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**Ms No.wjg/2007/009804****RAPID****COMMUNICATION****A red wine polyphenolic extract reduces the activation phenotype of cultured human liver myofibroblasts**

Running title: Wine extract decreases liver myofibroblasts activity

Véronique Neaud, Jean Rosenbaum

Véronique Neaud, Jean Rosenbaum, INSERM, U889, Bordeaux, F-33076 France;  
University Victor Segalen Bordeaux 2, Bordeaux, F-33076 France.**Author contributions:**

V.N. performed the experiments; J.R. designed the study and wrote the paper.

**Supported** in part by a grant from Institut Vin et Santé**Correspondence to:** Jean Rosenbaum, INSERM, U889, Université Victor Segalen  
Bordeaux 2, 146 rue Léo Saignat, 33076 Bordeaux, France. [jean.rosenbaum@gref.u-bordeaux2.fr](mailto:jean.rosenbaum@gref.u-bordeaux2.fr)

Telephone : +33-5- 575571594; Fax : +33-5-56514077

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**ABBREVIATIONS USED**

extracellular matrix (ECM); alpha smooth muscle actin (ASMA); matrix metalloproteinase-2 (MMP-2); tissue inhibitor of MMP-1 (TIMP-1); red wine polyphenolic extracts (RWPE); Fetal calf serum (FCS); Epidermal growth factor (EGF); platelet-derived growth factor BB (PDGF-BB); 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT).

## ABSTRACT

AIM : In order to define the potential anti-fibrotic effects of red wine polyphenolis, we tested the effect of a standardized red wine polyphenolic extract (RWPE) on the phenotype of human liver myofibroblasts in culture.

METHODS: Human myofibroblasts grown from liver explants were used in this study. Cell proliferation was measured with the MTT assay. Signaling events were analyzed by western blot with phospho-specific antibodies. Matrix-metalloproteinase activity was measured with gel zymography.

RESULTS: We found that cell proliferation was dose-dependently decreased by up to 90% by RWPE while cell viability was not affected. Exposure to RWPE also greatly decreased the phosphorylation of ERK1/ERK2 and Akt in response to stimulation by the mitogenic factor PDGF-BB. Finally, RWPE affected extracellular matrix remodeling by decreasing the secretion by myofibroblasts of matrix-metalloproteinase-2 and of tissue inhibitor of matrix-metalloproteinases-1.

CONCLUSION: Altogether, RWPE decreases the activation state of liver myofibroblasts. The identification of the active compounds in RWPE could offer new therapeutic strategies against liver fibrosis.

Key words : liver fibrosis, myofibroblasts, hepatic stellate cells, wine, phosphorylation, proliferation

## INTRODUCTION

Liver fibrosis is a serious health problem worldwide. It is a complication of most chronic liver diseases whether due to excessive alcohol consumption, chronic viral hepatitis B or C, non alcoholic steatohepatitis, hemochromatosis or others. The pathophysiology of liver fibrosis has been extensively studied (recently reviewed in <sup>[1, 2]</sup>). Whatever the initial insult, the abundant extracellular matrix (ECM) characteristic of liver fibrosis is synthesized by myofibroblastic cells. Myofibroblasts are mostly

absent from the normal liver but at least two types of resident liver cells can be differentiated into myofibroblasts during liver disease : hepatic stellate cells, and portal fibroblasts [3]. Myofibroblastic differentiation is characterized by a high rate of cell proliferation and of ECM synthesis and by cytoskeletal changes, notably expression of alpha smooth muscle actin (ASMA) that confers contractile properties to the cells [4]. In addition, degradation of the normal liver ECM results from an increased secretion of the enzyme matrix metalloproteinase-2 (MMP-2) by myofibroblasts, while the proteolytic degradation of the abnormal ECM is inhibited due to a high level synthesis of a MMP inhibitor, tissue inhibitor of MMP-1 (TIMP-1) [1, 2]. In the recent years, a series of natural products were shown to be of potential benefit against liver fibrosis [5, 6]. For instance, we and others found that a polyphenolic component of red wine, *trans*-resveratrol, was able to strongly deactivate liver fibrogenic cells [7, 8], while a related molecule, *trans*-piceid, was ineffective [8]. However, red wine contains many other polyphenolic substances, and red wine polyphenolic extracts (RWPE) showed many interesting biological effects in other settings, notably in the prevention of experimental atherosclerosis [9]. One of the mechanisms postulated in this context is related to the inhibitory effect of RWPE on the proliferation of vascular smooth muscle cells [10]. Since vascular smooth muscle cells share many characteristics with myofibroblasts, we tested the hypothesis that RWPE would affect the phenotypic characteristics of human liver myofibroblasts, especially those related to their pro-fibrogenic activity.

## **MATERIALS AND METHODS**

### ***Materials***

Preparation and characterization of the polyphenolic extract (RWPE) from a red French wine (Corbières, A. O. C.) was as described previously [11, 12]. One liter of red wine produced 2.9 g of extract, which contained 471 mg/g total phenolic compounds expressed as gallic acid. Phenolic levels in the extract were obtained according to HPLC analysis procedure. In particular, the extract contained 8.6 mg/g catechin, 8.7 mg/g epicatechin, dimers (B1: 6.9 mg/g; B2: 8.0 mg/g; B3: 20.7 mg/g; and B4: 0.7

mg/g), anthocyanins (malvidin-3-glucoside: 11.7 mg/g; peonidin-3-glucoside: 0.66 mg/g; and cyanidin-3-glucoside: 0.06 mg/g), and phenolic acids (gallic acid: 5.0 mg/g; caffeic acid 2.5 mg/g; and caftaric acid: 12.5 mg/g). Stock solutions (1 mg/ml) were prepared in distilled water containing 1 % ethanol and further diluted in culture medium. All dilutions were adjusted so as to contain 0.1% ethanol, and medium with 0.1% ethanol was used as a control.

Fetal calf serum (FCS) was from Dutscher (Brumath, France) and human serum from Etablissement Français du Sang (Bordeaux, France). Epidermal growth factor (EGF) was from Peprotech (Tebu, Le Perray en Yvelines, France) and recombinant platelet-derived growth factor BB (PDGF-BB) was from Eurobio (les Ulis, France). Phospho Thr308 AKT-1 and AKT-1 antibodies, phospho-ERK1/ERK2 and total ERK antibodies were from Cell Signaling Technology (Ozyme, Saint Quentin Yvelines, France). ASMA and vimentin antibodies were from Dako (Trappes, France). The TIMP antibody was from Santa Cruz Biotechnologies (Santa Cruz, CA). Beta-actin antibody was from Sigma (Saint Quentin Fallavier, France). IRDye 680 and IRDye 800 conjugated secondary antibodies, Odyssey Blocker and Odyssey infrared imaging system were from LI-COR Biosciences (ScienceTec, Les Ulis, France).

### ***Cell isolation and culture***

Human hepatic myofibroblasts were obtained from explants of non-tumor liver resected during partial hepatectomy and characterized as described previously <sup>[13, 14]</sup>. Specifically, the procedure, based on the selective growth advantage of myofibroblasts in the culture conditions used, allowed for 100% pure myofibroblasts population, as shown by positive staining for ASMA and vimentin, and negative staining for CD 68 (a Kupffer cell marker), von Willebrand factor (an endothelial cell marker) or cytokeratin (an epithelial cell marker). This procedure is in accordance with INSERM ethical regulation imposed by French legislation. Myofibroblasts were used between the 3<sup>rd</sup> and the 6<sup>th</sup> passage, and were grown in DMEM containing 5% FCS, 5% pooled human serum and 5 ng/mL EGF. EGF was removed from the medium at least 3 days before experiments.

### ***Cell proliferation assay***

Cells were seeded at a density of  $10^4$ /well in 24-well plates. On the following day, the medium was replaced by DMEM with 5% FCS and RWPE dilutions to be tested. After three or seven days, the medium was removed and the cells incubated with 1 mg/ml MTT (3-[4,5-dimethylthiazol-2-yl]-2,5 diphenyl tetrazolium bromide) for 2 h at 37°C [15]. The crystals were then dissolved with DMSO and the optical density was recorded at 540 nm. Results were expressed as proliferation index = (B-A)/(C-A) where A is the optical density recorded at Day 0, B is the optical density with the test compound and C is the optical density obtained in the control wells.

### ***Measure of cell DNA content***

Cells were grown to confluence in 24-well plates, serum-starved for 24 h, then incubated with RWPE. After 24 h, the cells were lysed with  $\text{NaH}_2\text{PO}_4/\text{Na}_2\text{HPO}_4$  50 mM, pH 7.4, NaCl 2M, EDTA 2 mM, and DNA content was measured as described [16].

### ***Western blot***

ERK and Akt phosphorylation was measured essentially as described previously [17]. Briefly, cells were grown to confluency and serum-starved for three days. They were then pre-incubated with RWPE in serum-free Waymouth medium for 1 h, then exposed to 20 ng/ml PDGF-BB for 10 min. At the end of the incubation, cell lysates were prepared in the presence of proteases and phosphatases inhibitors as described [18]. Proteins were measured with a Bio-Rad assay. Equivalent amounts of proteins were separated by SDS-PAGE, transferred to polyvinylidene difluoride membranes, and analyzed by two color Western blotting with antibodies to total-ERK and phospho-ERK, or phospho-Akt-1 and  $\beta$ -actin. Blots were blocked in Odyssey Blocker and incubated simultaneously with both primary antibodies, followed by both IR-labeled secondary antibodies. Signals were detected and quantified using the Odyssey infrared imaging system.

The expression of ASMA and of vimentin was measured in cell extracts from cells exposed for seven days to RWPE using Western blot as described [8].

The concentration of TIMP-1 in conditioned medium was also measured by Western blot. Confluent cells were incubated for two days with RWPE, the medium was collected, centrifuged and aliquots, normalized for DNA content of the cell layers, were analyzed by Western blot.

### **Zymography for MMP-2 activity**

The detection of MMP-2 in conditioned medium was performed by gelatin zymography [19], essentially as previously described [8, 20]. Briefly, cells were grown to confluence in 24-well plates, serum-starved for 24 h, then incubated with RWPE. The medium was collected, centrifuged and aliquots, normalized for DNA content of the cell layers, were analyzed on 8% SDS-PAGE gels containing 1 mg/ml gelatin. Following staining with Coomassie blue, MMP activity is detected as a white zone on a blue background.

### **Immunofluorescence detection of alpha-smooth muscle actin**

Cells were seeded at a density of  $10^4$ /well on glass coverslips in 24-well plates. On the following day, the medium was replaced by DMEM with 5% FCS and RWPE dilutions to be tested. After three days, cells were fixed with methanol at  $-20^{\circ}\text{C}$ , then incubated sequentially with an anti-ASMA monoclonal antibody and a Texas Red-conjugated secondary antibody. The slides were examined with a Zeiss Axioplan fluorescence microscope.

## **RESULTS**

### ***Effect of the RWPE on cell proliferation***

The RWPE dose-dependently inhibited the proliferation of myofibroblasts down to  $7.8 \pm 4.5\%$  of control at Day 3 and  $15.8 \pm 8.0$  at Day 7 (Fig. 1A-B). The concentration

that reduced growth by 50% was 50 µg/ml. A toxic effect of RWPE on cells could be ruled out because no morphological signs of toxicity nor cell detachment were observed (see also Fig. 2); furthermore, when confluent cells were exposed to a dose range of RWPE, there was no decrease in DNA content of the cell layers even at the highest concentrations (Fig. 1C).

### **Effect of the RWPE on expression of alpha-smooth muscle actin**

Expression of the cytoskeletal protein ASMA is hallmark of activated liver fibrogenic cells. We found that long term (up to seven days) exposure of liver myofibroblasts to RWPE did not affect ASMA expression. This was shown both by immunofluorescence and by Western blot (Fig. 2). In addition, the expression of another cytoskeletal protein, vimentin, which expression is independent of fibrogenic cell activation, was also unaffected by RWPE treatment (Fig. 2C).

### **Effect of the RWPE on the phosphorylation of MAPK and Akt**

In order to delineate the mitogenic pathways affected by RWPE, myofibroblasts were briefly exposed to PDGF-BB. PDGF-BB is major mediator of liver fibrogenic cell activation, as it stimulates notably their proliferation<sup>[13, 21]</sup> and migration<sup>[22, 23]</sup>, and is abundant in serum. We then examined the effect of RWPE on signalization pathways elicited by PDGF-BB. As expected, treatment with PDGF-BB induced a major increase in the phosphorylation of ERK1/ERK2 and of Akt. Exposure to RWPE greatly decreased the effect of PDGF-BB on the phosphorylation of both ERK1/ERK2 and Akt (Fig. 3).

### **Effect of the RWPE on MMP-2 and TIMP-1 expression**

A high level expression of the matrix remodeling enzyme MMP-2<sup>[24]</sup>, and of the inhibitor of extracellular matrix degradation, TIMP-1<sup>[25]</sup>, is characteristic of activated liver fibrogenic cells. Gelatin zymography showed a gelatinolytic band migrating at an apparent molecular weight of 72 kDa characteristic of MMP-2. The RWPE strikingly

and dose-dependently decreased the secretion of MMP-2 (Fig. 4A). It also greatly decreased TIMP-1 secretion, as assessed by Western blot (Fig. 4B).

## DISCUSSION

We show here that a standardized RWPE has striking effects on the phenotype of human liver myofibroblasts. This is shown by a decreased proliferation rate together with decreased secretion of MMP-2 and of TIMP-1. These effects are not the consequence of a direct toxicity of the extract on cells as shown by morphological examination and DNA content measurement. We investigated further the mechanism of the decreased proliferation rate and found that RWPE treatment largely abolished the phosphorylation of ERK1/ERK2 and of Akt induced by the mitogenic factor PDGF-BB. Previously, Iijima et al. showed that a RWPE decreased Akt but not ERK activation in vascular smooth muscle cells stimulated with PDGF [26]. The differences may be due to the fact that different RWPEs were used, or to a true cell specificity. For instance, despite the fact that myofibroblasts and smooth muscle cells are related cells, we found that they had a differential response to *trans*-resveratrol [8]. Some of the effects of the RWPE are reminiscent of those of *trans*-resveratrol, raising the possibility that RWPE effects may be due to this compound. However, resveratrol is present only at low concentration in wine (reviewed in [27]) and is unlikely to explain the effects of RWPE. In addition, whereas it does indeed decrease myofibroblasts proliferation, MMP-2 secretion [8] and Akt activation [28], it does not affect ERK activation in response to PDGF [28]. Furthermore, contrary to RWPE, it does decrease ASMA expression [8].

The RWPE effects observed in the present study are potentially of benefit against liver fibrosis, if they held true in the *in vivo* situation. This seems obvious for the reduced cell proliferation that will decrease the number of ECM-producing cells. Notably, the drastic effect on TIMP-1 secretion may have a major implication since TIMP-1 overexpression is considered one of the main determinants of liver fibrosis through the inhibition of ECM-degrading enzymes activity [29, 30].

Thus, although excessive wine consumption is one of the major causes of chronic

liver diseases, wine itself may unexpectedly contain potent anti-fibrotic compounds. The identification of the active compounds in RWPE could offer new therapeutic strategies against liver fibrosis.

## **ACKNOWLEDGMENT**

We thank PL Teissedre for providing the standardized RWPE.

## **COMMENTS(in 250 words)**

### ***Background***

Liver fibrosis is a worldwide problem, as it complicates all chronic liver diseases. There is no established treatment for liver fibrosis.

### ***Research frontiers***

A series of natural products have shown beneficial effects on liver fibrosis in cell culture and animal models. In addition, red wine polyphenols were shown to reduce the proliferation of vascular smooth muscle cells, a cell type closely related to liver fibrogenic cells.

### ***Innovations and breakthroughs***

We found that a standardized red wine polyphenolic extract grtaly decreased the proliferation of human liver fibrogenic cells. It also reduced their synthesis of matrix-metalloproteinase-2 and of the tissue inhibitor of matrix metalloproteinase-1, thus suggesting that it could affect the cell ability to remodel the extracellular matrix.

### ***Applications***

The identification of the active compound(s) in the extract could lead to in vivo testing of its anti-fibrotic activity in liver disease.

### ***Terminology***

Liver fibrosis : a common complication of most chronic liver diseases, where an excess of extracellular matrix components are deposited in the liver

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## FIGURE LEGENDS

### **Figure 1. RWPE decreases myofibroblasts proliferation**

Myofibroblasts were grown for either three (A) or seven days (B) in the presence of the indicated concentrations of RWPE. Cell numbers were estimated using the MTT assay and the proliferation index was calculated as described in Materials and Methods. Results are the mean  $\pm$  1 SD of 4 independent experiments conducted in quadruplicate. The effect of RWPE was highly significant using ANOVA ( $p < 0.0001$ ). In control experiments, myofibroblasts were exposed to RWPE for 24 h and the DNA content of the cell layer was measured (C). Results are expressed as the percentage of the values in treated cells as compared to cells treated with the solvent alone and are the mean  $\pm$  1 SD of 3 independent experiments conducted in quadruplicate. There was no significant differences between conditions.

### **Figure 2. Effect of the RWPE on expression of alpha-smooth muscle actin**

A- Myofibroblasts were incubated for seven days in the absence of RWPE (A) or with 50 (B), 75 (C), or 100 (D)  $\mu\text{g/ml}$  RWPE. They were then stained with an antibody to ASMA and a secondary fluorescent antibody. Nuclei were labelled with DAPI. Note that ASMA staining is not significantly different between conditions (whereas the differences in cell density reflect the anti-proliferative action of RWPE). Note also that the morphology of the cells is well preserved at every RWPE concentration. Aliquots of cell extracts grown in the same conditions were analyzed by Western blot (E) simultaneously for ASMA (A) and vimentin (V). F- The signals were quantified as described in Materials and Methods. The graph shows the mean of 2 separate experiments. (squares : actin; triangles : vimentin).

### **Figure 3. Effect of the RWPE on the phosphorylation of MAPK and Akt**

A-Myofibroblasts were pre-incubated for 1 h with the indicated concentrations of

RWPE (in  $\mu\text{g/ml}$ ) or solvent, then exposed for 10 min to 20 ng/ml PDGF-BB or buffer. Identical amounts of cell extracts were analyzed by Western blot with antibodies to phospho-ERK1/ERK2 (top panel) and to total ERKs (bottom panel). The picture is representative of 3 experiments.

B- Quantitative analysis of the experiment shown in A. The activation index refers to the ratio between the levels of phospho-ERK to those of total ERK.

C- Same as in A except that the blot was labelled with an anti-phospho-Akt antibody (top panel) and an antibody to  $\beta$ -actin (bottom panel).

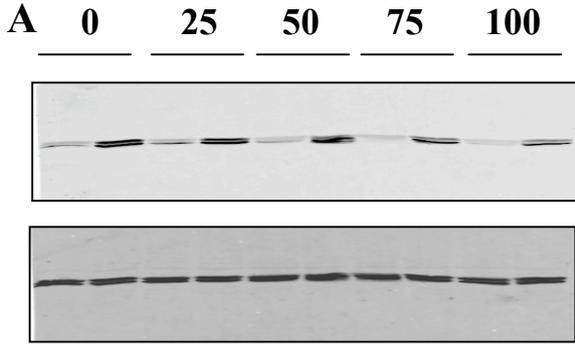
D- Quantitative analysis of the experiment shown in C. The activation index refers to the ratio between the levels of phospho-Akt to those of  $\beta$ -actin.

#### **Figure 4. Effect of the RWPE on MMP-2 and TIMP-1 expression**

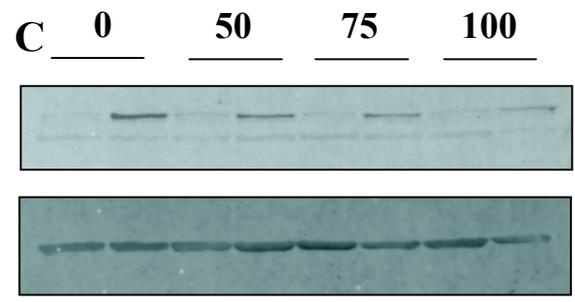
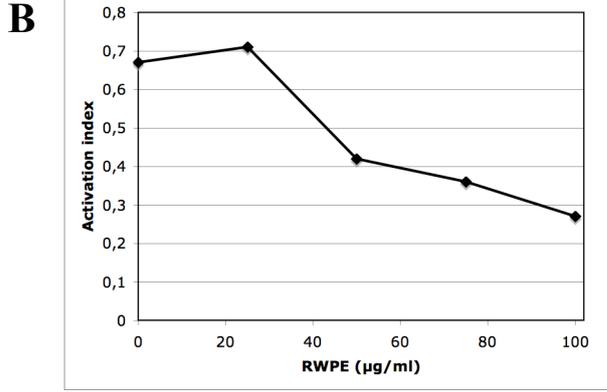
A- Myofibroblasts were incubated for 24 h with the indicated concentrations of RWPE (in  $\mu\text{g/ml}$ ). Aliquots of conditioned medium normalized for the DNA content of the cell monolayers were analyzed on gelatin-containing gels (A). The white bands on the dark background indicate gelatinolytic activity.

Other aliquots were analyzed by Western blot with an antibody against TIMP-1 (B). Another experiment gave similar results.

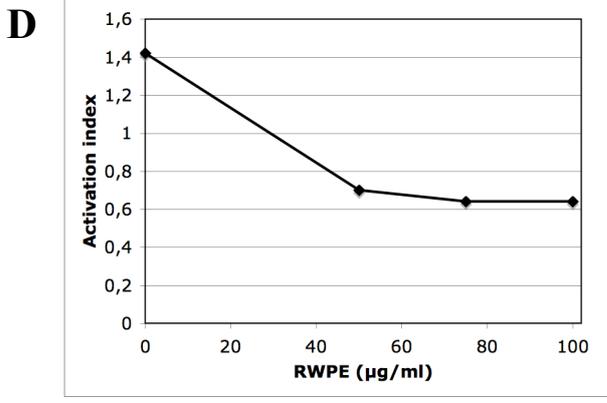
C- Quantitative analysis of the experiments. MMP-2 : mean of duplicate samples; TIMP-1 : mean of 2 independent experiments.

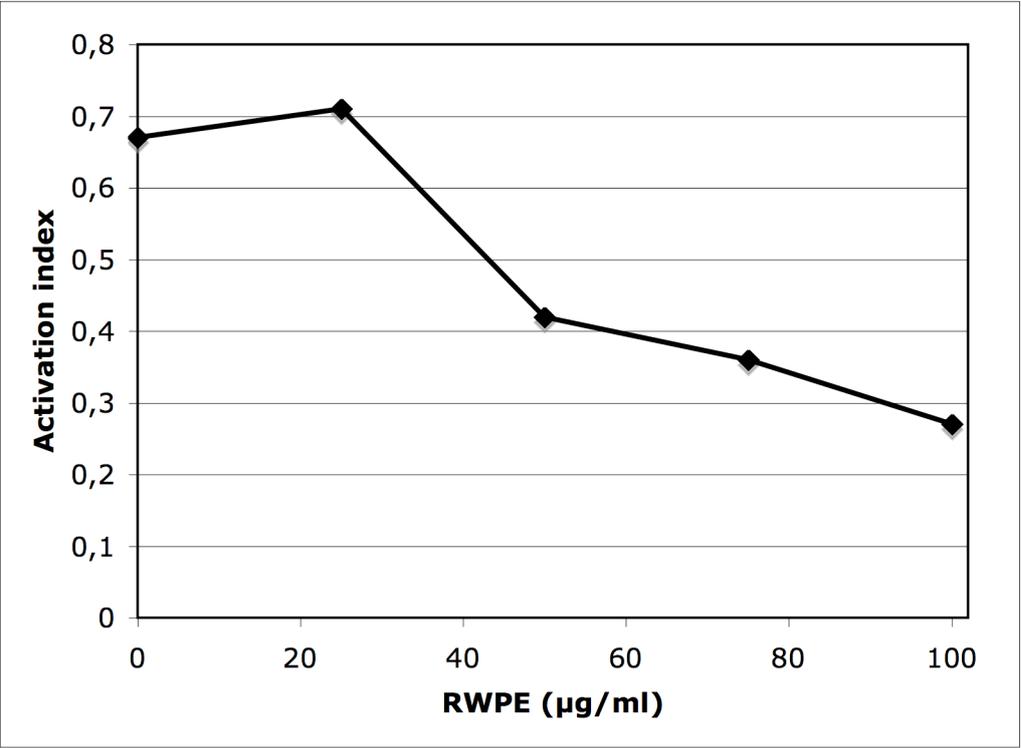


PDGF . + . + . + . + . +



PDGF . + . + . + . + . +





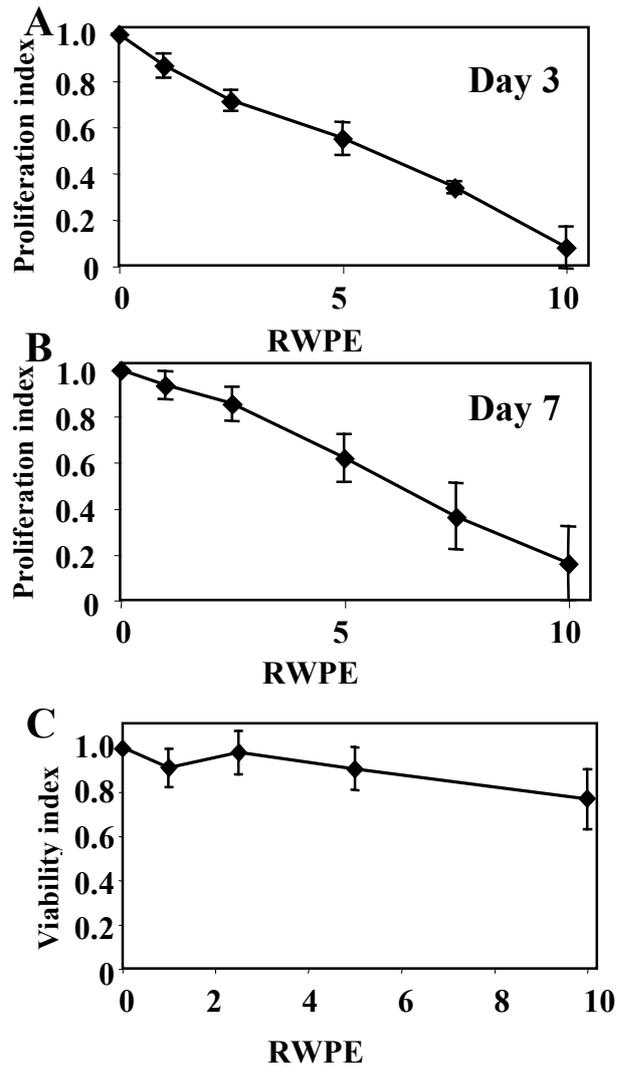
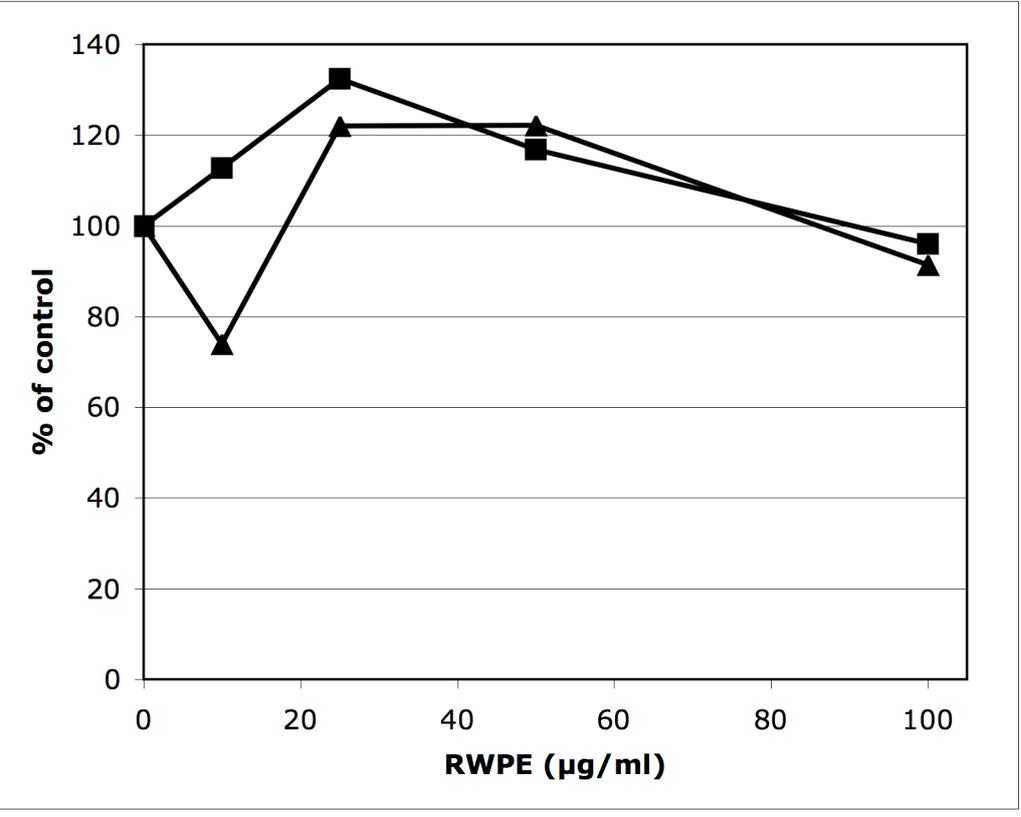


Figure 1



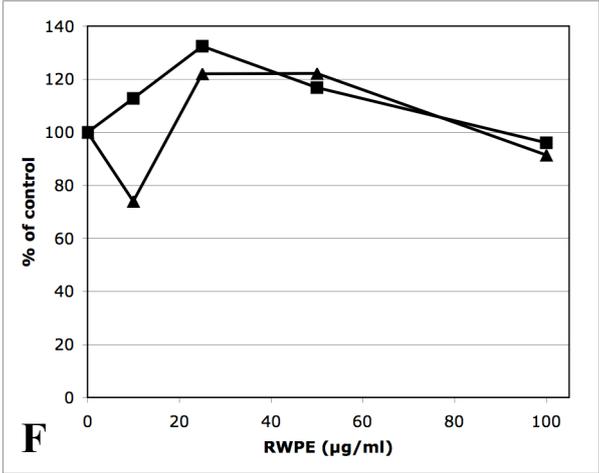
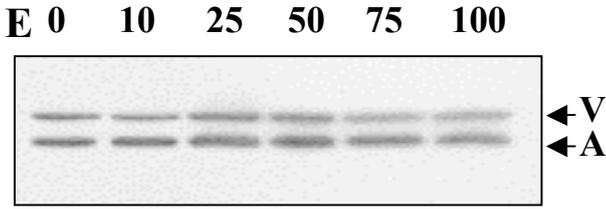
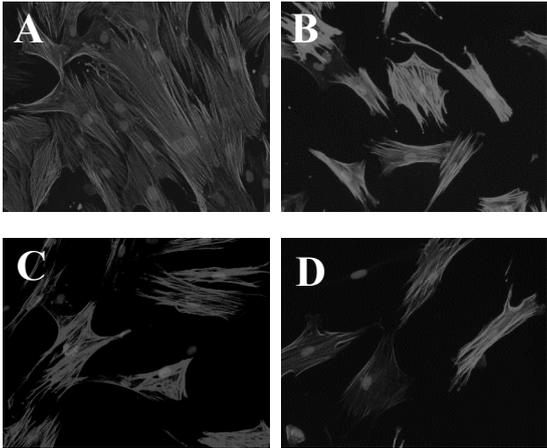
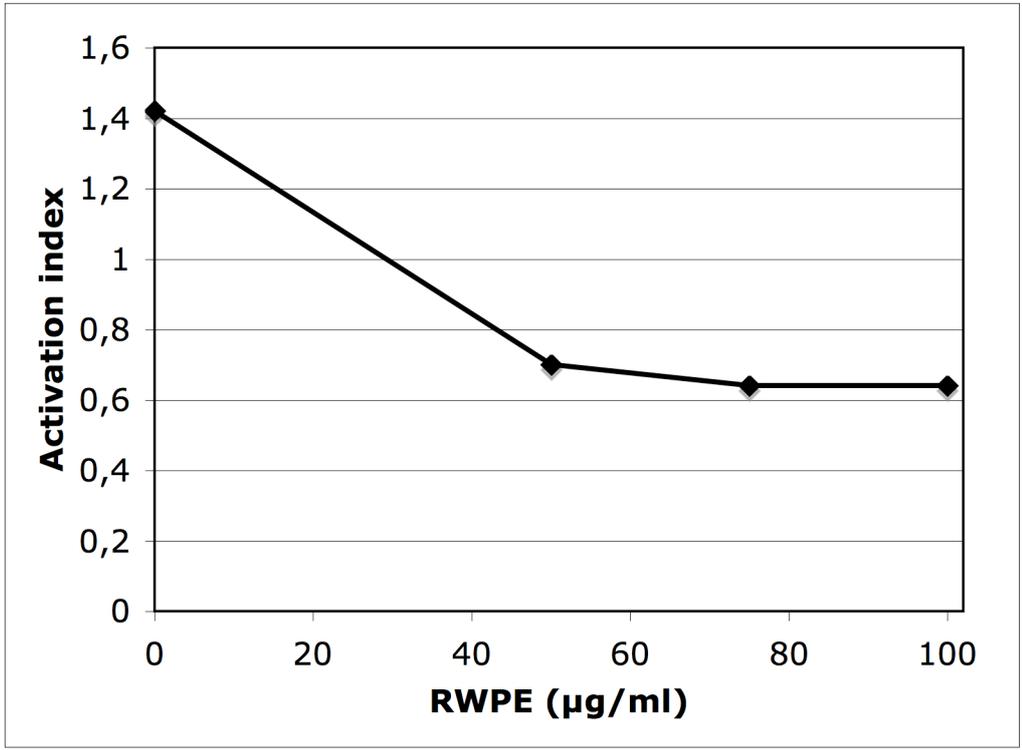
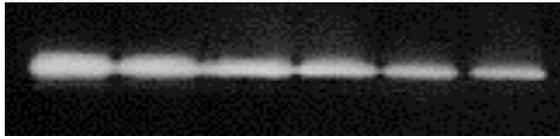


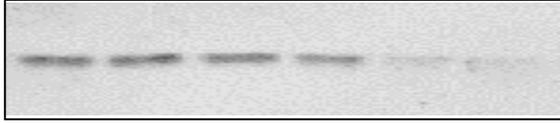
Figure 2



**A** 0 10 25 50 75 100



**B** 0 10 25 50 75 100



**C**

