

## Supplementary Figures

**Supplementary Figure S1. Immunohistochemical analysis of the inflammatory response in IHCA.** This IHCA overexpressed SAA (**a**) and CRP (**b**) with a sharp demarcation from the non-tumoural liver (NTL). Only hepatocytes of the IHCA expressed SAA or CRP; inflammatory infiltrates are negative for these acute phase inflammation proteins (asterix). Inflammatory infiltrates are made predominantly of lymphocytes ( $CD45^+$ ), localized around vascular axis inside the tumour (**c**); T lymphocytes ( $CD3^+$ ) with  $CD4/CD8$  (**d-f**) ratio around 2/1 are more numerous than B lymphocytes ( $CD20^+$ ) (**g**), intermingled with  $CD68^+$  histiocytes (**h**).

**Supplementary Figure S2. Example of somatic DNA alterations identified in exon 6 of the *IL6ST* gene.** Boundaries of the deletions are indicated by an arrow; mutated sequences are represented below the *IL6ST* wild-type sequence. **a**, Forward sequence of one representative case of mutated IHCA (T) and its matching non-tumour DNA (N) showing the somatic origin of the most frequent deletion identified; the nucleotide sequence deleted in tumour 382 is boxed. Amino acids (AA) and nucleotides (NT) are numbered according to *IL6ST* cDNA ORF. **b**, cDNA sequence analysis of two cases of mutated IHCA (T), showing expression of both the wild type and the mutant allele. **c**, Individual 770 is heterozygous for the polymorphism rs3729960 (697 G>C leading to G148R) located in exon 5 (arrow). The same ratio of allele is observed in normal and tumour sample both at the DNA and cDNA level. As this polymorphism is amplified in the same cDNA fragment than the mutated exon 6, these results show that the wild-type and the mutated allele are expressed at the same level in IHCA.

**Supplementary Figure S3. Comparison of expression profiles between gp130 mutated inflammatory HCA and gp130 non-mutated IHCA.** Comparison was carried out for the 24 genes validated by quantitative RT-PCR that were significantly differentially expressed between IHCA and non-tumour livers. Seven gp130 mutated IHCA (in black) were compared to 7 gp130 non-mutated IHCA (in grey). Results are expressed as mean  $\pm$  SD; \* difference between groups at  $P<0.05$  (two-tailed Mann-Whitney test).

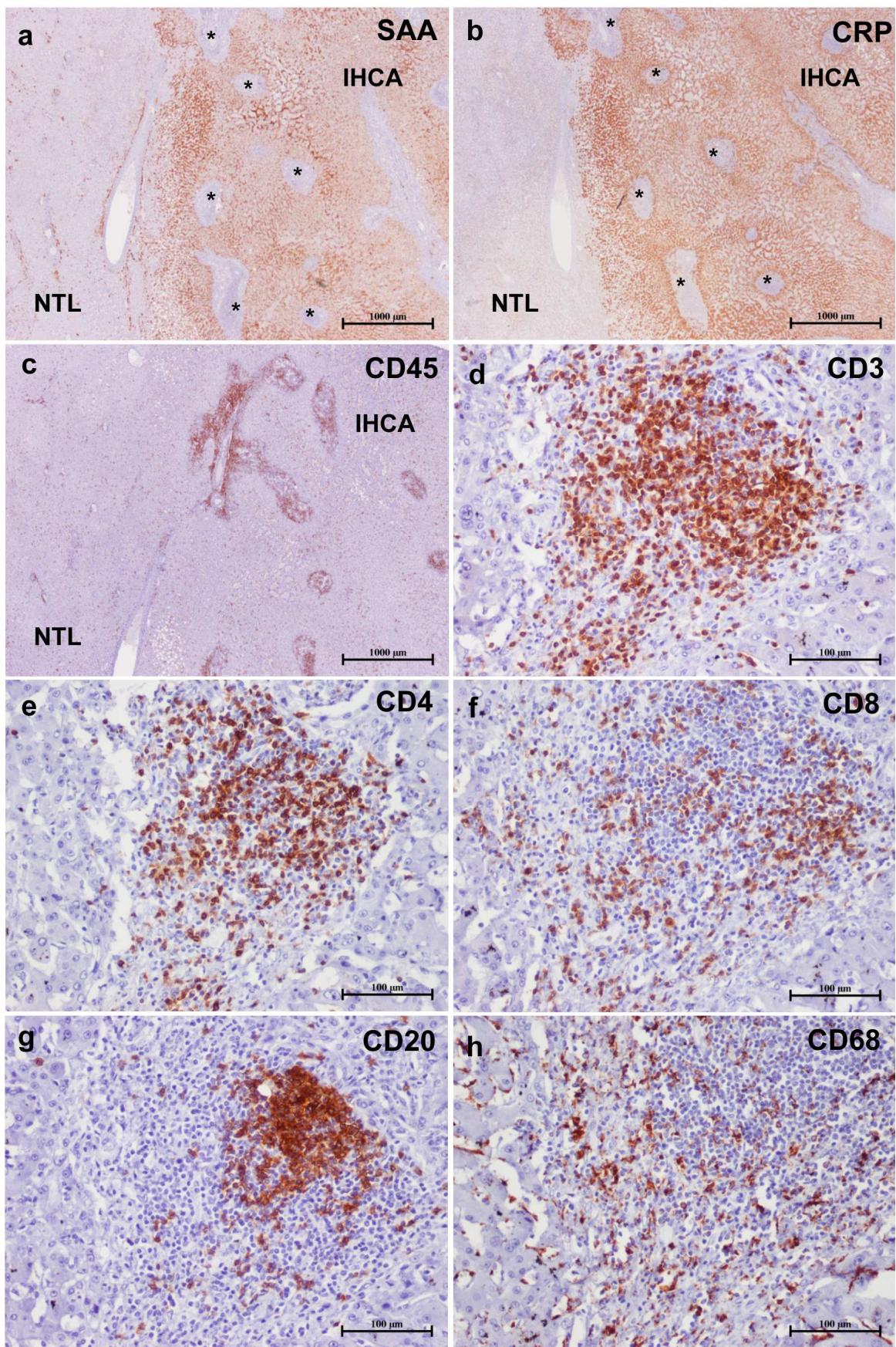
**Supplementary Figure S4. Analysis of gp130 and p-STAT3 expression in inflammatory HCA.** **a**, *IL6ST* mRNA level in 6 non-tumour livers (NTL) was compared with that of 7 gp130-mutated IHCA (M) and 7 gp130-non-mutated IHCA (NM), using qRT-PCR. Results are expressed as mean-fold difference  $\pm$  SD, relative to NTL; ns: non significant (two-tailed Mann-Whitney test). **b**, Western-blotting analysis of gp130 protein level in 3 non-tumour livers (NTL), 2 gp130-mutated IHCA and 7 gp130-non-mutated IHCA. Red ponceau staining was used as loading control. **c**, **d**, Immunohistochemical analysis of gp130 showing

overexpression of the protein in both gp130-mutated (**c**) and gp130-non-mutated (**d**) IHCA compared to their adjacent non-tumour liver parenchyma (NTL). **e, f, g**, Immunohistochemistry of phospho-STAT3 (tyr705) in gp130-mutated (**e**) and non-mutated (**f**) IHCA showing an intense nuclear staining in tumour cells while p-STAT3 is undetectable in non-tumour liver (NTL) (**g**).

**Supplementary Figure S5. mRNA expression of *IL11* and *IL11RA* in inflammatory HCA and transfected Hep3B cells.** **a**, *IL11* and *IL11RA* mRNA levels were assessed using qRT-PCR in 6 non-tumour livers (NTL), 7 gp130-mutated IHCA (M) and 7 gp130-non-mutated IHCA (NM). Results are expressed as mean-fold difference  $\pm$  SD, relative to NTL (mean NTL=1) \*, \*\*, difference between groups at  $P<0.05$  and 0.01, respectively (two-tailed Mann-Whitney test). **b**, qRT-PCR analysis of *IL11* mRNA expression in Hep3B cells transfected either with Wt gp130 or with four IHCA mutants of gp130 ( $\Delta$ ) (Y186\_Y190del (Y); K173\_D177del (K); S187\_Y190del (S) and V184\_Y186del, S187A (V)), as described in Fig3. Experiments were performed in triplicate and expressed as mean  $\pm$  SD in comparison to empty plasmid-transfected and untreated cells (mean=1). ns: non significant (two-tailed t-test).

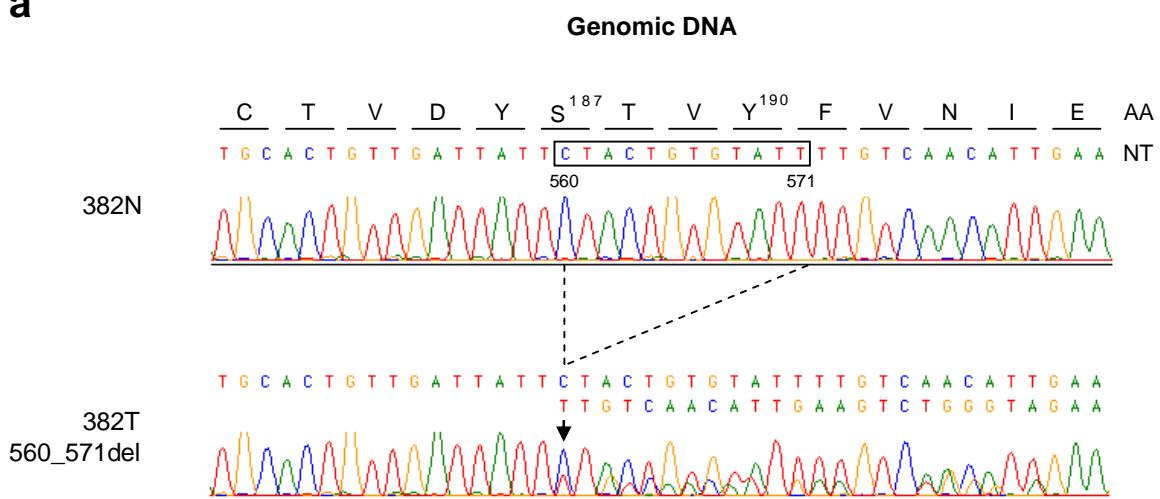
**Supplementary Figure S6. Western blotting analysis of ERK1/2 activation in inflammatory HCA and Hep3B cells.** **a**, Expression of the phosphorylated forms of ERK1 and ERK2 was compared between 10 IHCA (including 4 cases mutated for gp130) and 6 non-tumour livers (NTL). Red ponceau staining was used as loading control. **b**, Expression of phospho-ERK1/2 was compared between Hep3B cells transfected with the wild-type (Wt) or the mutant ( $\Delta S$ ) gp130 (S187\_Y190del). As a control cells were transfected with empty plasmid and untreated or stimulated with 100 ng/ $\mu$ l of IL6.

## Supplementary Figure S1

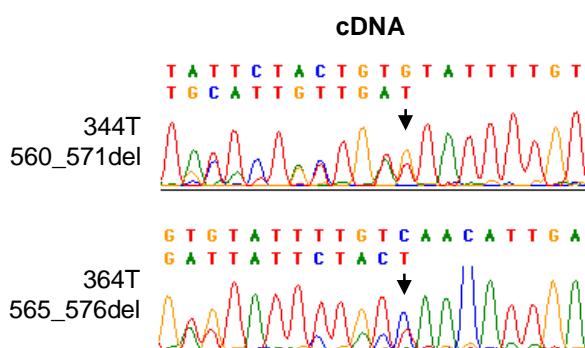


## Supplementary Figure S2

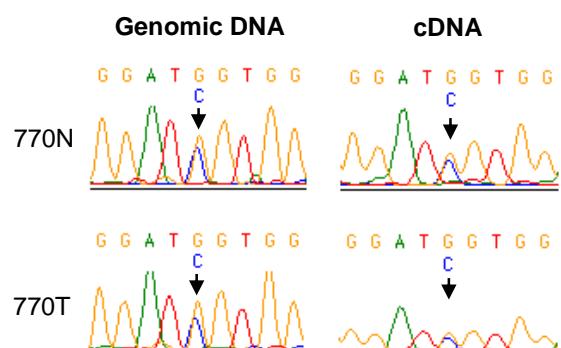
**a**



**b**

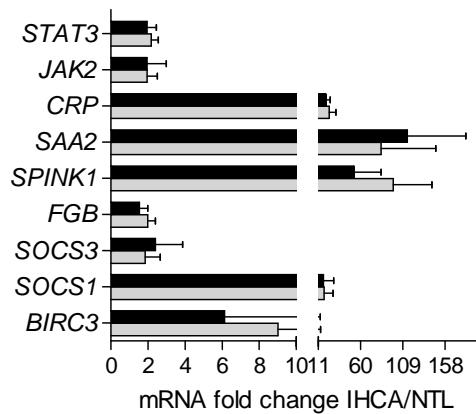


**c**

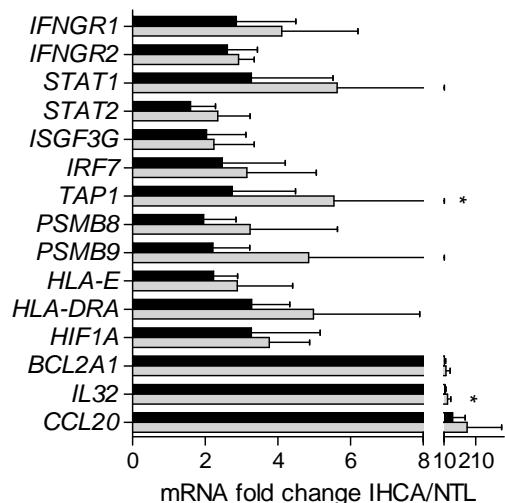


## Supplementary Figure S3

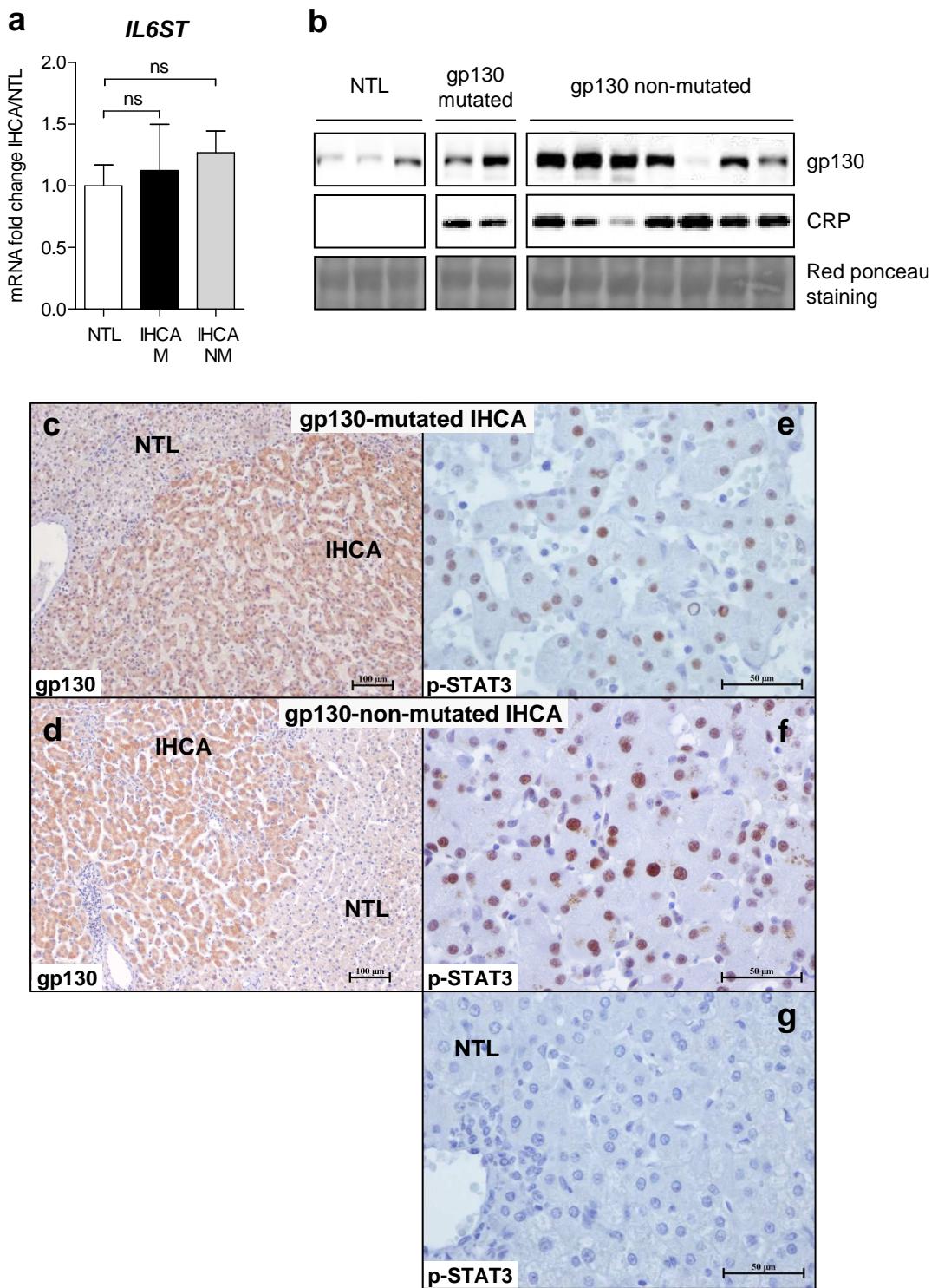
Interleukin-6 pathway



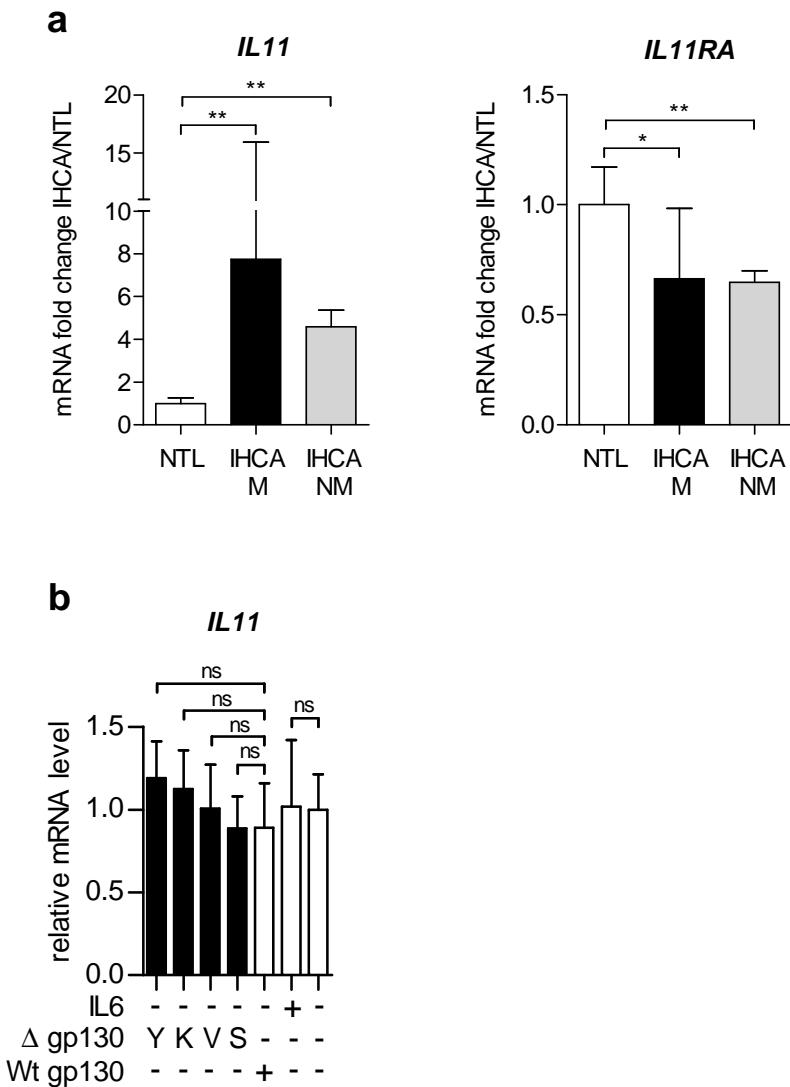
Interferon pathway



## Supplementary Figure S4

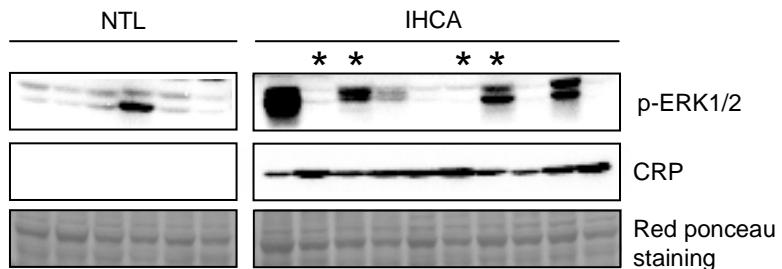


## Supplementary Figure S5



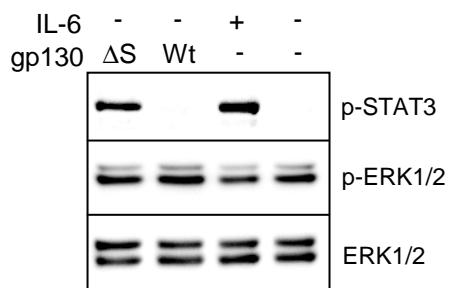
## Supplementary Figure S6

**a**



\* IHCA gp130-mutated

**b**











Hs.30246	ENSG00000117479	SLC19A2	-3.4	4.8E-04
Hs.524085	ENSG00000036672	USP2	-3.5	6.6E-06
Hs.203717	ENSG00000115414	FN1	-3.5	9.4E-04
Hs.144845	ENSG00000129151	BBOX1	-3.5	1.0E-11
Hs.479898	ENSG00000109193	SULT1E1	-3.5	5.9E-04
Hs.439056	ENSG00000198470	CYP2A6	-3.9	0.0E+00
Hs.485572	ENSG00000120833	SOCS2	-4.3	1.9E-12
Hs.463375	ENSG00000198740	ZNF652	-4.4	3.5E-06
Hs.839	ENSG00000099769	IGFALS	-4.4	2.9E-04
Hs.124112	ENSG00000167165	UGT1A6	-4.4	2.5E-03
Hs.505662	ENSG00000135447	PPP1R1A	-4.5	0.0E+00
Hs.72912	ENSG00000140465	CYP1A1	-4.7	2.6E-14
Hs.87889	ENSG00000100697	DICER1	-4.9	1.9E-03
Hs.127648	ENSG00000173930	SLCO4C1	-5.0	8.9E-04
Hs.144914	ENSG00000124713	GNMT	-5.3	6.3E-04
Hs.169948	ENSG00000177272	KCNA3	-5.4	2.0E-14
Hs.418167	ENSG00000163631	ALB	-5.7	3.0E-11
Hs.150595	ENSG00000095596	CYP26A1	-5.7	1.2E-05
Hs.75678	ENSG00000125740	FOSB	-5.9	3.6E-14
Hs.127826	ENSG00000187266	EPOR	-5.9	2.2E-06
Hs.224171	ENSG00000108515	ENO3	-6.1	0.0E+00
Hs.534364	ENSG00000127831	VIL1	-6.6	4.4E-16
Hs.523332	ENSG00000065154	OAT	-6.8	2.7E-04
Hs.272499	ENSG00000100867	DHRS2	-7.0	1.6E-11
Hs.107474	ENSG00000138386	NAB1	-8.5	1.6E-04
Hs.489786	ENSG00000001626	CFTR	-9.2	8.6E-06
Hs.98280	ENSG00000080709	KCNN2	-11.3	5.7E-05
Hs.351306	ENSG00000168679	SLC16A4	-14.9	1.6E-04
Hs.724	ENSG00000126351	THRA	-17.3	1.0E-07
Hs.234775	ENSG00000171119	NRTN	-19.4	6.7E-10
Hs.307734	ENSG00000196549	MME	-20.4	3.3E-05
Hs.282409	ENSG00000165841	CYP2C19	-21.6	1.8E-05

## Supplementary Table S2. Gene Ontology (GO) analysis

List of GO terms significantly enriched in genes over-expressed in inflammatory HCA, obtained using the web-based tool GOTree Machine and the hypergeometric test. GO terms related to inflammation and immune response are indicated in red.

**O:** Observed gene number in the GO category (referred to genes over-expressed in inflammatory HCA compared to non-tumour livers)

**E:** Expected gene number in the GO category (referred to the whole content of the HG-U133A Affymetrix array)

**R:** Ratio of enrichment for the GO category (O/E)

**P-value:** Significance of enrichment for the GO category

GO categories were considered significantly enriched when the *P*-value was less than 0.01

	O	E	R	P-value
<i>In biological process</i>				
protein kinase cascade	13	5.5	2.36	3E-03
JAK-STAT cascade	5	0.66	7.58	4E-04
regulation of JAK-STAT cascade	4	0.17	23.53	1E-05
negative regulation of JAK-STAT cascade	2	0.1	20	3E-03
negative regulation of tyrosine phosphorylation of STAT protein	2	0.1	20	3E-03
negative regulation of tyrosine phosphorylation of Stat3 protein	2	0.1	20	3E-03
positive regulation of JAK-STAT cascade	2	0.07	28.57	2E-03
positive regulation of tyrosine phosphorylation of STAT protein	2	0.07	28.57	2E-03
regulation of tyrosine phosphorylation of STAT protein	4	0.17	23.53	1E-05
regulation of tyrosine phosphorylation of Stat3 protein	2	0.12	16.67	6E-03
tyrosine phosphorylation of STAT protein	4	0.24	16.67	6E-05
tyrosine phosphorylation of Stat3 protein	2	0.12	16.67	6E-03
negative regulation of protein metabolism	6	1.36	4.41	2E-03
regulation of phosphorus metabolism	4	0.68	5.88	4E-03
regulation of amino acid metabolism	4	0.41	9.76	6E-04
negative regulation of amino acid metabolism	2	0.1	20	3E-03
negative regulation of protein amino acid phosphorylation	2	0.1	20	3E-03
regulation of phosphate metabolism	4	0.68	5.88	4E-03
negative regulation of phosphate metabolism	2	0.15	13.33	8E-03
negative regulation of phosphorylation	2	0.15	13.33	8E-03
negative regulation of peptidyl-tyrosine phosphorylation	2	0.1	20	3E-03
regulation of phosphorylation	4	0.61	6.56	3E-03
peptidyl-amino acid modification	5	1.19	4.2	6E-03
peptidyl-tyrosine modification	5	0.51	9.8	1E-04
peptidyl-tyrosine phosphorylation	4	0.46	8.7	1E-03
regulation of protein amino acid phosphorylation	4	0.39	10.26	5E-04
regulation of peptidyl-tyrosine phosphorylation	4	0.36	11.11	4E-04
organismal physiological process	61	35.54	1.72	6E-06
immune system process	38	12.02	3.16	1E-10
antigen processing and presentation	11	0.88	12.5	5E-10
antigen processing and presentation of endogenous antigen	6	0.41	14.63	2E-06
antigen processing and presentation of endogenous peptide antigen	6	0.36	16.67	8E-07
antigen processing and presentation of endogenous peptide antigen via MHC class I	6	0.36	16.67	8E-07
antigen processing and presentation of exogenous antigen	5	0.27	18.52	3E-06
antigen processing and presentation of exogenous peptide antigen	5	0.27	18.52	3E-06
antigen processing and presentation of exogenous peptide antigen via MHC class II	5	0.27	18.52	3E-06
antigen processing and presentation of peptide antigen	11	0.68	16.18	2E-11
antigen processing and presentation of peptide antigen via MHC class I	6	0.36	16.67	8E-07
antigen processing and presentation of peptide antigen via MHC class II	5	0.27	18.52	3E-06
antigen processing and presentation of peptide or polysaccharide antigen via MHC class II	5	0.27	18.52	3E-06
immune response	27	9.59	2.82	9E-07
response to stimulus	49	28.25	1.73	6E-05
response to external stimulus	20	9.49	2.11	1E-03
response to stress	31	15.4	2.01	1E-04
response to wounding	19	7.06	2.69	8E-05
physiological response to stimulus	47	22.38	2.1	5E-07
physiological response to wounding	19	6.81	2.79	5E-05
physiological defense response	16	7.18	2.23	2E-03
inflammatory response	15	5.04	2.98	2E-04
acute-phase response	4	0.58	6.9	2E-03
response to biotic stimulus	13	4.96	2.62	1E-03
response to other organism	12	4.01	2.99	7E-04
response to virus	7	1.41	4.96	5E-04
platelet activation	3	0.32	9.38	3E-03
carbohydrate biosynthesis	7	1.65	4.24	1E-03
alcohol biosynthesis	4	0.51	7.84	1E-03
monosaccharide biosynthesis	4	0.51	7.84	1E-03
hexose biosynthesis	4	0.51	7.84	1E-03
fucose biosynthesis	2	0.05	40	6E-04
L-fucose biosynthesis	2	0.05	40	6E-04
GDP-L-fucose biosynthesis	2	0.05	40	6E-04
de novo' GDP-L-fucose biosynthesis	2	0.05	40	6E-04
fucose metabolism	3	0.29	10.34	3E-03
L-fucose metabolism	3	0.24	12.5	2E-03
nucleotide-sugar biosynthesis	3	0.07	42.86	1E-05
nucleotide-sugar metabolism	3	0.24	12.5	2E-03
GDP-L-fucose metabolism	2	0.05	40	6E-04
GDP-mannose metabolism	2	0.05	40	6E-04
developmental maturation	4	0.46	8.7	1E-03
cell maturation	2	0.07	28.57	2E-03
myeloid cell differentiation	5	0.83	6.02	1E-03
erythrocyte differentiation	4	0.32	12.5	2E-04
erythrocyte development	2	0.07	28.57	2E-03
erythrocyte maturation	4	0.46	8.7	1E-03

<b>anatomical structure development</b>	39	26.1	1.49	6E-03
cell development	8	2.51	3.19	4E-03
cellular morphogenesis during differentiation	5	1.05	4.76	4E-03
cell projection morphogenesis	5	0.97	5.15	3E-03
neuron development	5	1.24	4.03	8E-03
neuron morphogenesis during differentiation	5	0.97	5.15	3E-03
neurite development	5	1.07	4.67	4E-03
neurite morphogenesis	5	0.97	5.15	3E-03
axonogenesis	5	0.92	5.43	2E-03
<b>cell adhesion</b>	24	11.22	2.14	3E-04
regulation of cell adhesion	5	0.78	6.41	1E-03
positive regulation of cell adhesion	2	0.1	20	3E-03
<b>cell motility</b>	14	4.79	2.92	3E-04
cell migration	7	2.07	3.38	5E-03
<b>locomotion</b>	14	4.79	2.92	3E-04
<b>cell proliferation</b>	20	11.22	1.78	8E-03
<b>phospholipid catabolism</b>	2	0.15	13.33	8E-03
<b>localization of cell</b>	14	4.79	2.92	3E-04
<i>In molecular function</i>				
<b>interferon binding</b>	2	0.09	22.22	3E-03
interferon receptor activity	2	0.09	22.22	3E-03
interferon-gamma receptor activity	2	0.05	40	6E-04
interferon-gamma binding	2	0.05	40	6E-04
<b>obsolete molecular function</b>	11	0.61	18.03	5E-12
MHC class I receptor activity	6	0.38	15.79	1E-06
MHC class II receptor activity	6	0.26	23.08	7E-08
<b>enzyme inhibitor activity</b>	12	4.21	2.85	1E-03
protease inhibitor activity	7	2.26	3.1	7E-03
endopeptidase inhibitor activity	7	2.23	3.14	7E-03
<b>glutathione peroxidase activity</b>	2	0.09	22.22	3E-03
<b>erythropoietin receptor binding</b>	2	0.07	28.57	2E-03
<b>selenium binding</b>	2	0.12	16.67	5E-03
<b>aldehyde dehydrogenase [NAD(P)+] activity</b>	2	0.14	14.29	8E-03





**Supplementary Table S5. TaqMan® pre-designed gene expression assays used for quantitative RT-PCR analyses**

Assay ID	Gene Symbol	Gene Name
Hs99999901_s1	18S	Eukaryotic 18S rRNA
Hs00187845_m1	BCL2A1	BCL2-related protein A1
Hs00154109_m1	BIRC3	baculoviral IAP repeat-containing 3
Hs00355476_m1	CCL20	chemokine (C-C motif) ligand 20
Hs00265044_m1	CRP	C-reactive protein, pentraxin-related
Hs00170586_m1	FGB	fibrinogen beta chain
Hs00153153_m1	HIF1A	hypoxia-inducible factor 1, alpha subunit (basic helix-loop-helix transcription factor)
Hs00219575_m1	HLA-DRA	major histocompatibility complex, class II, DR alpha
Hs00428366_m1	HLA-E	major histocompatibility complex, class I, E
Hs00166223_m1	IFNGR1	interferon gamma receptor 1
Hs00194264_m1	IFNGR2	interferon gamma receptor 2 (interferon gamma transducer 1)
Hs00174148_m1	IL11	interleukin 11
Hs00234415_m1	IL11RA	interleukin 11 receptor, alpha
Hs00992441_m1	IL32	interleukin 32
Hs00174360_m1	IL6ST	interleukin 6 signal transducer (gp130, oncostatin M receptor)
Hs00185375_m1	IRF7	interferon regulatory factor 7
Hs00196051_m1	ISGF3G	interferon-stimulated transcription factor 3, gamma 48kDa
Hs00234567_m1	JAK2	Janus kinase 2 (a protein tyrosine kinase)
Hs00544758_m1	PSMB8	proteasome (prosome, macropain) subunit, beta type, 8 (large multifunctional peptidase 7)
Hs00544762_m1	PSMB9	proteasome (prosome, macropain) subunit, beta type, 9 (large multifunctional peptidase 2)
Hs01667582_m1	SAA2	serum amyloid A2
Hs00705164_s1	SOCS1	suppressor of cytokine signaling 1
Hs00269575_s1	SOCS3	suppressor of cytokine signaling 3
Hs00162154_m1	SPINK1	serine peptidase inhibitor, Kazal type 1
Hs00234829_m1	STAT1	signal transducer and activator of transcription 1, 91kDa
Hs00237139_m1	STAT2	signal transducer and activator of transcription 2, 113kDa
Hs00374280_m1	STAT3	signal transducer and activator of transcription 3 (acute-phase response factor)
Hs00388682_m1	TAP1	transporter 1, ATP-binding cassette, sub-family B (MDR/TAP)

**Supplementary Table S6. List of primers used in PCR and sequencing**

F: forward primer; R: reverse primer

Exon	Sequence primer 5'->3'	Primer Name
<b><i>IL6ST</i> genomic DNA amplification and sequencing</b>		
3	CACAGCTACAGCATGGAAACA	3F-IL6ST
	GCATGTAAGATGACTGTTACAACCAA	3R-IL6ST
4	TGACGGTTGAGGGACTCTACA	4F-IL6ST
	GCAGTCAATAGTCATGACAAAGTC	4R-IL6ST
5	CGCAAATGCCCTGGGTAGTA	5F-IL6ST
	AAAAATTCCAGCAAAATGACT	5R-IL6ST
6	TAGTGACCAGCAGTTATTCGAA	6F-IL6ST
	CGACTACAGTGTCAAATAAACCTCTCA	6R-IL6ST
7	TTGGAAAACATAATAAACCTTAGGA	7F-IL6ST
	GGGTTGAAAGAAGGACTATTGAA	7R-IL6ST
8	TGTTGGTTGCTTGTCA	8F-IL6ST
	TCACAAACTAAGAAGCAATGAGG	8R-IL6ST
9	GCAATTGGAGGGCAAGAGTA	9F-IL6ST
	AATGGAGCGAGACTCGGTTT	9R-IL6ST
10	AGAGATTGACATAATGGCATG	10F-IL6ST
	GTGTTTTTATGGGAGCCCTG	10R-IL6ST
11	TCACATGTTTGGAAATGACTG	11F-IL6ST
	AGTCGTTCAACGGACCAAA	11RIL6ST
12-13	TGCAGTTAAAAGAAAATTAAGTGG	12/13F-IL6ST
	GGTGTCTCTGAGGCCATA	12/13R-IL6ST
14	CAATGCCAAATATAGGCCAAA	14F-IL6ST
	CCACCAACCAAAGCCAGATA	14R-IL6ST
15	CAGTAAGTCTGGTGGGAAGTTG	15F-IL6ST
	TTGGGCTGAGAAGAAGACAGA	15R-IL6ST
16	TGGTTGGTTGTAAAGAGTCTTG	16F-IL6ST
	TTCAGCTTGAATATCTTATCTTT	16R-IL6ST
17	CTGGGAAGTTCAAGAGATGC	17F-IL6ST
	CTTGCCTGTACAGTCTGTGGT	17R-IL6ST
<b><i>IL6ST</i> cDNA amplification and sequencing</b>		
3-7	ACCCAATGGAAAAGCATGA	3F.RT-IL6ST
	TGCTGTCTTCAGGAGGAA	7R.RT-IL6ST
7-11	AGAACAGAGAATGCCCTG	7F.RT-IL6ST
	CCTCTTAAATAGGTGCGATGC	11R.RT-IL6ST
11-15	TCACAAGATGAAATCACAT	11F.RT-IL6ST
	TGACCACTGGGCAATATGAC	15R.RT-IL6ST
16-17	TTCTGTTAATAAGCGAGACC	16F.RT-IL6ST
	GGGCAAATGATCATCTTCAGAG	17R.RT-IL6ST
<b><i>IL6ST</i> cDNA sequencing</b>		
3-7	GAACAGCATCCAGTGTCA	3FintRT-IL6ST
	GTTCGCTTGCAATCA	7RintRT-IL6ST
7-11	GCTGTATGAAGGAAGATG	7FintRT-IL6ST
	TCAGTTTGTCGATTAACGTG	11RintRT-IL6ST
11-15	AGACTGGCAACAAAGAAGA	11FintRT-IL6ST
	CTGTGTATGCTGCCATT	15RintRT-IL6ST
16-17	GACCTGTTCAAAAGGAAA	16FintRT-IL6ST
	ATCAGTCGCAGCTCCAT	17RintRT-IL6ST
<b><i>IL6R</i> cDNA amplification and sequencing</b>		
1-5	GGCTCCTGCGGATGGGGC	5'UTRF.RT-IL6R
	GGGCTTGGCAGGTGACAC	703R.RT-IL6R
4-10	CCAGTAGTGTGGGAGCAA	583F.RT-IL6R
	CATTTTGCTGAGCCCCT	3'UTRR.RT-IL6R
<b><i>IL6R</i> cDNA sequencing</b>		
1-5	GAGGAGCCCCAGCTCTCC	342F-IL6R
1-3	CTCACCAAGAGCACAGCC	434R-IL6R
9-10	CGACAAGCCTCCCAGTGC	1051F-IL6R
5-10	AGTACGGCGGATGATGC	1207R-IL6R
<b><i>LIFR</i> cDNA amplification and sequencing</b>		
1-6	AAGGTGAGCCTCAGAACATC	5'UTRF.RT-LIFR
	GCAGTTGTATGCCAACATCAGT	6R.RT-LIFR
6-10	AGGTTTTCTCAAGATAAAAGTGA	6F.RT-LIFR
	CTTGAGGGACTGGCTTCTG	10R.RT-LIFR
10-15	TTTCGGATTGTTGTTCTACTGA	10F.RT-LIFR
	TTCTTAACCTTATGTCAGAACGACCT	15R.RT-LIFR
15-19	GGGAAGACATTCTGTGGA	15F.RT-LIFR
	CCCCTCTACAGGGTCATT	19R.RT-LIFR
19	GGGACTGCACAGGTTATTAC	19F.RT-LIFR
	CACATGAGAAAACCACAAACA	3'UTRR.RT-LIFR

LIFR cDNA sequencing		
1-6	AAAACAGGTCCCGTTCTTGTATCA GTCTGAGCCTACAAGTATCAC	5'UTRintFRT-LIFR 6RintRT-LIFR
6-10	TCTTGGCATTTACCAGGCAACT TCCTCATCTGATGAACACGAT	6FintRT-LIFR 10RintRT-LIFR
10-15	CGCTCCATGATTGGATATATAAG GGTTTCAGGCTCTGCATCAGA	10FintRT-LIFR 15RintRT-LIFR
19	TTACAGACCTCAGGCCAATG	19FintRT-LIFR
OSMR cDNA amplification and sequencing		
1-4	TGGATACAGGGAAAGGGAGAA GCTCCTCAAGTACTTTGAGACAA	3'UTRF.RT-OSMR 4R.RT-OSMR
4-10	GGAAGGGAAACAGATTCTG TGGATGGTTGTCTAGGTTT	4F.RT-OSMR 10R.RT-OSMR
8-14	TGGAGTGAATGGAGTGTC TGAATCAGCATCGAGGAGT	8F.RT-OSMR 14R.RT-OSMR
14-17	CAAAATTGACAACCGGAAG GTGCCAGCTGCTCTTCTT	14F.RT-OSMR 5'UTRR.RT-OSMR
SOCS3 genomic DNA amplification and sequencing		
2	AGGGTCCGGGCACTCAA TTCTTGCTTGTGCCATGT	2F-SOCS3 2R-SOCS3
JAK1 genomic DNA amplification and sequencing		
11	CAGCAGACGTTGGCTGTC CCCCAAGCTGCTCCATC	11F-JAK1 11R-JAK1
12	ATTGAGGACCCATTCCCAG GTTCCACTGGCTCCAGAAC	12F-JAK1 12R-JAK1
13	CTGGCTTCCCCCTAACAG ACAGCACCCAAGCTCCC	13F-JAK1 13R-JAK1
14	CTGTCGTAAGGGGATGAAG TCCCAGGGACACTCTGGTT	14F-JAK1 14R-JAK1
15	GGGACCAGGGTTGAAATAG CAGCCCTCTCTGCTGAC	15F-JAK1 15R-JAK1
16	TCACTGTTCTGGCTGCAGTG ACAACCTCTGGGAAGCCC	16F-JAK1 16R-JAK1
17-18	GGGGCTGAGAAGTTGTAGG TACGTGCAGAGAGTGGCG	17/18F-JAK1 17/18R-JAK1
19	GACTCTGTGTGTTCCGTGG TTGCACTGGCCTTTATGACC	19F-JAK1 19R-JAK1
20	CTTGCCTCTCCATGGTC CAAACCAATTACCCAGGACAG	20F-JAK1 20R-JAK1
21	TCTGCATTATTAGATGAAAAGTTAGC TCAGGCCAGAGGAATGAGAC	21F-JAK1 21R-JAK1
22	TCCTGTAATGTCGGATGGAG TTAACCAAGCAGAGGGATGG	22F-JAK1 22R-JAK1
23	ATTTCACCTTAATCTGTTGGC CAACTCAAGCTGCGAAAAC	23F-JAK1 23R-JAK1
24	TGAACTAGAGTTGATCCCCATTTC ACTGGGCATTTGGCAG	24F-JAK1 24R-JAK1
JAK2 genomic DNA amplification and sequencing		
12	CAAAGTTCAATGAGTTGACCCC TGCTAACATCTAACACAAGGTTGG	12F-JAK2 12R-JAK2
13	TGTATTTCTGTTCCACTTCGTT TTACAATTGAAAAGCTGCAC	13F-JAK2 13R-JAK2
14	GAGAAAGTGCATCTTATTATGGC CTGACACCTAGCTGTGATCCTG	14F-JAK2 14R-JAK2
15	TGTTTAGACTCCTACTCTGCTGTT CACAGACTATTACATGAATTGGC	15F-JAK2 15R-JAK2
16	TTCTCTTTAAATCTGTTTGGGG CCTTACACCACTGCCAAG	16F-JAK2 16R-JAK2
17-18	CCCCTCCAAAATAAAGAAAATAGG AGGGCCCAAATGACATCAAG	17/18F-JAK2 17/18R-JAK2
19	GACCA GTTGGTGGCGGG GAAGGCCCTGTCAGATTATGGG	19R-JAK2 19F-JAK2
20	TTTATTGTAATTGCTTGAAAAC TTACACAGGATGCCAACAC	20F-JAK2 20R-JAK2
21-22	CCCACGTGGACTATAACCATGAC CTGAAACGCAAATAGTTCAAAGC	21/22R-JAK2 21/22F-JAK2
23	GTAGATGTCAGCATAAACCGTATTG GCCATTGGTAGATGTC	23F-JAK2 23R-JAK2
24-25	CCATTGACTGGAGGAAATTGAG TCCACAGCAATGTGAAATATAAAAC	24/25F-JAK2 24/25R-JAK2

TYK2 genomic DNA amplification and sequencing		
12-13	GTGGGATGTGGCATCTCTCC	12/13F-TYK2
	CCACAGACACACCCCCCTCC	12/13R-TYK2
14	GTGTCCTGGAGGGAGGTG	14F-TYK2
	GGAGTGGGTATAAAGTCAGGGG	14R-TYK2
15	ACATGCCAACGTCCCTCCC	15F-TYK2
	TCTCGAACTCCTGGGCTCC	15R-TYK2
16-17	GAGCTCTGATTCTGCCTCTGG	17R-TYK2
	GTTGTCCTTTCACATGGCTG	16F-TYK2
18	CTGCCTCTGGGAACG	18F-TYK2
	GCTATAGGCATAACAGCAGGG	18R-TYK2
19	TTTGTGACTCCCAAGTGTGG	19F-TYK2
	CCAAACTCCTCCCGACC	19R-TYK2
20	GAGTGGTGCAGGGATTGG	20F-TYK2
	CACAAGGCCACACCCAC	20R-TYK2
21	CTCTGCTGGGCTCAAGGTAG	21F-TYK2
	CAGCCCAAGCTGAAGAGG	21R-TYK2
22	GCTGCTCAGGTCTGCC	22F-TYK2
	CTACTCCACCCCTGCCTGTC	22R-TYK2
23	GTCTCAGGTGAGGGCTTCAG	23F-TYK2
	TCTCTCCGTCTGTCCTGTC	23R-TYK2
24-25	CGTGCCTGCCCTTCATTG	24/25F-TYK2
	TTGGTTTCATCCTGGAGCAG	24/25R-TYK2
TYK2 genomic DNA sequencing		
22	CGAACCCCGCCCCCACTGA	22F-TYK2
	CCAAGTGACCCAGCACC	22R-TYK2