

Identification of the L,D-Transpeptidases Responsible for Attachment of the Braun Lipoprotein to *Escherichia coli* Peptidoglycan

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The Ldt_{fm} L,D-transpeptidase catalyzes peptidoglycan cross-linking in β -lactam-resistant mutants of *Enterococcus faecium*. Here, we show that in *Escherichia coli* Ldt_{fm} homologues are responsible for attachment of the Braun lipoprotein to murein indicating that evolutionary-related domains have been tailored to use mucopeptides or proteins as acyl acceptors in the L,D-transpeptidation reaction.

The peptidoglycan, which surrounds the bacterial cell, is polymerized from disaccharide-peptide subunits via formation of glycoside and peptide bonds. Variations in the structure of mature peptidoglycan involve mainly the sequence of the stem peptide and its mode of cross-linking (15). In *Escherichia coli*, cross-linking of L-Ala¹-D-Glu²-*meso*DAP³-D-Ala⁴-D-Ala⁵ (DAP: diaminopimelic acid) stem peptides occurs predominantly between the α -carboxyl of D-Ala⁴ of one subunit and the ϵ -amine of *meso*DAP³ of another subunit (4-3 cross-links). Cross-links involving two *meso*DAP³ (3-3 cross-links) are less abundant, representing 3 and 10 % of the total muropeptides content in the exponentially and stationary phases of growth, respectively (14). The 4-3 cross-links are formed by the D,D-transpeptidase activity of high-molecular weight penicillin-binding proteins (PBPs), while the enzymes responsible for the formation of 3-3 cross-links remain unknown in gram-negative bacteria. In gram-positive bacteria, an L,D-transpeptidase (Ldt_{fm}) has recently been shown to catalyze formation of 3-3 cross-links in a β -lactam-resistant mutant of *Enterococcus faecium* selected *in vitro* (11) (Fig. 1A). This enzyme confers resistance by by-passing the β -lactam-sensitive D,D-transpeptidase activity of PBPs (12). The catalytic domain of Ldt_{fm} is the first functionally characterized member of a conserved family of proteins designated ErfK-YcfS-YhnG or pfam 03734 in databases. Additional members of this family from *Enterococcus faecalis* and *Bacillus subtilis* were also recently shown to catalyze peptidoglycan cross-linking *in vitro* (10). In this study, we have investigated by multiple chromosomal deletions and genetic complementation the role of four Ldt_{fm} homologues from *E. coli*.

Four genes, named *erfK*, *ycfS*, *ynhG*, and *ybiS*, encoding proteins which contain a domain related to the catalytic domain of Ldt_{fm} are present in the *E. coli* genome (Fig. 2). Single and multiple deletions of the four genes were constructed in strain BW25113 by the procedure of Datsenko and Wanner (7). The peptidoglycan of the parental strain and of the quadruple mutant (BW25113 Δ 4) was extracted (8) from stationary phase cultures performed in Brain

Heart Infusion (BHI) broth at 37°C. Carbohydrates and proteins were digested with α -amylase and a mixture of proteases (pronase from Roche), respectively (8). The purified peptidoglycan was digested with muramidases and MurNAc was reduced to muramitol using sodium borohydride (2). The resulting muropeptides were separated by rp-HPLC on a C18 column and analyzed by mass spectrometry (2). The relative abundance of muropeptides was estimated by integration of the different peaks of the rp-HPLC profile. The dimer with a monoisotopic mass of 1,722.7, containing a *meso*DAP-*meso*DAP cross-link generated by L,D-transpeptidation, was detected in both strains (peak 5, Fig. 3) indicating that none of the four *Ldt_{im}* homologues was required for formation of 3-3 cross-links. The quadruple deletion suppressed formation of muropeptides eluting in peaks 8, 13, and 14 in the wild-type elution profile (Fig. 3). These muropeptides differed from the monomer eluting in peak 2 and from the dimers eluting in peaks 5 and 9 by a mass of 284.2 corresponding to the mass of the dipeptide Lys-Arg. Tandem mass spectrometry analysis of the monomer in peak 8 showed that Lys-Arg was linked via the ϵ -amine of Lys to the α -carboxyl of *meso*DAP³ of a disaccharide-tripeptide (Boxed in Fig. 1B). Thus, this muropeptide contains a C-terminal dipeptide fragment of the Braun lipoprotein, known to be covalently linked to the peptidoglycan as depicted in Fig. 1B (5). These data indicate that the enzymes responsible for anchoring the major outer membrane lipoprotein to the peptidoglycan of *E. coli* are related to the L,D-transpeptidases that catalyze formation of 3-3 cross-links in gram-positive bacteria. In addition, deletion of the four genes uncovered an additional unknown L,D-transpeptidase since the 3-3 cross-links persisted in the quadruple mutant.

Transcomplementation was performed to determine which of the four genes could restore attachment of the Braun lipoprotein to the peptidoglycan. The *erfK*, *ycfS*, *ynhG*, and *ybiS* genes were independently cloned into the pTrc99a expression vector (1) and the recombinant plasmids were introduced into *E. coli* BW25113 Δ 4. The transformants were grown in BHI

broth containing ampicillin (150 $\mu\text{g/ml}$) and expression of the cloned genes was induced with isopropyl- β -D-thiogalactopyranoside (0.01 mM) at an OD of 0.4 at 600 nm. Following further incubation at 37°C, peptidoglycan was extracted from stationary phase cultures and analyzed as described above for BW25113 Δ 4. Expression of three out of the four genes, *erfK*, *ycfS*, and *ybiS*, restored the covalent anchoring of the lipoprotein to the peptidoglycan (data not shown). Thus, complementation analysis revealed that ErfK, YcfS, and YbiS can independently catalyze the covalent anchoring of the Braun lipoprotein to the peptidoglycan. However, this function appears to be mainly performed by YbiS in the parental strain since deletion of the *ybiS* gene alone suppressed almost completely peak 8 whereas deletion of *erfK*, *ycfS*, and *ynhG*, alone or in combination, had no effect on this peak (data not shown).

The mucopeptide elution profiles of the various strains were also analyzed to determine whether additional functions could be associated with the Ldt_{fm} homologues. Expression of *ynhG* resulted in an increase of peaks 4 and 5, which correspond to mucopeptides containing a 3-3 cross-link (from 0.4 to 7.4 % for peak 4 and from 0.8 to 8.8 % for peak 5). These results suggest that YnhG can contribute to L,D-transpeptidation of peptidoglycan subunits together with the unknown L,D-transpeptidase uncovered by deletion of the four *ldt_{fm}* homologues (see above). Because of the existence of this additional enzyme, it is however not possible to exclude the possibility that the effect of *ynhG* expression on the abundance of 3-3 cross-links is only indirect. For example, formation of tripeptide from pentapeptide by YnhG, as recently shown for the L,D-transpeptidase from *E. faecalis* (10), may modify the relative abundance of the substrates for the D,D-transpeptidation and L,D-transpeptidation reactions.

In conclusion, we have shown that the L,D-transpeptidases for peptidoglycan cross-linking in gram-positive bacteria and for attachment of the Braun lipoprotein to peptidoglycan in *E. coli* belong to the same protein family. The two reactions are expected to involve similar acyl donor but distinct acyl acceptor substrates (Fig. 1). The L,D-transpeptidase of *E. faecium*

(Ldt_{fm}) cleaves the peptide bond between the third and fourth residue of a donor disaccharide-tetrapeptide and links the α -carboxyl of the third residue to the side chain amine at the third position of an acceptor disaccharide-tetrapeptide (Fig. 1A). *In vivo*, this reaction results in the formation of 3-3 cross-links between two stem peptides carried by adjacent glycan strands in the peptidoglycan layer. By analogy, ErfK, YcfS, and YbiS are expected to cleave the peptide bond between *meso*DAP³ and D-Ala⁴ in a donor disaccharide-tetrapeptide stem and to link the α -carboxyl of *meso*DAP³ to the side chain amine of the L-Lys residue located at the C-terminus of the Braun lipoprotein (Fig. 1B). *In vivo*, this reaction results in the anchoring of the C-terminus of the Braun lipoprotein to the peptidoglycan layer. The N-terminal Cys residue of the mature Braun lipoprotein is also modified by the addition of fatty acid residues that insert into the outer membrane. Thus, the Braun lipoprotein, which folds in a stable trimeric structure (16), is thought to contribute to the integrity of the outer envelope structure by connecting the outer membrane to the peptidoglycan (4), although neither the loss of the protein (9) nor its anchoring to the peptidoglycan layer (this work) lead to deleterious phenotypes (data not shown). In gram-positive bacteria surface proteins are anchored to peptidoglycan by sortases which cleave a peptide bond within a sorting signal and link the carboxyl of the C-terminal residue to the side chain at the third position of a disaccharide-peptide (Fig. 1C). In this reaction, the protein acts as the carbonyl donor and the disaccharide-peptide as the acceptor. Sortases and L,D-transpeptidases of the Ldt_{fm} family are structurally unrelated although both types of enzymes function with a catalytic Cys residue (11, 13).

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REFERENCES

1. **Amann, E., B. Ochs, and K. J. Abel.** 1988. Tightly regulated tac promoter vectors useful for the expression of unfused and fused proteins in *Escherichia coli*. *Gene* **69**:301-15.
2. **Arbeloa, A., J. E. Hugonnet, A. C. Sentilhes, N. Josseaume, L. Dubost, C. Monsempes, D. Blanot, J. P. Brouard, and M. Arthur.** 2004. Synthesis of mosaic peptidoglycan cross-bridges by hybrid peptidoglycan assembly pathways in gram-positive bacteria. *J. Biol. Chem.* **279**:41546-56.
3. **Biarrotte-Sorin, S., J. E. Hugonnet, V. Delfosse, J. L. Mainardi, L. Gutmann, M. Arthur, and C. Mayer.** 2006. Crystal structure of a novel beta-lactam-insensitive peptidoglycan transpeptidase. *J. Mol. Biol.* **359**:533-8.
4. **Braun, V.** 1975. Covalent lipoprotein from the outer membrane of *Escherichia coli*. *Biochim. Biophys. Acta.* **415**:335-77.
5. **Braun, V., and K. Rehn.** 1969. Chemical characterization, spatial distribution and function of a lipoprotein (murein-lipoprotein) of the *E. coli* cell wall. The specific effect of trypsin on the membrane structure. *Eur. J. Biochem.* **10**:426-38.
6. **Choi, D. S., H. Yamada, T. Mizuno, and S. Mizushima.** 1986. Trimeric structure and localization of the major lipoprotein in the cell surface of *Escherichia coli*. *J. Biol. Chem.* **261**:8953-7.
7. **Datsenko, K. A., and B. L. Wanner.** 2000. One-step inactivation of chromosomal genes in *Escherichia coli* K-12 using PCR products. *Proc. Natl. Acad. Sci. USA* **97**:6640-5.
8. **Glauner, B.** 1988. Separation and quantification of mucopeptides with high-performance liquid chromatography. *Anal. Biochem.* **172**:451-64.

- 155 9. **Hirota, Y., H. Suzuki, Y. Nishimura, and S. Yasuda.** 1977. On the process of
156 cellular division in *Escherichia coli*: a mutant of *E. coli* lacking a murein-lipoprotein.
157 Proc. Natl. Acad. Sci. USA **74**:1417-20.
- 158 10. **Magnet, S., A. Arbeloa, J. L. Mainardi, J. E. Hugonnet, M. Fourgeaud, L.**
159 **Dubost, A. Marie, V. Delfosse, C. Mayer, L. B. Rice, and M. Arthur.** 2007.
160 Specificity of L,D-transpeptidases from Gram-positive bacteria producing different
161 peptidoglycan chemotypes. J. Biol. Chem. (E-pub)
- 162 11. **Mainardi, J., Fourgeaud M, Hugonnet JE, Dubost L, Brouard JP, Ouazzani J,**
163 **Rice LB, Gutmann L, Arthur M.** 2005. A novel peptidoglycan cross-linking enzyme
164 for a beta-lactam-resistant transpeptidation pathway. J. Biol. Chem. **280**:38146-52.
- 165 12. **Mainardi, J. L., V. Morel, M. Fourgeaud, J. Cremniter, D. Blanot, R. Legrand,**
166 **C. Frehel, M. Arthur, J. Van Heijenoort, and L. Gutmann.** 2002. Balance between
167 two transpeptidation mechanisms determines the expression of beta-lactam resistance
168 in *Enterococcus faecium*. J. Biol. Chem. **277**:35801-7.
- 169 13. **Marraffini, L. A., A. C. Dedent, and O. Schneewind.** 2006. Sortases and the art of
170 anchoring proteins to the envelopes of gram-positive bacteria. Microbiol. Mol. Biol.
171 Rev. **70**:192-221.
- 172 14. **Pisabarro, A. G., M. A. de Pedro, and D. Vazquez.** 1985. Structural modifications
173 in the peptidoglycan of *Escherichia coli* associated with changes in the state of growth
174 of the culture. J. Bacteriol. **161**:238-42.
- 175 15. **Schleifer, K. H., and O. Kandler.** 1972. Peptidoglycan types of bacterial cell walls
176 and their taxonomic implications. Bacteriol. Rev. **36**:407-77.
- 177 16. **Shu, W., J. Liu, H. Ji, and M. Lu.** 2000. Core structure of the outer membrane
178 lipoprotein from *Escherichia coli* at 1.9 Å resolution. J. Mol. Biol. **299**:1101-12.

- 179 17. **Yakushi, T., T. Tajima, S. Matsuyama, and H. Tokuda.** 1997. Lethality of the
180 covalent linkage between mislocalized major outer membrane lipoprotein and the
181 peptidoglycan of *Escherichia coli*. J. Bacteriol. **179**:2857-62.

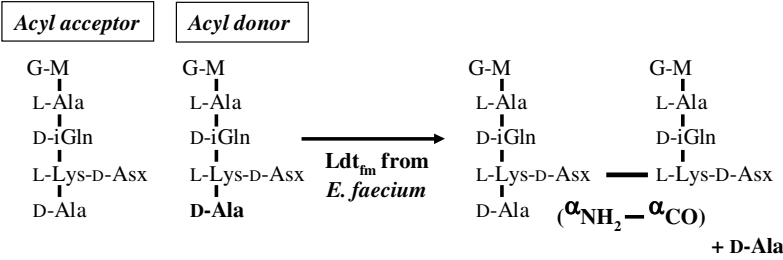
FIGURE LEGENDS

Figure 1. Transpeptidation reactions catalyzed by Ldt_{fm} from *E. faecium*, YbiS from *E. coli*, and by the sortases from *Staphylococcus aureus*. (A) Ldt_{fm} from *E. faecium* catalyzes formation of 3-3 cross-links between two peptidoglycan subunits. The third position of the stem peptide consists of a D-Asp or D-Asn residue (D-Asx) linked to the ε-amine of L-Lys via their β-carboxyl. The 3-3 cross-link connects the α-carboxyl of D-Ala in the donor to the α-amine of D-Asx in the acceptor. (B) Anchoring of the Braun lipoprotein to the peptidoglycan in *E. coli*. The Ser residue at position 2 of the mature lipoprotein is the critical residue of a sorting signal for addressing the protein to the outer membrane (17). Attachment of three fatty-acids (FA) to the N-terminal glyceryl-cysteine residue of the mature protein is responsible for its insertion into the outer membrane (6). The peptide bond formed by the YbiS L,D-transpeptidase links the α-carboxyl of *meso*DAP³ of a disaccharide-peptide to the side chain amine of the C-terminal residue of the Braun lipoprotein (Lys⁵⁸). Muropetide 8 (boxed) consists of a disaccharide-tripeptide substituted by the C-terminal dipeptide of the Braun lipoprotein (Lys⁵⁸-Arg⁵⁷) following cleavage of the Tyr⁵⁶-Arg⁵⁷ peptide bond by pronase during peptidoglycan preparation. (C) Sortases catalyze anchoring of proteins to the peptidoglycan of gram-positive bacteria. The StrA sortase from *S. aureus* cleaves the Thr-Gly peptide bond of the sorting signal (consensus sequence Leu-Pro-X-Thr-Gly; X, any amino acid) and links the α-carboxyl of Thr to the side chain amine at the third position of an acceptor disaccharide peptide (containing L-Lys substituted by a pentaglycine in this bacterium). Residues flanking the sorting signal are represented by dot lines. G-M, GlcNAc-MurNAc. D-iGln, D-isoglutamine.

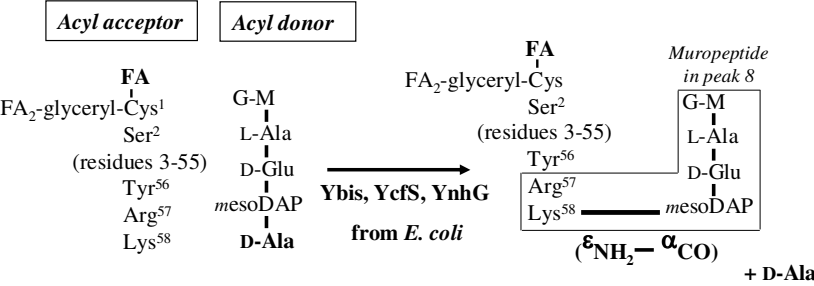
Figure 2. Structure of Ldt_{fm} and related proteins. (A) Domain composition of L,D-transpeptidases from *E. faecium* (Ldt_{fm}), *B. subtilis* (Ldt_{Bs}), and of the four homologues from *E. coli* (ErfK, YcfS, YnhG, and YbiS). The hatched box represents the putative membrane anchor of Ldt_{fm} (11). The boxes labelled I and II represent structural domains of Ldt_{fm} (3). Homologues of the catalytic domain of Ldt_{fm} are present in the other proteins (also labeled as II). LysM designates a putative peptidoglycan-binding module. The four proteins from *E. coli* contain a putative peptide signal (grey boxes). Numbers indicated amino acid positions. (B) Sequence alignment of the putative catalytic domains (domain II). Amino acids conserved in the six proteins or only in the four proteins of *E. coli* are indicated by stars and dots, respectively. The catalytic Cys residue of Ldt_{fm} is indicated by an arrow.

Figure 3. Peptidoglycan composition of the parental strain and of the quadruple mutant BW25113Δ4 obtained by deletion of the *erfK*, *ycfS*, *ynhG*, and *ybiS* genes. (A) rp-HPLC profiles of muropeptides obtained by digestion of the peptidoglycan by muramidases. (B) Identification of muropeptides in the main peaks by mass spectrometry. GM^R, *N*-acetyl-glucosamine linked to reduced *N*-acetyl-muramic acid; GM^A, *N*-acetyl-glucosamine linked to anhydro-*N*-acetyl-muramic acid; Di, dipeptide L-Ala-D-Glu; Tri, tripeptide L-Ala-D-iGlu-*meso*DAP; Tetra, tetrapeptide L-Ala-D-iGlu-*meso*DAP-D-Ala. The type of cross links (3-3 or 4-3) is indicated in parenthesis for dimers. ND, Not detected.

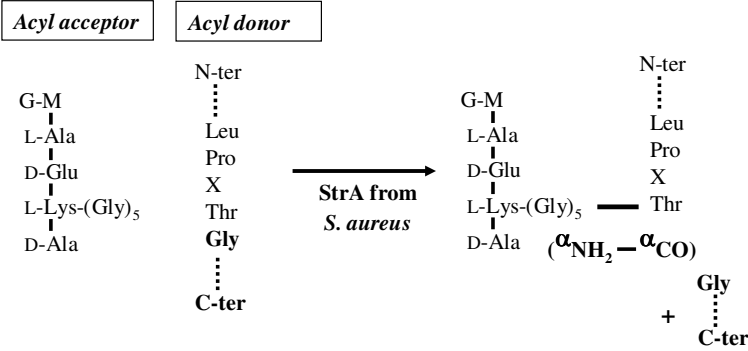
A



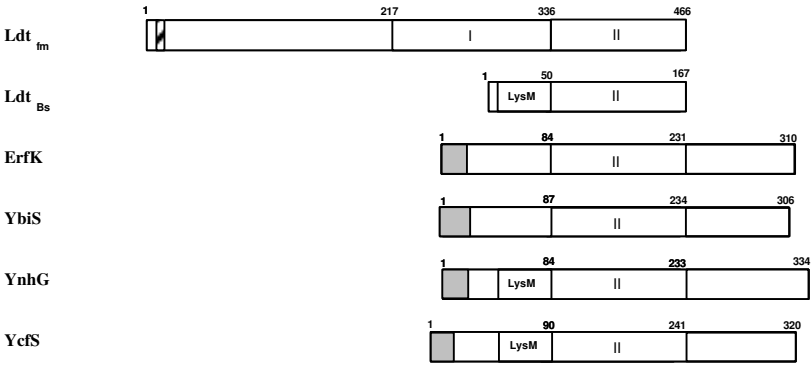
B



C



A



B

Sequence alignment of Ldt_{fm}, Ldt_{Bs}, erfK, ybiS, ynhG, and ycfS. The alignment shows conserved regions across the proteins, with residues numbered 10 to 60, 70 to 120, and 130 to 150. An arrow points to a specific residue in the ycfS sequence.

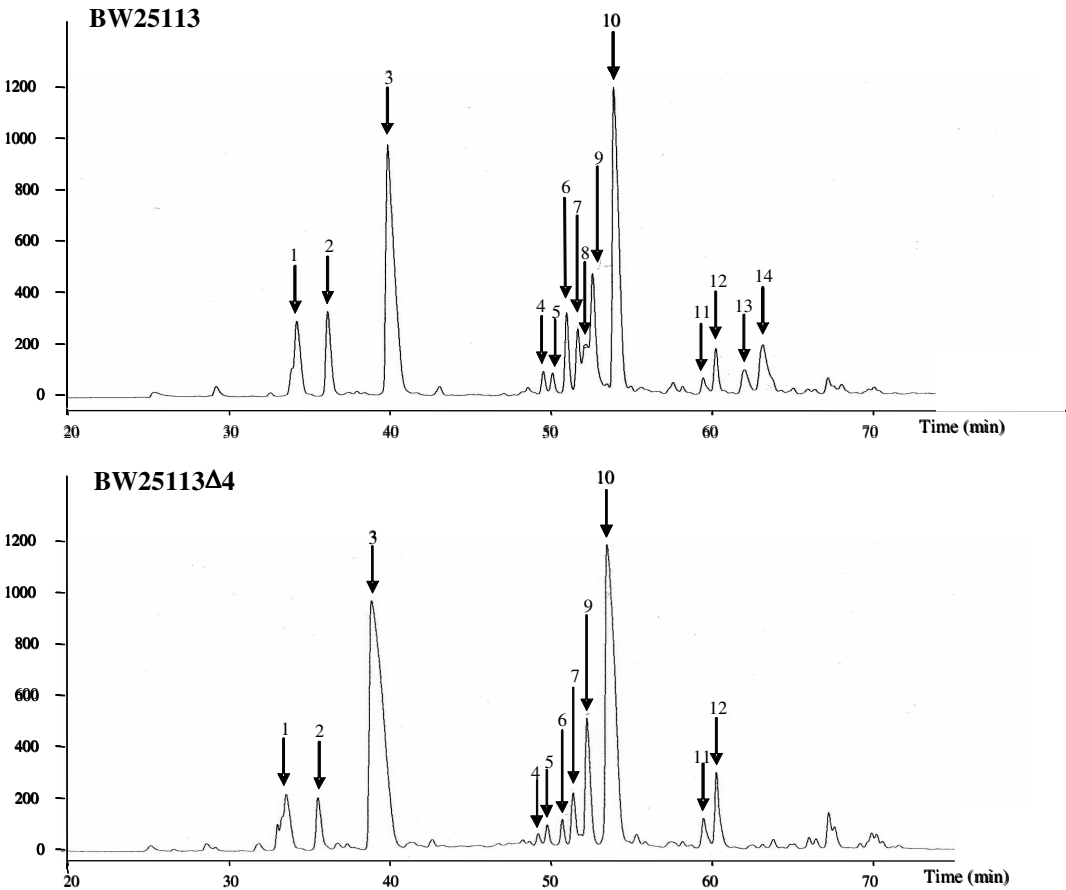
10 20 30 40 50 60
Ldt_{fm} --DHPLIEDT Y---IEVDLE NQHMMYYKDG KVALETID-IV SGKPTTPTPA --GVFYVWNK
Ldt_{Bs} LPDPYTIPTPH ----IAVSIG AKTLTILS-LN NRVMTYPIA VGKILTQTPT --GEFYIINR
erfK IPQQLILPDT VRKGIVVNVA EMRLYYPPD SNTVEVFPPIG IGQAGRETPT -NWVTIVERK
ybiS IPQQLILPDT VHEGIVINSA EMRLYYPPG TNTVIVLPPIG IGQLGKDTPI -NWTTKVERK
ynhG IPSQLLLPDA PRQGIIVNLA ELRLYYPPG ENIVQVYPIG IGLQGLETPV --METRVGQK
ycfS IPLQTLTPDA PREGIVINIA ELRLYYPPG KNSVTVPPIG IGQLGGDTLT PTMVTTVSDK
. * *

70 80 90 100 110 120
Ldt_{fm} EEDATLKGTN -----DDG -----TP YESPVNYWMP ID---WTGVG IHDSDWQPEY
Ldt_{Bs} QRNP-----G -----G ---PFGAYWL SLS--KQHYG IHGTNNPASI
erfK QEAPTWTPTP NTRREYAKRG ESPLPAVPAG PDNPMGLYAI YI---GRLYA IHGTNANFGI
ybiS KAGPTWTPTA KMHAEYRAAG EPLPAVPAG PDNPMGLYAI YI---GRLYA IHGTNANFGI
ynhG IPNPTWTPTA GIRQRSLEAG IKLPPVPAG PNNPLGRYAL RLAHNGEYL IHGTSAPDSV
ycfS RANPTWTPTA NIRARYKAQG IELPAVPAG LDNPMGHAI RLAAAYGGVYL LHGTNADFGI
. * *

130 140 150
Ldt_{fm} GGDLWKTRGS HGCINTPPSV MKELFGMVEK GTPVLVF
Ldt_{Bs} G-----KAVS KGCIRMHNDK VIELASIVPN GTRVTIN
erfK G-----LRVS QGCIRLRNDD IKYLFDNVPV GTRVQII
ybiS G-----LRVS HGCVRLRNED IKFLFEKVPV GTRVQFI
ynhG G-----LRVS SGCIRMNAPD IKALFSSVRT GTPVKVI
ycfS G-----MRVS SGCIRLRDDD IKTLSQVTP GTKVNII
* * * * *

↑

A



B

Peak	Oligomer	Muropeptide (cross-link)	Monoisotopic mass		
			Calculated	Observed for BW25113	Observed for BW25113Δ4
1	monomer	GM ^R -Di	698.29	698.29	698.29
1	monomer	GM ^R -Tri	870.37	870.38	870.38
2	monomer	GM ^R -Tri -Gly	927.39	927.37	927.37
3	monomer	GM ^R -Tetra	941.41	941.38	941.38
4	dimer	GM ^R -Tri-Gly / GM ^R -Tri (3-3)	1,779.75	1,779.80	1,779.80
5	dimer	GM ^R -Tri / GM ^R -Tri (3-3)	1,722.73	1,722.78	1,722.78
6	dimer	GM ^R -Tri -Gly / GM ^R -Tetra (3-3 or 4-3)	1,850.79	1,850.74	1,850.74
7	dimer	GM ^R -Tetra / Tetra (4-3)	1,384.59	1,384.64	1,384.63
8	monomer	GM ^R -Tri -L-Lys-L-Arg	1,154.57	1,154.57	ND
9	dimer	GM ^R -Tetra / GM ^R -Tri (3-3 or 4-3)	1,793.77	1,793.86	1,793.79
10	dimer	GM ^R -Tetra / GM ^R -Tetra (4-3)	1,864.80	1,864.74	1,864.85
11	trimer	GM ^R -Tetra / GM ^R -Tetra / GM ^R -Tetra (4-3)	2,788.20	2,788.13	2,788.26
12	monomer	GM ^A -Tetra	921.38	921.36	921.36
13	dimer	GM ^R -Tri / GM ^R -Tri -L-Lys-L-Arg (3-3)	2,006.93	2,006.98	ND
14	dimer	GM ^R -Tetra / GM ^R -Tri -L-Lys-L-Arg (4-3)	2,077.96	2,078.02	ND