles
16 are summarized in table III by molecule from the most recent to the oldest one.^{[8, 79, 84,}
17 ^{87-89, 122, 132]} Of note, five were prospective study and interestingly found a significant
18 relationship between higher intracellular concentrations and virological response.^{[8, 84,}
19 ^{87, 88, 132]} The correlation with plasma concentration was not always studied but was
20 mainly not or less significant. The three studies with non significant results were
21 cross-sectional studies not designed for that purpose.^[88, 89, 122] We start as in table II
22 by the results on NRTIs and then on PIs, there is no such study for NNRTIs. With
23 nucleoside analogues, it is important to notice that the relevant determinant of
24 pharmacodynamic response is the ratio between drug triphosphate and endogenous
25 nucleoside triphosphates rather than the absolute intracellular concentration.^[133]
26 The study by Moore et al.,^[87] was a substudy of the ACTG 862, a prospective trial
27 where naïve patients were starting a dual NRTI therapy. A significant correlation
28 between change in viral load between week 0 and week 28 and intracellular
29 concentrations was found for 3TC-TP ($R^2= 0.62$) in the 39 patients receiving either
30 3TC-ZDV or 3TC-d4T, and for ZDV-TP ($R^2= 0.28$) in the 10 patients receiving 3TC-
31 ZDV. No significant correlation was found for d4T-TP in the 15 patients receiving
32 3TC-d4T and no significant relationship was found for any drug when studying
33 change in CD4 cells. The authors did not study the relationships between efficacy

1 and plasma concentration but showed a very large interpatient variability in the
2 intracellular to plasma concentration ratio. The authors also showed that there was
3 an important increase of intracellular concentration between first dose and week 28
4 only for 3TC-TP.

5 The study of Aweeka et al.,^[122] was performed in HCV or HBV co-infected HIV
6 patients and its primary objective was to study the influence of ribavirin on ZDV
7 plasma and intracellular concentrations by measuring the AUC in patients before and
8 after introduction of ribavirin. Different regimens were allowed and all patients had to
9 receive ZDV for at least 4 weeks. In the cross-sectional analysis performed in 13
10 patients before receiving ribavirin, no significant relationships was found between
11 AUC of ZDV-TP and the CD4 cell count measured the same day.

12 The studies by Anderson et al.,^[8] and Fletcher et al.,^[84] are substudies of the well
13 known concentration-controlled randomised trial by Fletcher et al.,^[134]. In this trial
14 patients received a tritherapy with ZDV, 3TC and IDV either with a fixed dose or with
15 a dose adapted to get trough concentrations in a define range. Intracellular
16 concentrations of ZDV-TP and 3TC-TP were measured 2 h after dose administration
17 at the three pharmacokinetics visits scheduled at weeks 2, 26 and 52 and at variable
18 time post dose at the nine bimontly visits. For each patient the median intracellular
19 concentration from all measurements was considered for the analysis of the link with
20 efficacy. Unfortunately the authors did not analyse intrapatient variability of these
21 concentrations.

22 In their first study^[84], only 8 patients were studied and the efficacy criteria were the
23 change between baseline and week 24 HIV RNA or the percent change between
24 baseline and week 24 CD4 cell count. For ZDV-TP a significant correlation with
25 intracellular concentrations was found both for changes in HIV RNA ($R^2=0.54$) and
26 CD4 cells ($R^2=0.84$). For 3TC-TP a significant correlation with intracellular
27 concentrations was found for changes in HIV RNA ($R^2=0.79$) but not for CD4 cells
28 ($P=0.07$, $R^2=0.44$). There was no significant correlation between efficacy and steady-
29 state plasma concentration of these drugs.

30 In the article by Anderson et al.,^[8], 33 patients were studied; the analysis was very
31 thorough with various efficacy endpoints and with several multivariate analyses. After
32 a first simple correlation study, the authors defined thresholds for intracellular
33 concentrations as the first quartiles, which are of 30 fmol/ 10^6 cells for ZDV-TP and of
34 7017 fmol/ 10^6 cells for 3TC-TP. They then studied the impact on several efficacy

1 endpoints of having a median intracellular concentration below or above those
2 thresholds. The first efficacy endpoint was the time to plasma HIV RNA less than 50
3 cp/mL, using a survival analysis. For ZDV-TP, the median time to less than 50 cp/mL
4 is significantly reduced in patients with ZDV-TP above the threshold than in patients
5 below as shown.^[8] A significant relationship was also found for 3TC-TP but only ZDV-
6 TP remained in the multivariate analysis. The second efficacy endpoints were the
7 virological status (< 50 cp/mL) at week 24 and at week 52 after starting ARVs drug
8 regimen. For ZDV-TP, 92% of patients with concentrations above threshold were
9 undetectable at week 24, this proportion was significantly lower (44%, P=0.009) for
10 patients below the threshold; but this relationship was not significant at week 52. For
11 3TC-TP, 96% of patients with concentrations above threshold were undetectable at
12 week 24, proportion that was significantly higher than for patients below the threshold
13 (37.5 %, P=0.002); similarly at week 52, 91.7% of patients above threshold were
14 undetectable which is significantly higher than for patients below the threshold (25%,
15 P=0.0008). Another efficacy endpoint was the time to rebound (two HIV RNA greater
16 than 50 cp/mL), this time was shorter for patients with ZDV-TP and 3TC-TP lower
17 threshold; in the multivariate analysis only 3TC-TP remains significant (P=0.009). It is
18 interesting to note that ZDV-TP was mostly associated with the initial viral load
19 decrease whereas 3TC-TP with sustained response. No significant relationship was
20 found between CD4 cell count and intracellular concentrations. However, the authors
21 did not study the change in CD4 cell count but performed cross-sectional analyses
22 looking for correlation at week 24 and week 52 between CD4 cell counts and
23 corresponding intracellular concentrations. Link between plasma concentrations of
24 ZDV and 3TC and efficacy was not studied and no significant relationships were
25 found with IDV concentrations. The authors also found that intracellular
26 concentrations were significantly higher in female than in male: 2.3 folds for ZDV-TP
27 (P<0.0001) and 1.6 fold for 3TC-TP (P<0.0001), whereas no influence of gender was
28 found on plasma concentrations. These results suggest NRTI phosphorylation
29 differences between genders.

30 The first ever published article that analyzed the link between intracellular
31 concentration and efficacy was the pharmacokinetic trial reported by Stretcher et
32 al.,^[132] where ZDV was given five times a day as a single therapy (500 mg/day) in 21
33 patients followed during 24 weeks. AUC of total ZDV-phosphates (ZDV-P) was
34 evaluated at week 4 and week 24 from five samples. Efficacy was analysed mainly

1 through CD4 cell count, %CD4 cells and CD4/CD8 ratio. Unfortunately, in the
2 analysis of the correlation of ZDV-P AUC and immunological efficacy, the authors
3 pooled the observations made at week 4 and week 24, not taking into account the
4 correlation induced by the repetition within patients. Here, only results on the
5 analyses performed separately at week 4 and week 24 are reported. At week 4, the
6 authors found a significant correlation of ZDV-P AUC both with %CD4 cells change
7 from baseline ($R^2=0.06$, $P=0.029$) and with CD4/CD8 change from baseline ($R^2=0.06$,
8 $P=0.028$) but not with the value measured at week 4. These correlations were no
9 longer significant at week 24. No significant correlation was found with plasma AUC.
10 With respect to studies with PIs, the main objective of the prospective trial reported
11 by Lamotte et al.,^[89] was to investigate the concept of a once daily administration of
12 the new galenic formulation soft gel capsule of SQV in association with RTV in PI-
13 experienced HIV patients. The evaluation of the link between SQV intracellular
14 concentrations and virological efficacy in 13 patients was explored as one of the
15 secondary objectives. No significant correlation was obtained between trough SQV
16 intracellular concentration at weeks 2, 4 or 12 and variation of plasma HIV RNA
17 between week 0 and week 12. No significant correlation was found also for plasma
18 SQV concentrations.
19 The main objective of the prospective study reported by Breilh et al.,^[88] was the
20 impact on virological success, defined as HIV less than 50 cp/mL, of intracellular and
21 plasma trough concentrations of LPV in 38 patients receiving LPV/RTV based
22 regimen. They found that trough intracellular concentrations of LPV at week 4 were
23 significantly higher (12.7 $\mu\text{g/mL}$) in patients achieving virological success before
24 week 4 than in others (4.8 $\mu\text{g/mL}$) ($P<0.0002$). Similarly intracellular concentrations of
25 LPV at week 24 were significantly higher (10.5 $\mu\text{g/mL}$) in patients achieving
26 virological success before week 24 than in others (4.6 $\mu\text{g/mL}$) ($P<0.002$).^[88]
27 Virological success was also significantly associated with higher plasma trough
28 concentration at week 4 ($P=10^{-5}$) and 24 ($P=0.05$) and with the genotype inhibitory
29 quotient at week 4 ($P=10^{-6}$) and 24 ($P=0.0004$). In a multivariate analysis of
30 virological success at week 24, the authors found the effect of baseline LPV
31 mutations, plasma concentration at week 4 and intracellular concentration at week
32 24. The authors defined thresholds of 4 $\mu\text{g/mL}$ and 8 $\mu\text{g/mL}$ for plasma and
33 intracellular concentration, respectively; they suggested combining plasma and

1 intracellular concentration of LPV for therapeutic drug monitoring. The authors
2 derived cellular accumulation ratio but it was not used in the analysis of link with
3 efficacy.

4 The cross-sectional study by Chaillou et al.,^[79] included 49 patients with antiretroviral
5 combinations containing various protease inhibitors. The first objective was to study
6 the relationship between MDR-1 gene expression and intracellular PI concentrations
7 and then to evaluate the correlation of PI intracellular concentrations with virological
8 response. Efficacy was defined as undetectable HIV RNA load (<40 cp/mL) at day of
9 study. As various PIs were analyzed, to normalize concentrations authors studied the
10 influence of the ratio of intracellular to plasma concentrations that they defined as
11 accumulation but which is not a measure showing the amount of drug in body. They
12 did not found any significant correlation on the main PI. The only parameter
13 significantly linked with efficacy was the intracellular presence of RTV (P=0.04). For
14 the 19 patients receiving RTV as a booster, patients with undetectable HIV viral load
15 had significantly higher RTV intracellular accumulation than patients with detectable
16 HIV RNA (P=0.029).

17 In conclusion, in prospective studies well designed and with a reasonable number of
18 patients, all authors found a significant correlation between virological efficacy and
19 intracellular concentrations of NRTIs with no influence of plasma concentration. For
20 PIs, there is only one well-designed prospective trial on LPV, which found both the
21 influence of trough plasma and intracellular concentrations at different week after
22 treatment initiation. From these results, it is difficult to know whether the primary
23 association is with plasma or intracellular concentrations. These findings obtained in
24 only 38 patients should be confirmed by other studies.

25

26 **5.4 Relationship between intracellular concentrations and toxicity**

27 ARVs are known to produce important adverse effects which are the main drawback
28 of HAART.^[135] The toxicity related to ARVs is indeed an important cause of poor
29 compliance, which is in return the main cause of treatment failure.^[136] Besides, a viral
30 rebound can be associated to the acquisition of mutation resistances by the virus,
31 which can critically penalize the choice of the subsequent therapeutic strategy.^[137-139]
32 Most toxicities displayed by ARVs are typical of a pharmacological class excepted for
33 NRTIs which trend to have their own toxicities. For instance, PIs are known to induce

1 digestive troubles and metabolic disorders, such as hyperlipidaemia, insulin
2 resistance, diabetes mellitus, peripheral lipodystrophy, central adiposity^[140-143]. NRTIs
3 can be responsible for lipodystrophy^[144], neuropathy^[145] (ddl, d4T), myopathy^[146],
4 pancreatitis^[147] (ddl,d4T), anaemia and neutropenia^[148],(ZDV), renal impairment and
5 fanconi syndrome^[149] (TFV), hepatic steatosis, and lactic acidosis^[144]; whereas
6 NNRTIs can provide neuropsychological disorders for EFV^[150] and skin or hepatic
7 toxicity for NVP^[151-153].

8 To date, the mechanisms leading to these toxicities are not perfectly understood, but,
9 the main hypotheses highly suggest interferences with some cellular endogenous
10 processes. For example, PIs could alter adipose tissue and lipid metabolism by
11 inhibiting the heterodimeric nuclear receptor complex composed of peroxisome
12 proliferator activated receptor γ and the retinoid X receptor, the cellular retinoic acid-
13 binding protein, and the synthesis of cis-9-retinoic acid.^[154] PIs could also inhibit the
14 degradation of the sterol element-binding proteins which regulate the transcription of
15 the LDL receptor gene.^[155, 156] Diabetes mellitus induced by PIs could be secondary
16 to the direct inhibition of GLUT4, a transporter that mediates the cellular uptake of
17 glucose stimulated by insulin.^[157] Similarly, adverse effects due to NRTIs are thought
18 to be related to mitochondrial damages, which are a consequence of NRTIs ability to
19 inhibit the mitochondrial DNA polymerase.^[158]

20 Despite these elements, the possible relationship between intracellular concentration
21 of ARVs and their related toxicity was to date investigated for four molecules (ZDV,
22 3TC, TDF and EFV) only.

23 First, Stretcher et al.,^[132] evaluated in 13 naïve patients if the intracellular
24 concentration of ZDV-P in PBMCs was related to some markers of ZDV-induced
25 toxicity. A negative correlation was found between intracellular ZDV-P and the
26 decrease in haemoglobin from its baseline level. It is noteworthy that the authors did
27 not find in this study a significant correlation between plasma ZDV and intracellular
28 ZDV-P, and did not investigate the possible relationship between plasma ZDV and
29 haemoglobin decrease. However, other studies evidenced an association between
30 plasma ZDV and anaemia^[159, 160], so the relative strength of the association between
31 haemoglobin decrease and plasma ZDV compared to intracellular ZDV-P is still
32 unknown. A different result was nevertheless found in both adults and children.
33 Indeed, in a study performed by Anderson et al.,^[8] on 33 naïve adult patients
34 receiving a ZDV-3TC-IDV regimen, no difference in the intracellular concentrations of

1 ZDV-TP and 3TC-TP was observed between the 14 patients who experienced at
2 least a grade I biological event and the 19 patients who did not. A similar result was
3 found in 49 neonates, as the proportion of observed haematological toxicities was not
4 related to the intracellular concentrations of ZDV-TP and 3TC-TP.^[99] However,
5 differences between the pharmacokinetic criteria and the pharmacodynamic
6 endpoints used (see table IV^[8, 90, 99, 132]) might explain the inconsistency between
7 these studies. There are also *in vitro* data suggesting ZDV-MP is the culprit of ZDV-
8 related anemia^[161], so toxicity relationships with ZDV-TP may not be relevant.
9 Among NNRTIs, in a study performed on 55 patients, Rotger et al.,^[90] found a
10 significant correlation between the intracellular concentration of EFV and the risk of
11 mood disorders. No significant correlation was found with the risk of sleep disorders
12 and fatigue. In contrast with other studies^[162, 163], no significant association was
13 observed between the plasma concentration of EFV and the neuropsychological
14 trouble that were investigated. Once again, methodological differences penalize the
15 comparison between studies.
16 Last, Izzedine et al.,^[64] found, without measuring the intracellular concentration of
17 TFV-DP, that the risk to develop a renal proximal tubulopathy with TDF-containing
18 treatments was significantly associated with genetic variants in the gene coding for
19 MRP2, a transporter involved in TVF efflux from tubular cells to the urine. Since
20 these variants are thought to be associated with a reduced activity of the transporter,
21 this result could indicate that an accumulation in the tubular cell due to an altered
22 MRP2-based efflux might be responsible for TDF-induced toxicity.^[64] However, for
23 obvious reasons, TFV concentration in the tubular cells could not be investigated, so
24 this mechanistic explanation remains speculative, as are the role of mitochondria and
25 TFV-DP in TDF toxicity.
26 More generally, the impossibility to investigate cellular concentration of ARVs in the
27 tissues targeted by their toxicity is a major weakness of these studies. However, it is
28 interesting to note the neuropsychological effect of efavirenz or the anaemia induced
29 by ZDV are not explained by the diffusion of these compounds to the PBMCs.
30 Significant correlations indicate that concentration in PBMCs possibly reflects the
31 diffusion of these drugs to other tissues like central nervous system or bone marrow.
32 Measuring the concentrations of ARVs in PBMCs could therefore be an interesting
33 tool to predict and consequently to prevent the appearance of toxicities related to
34 HAART in a clinical setting. Further studies are therefore warranted to validate

1 PBMCs as a reliable model for investigating the relationship between the intracellular
2 concentration of ARVs and their toxicity.

3

4 **6 Conclusion**

5 In conclusion, intracellular concentrations of ARVs play a major role in their efficacy
6 and toxicity and are influenced by numerous factors. Although measurement of
7 intracellular concentrations needs standardisation, this review demonstrates that
8 relationships between intracellular concentrations of ARVs and their efficacy have
9 been evidenced. Such relationships should be interpreted with caution as intracellular
10 concentrations reflect the total amount of drug within the cell and not the effective
11 unbound fraction. The number of clinical studies in that area is however rather
12 limited, most studies being small and not always adequately designed. Improved
13 techniques measuring relevant intracellular concentrations, improved knowledge on
14 which cells would be the best surrogate marker on antiretroviral therapy including
15 reservoirs as well as larger and prospectively designed clinical studies are needed to
16 further investigate the links between intracellular concentrations and clinical
17 endpoints.

18

19

20

1 References

- 2 1. Carpenter CC, Cooper DA, Fischl MA, et al. Antiretroviral therapy in adults:
3 updated recommendations of the International AIDS Society-USA Panel. *Jama* 2000
4 Jan 19;283 (3): 381-90
- 5 2. Hammer SM, Saag MS, Schechter M, et al. Treatment for adult HIV infection:
6 2006 recommendations of the International AIDS Society-USA panel. *Jama* 2006
7 Aug 16;296 (7): 827-43
- 8 3. Chen LF, Hoy J, Lewin SR. Ten years of highly active antiretroviral therapy for
9 HIV infection. *Med J Aust* 2007 Feb 5;186 (3): 146-51
- 10 4. Stebbing J, Gazzard B, Douek DC. Where does HIV live? *N Engl J Med* 2004
11 Apr 29;350 (18): 1872-80
- 12 5. Guidelines for the use of antiretroviral agents in HIV-1 infected adults and
13 adolescents. Department of health and human services (DHHS).
14 <http://aidsinfo.nih.gov/guidelines> [accessed 2008 dec 1]
- 15 6. Yeni P. Prise en charge médicale des personnes infectées par le VIH :
16 Recommandations du groupe d'experts. Paris: Flammarion, 2008
- 17 7. Rockstroh JK, Bhagani S, Benhamou Y, et al. European AIDS Clinical Society
18 (EACS) guidelines for the clinical management and treatment of chronic hepatitis B
19 and C coinfection in HIV-infected adults. *HIV Med* 2008 Feb;9 (2): 82-8
- 20 8. Anderson PL, Kakuda TN, Kawle S, et al. Antiviral dynamics and sex
21 differences of zidovudine and lamivudine triphosphate concentrations in HIV-infected
22 individuals. *Aids* 2003 Oct 17;17 (15): 2159-68
- 23 9. Hennessy M, Clarke S, Spiers JP, et al. Intracellular indinavir
24 pharmacokinetics in HIV-infected patients: comparison with plasma
25 pharmacokinetics. *Antivir Ther* 2003 Jun;8 (3): 191-8
- 26 10. Rousseau FS, Kahn JO, Thompson M, et al. Prototype trial design for rapid
27 dose selection of antiretroviral drugs: an example using emtricitabine (Coviracil). *J*
28 *Antimicrob Chemother* 2001 Oct;48 (4): 507-13
- 29 11. Becher F, Landman R, Mboup S, et al. Monitoring of didanosine and stavudine
30 intracellular trisphosphorylated anabolite concentrations in HIV-infected patients.
31 *Aids* 2004 Jan 23;18 (2): 181-7
- 32 12. Stretcher BN, Pesce AJ, Frame PT, et al. Pharmacokinetics of zidovudine
33 phosphorylation in peripheral blood mononuclear cells from patients infected with
34 human immunodeficiency virus. *Antimicrob Agents Chemother* 1994 Jul;38 (7): 1541-
35 7
- 36 13. Stretcher BN, Pesce AJ, Hurtubise PE, et al. Pharmacokinetics of zidovudine
37 phosphorylation in patients infected with the human immunodeficiency virus. *Ther*
38 *Drug Monit* 1992 Aug;14 (4): 281-5
- 39 14. MacArthur RD, Novak RM. Reviews of anti-infective agents: maraviroc: the
40 first of a new class of antiretroviral agents. *Clin Infect Dis* 2008 Jul 15;47 (2): 236-41
- 41 15. Patel IH, Zhang X, Nieforth K, et al. Pharmacokinetics, pharmacodynamics
42 and drug interaction potential of enfuvirtide. *Clin Pharmacokinet* 2005;44 (2): 175-86
- 43 16. Ray AS, Olson L, Fridland A. Role of purine nucleoside phosphorylase in
44 interactions between 2',3'-dideoxyinosine and allopurinol, ganciclovir, or tenofovir.
45 *Antimicrob Agents Chemother* 2004 Apr;48 (4): 1089-95
- 46 17. Anderson PL, Kakuda TN, Lichtenstein KA. The cellular pharmacology of
47 nucleoside- and nucleotide-analogue reverse-transcriptase inhibitors and its
48 relationship to clinical toxicities. *Clin Infect Dis* 2004 Mar 1;38 (5): 743-53

- 1 18. Moyle G. Toxicity of antiretroviral nucleoside and nucleotide analogues: is
2 mitochondrial toxicity the only mechanism? *Drug Saf* 2000 Dec;23 (6): 467-81
- 3 19. Barry M, Mulcahy F, Merry C, et al. Pharmacokinetics and potential
4 interactions amongst antiretroviral agents used to treat patients with HIV infection.
5 *Clin Pharmacokinet* 1999 Apr;36 (4): 289-304
- 6 20. Iwamoto M, Wenning LA, Petry AS, et al. Safety, tolerability, and
7 pharmacokinetics of raltegravir after single and multiple doses in healthy subjects.
8 *Clin Pharmacol Ther* 2008 Feb;83 (2): 293-9
- 9 21. Correll T, Klibanov OM. Integrase inhibitors: a new treatment option for
10 patients with human immunodeficiency virus infection. *Pharmacotherapy* 2008
11 Jan;28 (1): 90-101
- 12 22. Iwamoto M, Wenning LA, Mistry GC, et al. Atazanavir modestly increases
13 plasma levels of raltegravir in healthy subjects. *Clin Infect Dis* 2008 Jul 1;47 (1): 137-
14 40
- 15 23. Cooper CL, van Heeswijk RP, Gallicano K, et al. A review of low-dose ritonavir
16 in protease inhibitor combination therapy. *Clin Infect Dis* 2003 Jun 15;36 (12): 1585-
17 92
- 18 24. Moyle GJ, Back D. Principles and practice of HIV-protease inhibitor
19 pharmacoenhancement. *HIV Med* 2001 Apr;2 (2): 105-13
- 20 25. Becher F, Pruvost A, Goujard C, et al. Improved method for the simultaneous
21 determination of d4T, 3TC and ddI intracellular phosphorylated anabolites in human
22 peripheral-blood mononuclear cells using high-performance liquid
23 chromatography/tandem mass spectrometry. *Rapid Commun Mass Spectrom*
24 2002;16 (6): 555-65
- 25 26. Pruvost A, Becher F, Bardouille P, et al. Direct determination of
26 phosphorylated intracellular anabolites of stavudine (d4T) by liquid
27 chromatography/tandem mass spectrometry. *Rapid Commun Mass Spectrom*
28 2001;15 (16): 1401-8
- 29 27. Data on file, Compain S, Benech H, Grassi J, et al., 2008
- 30 28. Durand-Gasselín L, Da Silva D, Benech H, et al. Evidence and possible
31 consequences of the phosphorylation of nucleoside reverse transcriptase inhibitors in
32 human red blood cells. *Antimicrob Agents Chemother* 2007 Jun;51 (6): 2105-11
- 33 29. Colombo S, Beguin A, Telenti A, et al. Intracellular measurements of anti-HIV
34 drugs indinavir, amprenavir, saquinavir, ritonavir, nelfinavir, lopinavir, atazanavir,
35 efavirenz and nevirapine in peripheral blood mononuclear cells by liquid
36 chromatography coupled to tandem mass spectrometry. *J Chromatogr B Analyt*
37 *Technol Biomed Life Sci* 2005 May 25;819 (2): 259-76
- 38 30. Almond LM, Edirisinghe D, Dalton M, et al. Intracellular and plasma
39 pharmacokinetics of nevirapine in human immunodeficiency virus-infected
40 individuals. *Clin Pharmacol Ther* 2005 Aug;78 (2): 132-42
- 41 31. Benech H, Theodoro F, Herbet A, et al. Peripheral blood mononuclear cell
42 counting using a DNA-detection-based method. *Anal Biochem* 2004 Jul 1;330 (1):
43 172-4
- 44 32. Gao WY, Cara A, Gallo RC, et al. Low levels of deoxynucleotides in peripheral
45 blood lymphocytes: a strategy to inhibit human immunodeficiency virus type 1
46 replication. *Proc Natl Acad Sci U S A* 1993 Oct 1;90 (19): 8925-8
- 47 33. Traut TW. Physiological concentrations of purines and pyrimidines. *Mol Cell*
48 *Biochem* 1994 Nov 9;140 (1): 1-22

- 1 34. Almond LM, Hoggard PG, Edirisinghe D, et al. Intracellular and plasma
2 pharmacokinetics of efavirenz in HIV-infected individuals. *J Antimicrob Chemother*
3 2005 Oct;56 (4): 738-44
- 4 35. Rodriguez Orengo JF, Santana J, Febo I, et al. Intracellular studies of the
5 nucleoside reverse transcriptase inhibitor active metabolites: a review. *P R Health*
6 *Sci J* 2000 Mar;19 (1): 19-27
- 7 36. Rodriguez JF, Rodriguez JL, Santana J, et al. Simultaneous quantitation of
8 intracellular zidovudine and lamivudine triphosphates in human immunodeficiency
9 virus-infected individuals. *Antimicrob Agents Chemother* 2000 Nov;44 (11): 3097-100
- 10 37. Moore JD, Valette G, Darque A, et al. Simultaneous quantitation of the 5'-
11 triphosphate metabolites of zidovudine, lamivudine, and stavudine in peripheral
12 mononuclear blood cells of HIV infected patients by high-performance liquid
13 chromatography tandem mass spectrometry. *J Am Soc Mass Spectrom* 2000 Dec;11
14 (12): 1134-43
- 15 38. King T, Bushman L, Kiser J, et al. Liquid chromatography-tandem mass
16 spectrometric determination of tenofovir-diphosphate in human peripheral blood
17 mononuclear cells. *J Chromatogr B Analyt Technol Biomed Life Sci* 2006 Nov 7;843
18 (2): 147-56
- 19 39. Robbins BL, Poston PA, Neal EF, et al. Simultaneous measurement of
20 intracellular triphosphate metabolites of zidovudine, lamivudine and abacavir
21 (carbovir) in human peripheral blood mononuclear cells by combined anion exchange
22 solid phase extraction and LC-MS/MS. *J Chromatogr B Analyt Technol Biomed Life*
23 *Sci* 2007 May 1;850 (1-2): 310-7
- 24 40. Pruvost A, Becher F, Bardouille P, et al. Direct determination of
25 phosphorylated intracellular anabolites of stavudine (d4T) by liquid
26 chromatography/tandem mass spectrometry. *Rapid Commun Mass Spectrom*
27 2007;21 (13): 2167
- 28 41. Becher F, Schlemmer D, Pruvost A, et al. Development of a direct assay for
29 measuring intracellular AZT triphosphate in humans peripheral blood mononuclear
30 cells. *Anal Chem* 2002 Aug 15;74 (16): 4220-7
- 31 42. Pruvost A, Theodoro F, Agrofoglio L, et al. Specificity enhancement with LC-
32 positive ESI-MS/MS for the measurement of nucleotides: application to the
33 quantitative determination of carbovir triphosphate, lamivudine triphosphate and
34 tenofovir diphosphate in human peripheral blood mononuclear cells. *J Mass*
35 *Spectrom* 2008 Feb;43 (2): 224-33
- 36 43. King T, Bushman L, Anderson PL, et al. Quantitation of zidovudine
37 triphosphate concentrations from human peripheral blood mononuclear cells by anion
38 exchange solid phase extraction and liquid chromatography-tandem mass
39 spectroscopy; an indirect quantitation methodology. *J Chromatogr B Analyt Technol*
40 *Biomed Life Sci* 2006 Feb 2;831 (1-2): 248-57
- 41 44. Compain S, Durand-Gasselien L, Grassi J, et al. Improved method to quantify
42 intracellular zidovudine mono- and triphosphate in peripheral blood mononuclear
43 cells by liquid chromatography-tandem mass spectrometry. *J Mass Spectrom* 2007
44 Mar;42 (3): 389-404
- 45 45. Chambers E, Wagrowski-Diehl DM, Lu Z, et al. Systematic and
46 comprehensive strategy for reducing matrix effects in LC/MS/MS analyses. *J*
47 *Chromatogr B Analyt Technol Biomed Life Sci* 2007 Jun 1;852 (1-2): 22-34
- 48 46. Becher F, Pruvost A, Gale J, et al. A strategy for liquid
49 chromatography/tandem mass spectrometric assays of intracellular drugs:

- 1 application to the validation of the triphosphorylated anabolite of antiretrovirals in
2 peripheral blood mononuclear cells. *J Mass Spectrom* 2003 Aug;38 (8): 879-90
- 3 47. Pelerin H, Compain S, Duval X, et al. Development of an assay method for the
4 detection and quantification of protease and non-nucleoside reverse transcriptase
5 inhibitors in plasma and in peripheral blood mononuclear cells by liquid
6 chromatography coupled with ultraviolet or tandem mass spectrometry detection. *J*
7 *Chromatogr B Analyt Technol Biomed Life Sci* 2005 May 5;819 (1): 47-57
- 8 48. Jemal M, Rao S, Gatz M, et al. Liquid chromatography-tandem mass
9 spectrometric quantitative determination of the HIV protease inhibitor atazanavir
10 (BMS-232632) in human peripheral blood mononuclear cells (PBMC): practical
11 approaches to PBMC preparation and PBMC assay design for high-throughput
12 analysis. *J Chromatogr B Analyt Technol Biomed Life Sci* 2003 Oct 5;795 (2): 273-89
- 13 49. Rouzes A, Berthoin K, Xuereb F, et al. Simultaneous determination of the
14 antiretroviral agents: amprenavir, lopinavir, ritonavir, saquinavir and efavirenz in
15 human peripheral blood mononuclear cells by high-performance liquid
16 chromatography-mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci*
17 2004 Dec 25;813 (1-2): 209-16
- 18 50. Ehrhardt M, Mock M, Haefeli WE, et al. Monitoring of lopinavir and ritonavir in
19 peripheral blood mononuclear cells, plasma, and ultrafiltrate using a selective and
20 highly sensitive LC/MS/MS assay. *J Chromatogr B Analyt Technol Biomed Life Sci*
21 2007 May 1;850 (1-2): 249-58
- 22 51. Azoulay S, Nevers MC, Creminon C, et al. An enzyme immunoassay for the
23 quantification of plasma and intracellular lopinavir in HIV-infected patients. *J Immunol*
24 *Methods* 2004 Dec;295 (1-2): 37-48
- 25 52. Roucairol C, Azoulay S, Nevers MC, et al. Quantitative immunoassay to
26 measure plasma and intracellular atazanavir levels: analysis of drug accumulation in
27 cultured T cells. *Antimicrob Agents Chemother* 2007 Feb;51 (2): 405-11
- 28 53. Ford J, Khoo SH, Back DJ. The intracellular pharmacology of antiretroviral
29 protease inhibitors. *J Antimicrob Chemother* 2004 Dec;54 (6): 982-90
- 30 54. Boffito M, Back DJ, Blaschke TF, et al. Protein binding in antiretroviral
31 therapies. *AIDS Res Hum Retroviruses* 2003 Sep;19 (9): 825-35
- 32 55. Cropp CD, Yee SW, Giacomini KM. Genetic variation in drug transporters in
33 ethnic populations. *Clin Pharmacol Ther* 2008 Sep;84 (3): 412-6
- 34 56. Minuesa G, Purcet S, Erkizia I, et al. Expression and functionality of anti-
35 human immunodeficiency virus and anticancer drug uptake transporters in immune
36 cells. *J Pharmacol Exp Ther* 2008 Feb;324 (2): 558-67
- 37 57. Kock K, Grube M, Jedlitschky G, et al. Expression of adenosine triphosphate-
38 binding cassette (ABC) drug transporters in peripheral blood cells: relevance for
39 physiology and pharmacotherapy. *Clin Pharmacokinet* 2007;46 (6): 449-70
- 40 58. Pan G, Giri N, Elmquist WF. Abcg2/Bcrp1 mediates the polarized transport of
41 antiretroviral nucleosides abacavir and zidovudine. *Drug Metab Dispos* 2007 Jul;35
42 (7): 1165-73
- 43 59. Weiss J, Rose J, Storch CH, et al. Modulation of human BCRP (ABCG2)
44 activity by anti-HIV drugs. *J Antimicrob Chemother* 2007 Feb;59 (2): 238-45
- 45 60. St-Pierre MV, Ugele B, Gambling L, et al. Mechanisms of drug transfer across
46 the human placenta-a workshop report. *Placenta* 2002 Apr;23 Suppl A: S159-64
- 47 61. Pastor-Anglada M, Cano-Soldado P, Molina-Arcas M, et al. Cell entry and
48 export of nucleoside analogues. *Virus Res* 2005 Feb;107 (2): 151-64

- 1 62. Hoggard PG, Back DJ. Intracellular pharmacology of nucleoside analogues
2 and protease inhibitors: role of transporter molecules. *Curr Opin Infect Dis* 2002
3 Feb;15 (1): 3-8
- 4 63. Strazielle N, Belin MF, Gherzi-Egea JF. Choroid plexus controls brain
5 availability of anti-HIV nucleoside analogs via pharmacologically inhibitable organic
6 anion transporters. *Aids* 2003 Jul 4;17 (10): 1473-85
- 7 64. Izzedine H, Hulot JS, Villard E, et al. Association between ABCC2 gene
8 haplotypes and tenofovir-induced proximal tubulopathy. *J Infect Dis* 2006 Dec 1;194
9 (11): 1481-91
- 10 65. Anderson PL, Zheng JH, King T, et al. Concentrations of zidovudine- and
11 lamivudine-triphosphate according to cell type in HIV-seronegative adults. *Aids* 2007
12 Sep 12;21 (14): 1849-54
- 13 66. Fletcher C, King T, Bushman L, et al. Compartmental kinetics of intracellular
14 tenofovir [abstract n° 754]. Conference on retroviruses and Opportunistic Infections
15 2008 Feb 3-6; Boston (MA). 341
- 16 67. Arner ES, Valentin A, Eriksson S. Thymidine and 3'-azido-3'-deoxythymidine
17 metabolism in human peripheral blood lymphocytes and monocyte-derived
18 macrophages. A study of both anabolic and catabolic pathways. *J Biol Chem* 1992
19 Jun 5;267 (16): 10968-75
- 20 68. Gavegnano C, Fromentin E, RF SchinaziCenter for AIDS Research. Lower
21 levels of nucleoside analog triphosphates in primary human macrophages compared
22 to human lymphocytes could impair potency of antiretroviral drugs in human viral
23 reservoirs. HIV DART; Puerto Rico (United States). 2008 Dec 9-12
- 24 69. Perno CF, Svicher V, Schols D, et al. Therapeutic strategies towards HIV-1
25 infection in macrophages. *Antiviral Res* 2006 Sep;71 (2-3): 293-300
- 26 70. Stormer E, von Moltke LL, Perloff MD, et al. Differential modulation of P-
27 glycoprotein expression and activity by non-nucleoside HIV-1 reverse transcriptase
28 inhibitors in cell culture. *Pharm Res* 2002 Jul;19 (7): 1038-45
- 29 71. Cascorbi I. Role of pharmacogenetics of ATP-binding cassette transporters in
30 the pharmacokinetics of drugs. *Pharmacol Ther* 2006 Nov;112 (2): 457-73
- 31 72. Anderson PL, Lamba J, Aquilante CL, et al. Pharmacogenetic characteristics
32 of indinavir, zidovudine, and lamivudine therapy in HIV-infected adults: a pilot study.
33 *J Acquir Immune Defic Syndr* 2006 Aug 1;42 (4): 441-9
- 34 73. Meaden ER, Hoggard PG, Newton P, et al. P-glycoprotein and MRP1
35 expression and reduced ritonavir and saquinavir accumulation in HIV-infected
36 individuals. *J Antimicrob Chemother* 2002 Oct;50 (4): 583-8
- 37 74. Ho RH, Kim RB. Transporters and drug therapy: implications for drug
38 disposition and disease. *Clin Pharmacol Ther* 2005 Sep;78 (3): 260-77
- 39 75. Telenti A, Zanger UM. Pharmacogenetics of anti-HIV drugs. *Annu Rev*
40 *Pharmacol Toxicol* 2008;48: 227-56
- 41 76. La Porte CJ, Li Y, Beique L, et al. The effect of ABCB1 polymorphism on the
42 pharmacokinetics of saquinavir alone and in combination with ritonavir. *Clin*
43 *Pharmacol Ther* 2007 Oct;82 (4): 389-95
- 44 77. Saitoh A, Singh KK, Powell CA, et al. An MDR1-3435 variant is associated
45 with higher plasma nelfinavir levels and more rapid virologic response in HIV-1
46 infected children. *Aids* 2005 Mar 4;19 (4): 371-80
- 47 78. Ford J, Cornforth D, Hoggard PG, et al. Intracellular and plasma
48 pharmacokinetics of nelfinavir and M8 in HIV-infected patients: relationship with P-
49 glycoprotein expression. *Antivir Ther* 2004 Feb;9 (1): 77-84

- 1 79. Chaillou S, Durant J, Garraffo R, et al. Intracellular concentration of protease
2 inhibitors in HIV-1-infected patients: correlation with MDR-1 gene expression and low
3 dose of ritonavir. *HIV Clin Trials* 2002 Nov-Dec;3 (6): 493-501
- 4 80. Kiser JJ, Aquilante CL, Anderson PL, et al. Clinical and genetic determinants
5 of intracellular tenofovir diphosphate concentrations in HIV-infected patients. *J Acquir*
6 *Immune Defic Syndr* 2008 Mar 1;47 (3): 298-303
- 7 81. Dumond JB, Reddy YS, Troiani L, et al. Differential Extracellular and
8 Intracellular Concentrations of Zidovudine and Lamivudine in Semen and Plasma of
9 HIV-1-Infected Men. *J Acquir Immune Defic Syndr* 2008 Mar 20:
- 10 82. Hawkins T, Veikley W, St Claire RL, 3rd, et al. Intracellular pharmacokinetics
11 of tenofovir diphosphate, carbovir triphosphate, and lamivudine triphosphate in
12 patients receiving triple-nucleoside regimens. *J Acquir Immune Defic Syndr* 2005
13 Aug 1;39 (4): 406-11
- 14 83. Hoggard PG, Lloyd J, Khoo SH, et al. Zidovudine phosphorylation determined
15 sequentially over 12 months in human immunodeficiency virus-infected patients with
16 or without previous exposure to antiretroviral agents. *Antimicrob Agents Chemother*
17 2001 Mar;45 (3): 976-80
- 18 84. Fletcher CV, Kawle SP, Kakuda TN, et al. Zidovudine triphosphate and
19 lamivudine triphosphate concentration-response relationships in HIV-infected
20 persons. *Aids* 2000 Sep 29;14 (14): 2137-44
- 21 85. Pruvost A, Negrodo E, Benech H, et al. Measurement of intracellular
22 didanosine and tenofovir phosphorylated metabolites and possible interaction of the
23 two drugs in human immunodeficiency virus-infected patients. *Antimicrob Agents*
24 *Chemother* 2005 May;49 (5): 1907-14
- 25 86. Sankatsing SU, Hoggard PG, Huitema AD, et al. Effect of mycophenolate
26 mofetil on the pharmacokinetics of antiretroviral drugs and on intracellular nucleoside
27 triphosphate pools. *Clin Pharmacokinet* 2004;43 (12): 823-32
- 28 87. Moore JD, Acosta EP, Johnson VA, et al. Intracellular nucleoside triphosphate
29 concentrations in HIV-infected patients on dual nucleoside reverse transcriptase
30 inhibitor therapy. *Antivir Ther* 2007;12 (6): 981-6
- 31 88. Breilh D, Pellegrin I, Rouzes A, et al. Virological, intracellular and plasma
32 pharmacological parameters predicting response to lopinavir/ritonavir (KALEPHAR
33 study). *Aids* 2004 Jun 18;18 (9): 1305-10
- 34 89. Lamotte C, Landman R, Peytavin G, et al. Once-daily dosing of saquinavir
35 soft-gel capsules and ritonavir combination in HIV-1-infected patients (IMEA015
36 study). *Antivir Ther* 2004 Apr;9 (2): 247-56
- 37 90. Rotger M, Colombo S, Furrer H, et al. Influence of CYP2B6 polymorphism on
38 plasma and intracellular concentrations and toxicity of efavirenz and nevirapine in
39 HIV-infected patients. *Pharmacogenet Genomics* 2005 Jan;15 (1): 1-5
- 40 91. Flynn PM, Rodman J, Lindsey JC, et al. Intracellular pharmacokinetics of once
41 versus twice daily zidovudine and lamivudine in adolescents. *Antimicrob Agents*
42 *Chemother* 2007 Oct;51 (10): 3516-22
- 43 92. Aweeka FT, Rosenkranz SL, Segal Y, et al. The impact of sex and
44 contraceptive therapy on the plasma and intracellular pharmacokinetics of
45 zidovudine. *Aids* 2006 Sep 11;20 (14): 1833-41
- 46 93. Rodman JH, Flynn PM, Robbins B, et al. Systemic pharmacokinetics and
47 cellular pharmacology of zidovudine in human immunodeficiency virus type 1-
48 infected women and newborn infants. *J Infect Dis* 1999 Dec;180 (6): 1844-50
- 49 94. Barry MG, Khoo SH, Veal GJ, et al. The effect of zidovudine dose on the
50 formation of intracellular phosphorylated metabolites. *AIDS* 1996 Oct;10 (12): 1361-7

- 1 95. Kiser JJ, Fletcher CV, Flynn PM, et al. Pharmacokinetics of antiretroviral
2 regimens containing tenofovir disoproxil fumarate and atazanavir-ritonavir in
3 adolescents and young adults with human immunodeficiency virus infection.
4 *Antimicrob Agents Chemother* 2008 Feb;52 (2): 631-7
- 5 96. Djabarouti S, Breilh D, Pellegrin I, et al. Intracellular and plasma efavirenz
6 concentrations in HIV-infected patients switching from successful protease inhibitor-
7 based highly active antiretroviral therapy (HAART) to efavirenz-based HAART
8 (SUSTIPHAR Study). *J Antimicrob Chemother* 2006 Nov;58 (5): 1090-3
- 9 97. Ford J, Boffito M, Wildfire A, et al. Intracellular and plasma pharmacokinetics
10 of saquinavir-ritonavir, administered at 1,600/100 milligrams once daily in human
11 immunodeficiency virus-infected patients. *Antimicrob Agents Chemother* 2004 Jul;48
12 (7): 2388-93
- 13 98. Crommentuyn KM, Mulder JW, Mairuhu AT, et al. The plasma and intracellular
14 steady-state pharmacokinetics of lopinavir/ritonavir in HIV-1-infected patients. *Antivir
15 Ther* 2004 Oct;9 (5): 779-85
- 16 99. Durand-Gasselin L, Pruvost A, Dehee A, et al. High levels of zidovudine (AZT)
17 and its intracellular phosphate metabolites in AZT- and AZT-lamivudine-treated
18 newborns of human immunodeficiency virus-infected mothers. *Antimicrob Agents
19 Chemother* 2008 Jul;52 (7): 2555-63
- 20 100. Wang LH, Begley J, St Claire RL, 3rd, et al. Pharmacokinetic and
21 pharmacodynamic characteristics of emtricitabine support its once daily dosing for
22 the treatment of HIV infection. *AIDS Res Hum Retroviruses* 2004 Nov;20 (11): 1173-
23 82
- 24 101. Harris M, Back D, Kewn S, et al. Intracellular carbovir triphosphate levels in
25 patients taking abacavir once a day. *Aids* 2002 May 24;16 (8): 1196-7
- 26 102. Havlir DV, Tierney C, Friedland GH, et al. In vivo antagonism with zidovudine
27 plus stavudine combination therapy. *J Infect Dis* 2000 Jul;182 (1): 321-5
- 28 103. Hoggard P, Khoo S, Barry M, et al. Intracellular metabolism of zidovudine and
29 stavudine in combination. *J Infect Dis* 1996 Sep;174 (3): 671-2
- 30 104. Hoggard PG, Kewn S, Barry MG, et al. Effects of drugs on 2',3'-dideoxy-2',3'-
31 didehydrothymidine phosphorylation in vitro. *Antimicrob Agents Chemother* 1997
32 Jun;41 (6): 1231-6
- 33 105. Pruvost A, Negredo E, Theodoro F, et al. A pilot pharmacokinetic study in HIV
34 infected patient receiving tenofovir disoproxil fumarate (TDF): Investigation of
35 systemic and intracellular interaction between TDF and abacavir, lamivudine or
36 lopinavir/ritonavir. *Antimicrob Agents Chemother* 2009 Mar 9:
- 37 106. Gallant JE, Rodriguez AE, Weinberg WG, et al. Early virologic nonresponse to
38 tenofovir, abacavir, and lamivudine in HIV-infected antiretroviral-naive subjects. *J
39 Infect Dis* 2005 Dec 1;192 (11): 1921-30
- 40 107. Kiser JJ, Carten ML, Aquilante CL, et al. The effect of lopinavir/ritonavir on the
41 renal clearance of tenofovir in HIV-infected patients. *Clin Pharmacol Ther* 2008
42 Feb;83 (2): 265-72
- 43 108. Hoggard PG, Sales SD, Phiboonbanakit D, et al. Influence of prior exposure to
44 zidovudine on stavudine phosphorylation in vivo and ex vivo. *Antimicrob Agents
45 Chemother* 2001 Feb;45 (2): 577-82
- 46 109. Katlama C, Valantin MA, Matheron S, et al. Efficacy and tolerability of
47 stavudine plus lamivudine in treatment-naive and treatment-experienced patients
48 with HIV-1 infection. *Ann Intern Med* 1998 Oct 1;129 (7): 525-31

- 1 110. Murphy MD, O'Hearn M, Chou S. Fatal lactic acidosis and acute renal failure
2 after addition of tenofovir to an antiretroviral regimen containing didanosine. Clin
3 Infect Dis 2003 Apr 15;36 (8): 1082-5
- 4 111. Masia M, Gutierrez F, Padilla S, et al. Didanosine-associated toxicity: a
5 predictable complication of therapy with tenofovir and didanosine? J Acquir Immune
6 Defic Syndr 2004 Apr 1;35 (4): 427-8
- 7 112. Karrer U, Ledergerber B, Furrer H, et al. Dose-dependent influence of
8 didanosine on immune recovery in HIV-infected patients treated with tenofovir. Aids
9 2005 Nov 18;19 (17): 1987-94
- 10 113. Barreiro P, Soriano V. Suboptimal CD4 gains in HIV-infected patients
11 receiving didanosine plus tenofovir. J Antimicrob Chemother 2006 May;57 (5): 806-9
- 12 114. Perez-Elias MJ, Moreno S, Gutierrez C, et al. High virological failure rate in
13 HIV patients after switching to a regimen with two nucleoside reverse transcriptase
14 inhibitors plus tenofovir. Aids 2005 Apr 29;19 (7): 695-8
- 15 115. Tung MY, Mandalia S, Bower M, et al. The durability of virological success of
16 tenofovir and didanosine dosed at either 400 or 250 mg once daily. HIV Med 2005
17 May;6 (3): 151-4
- 18 116. Holdich T, Shiveley LA, Sawyer J. Effect of Lamivudine on the plasma and
19 intracellular pharmacokinetics of apricitabine, a novel nucleoside reverse
20 transcriptase inhibitor, in healthy volunteers. Antimicrob Agents Chemother 2007
21 Aug;51 (8): 2943-7
- 22 117. Frank I, Bosch RJ, Fiscus S, et al. Activity, safety, and immunological effects
23 of hydroxyurea added to didanosine in antiretroviral-naive and experienced HIV type
24 1-infected subjects: a randomized, placebo-controlled trial, ACTG 307. AIDS Res
25 Hum Retroviruses 2004 Sep;20 (9): 916-26
- 26 118. Bakshi RP, Hamzeh F, Frank I, et al. Effect of hydroxyurea and dideoxyinosine
27 on intracellular 3'-deoxyadenosine-5'-triphosphate concentrations in HIV-infected
28 patients. AIDS Res Hum Retroviruses 2007 Nov;23 (11): 1360-5
- 29 119. Margolis D, Heredia A, Gaywee J, et al. Abacavir and mycophenolic acid, an
30 inhibitor of inosine monophosphate dehydrogenase, have profound and synergistic
31 anti-HIV activity. J Acquir Immune Defic Syndr 1999 Aug 15;21 (5): 362-70
- 32 120. Dixit NM, Perelson AS. The metabolism, pharmacokinetics and mechanisms
33 of antiviral activity of ribavirin against hepatitis C virus. Cell Mol Life Sci 2006 Apr;63
34 (7-8): 832-42
- 35 121. Rodriguez-Torres M, Torriani FJ, Soriano V, et al. Effect of ribavirin on
36 intracellular and plasma pharmacokinetics of nucleoside reverse transcriptase
37 inhibitors in patients with human immunodeficiency virus-hepatitis C virus coinfection:
38 results of a randomized clinical study. Antimicrob Agents Chemother 2005 Oct;49
39 (10): 3997-4008
- 40 122. Aweeka FT, Kang M, Yu JY, et al. Pharmacokinetic evaluation of the effects of
41 ribavirin on zidovudine triphosphate formation: ACTG 5092s Study Team. HIV Med
42 2007 Jul;8 (5): 288-94
- 43 123. Sim SM, Hoggard PG, Sales SD, et al. Effect of ribavirin on zidovudine
44 efficacy and toxicity in vitro: a concentration-dependent interaction. AIDS Res Hum
45 Retroviruses 1998 Dec 20;14 (18): 1661-7
- 46 124. Vogt MW, Hartshorn KL, Furman PA, et al. Ribavirin antagonizes the effect of
47 azidothymidine on HIV replication. Science 1987 Mar 13;235 (4794): 1376-9
- 48 125. Balzarini J, Lee CK, Herdewijn P, et al. Mechanism of the potentiating effect of
49 ribavirin on the activity of 2',3'-dideoxyinosine against human immunodeficiency virus.
50 J Biol Chem 1991 Nov 15;266 (32): 21509-14

- 1 126. Bani-Sadr F, Carrat F, Pol S, et al. Risk factors for symptomatic mitochondrial
2 toxicity in HIV/hepatitis C virus-coinfected patients during interferon plus ribavirin-
3 based therapy. *J Acquir Immune Defic Syndr* 2005 Sep 1;40 (1): 47-52
- 4 127. Moreno A, Quereda C, Moreno L, et al. High rate of didanosine-related
5 mitochondrial toxicity in HIV/HCV-coinfected patients receiving ribavirin. *Antivir Ther*
6 2004 Feb;9 (1): 133-8
- 7 128. Ford J, Boffito M, Maitland D, et al. Influence of atazanavir 200 mg on the
8 intracellular and plasma pharmacokinetics of saquinavir and ritonavir 1600/100 mg
9 administered once daily in HIV-infected patients. *J Antimicrob Chemother* 2006
10 Nov;58 (5): 1009-16
- 11 129. DiCenzo R, Frerichs V, Larpanichpoonphol P, et al. Effect of quercetin on the
12 plasma and intracellular concentrations of saquinavir in healthy adults.
13 *Pharmacotherapy* 2006 Sep;26 (9): 1255-61
- 14 130. Khoo SH, Hoggard PG, Williams I, et al. Intracellular accumulation of human
15 immunodeficiency virus protease inhibitors. *Antimicrob Agents Chemother* 2002
16 Oct;46 (10): 3228-35
- 17 131. Vispo E, Barreiro P, Pineda JA, et al. Low response to pegylated interferon
18 plus ribavirin in HIV-infected patients with chronic hepatitis C treated with abacavir.
19 *Antivir Ther* 2008;13 (3): 429-37
- 20 132. Stretcher BN, Pesce AJ, Frame PT, et al. Correlates of zidovudine
21 phosphorylation with markers of HIV disease progression and drug toxicity. *Aids*
22 1994 Jun;8 (6): 763-9
- 23 133. Back DJ, Burger DM, Flexner CW, et al. The pharmacology of antiretroviral
24 nucleoside and nucleotide reverse transcriptase inhibitors: implications for once-daily
25 dosing. *J Acquir Immune Defic Syndr* 2005 Aug 1;39 Suppl 1: S1-23, quiz S4-5
- 26 134. Fletcher CV, Anderson PL, Kakuda TN, et al. Concentration-controlled
27 compared with conventional antiretroviral therapy for HIV infection. *Aids* 2002 Mar
28 8;16 (4): 551-60
- 29 135. Drechsler H, Powderly WG. Switching effective antiretroviral therapy: a review.
30 *Clin Infect Dis* 2002 Nov 15;35 (10): 1219-30
- 31 136. Kelly M. Induction-maintenance antiretroviral strategies to reduce long-term
32 toxicity. *J HIV Ther* 2003 Feb;8 (1): 11-4
- 33 137. Maggiolo F, Airoidi M, Kleinloog HD, et al. Effect of adherence to HAART on
34 virologic outcome and on the selection of resistance-conferring mutations in NNRTI-
35 or PI-treated patients. *HIV Clin Trials* 2007 Sep-Oct;8 (5): 282-92
- 36 138. Harrigan PR, Hogg RS, Dong WW, et al. Predictors of HIV drug-resistance
37 mutations in a large antiretroviral-naïve cohort initiating triple antiretroviral therapy. *J*
38 *Infect Dis* 2005 Feb 1;191 (3): 339-47
- 39 139. Braithwaite RS, Shechter S, Roberts MS, et al. Explaining variability in the
40 relationship between antiretroviral adherence and HIV mutation accumulation. *J*
41 *Antimicrob Chemother* 2006 Nov;58 (5): 1036-43
- 42 140. Calza L, Manfredi R, Chiodo F. Dyslipidaemia associated with antiretroviral
43 therapy in HIV-infected patients. *J Antimicrob Chemother* 2004 Jan;53 (1): 10-4
- 44 141. Bodasing N, Fox R. HIV-associated lipodystrophy syndrome: description and
45 pathogenesis. *J Infect* 2003 Apr;46 (3): 149-54
- 46 142. Perez-Molina JA, Domingo P, Martinez E, et al. The role of efavirenz
47 compared with protease inhibitors in the body fat changes associated with highly
48 active antiretroviral therapy. *J Antimicrob Chemother* 2008 Aug;62 (2): 234-45

- 1 143. Mallewa JE, Wilkins E, Vilar J, et al. HIV-associated lipodystrophy: a review of
2 underlying mechanisms and therapeutic options. *J Antimicrob Chemother* 2008 Jun
3 18:
- 4 144. Nolan D, Mallal S. Complications associated with NRTI therapy: update on
5 clinical features and possible pathogenic mechanisms. *Antivir Ther* 2004 Dec;9 (6):
6 849-63
- 7 145. Dragovic G, Jevtovic D. Nucleoside reverse transcriptase inhibitor usage and
8 the incidence of peripheral neuropathy in HIV/AIDS patients. *Antivir Chem*
9 *Chemother* 2003 Sep;14 (5): 281-4
- 10 146. Sinnwell TM, Sivakumar K, Soueidan S, et al. Metabolic abnormalities in
11 skeletal muscle of patients receiving zidovudine therapy observed by ³¹P in vivo
12 magnetic resonance spectroscopy. *J Clin Invest* 1995 Jul;96 (1): 126-31
- 13 147. Yarchoan R, Pluda JM, Thomas RV, et al. Long-term toxicity/activity profile of
14 2',3'-dideoxyinosine in AIDS or AIDS-related complex. *Lancet* 1990 Sep 1;336
15 (8714): 526-9
- 16 148. Brogan KL, Zell SC. Hematologic toxicity of zidovudine in HIV-infected
17 patients. *Am Fam Physician* 1990 May;41 (5): 1521-8
- 18 149. Gupta SK. Tenofovir-associated Fanconi syndrome: review of the FDA
19 adverse event reporting system. *AIDS Patient Care STDS* 2008 Feb;22 (2): 99-103
- 20 150. Clifford DB, Evans S, Yang Y, et al. Impact of efavirenz on neuropsychological
21 performance and symptoms in HIV-infected individuals. *Ann Intern Med* 2005 Nov
22 15;143 (10): 714-21
- 23 151. Martin-Carbonero L, Nunez M, Gonzalez-Lahoz J, et al. Incidence of liver
24 injury after beginning antiretroviral therapy with efavirenz or nevirapine. *HIV Clin*
25 *Trials* 2003 Mar-Apr;4 (2): 115-20
- 26 152. Patel SM, Johnson S, Belknap SM, et al. Serious adverse cutaneous and
27 hepatic toxicities associated with nevirapine use by non-HIV-infected individuals. *J*
28 *Acquir Immune Defic Syndr* 2004 Feb 1;35 (2): 120-5
- 29 153. Knobel H, Guelar A, Montero M, et al. Risk of side effects associated with the
30 use of nevirapine in treatment-naive patients, with respect to gender and CD4 cell
31 count. *HIV Med* 2008 Jan;9 (1): 14-8
- 32 154. Carr A, Samaras K, Chisholm DJ, et al. Pathogenesis of HIV-1-protease
33 inhibitor-associated peripheral lipodystrophy, hyperlipidaemia, and insulin resistance.
34 *Lancet* 1998 Jun 20;351 (9119): 1881-3
- 35 155. Andre P, Groettrup M, Klenerman P, et al. An inhibitor of HIV-1 protease
36 modulates proteasome activity, antigen presentation, and T cell responses. *Proc Natl*
37 *Acad Sci U S A* 1998 Oct 27;95 (22): 13120-4
- 38 156. Sakai J, Rawson RB. The sterol regulatory element-binding protein pathway:
39 control of lipid homeostasis through regulated intracellular transport. *Curr Opin*
40 *Lipidol* 2001 Jun;12 (3): 261-6
- 41 157. Murata H, Hruz PW, Mueckler M. The mechanism of insulin resistance caused
42 by HIV protease inhibitor therapy. *J Biol Chem* 2000 Jul 7;275 (27): 20251-4
- 43 158. Moyle G. Mechanisms of HIV and nucleoside reverse transcriptase inhibitor
44 injury to mitochondria. *Antivir Ther* 2005;10 Suppl 2: M47-52
- 45 159. Capparelli EV, Englund JA, Connor JD, et al. Population pharmacokinetics
46 and pharmacodynamics of zidovudine in HIV-infected infants and children. *J Clin*
47 *Pharmacol* 2003 Feb;43 (2): 133-40
- 48 160. Mentré F, Escolano S, Diquet B, et al. Clinical pharmacokinetics of zidovudine:
49 inter and intraindividual variability and relationship to long term efficacy and toxicity.
50 *Eur J Clin Pharmacol* 1993;45 (5): 397-407

- 1 161. Sales SD, Hoggard PG, Sunderland D, et al. Zidovudine phosphorylation and
2 mitochondrial toxicity in vitro. *Toxicol Appl Pharmacol* 2001 Nov 15;177 (1): 54-8
- 3 162. Csajka C, Marzolini C, Fattinger K, et al. Population pharmacokinetics and
4 effects of efavirenz in patients with human immunodeficiency virus infection. *Clin*
5 *Pharmacol Ther* 2003 Jan;73 (1): 20-30
- 6 163. Gutierrez F, Navarro A, Padilla S, et al. Prediction of neuropsychiatric adverse
7 events associated with long-term efavirenz therapy, using plasma drug level
8 monitoring. *Clin Infect Dis* 2005 Dec 1;41 (11): 1648-53
- 9
- 10

1 **Tables**2
3**Table I.** Summary of pharmacokinetics parameters of available antiretroviral drugs adapted from [6, 8-13]

		Bioavailability (%)	t _{max} (h)	Protein Binding (%)	Elimination Pathway	Plasma t _{1/2} (h)	Intracellular t _{1/2} (h)
Entry Inhibitors							
Enfuvirtide	T20	70 (SC)	7	97	Peptidases -> amino acids	3 - 8	
Maraviroc		25-35%	2	76	25% renal + CYP3A	13	
Nucleosidic Reverse Transcriptase Inhibitors (NRTI)							
Zidovudine	ZDV	60	1	20	20% renal + 80% glucuronidation UGT2B7	1 - 1.5	7-11
Didanosine	ddl	40	1	< 5	50% renal	1-2	15 - 20
Stavudine	d4T	80	1	< 5	80% renal	1 - 1.5	7
Lamivudine	3TC	80	1	< 5	80% renal	2-3	22
Abacavir	ABC	75	1	49	< 5% renal + liver biotransformation	0,8 - 1,5	21
Tenofovir	TFV	40	2-3	< 10	80% renal	14	150-180
Emtricitabine	FTC	90	1	< 5	80% renal	9	39
Non Nucleosidic Reverse Transcriptase Inhibitors (NNRTI)							
Efavirenz	EFV	50	2-5	99.5	< 1% renal + CYP2B6	50	
Nevirapine	NVP	90	4	60	< 15% renal + CYP2B6+3A4	25-30	
Integrase Inhibitor							
Raltegravir		ND	3	83	< 5% renal + UGT1A1	9	
Protease Inhibitors (PI)							
Saquinavir	SQV	4-10	1-2	97	< 5% renal + CYP3A	5	
Indinavir	IDV	60	1	60	10% renal + CYP3A	1.5 - 2	2
Ritonavir	RTV	70	3	99	< 5% renal + CYP3A	3-5	
Nelfinavir	NFV	60-80	3	98	<5% renal + CYP3A	5-6	
Lopinavir/r	LPV/r	ND	5	99	< 5% renal + CYP3A	5-6	
Amprenavir	APV	30-90	2	90	< 5% renal + CYP3A	7-12	
Atazanavir	ATV	ND	2	86	< 10% renal + CYP3A	7	
Darunavir	DRV	ND	1-4	94	< 5% renal + CYP3A	10 -15	
Tipranavir	TPV	ND	3	99	< 5% renal + CYP3A	6 (single dose)	

4 **Abbreviations:** SC: subcutaneous administration; r = ritonavir low-dose; CYP = P450 cytochrome; UGT=UDP-glucuronosyltransferase

5

Table II. Intracellular pharmacokinetics of antiretrovirals and relationship with plasma pharmacokinetics

Drug	Dose ^a	Design			Plasma and intracellular pharmacokinetics				Correlation	Ref
		N	Plasma (n / v)	Intracellular (n / v)	Parameters ^b	Plasma	Intracellular	Ratio ^c		
NRTI										
Zidovudine (ZDV)										
8 mg / kg / day	49	1 / 2	1 / 2	C _{trough}	0.00 – 1.29 µg/mL	16 – 385 fmol/10 ⁶ cells	NR	Yes (R ² = NR)	[99]	
300 mg bid	14	8 / 1	3 / 3	AUC _{0-12h}	1.8 – 1.7 h.µg/mL	1241 – 2172 h.fmol/10 ⁶ cells	NR	No	[81]	
300 mg bid	15	2 / 2	2 / 2	C _{1h}	0.01 – 0.96 µg/mL	4 – 53 fmol/10 ⁶ cells	9 10 ⁻⁶ – 4.2 10 ⁻³	NR	[87]	
				C _{4h}	0.01 – 1.08 µg/mL	5.5 – 50.7 fmol/10 ⁶ cells	9 10 ⁻⁶ – 3.4 10 ⁻³	NR		
300 mg bid	26	-	5 / 1	C _{trough}	-	7.7 – 23.6 fmol/10 ⁶ cells	-	-	[91]	
600 mg qd				C _{trough}	-	6.4 – 12.9 fmol/10 ⁶ cells	-	-		
	27			C _{24h}	-	4.9 – 13.2 fmol/10 ⁶ cells	-	-		
200 mg tid	38	7 / 2	4 / 2	AUC _{0-8h}					[92]	
				Women	0.82 – 1.34 h.µg/mL	240 – 670 h.fmol/10 ⁶ cells	NR	NR		
				Men	0.88 – 1.35 h.µg/mL	520 – 1150 h.fmol/10 ⁶ cells	NR	NR		
300 mg tid	33	-	2 / 3	C _{trough} – C _{2h}	-	35 – 64 fmol/10 ⁶ cells	-	-	[72]	
300 mg bid	23	3 / 7	3 / 7	AUC _{0-2h}			NR	Yes (R ² = NR)	[83]	
				Week 0	0.99 ± 0.71 h.µg/mL	110 ± 80 h.fmol/10 ⁶ cells				
				Week 12	4.21 ± 1.52 h.µg/mL	130 ± 110 h.fmol/10 ⁶ cells				
				Week 48	1.12 ± 0.41 h.µg/mL	130 ± 120 h.fmol/10 ⁶ cells				
200 / 300 mg bid ^d	8	1-2 / 12	1-2 / 12	C _{2h} – C _{8h}	0.18 – 0.32 µg/mL	13.8 – 96.4 fmol/10 ⁶ cells	NR	No	[84]	
				CL/F	1.16 – 2.53 L/h/kg	NR	-	-		

iv 1 / 2 mg / kg / h ^e	28 ^f	>1 / 1	>1 / 1	C _{delivery}	0.19 – 3.66 µg/mL	11 – 127 fmol/10 ⁶ cells	NR	No	[93]
				CL/F	0.07 – 0.89 L/h/kg	NR	-	-	
100 mg qd	10	6 / 1	6 / 1	AUC _{0-12h}	0.38 ± 0.13 h.µg/mL	420 ± 420 fmol/10 ⁶ cells	NR	No	[94]
300 mg bid				AUC _{0-12h}	1.22 ± 0.21 h.µg/mL	610 ± 810 fmol/10 ⁶ cells			
500 mg qd	21	6 / 2	6 / 2	AUC _{0-8h}			NR	No	[12]
				Week 4	0.71 ± 0.31 h.µg/mL	3290 ± 970 h.fmol/10 ⁶ cells			
				Week ≥24	0.79 ± 0.41 h.µg/mL	2160 ± 1090 h.fmol/10 ⁶ cells			
500 mg qd	6	6 / 1	6 / 1	AUC _{0-8h}	0.74 ± 0.27 h.µg/mL	4200 ± 2720 h.fmol/10 ⁶ cells	NR	No	[13]
Didanosine (ddl)									
400 mg qd	16	-	4 / 1	C _{trough} – C _{4h}	-	3.8 – 13.3 fmol/10 ⁶ cells	-	-	[85]
400 / 250 mg bid	28	1 / 1	1 / 1	C _{2.5h} – C _{28.5h}	0.00 – 0.16 µg/mL	0 – 23 fmol/10 ⁶ cells	NR	No	[11]
Stavudine (d4T)									
40 mg bid	19	2 / 2	2 / 2	C _{1h}	0.04 – 1.39 µg/mL	3 – 25 fmol/10 ⁶ cells	5 10 ⁻⁶ – 6.5 10 ⁻⁵	NR	[87]
				C _{4h}	0.04 – 0.73 µg/mL	3.2 – 18.5 fmol/10 ⁶ cells	7 10 ⁻⁶ – 2.4 10 ⁻⁴	NR	
40 / 30 mg tid	28	1 / 1	1 / 1	C _{2.5h} – C _{28.5h}	0.04 – 0.67 µg/mL	0 – 99 fmol/10 ⁶ cells	NR	Yes (R ² = 0.46)	[11]
Lamivudine (3TC)									
300 mg qd	15	4 / 1	4 / 1	AUC _{0-24h}	3.71 – 7.14 h.µg/mL	26910 – 80810 h.fmol/10 ⁶ cells	-	-	[105]
				C _{max}	1.66 – 2.59 µg/mL	-	-	-	
				C _{trough}	0.055 – 0.18 µg/mL	6000 – 11460 fmol/10 ⁶ cells	-	-	
4 mg / kg / day	49	1 / 2	1 / 2	C _{trough}	0.00 – 1.16 µg/mL	570 – 38900 fmol/10 ⁶ cells	NR	Yes (R ² = NR)	[99]
150 mg bid	14	8 / 1	3 / 3	AUC _{0-12h}	4.3 – 6.2 h.µg/mL	78.6 – 164.9 h.fmol/10 ⁶ cells	NR	NR	[81]
150 mg bid	41	2 / 2	2 / 2	C _{1h}	0.06 – 1.42 µg/mL	42 – 4579 fmol/10 ⁶ cells	0.01 – 1.17	NR	[87]
				C _{4h}	0.02 – 1.81 µg/mL	200 – 10730 fmol/10 ⁶ cells	0.06 – 41.58	NR	

300 mg qd	25	-	4 / 1	C _{trough}	-	0.7 10 ⁶ – 2.4 10 ⁶ fmol/10 ⁶ cells	-	-	[91]
				C _{24h}	-	1.0 10 ⁶ – 2.5 10 ⁶ fmol/10 ⁶ cells	-	-	
150 mg bid	27			C _{trough}	-	0.7 10 ⁶ – 4.0 10 ⁶ fmol/10 ⁶ cells	-	-	
300 mg qd	13	-	1-2 / 6	C _{trough}	-	7060 – 11600 fmol/10 ⁶ cells	-	-	[82]
150 mg bid	32	-	2 / 3	C _{trough} – C _{2h}	-	7252 – 9313 fmol/10 ⁶ cells	-	-	[72]
150 mg bid ^e	8	1-2 / 12	1-2 / 12	C _{2h} – C _{8h}	0.43 – 0.69 µg/mL	2352 – 13024 fmol/10 ⁶ cells	NR	Yes	[84]
				CL/F	0.30 – 0.53 µg/mL	NR	-	(R ² = 0.66)	
Abacavir (ABC)									
300 mg bid	12	4 / 1	4 / 1	AUC _{0-24h}	2.25 – 7.80 h.µg/mL	656 – 2234 h.fmol/10 ⁶ cells	-	-	[105]
				C _{max}	0.95 – 3.72 µg/mL	-	-	-	
				C _{trough}	0.03 – 0.18 µg/mL	98.3 – 472.8 fmol/10 ⁶ cells	-	-	
600 mg qd	8	-	1-2 / 6	C _{trough}	-	88 – 200 fmol/10 ⁶ cells	-	-	[82]
300 mg bid	9	8 / 1	4 / 1	C _{trough}	NQ	NQ	-	-	[86]
				C _{max}	1.48 – 2.35 µg/mL	-	-	-	
600 mg qd	5	-	8 / 1	C _{trough}	-	127 – 575 fmol/10 ⁶ cells	-	-	[101]
				C _{1h}	-	107 – 670 fmol/10 ⁶ cells	-	-	
				C _{12h}	-	188 fmol/10 ⁶ cells	-	-	
				C _{14h} – C _{16h}	-	101 – 548 fmol/10 ⁶ cells	-	-	
				C _{18h}	-	113 – 648 fmol/10 ⁶ cells	-	-	
				C _{20h}	-	80 – 660 fmol/10 ⁶ cells	-	-	
				C _{22h}	-	105 – 201 fmol/10 ⁶ cells	-	-	
				C _{24h}	-	62 – 354 fmol/10 ⁶ cells	-	-	
Tenofovir (TDF)									
300 mg qd	27	4 / 1	4 / 1	AUC _{0-24h}	0.45 – 1.34 h.µg/mL	476.4 – 1386 h.fmol/10 ⁶ cells	-	-	[105]

				C_{max}	0.19 – 0.45 µg/mL	-	-		
				C_{trough}	0.03 – 0.119 µg/mL	116.5 – 376.5 fmol/10 ⁶ cells	-		
300 mg qd	7	-	1-2 / 6	C_{trough}	-	85 – 110 fmol/10 ⁶ cells	-	-	[82]
300 mg qd	8	-	4 / 1	$C_{trough} - C_{4h}$	-	129 – 373 fmol/10 ⁶ cells	-	-	[85]
300 mg qd	22	7 / 1	3 / 1	AUC_{0-24h}	0.002 – 0.003 h.µg/mL	NR	NR	No	[95]
				C_{1h}	NR	71 – 130 fmol/10 ⁶ cells	-	-	
				C_{4h}	NR	68 – 130 fmol/10 ⁶ cells	-	-	
				C_{24h}	0.052 – 0.068 h.µg/mL	70 – 123 fmol/10 ⁶ cells	NR	NR	

Emtricitabine (FTC)

25 mg bid	9	8 / 1	5-2 / 2	C_{1h}	NR	0 – 1125 fmol/10 ⁶ cells	-	-	[10,
				C_{4h}	NR	0 – 1450 fmol/10 ⁶ cells	-	-	100]
200 mg qd	8			C_{1h}	NR	0 – 2125 fmol/10 ⁶ cells	-	-	
				C_{4h}	NR	0 – 2250 fmol/10 ⁶ cells	-	-	
100 mg bid	8			C_{1h}	NR	0 – 4250 fmol/10 ⁶ cells	-	-	
				C_{4h}	NR	0 – 4625 fmol/10 ⁶ cells	-	-	
100 mg qd	8			C_{1h}	NR	0 – 2675 fmol/10 ⁶ cells	-	-	
				C_{4h}	NR	0 – 4000 fmol/10 ⁶ cells	-	-	
200 mg bid	8			C_{1h}	NR	0 – 4000 fmol/10 ⁶ cells	-	-	
				C_{4h}	NR	0 – 4100 fmol/10 ⁶ cells	-	-	

NNRTI

Efavirenz (EFV)

600 mg qd	49	1 / 2	1 / 2	C_{12h}					[96]
				Week 4	1.6 – 3.1 µg/mL	2.8 – 11.5 µg/mL	2.2 – 5.4	No	
				Week 24	1.4 – 2.5 µg/mL	3.9 – 8.8 µg/mL	0.5 – 1.8	No	

600 mg qd	10	5 / 1	5 / 1	AUC _{0-24h}	36.8 – 131.9 h.µg/mL	29.8 – 176.9 h.fmol/10 ⁶ cells	0.7 – 3.3	Yes (R ² = 0.59)	[34]
600 mg qd	55	1 / 1	1 / 1	AUC	7.9 – 63.1 h.µg/mL	6.3 – 794.3 h.µg/mL	NR	Yes (R ² = 0.24)	[90]
Nevirapine (NVP)									
200 mg bid	10	5 / 1	5 / 1	AUC _{0-12h}	0.05 – 0.09 h.µg/mL	0.05 – 2.9 h.µg/mL	0 – 0.05	Yes (R ² = 0.62)	[30]
400 mg bid	10	1 / 1	1 / 1	AUC	39.8 – 398.1 h.µg/mL	0.82 – 46.0 h.µg/mL	NR	NR	[90]
PI									
Saquinavir (SQV)									
1600 mg qd (+100 mg RTV)	12	4 / 1	4 / 1	AUC _{0-24h}	5.7 – 39.3 h.µg/mL	24.7 – 114.6 h.µg/mL	1.5 – 6.7	Yes (R ² = 0.63)	[97]
1600 mg qd (+100 mg RTV)	13	1 / 3	1 / 3	C _{24h}			1.1 – 8.7	Yes (R ² = 0.31)	[89]
				Week 2	0.04 – 1.43 µg/mL	0.15 – 0.79 µg/mL			
				Week 4	0.04 – 1.97 µg/mL	0.11 – 0.84 µg/mL			
600 mg tid (+100 mg RTV bid)	9	2 / 1	2 / 1	C _{trough}	NR	NR	0 – 50	NR	[79]
				C _{max}	NR	NR	0 – 20	NR	
Indinavir (IDV)									
800 mg tid	10	6 / 1	6 / 1	AUC _{0-8h}	25.1 ± 4.2 h.µg/mL	7.6 ± 1.0 h.µg/mL	NR	No	[9]
800 mg bid (+100 mg RTV)	19	2 / 1	2 / 1	C _{trough}	10.7 ± 1.3 µg/mL	3.2 ± 0.7 µg/mL	NR		[79]
				C _{trough}	NR	NR	0 – 20	NR	
				C _{max}	NR	NR	1 – 5	NR	
400 mg bid (+400 mg RTV)				C _{trough}	NR	NR	2.5 – 75	NR	
				C _{max}	NR	NR	2.5 – 13	NR	
800 mg tid				C _{trough}	NR	NR	0 – 0	NR	
				C _{max}	NR	NR	0 – 7.5	NR	

Ritonavir (RTV)										
100 mg bid	11	14 / 1	4 / 1	AUC _{0-12h}	2.7 – 4.1 h.µg/mL	9.1 – 14.2 h.µg/mL	3.2 – 7.7	NR	[98]	
100 mg qd	12	4 / 1	4 / 1	AUC _{0-24h}	1.5 – 14.6 h.µg/mL	3.2 – 13.7 h.µg/mL	0.8 – 4.2	No	[97]	
400 / 100 mg bid	22	2 / 1	2 / 1	C _{trough}	NR	NR	0 – 3.1	NR	[79]	
Nelfinavir (NVF)										
1250 mg bid	12	5 / 1	5 / 1	AUC _{0-12h}	5.6 – 50.8 h.µg/mL	5.1 – 60.8 h.µg/mL	NR	Yes (R ² = 0.45)	[78]	
750 mg tid	12	2 / 1	2 / 1	C _{trough}	NR	NR	NR	Yes	[79]	
				C _{trough}	NR	NR	0 – 37.5	NR		
				C _{max}	NR	NR	0 – 11.3	NR		
Lopinavir (LPV)										
400 / 533 mg bid (+100 / 133 mg RTV)	38	1 / 2	1 / 2	C _{trough} Week 4	0.7 – 5.7 µg/mL	2.9 – 29.0 µg/mL	1.9 – 3.8	Yes (R ² = 0.72)	[88]	
400 mg bid (+100 mg RTV)	11	14 / 1	4 / 1	AUC _{0-12h}	61.8 – 82.8 h.µg/mL	63.1 – 113.8 h.µg/mL	0.7 – 2.1	NR	[98]	

1
2
3
4
5
6
7
8
9

^a All studies are performed at steady state

^b Values of the pharmacokinetics parameters as published in the original article: range, IQR or mean ± SD

^c Ratio defined by intracellular/plasma concentration

^d Individualized regimen after the second visit

^e Followed by a continuous infusion of 1 mg/kg/h until delivery

^f Pregnant women

Abbreviations: AUC = area under the concentration time curve; bid = twice a day; C_{max} = maximum concentration; CL/F = clearance; C_{trough} = trough concentration; NNRTI = Non nucleoside analog inhibitors of reverse transcriptase; N = number of patients; n / v = number of samples per visit / number of visits; NRTI = Nucleoside

1 and nucleotide analog inhibitors of reverse transcriptase; NR = not reported in the article; NQ = not quantifiable; PI = protease inhibitors; qd = once a day; tid = three
2 times a day.
3

1 **Table III.** Relationships between intracellular concentrations and efficacy of antiretroviral agents in patients

2

Study	Primary objective (yes/no)	Intracellular moieties	Dosage regimen	Patients	Studied parameters from intracellular concentrations	Efficacy criterion	Results ^a
Type of trial							
NRTI							
Moore et al. ^[87]	Yes Substudy of clinical trial	3TC-TP	3TC 150 mg + ZDV 300 mg bid or 3TC 150 mg + d4T 40 mg bid	39 naïve	W28: Average of 1h and 4h conc. post dose	Change between W0 and W24 in: (1) Log Plasma HIV RNA (2) CD4 cells	(1) P < 0.02 (2) NS
		ZDV-TP	ZDV 300 mg + 3TC 150 mg bid	10 naïve			(1) P < 0.02 (2) NS
		d4T-TP	d4T 40 mg + 3TC 150 mg bid	15 naïve			(1) NS (2) NS
Aweeka et al. ^[122]	No Cross-sectional analysis	ZDV -TP	Any regimen containing ZDV	13 HCV or HBV co-infected patients with stable regimen > 4 weeks	AUC (NCA) from 5 samples: pre dosing and 1,4 ,6 & 8h post dosing	CD4 cell count at time of pharmacokinetic sampling	NS
Anderson et al. ^[8]	Yes Substudy of clinical trial	ZDV-TP	ZDV 300 mg bid + 3TC 150 mg bid + IDV 800 mg tid or Concentration-Controlled ZDV-3TC-IDV regimen	33 naïve	Median conc. above threshold (yes/no) from samples 2h post dose at W2, 28 and 56 and at 2 to 8 h post dose at 9 visits from W8 to W80	(1) Time to reach less than 50cp/mL of HIV RNA (2) Undetectable HIV RNA (< 50 cp/mL) at W24 and W52 (3) Time to loss of virological response in patients achieving undetectable (4) CD4 level at W24 and W48	(1) P = 0.01 (2) W24: P = 0.009 W52: NS (3) P = 0.02 (4) NS
		3TC-TP					Thresholds: ZDV-TP: 30 fmol/10 ⁶ 3TC-TP: 7017 fmol/10 ⁶

Fletcher et al. ^[84]	Yes Substudy of clinical trial	ZDV-TP 3TC-TP	ZDV 300 mg bid + 3TC 150 mg bid + IDV 800 mg tid or Concentration-Controlled ZDV-3TC-IDV regimen	8 naïve	Median conc. from samples 2h post dose at W2, 28 and 56 and at 2 to 8 h post dose at 9 visits from W8 to W80	Change between W0 and W24 in (1) log HIV RNA (2) CD4 cell	(1) P = 0.03 (2) P = 0.001 (1) P = 0.003 (2) NS
Stretcher et al. ^[132]	Yes PK trial	ZDV-P	ZDV only: 800 mg /day 100mg every 4h while awake	21 naïve of ZDV	AUC (NCA) from 6 samples: pre dosing and 1, 2, 4, 6 & 8h post dosing at W4 and >W24	Change between W0 and W4 or W0 and W24 of (1) % CD4 (2) CD4 / CD8 ratio	(1) W4: P = 0.029 W24: NS (2) W4: P = 0.028 W24: NS
PI							
Lamotte et al. ^[89]	No Secondary objective of clinical trial	SQV	SQV1600 mg + RTV 100 mg qd + 2 or 3 NRTI/NNRTI	13 naïve	C _{trough} (24h post dose) at W2, W4 or W12	Change in Plasma HIV RNA between W0 and W12	NS for all dates of trough measurements
Breilh et al. ^[88]	Yes Observational study	LPV	LPV 400 mg + RTV 100 mg bid + 2 or 3 NRTI/NNRTI	38 naïve of LPV	C _{trough} (12h post dose) at W4 and W24	Virological success: (1) HIV RNA < 50 cp/mL before W4 (2) HIV RNA < 50 cp/mL before W4 and during all follow up until W24	(1) P < 0.0002 (2) P < 0.002
Chaillou et al. ^[79]	No Secondary objective Cross-sectional trial	NFV, IDV, APV, SQV, RTV	NFV 750 mg tid or IDV 800 mg tid or APV 1200 mg bid or IDV 800 mg + RTV 100 mg bid or SQV 600mg + RTV 100mg bid or APV 600 mg + RTV 100 mg bid or IDV 400 mg + RTV 400 mg bid + 2 or 3 NRTI/NNRTI	49 experienced patients	Ratio of intracellular to plasma C _{trough} and C _{max} (1.5 to 3 h post dose) at day of study: (1) main PI (2) RTV	Undetectable HIV RNA (<40 cp/mL) at day of study	(1) NS for PI ratio (2) P = 0.04 for presence of intracellular RTV P = 0.029 with RTV ratio in 28 patients receiving RTV

1
2^a Significant results always associated better efficacy with higher intracellular concentrations

1 **Abbreviations:** APV = amprenavir; AUC = area under the concentration time curve; bid = twice a day; C_{max} = maximum concentration; C_{trough} = trough
2 concentration; d4T = stavudine; d4T-TP = stavudine triphosphate; IDV = indinavir; LPV = lopinavir ; NCA= non compartmental analysis; NFV = nelfinavir;
3 NRTI = Nucleoside and nucleotide analog inhibitors of reverse transcriptase; NNRTI = Non nucleoside analog inhibitors of reverse transcriptase; NS = non-
4 significant ; PI = protease inhibitors; qd = once a day ; PK = pharmacokinetic; RTV = ritonavir ; SQV = saquinavir; tid = three times a day; ZDV = zidovudine;
5 ZDV-P= total zidovudine phosphates; ZDV-TP = zidovudine triphosphate; 3TC = lamivudine ; 3TC -TP= lamivudine triphosphate; W = week.

6

7

1
2

Table IV. Relationships between intracellular concentrations and toxicity of antiretroviral agents in patients

Study	Primary objective	Intracellular moieties	Dosage regimen	Patients	Studied parameters from intracellular concentrations	Toxicity criterion	Results ^a
NRTI							
Anderson et al. ^[8]	Yes Substudy of clinical trial	ZDV-TP 3TC-TP	ZDV 300 mg bid + 3TC 150 mg bid + IDV 800 mg tid or Concentration-Controlled ZDV-3TC-IDV regimen	33 naïve	Median conc. above threshold (yes/no) from samples 2h post dose at W2, 28 and 56 and at 2 to 8 hr post dose at 9 visits from W8 to W80 Thresholds: ZDV-TP: 30 fmol/10 ⁶ 3TC-TP: 7017 fmol/10 ⁶	Apparition of a grade I laboratory event (hemoglobin, absolute neutrophil count, aspartate, aminotransferase/alanine)	NS
Stretcher et al. ^[132]	Yes PK trial	ZDV-P	ZDV only: 800 mg / day 100mg every 4 h while awake	21 naïve	AUC (NCA) from 6 samples: pre dosing and 1, 2, 4, 6 & 8h post dosing at W4 and W24	Change between W0 and W4 or W0 and W24 of (1) neutrophils (2) red blood cells (3) hemoglobin	(1) NS (2) NS (3) P = NS
Durand-Gasselín et al. ^[99]	Yes PK trial	ZDV-TP 3TC-TP	ZDV (8 mg/kg/day in 4 daily doses) ± 3TC(4 mg/kg/day in 2 daily doses)	49 neonates	Single point concentration (time of sampling not reported)	Proportions of the hematological toxicity grade between neonates with intracellular concentrations above or below the observed median	NS
NNRTI							
Rotger et al. ^[90]	Yes PK Trial	EFV	EFV+ZDV+3TC EFV+ABC+3TC EFV+d4T+ddl +/- PI (doses not provided)	55	Intracellular AUC obtained by Bayesian estimation (number of samples per patient and sampling times not provided)	Presence of grade I to IV of (1) sleep disorder (2) mood disorder (3) fatigue	(1) NS (2) P = 0.02 (3) NS

3
4
5
6
7
8
9

^a Significant results always associated increased risk of toxicity with higher intracellular concentrations

Abbreviations: ABC = abacavir; AUC = area under the concentration time curve; bid = two times a day; tid = three times a day; ddl = didanosine; d4T = stavudine; EFV = efavirenz; IDV = indinavir; PI = protease inhibitors; PK = pharmacokinetic; NCA = non compartmental analysis; NS = non significant; ZDV = zidovudine; ZDV-P= total zidovudine phosphates; ZDV-TP = zidovudine triphosphate; 3TC = lamivudine; 3TC -TP= lamivudine triphosphate; W = week.

Figure 1

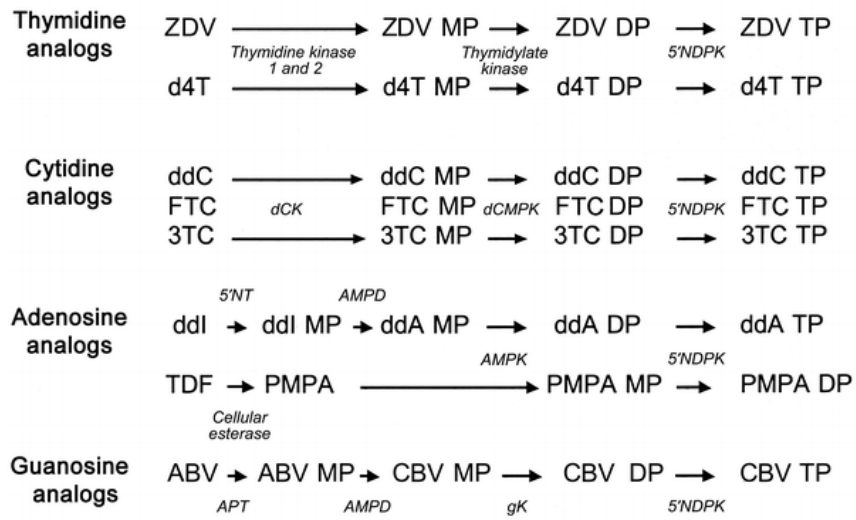


Figure 2

