



HAL
open science

Regulation of Bcl-2-family proteins in myeloma cells by three myeloma survival factors: interleukin-6, interferon-alpha and insulin-like growth factor 1.

Michel Jourdan, John de Vos, Nadir Mechti, Bernard Klein

► To cite this version:

Michel Jourdan, John de Vos, Nadir Mechti, Bernard Klein. Regulation of Bcl-2-family proteins in myeloma cells by three myeloma survival factors: interleukin-6, interferon-alpha and insulin-like growth factor 1.. Cell Death and Differentiation, 2000, 7 (12), pp.1244-52. 10.1038/sj.cdd.4400758 . inserm-00130981

HAL Id: inserm-00130981

<https://inserm.hal.science/inserm-00130981>

Submitted on 14 Feb 2007

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

**REGULATION OF BCL-2-FAMILY PROTEINS IN MYELOMA CELLS BY THREE
MYELOMA SURVIVAL FACTORS: INTERLEUKIN-6, INTERFERON-ALPHA AND
INSULIN-LIKE GROWTH FACTOR 1**

Michel Jourdan¹, John De Vos¹, Nadir Mechti¹, Bernard Klein^{1,2}

Running title: Bcl-2-family proteins in myeloma cells

¹INSERM U475 and Unit for Cellular Therapy, CHU Montpellier, 34197 Montpellier, France

²corresponding author: Bernard Klein, INSERM U475, 99 rue Puech Villa, 34197 Montpellier, France. tel: 33-(0)4 67 63 61 25;
fax: 33-(0)4 67 04 18 63; e-mail: klein@montp.inserm.fr

Summary

As survival regulation is a key process in multiple myeloma biology, we have studied the Bcl-2 family proteins that can be regulated by 3 myeloma cell survival factors: interleukin-6 (IL-6), interferon- α (IFN- α) and insulin-like growth factor 1 (IGF-1). Eleven myeloma cell lines, whose survival and proliferation are dependent on addition of IL-6, variably expressed 10 anti-apoptotic or pro-apoptotic proteins of the Bcl-2-family. When myeloma cells from 4 cell lines were IL-6 starved and activated with IL-6 or IFN- α , we observed that only Mcl-1 expression was up-regulated with myeloma cell survival induction. Nor was obvious regulation of these 10 pro-apoptotic or anti-apoptotic proteins found with IGF-1, another potent myeloma cell survival factor. Our results indicate that the myeloma cell survival activity of IL-6 linked to Bcl-xL regulation cannot be generalized and emphasize that Mcl-1 is the main target of IL-6 and IFN- α stimulation. However, other changes in the activity of the Bcl-2 protein family or other apoptosis regulators must be identified to elucidate the IGF-1 action mechanism.

Key words: myeloma, IL-6, IFN, IGF, apoptosis

Abbreviations: BH, Bcl-2 homology domain; BSA, bovine serum albumin; HMCL, human myeloma cell line; IGF-1, insulin-like growth factor 1; IL-6, interleukin-6; IFN- α , interferon-alpha; PTP, permeability transition pore

Introduction

Multiple myeloma is a B cell neoplasia characterized by the bone marrow accumulation of clonal malignant plasma cells. In the majority of patients, tumor cells do not proliferate or proliferate very slowly^{1,2}. Thus, a long survival of myeloma cells can explain the tumor growth. Interestingly, normal plasma cells can also survive for several months *in vivo*³. Medullary stromal cells are probably critical in promoting this survival of normal or malignant plasma cells *in vivo*. Indeed, normal plasma cells generated *in vitro* rapidly die but medullary stromal cells can trigger their survival *in vitro*⁴. Malignant plasma cells from patients with chronic disease rapidly apoptose as soon as they are separated from stromal cells⁵. However, the survival factors produced by stromal cells are poorly identified. IL-6 and cytokines of the IL-6 family⁶⁻⁸ are important growth factors but are not sufficient to prevent apoptosis of purified myeloma cells⁵. IFN- α ^{9,10} and IGF-1¹¹⁻¹³ are also myeloma cell survival factors.

Induction of apoptosis involves a family of death proteases, called caspases, which are activated in a proteolytic cascade to execute the cell death program. Caspase activation is controlled by a growing family of anti-apoptotic and pro-apoptotic proteins (see Gross et al.¹⁴ and Adams et al.¹⁵ for review). These proteins possess 1 to 4 conserved motifs, called Bcl-2 homology domains (BH-1 to BH-4) and may form homo- or hetero-dimers. In numerous cells, the survival activity of growth factors is determined in part by the relative concentrations of pro- and anti-apoptotic Bcl-2-family members. The most widely studied anti-apoptotic proteins are Bcl-2 and Bcl-xL with four BH domains and a transmembrane domain that allows their location to the outer mitochondrial membrane. They are thought to regulate the permeability transition pore (PTP) and block both the mitochondrial exit of cytochrome *c* and the activation of procaspase 9 by APAF-1-cytochrome *c* complexes. Like Bcl-2, Mcl-1 has four BH domains and a transmembrane domain and may function as Bcl-2 and Bcl-xL, but this

has yet to be proved¹⁴. Bag-1 does not belong to the Bcl-2-family proteins but has an anti-apoptotic activity probably by increasing Bcl-2, Bcl-xL or Mcl-1 activity¹⁶. Bfl-1/A1 has 4 BH domains but no transmembrane domain. There are also numerous pro-apoptotic proteins. The most thoroughly studied are Bax and Bak, which have three BH domains and a transmembrane domain and are translocated to the mitochondrial membrane when an apoptotic signal is delivered. They are believed to deliver apoptosis by opening the PTP, thus affecting the electrical potential gradient of the mitochondria and favoring cytochrome c release. They can also dimerize with anti-apoptotic proteins, thus encouraging PTP to open and APAF-1 to activate. Roughly eight pro-apoptotic proteins have only a BH3 domain with or without a transmembrane domain. Bid and Bad are the most widely studied and are mostly competitors of anti-apoptotic proteins^{14,15}.

Presently, only a few members of the Bcl-2 family have been studied in myeloma cells. Several groups have shown a high expression of Bcl-2 in myeloma cell lines, in primary myeloma cells as well as in normal plasma cells^{17,18}. This high Bcl-2 expression in myeloma cells did not correlate with patient survival^{19,20} and even inversely correlated with the plasma cell proliferation *in vivo*²¹. IL-6, a major myeloma cell survival factor, did not affect Bcl-2 expression²²⁻²⁴. Bcl-xL was also expressed in a majority of myeloma cell lines or primary myeloma cells^{20,21}. In the U266 and MDN myeloma cell lines, an increase in Bcl-xL expression was recently shown to be associated with myeloma cell survival induced by IL-6^{24,25}. However, IL-6 did not regulate Bcl-xL expression in the RPMI 8226 myeloma cell line although it blocked apoptosis induced by dexamethasone in this cell line²⁶. Recently, Puthier et al. found that IL-6 regulated the Mcl-1 anti-apoptotic protein in the MDN IL-6-dependent cell line²⁴. Among the pro-apoptotic proteins, Bax expression was documented in primary myeloma cells²⁰ and in myeloma cell lines²². Bad was weakly expressed in myeloma cell lines and its phosphorylation by IL-6 was not detected²⁷. Finally, Bcl-xS was not detected in myeloma cells^{20,21}.

Study of the mechanisms controlling malignant plasma cell survival is limited by the difficulty in obtaining a sufficient number of tumor cells from patients. For this reason, the few studies dealing with this topic have used either cells or cell lines obtained from patients with terminal disease and extramedullary proliferation. From these patients, we obtained several myeloma cell lines whose survival and proliferation require adding exogenous IL-6. These cell lines very closely resemble the myeloma stem cells that are present in patients with terminal disease, as previously discussed²⁸. Using these cell lines, we have shown that IFN- α is also a potent survival factor⁹ and that IGF-1 is a survival and proliferation factor¹³. In the current study, we investigated the expression of 10 Bcl-2-family proteins in 11 myeloma cell lines whose survival requires adding an exogenous cytokine. We also studied how IL-6, IFN- α or IGF-1 regulates these various proteins in 4 cell lines. We report that Mcl-1 was the only one of the 10 Bcl-2-family proteins regulated by IL-6 and IFN- α . None of these 10 proteins was regulated by IGF-1.

Results

Comparative analysis of the survival activity of IL-6, IFN- α and IGF-1 on myeloma cells

We used myeloma cell lines whose survival and proliferation are dependent on addition of IL-6. In previous studies, we have shown that IFN- α ⁹ and IGF-1¹³ are also survival factors for these cell lines. We used serum-free culture conditions to avoid the confounding effects of IGF-1 and IGF-binding proteins present in serum and to compare the myeloma cell survival activity of IL-6, IFN- α and IGF-1. Detailed results of one representative experiment with the XG-13 HMCL are outlined in Figure 1 and the mean results of 3 to 6 experiments obtained with four HMCL are displayed in Table 1. When myeloma cells of the four cell lines were IL-6 and FCS starved, a high percentage of cells bound annexin V (Table 1). On the other hand, annexin V⁺ cells had a DNA profile of apoptotic cells. It should be noted that during the first 24 hours of culture in serum free conditions, more than 85% of the cells failed to incorporate trypan blue, whereas a high percentage of cells was stained with annexin V. We have confirmed that these annexin V⁺ myeloma cells were apoptotic cells, in agreement with previous findings²⁹. Indeed, when annexin V⁺ and annexin V⁻ cells were sorted and labeled with propidium iodide, annexin V⁻ cells had a PI fluorescence profile of viable cells with a double peak characteristic of cells in the G1 phase and of cells in the G2 and M phases (Figure 2).

Thus, using annexin V staining, we found that 69% of the XG-13 cells were apoptotic after 1 day of culture without FCS and exogenous cytokines in the experiment outlined in Figure 1. Adding IL-6 partially but significantly ($P = .011$) protected from apoptosis (Table 1). IFN- α and IGF-1 was as potent as IL-6 to reduce apoptosis (Figure 1 and Table 1, $P = .009$ and $P = .001$ respectively). Similar results were obtained with 3 other myeloma cell lines: XG-1, XG-2 and XG-6 (Table 1). In previous papers we showed that the cytokine concentrations used here were optimal for inducing the

survival of these myeloma cell lines and that no significant further protection was found when either IL-6 and IFN- α^9 or IL-6 and IGF-1 were added¹³.

Expression of Bcl-2-family proteins in myeloma cells

In order to identify the anti-apoptotic and pro-apoptotic proteins potentially involved in the control of apoptosis, we investigated the expression of Bcl-2-family members in a panel of 11 IL-6-dependent myeloma cell lines harvested during the exponential growth phase. Figure 3A illustrates the results from Western blot analyses of anti-apoptotic proteins. For each cell line, the same amount of proteins were loaded per lane. All cell lines expressed a similar level of Mcl-1. Bcl-2 and Bcl-xL were variably expressed. Bcl-2 was weakly expressed in 4 out of the 11 cell lines (XG-1, XG-3, XG-7 and XG-14) and highly expressed in XG-5. Bcl-xL was weakly expressed in 2 cell lines (XG-3 and XG-13). Bag-1 was also detected in all cell lines at a variable level. Bfl-1/A1 was found in XG-12 only. Concerning the pro-apoptotic proteins (Figure 3B), Bax was weakly expressed in XG-1, XG-2, XG-5, XG-13 and XG-14. Bak and Bid were variably expressed by all cell lines excepted for XG-7, which was negative for Bid. Finally, Bcl-xS and Bad could not be detected in the 11 cell lines (results not shown).

Regulation of Bcl-2-family protein expression by IL-6, IFN- α or IGF-1

We investigated the proteins that were regulated by IL-6, IFN- α or IGF-1. Cells were IL-6 and FCS starved for one hour and stimulated by IL-6, IFN- α , IGF-1 or no cytokine for several hours. Detailed results for XG-6 are shown in Figure 4. IL-6 and FCS starvation for 2 to 24 hours resulted in a progressive disappearance of Mcl-1 whereas the levels of Bcl-xL and Bcl-2 were not reproducibly affected in three separate experiments. This Mcl-1 disappearance was associated at 6 and 24 hours with an increased percentage of annexin V positive cells, whereas the myeloma cell count remained constant and the percentage of viable cells as assayed by trypan blue

HAL author manuscript inserm-00130981, version 1

exclusion remained high (Figure 4). Adding IL-6 reversed Mcl-1 disappearance. IL-6 slightly increased Bcl-2 and Bcl-xL levels in the experiment outlined in Figure 4. However, this slight increase was not reproduced in other separate experiments. Adding IFN- α also resulted in a dramatic up-regulation of Mcl-1 without reproducibly affecting the levels of Bcl-xL. Bcl-2 was slightly up-regulated by IFN- α in other separate experiments. Finally, we found no IGF-1 up-regulation of any of the anti-apoptotic proteins tested (Mcl-1, Bcl-xL, Bcl-2 or Bag-1). In these experiments, actin expression was used as a loading control for cells cultured for the first six hours since its expression remained stable. However, a decrease in actin expression was detected after 24 hours of culture with no cytokine, which fits its identification as a caspase target³⁰. Accordingly, we used MAPK (ERK1 and ERK2) expression as a loading protein control for experiments shown in Figure 4 because its expression did not vary with the level of apoptosis. Results obtained with XG-6 were further confirmed with 3 other cell lines (XG-1, XG-2 and XG-13). As shown in Figure 5, Mcl-1 expression is highly and rapidly regulated in the myeloma cell lines by the 2 myeloma survival factors, IL-6 and IFN- α , whereas Bcl-xL levels were not significantly affected. As for the XG-6 cells IFN- α slightly increased Bcl-2 in XG-1 cells and IGF-1 did not affect the levels of Mcl-1, Bcl-2 or Bcl-xL in XG-1 and XG-2 HMCLs. A slight up-regulation of Mcl-1 by IGF-1 was found in XG-13 cells in one experiment (Figure 5); however, this was not reproducible in 2 other separate experiments.

This lack of Bcl-xL regulation by IL-6 or IFN- α contradicts previous studies suggesting that IL-6 may promote myeloma cell survival by up-regulating Bcl-xL expression in the U266²⁵ or the MDN²⁴ myeloma cell lines. In our experiments we used serum-free culture conditions. However, we also found that only Mcl-1 and not Bcl-xL was reproducibly increased by IL-6 when myeloma cells were IL-6 and FCS starved and then cultured with FCS (Figure 6). FCS alone increased MCL1 expression (Figure 6) in agreement with its myeloma cell survival activity (unpublished observations). As Bcl-xL

is known to have a longer half-life than Mcl-1, XG-6 myeloma cells were IL-6 starved for 24 hours in presence of FCS and cultured with or without IL-6 for 96 hours. As shown in Figure 7, IL-6 starvation induced apoptosis in myeloma cells and accordingly to our previous results in serum-free culture conditions, we found a strong decrease in Mcl-1 expression following IL-6 withdrawal that was abolished by adding IL-6. On the contrary, we found no decrease or increase of Bcl-xL levels. Same results were obtained with the XG-1 and XG-2 myeloma cell lines in long-term culture conditions (96 hours, results not shown). Finally, to confirm the lack of Bcl-xL regulation by either IL-6 or IFN- α , we used a second different anti-Bcl-xL antibody that yielded similar results (results not shown). In addition, we checked that these two antibodies specifically recognized recombinant Bcl-xL (results not shown).

For the pro-apoptotic proteins, we found no regulation or induction of Bax, Bak (Figure 4), Bid, Bcl-xS and Bad expression (results not shown).

Differential activation of Stat3 by IL-6, IFN- α and IGF-1.

We previously reported that IL-6 and IFN- α , unlike IGF-1, activated Stat3 in XG-1 and XG-2 cell lines^{9,13}. We confirmed in the present study that Stat3 was rapidly phosphorylated after activation by IL-6 and IFN- α in 2 other cell lines, XG-6 and XG-13 (Figure 8). Stat3 phosphorylation lasted for more than 6 hours. As expected, IGF-1 did not induce Stat3 phosphorylation in these two cell lines (Figure 8). However, IL-6, IFN- α and IGF-1 induced ERK1 and ERK2 phosphorylation.

Discussion

As the survival regulation of tumor cells is a key process in multiple myeloma biology, the aim of the current study was to identify the anti-apoptotic and pro-apoptotic proteins of Bcl-2-family proteins that were regulated in myeloma cells by three survival factors: IL-6, IFN- α and IGF-1. We used myeloma cell lines whose survival and proliferation are dependent on addition of exogenous growth factors²⁸.

We used serum-free culture conditions to avoid the interference of serum proteins with these growth factors, in particular the IGF binding proteins. In addition, FCS contains unidentified survival factors that might mask the effect of recombinant cytokines. In serum-free culture conditions, the level of apoptosis was more or less marked depending on the cell lines and on various parameters such as initial cell count and viability. Apoptosis was measured by the binding of annexin V due to the exposure of phosphatidylcholine outside the cell. Sorting the annexin V⁺ and annexin V⁻ cells and staining DNA with propidium iodide, the annexin V⁻ cells were proved to have the DNA profile of viable cells, unlike annexin V⁺ cells. We also found apoptosis induction in myeloma cells after IL-6 starvation by internucleosomal cleavage of DNA (results not shown). For all the cell lines, adding IL-6, IFN- α or IGF-1 significantly reduced the apoptosis and the effect was even more pronounced after 24 hours of culture. The three cytokines yielded a significant level of survival and had no additive survival effects when combined.

We investigated 10 proteins of the Bcl-2 family that were expressed in a panel of 11 myeloma cells during exponential growth. Bcl-2, Bcl-xL and Mcl-1 anti-apoptotic proteins were detected in all cell lines. Their expressions varied according to the different cell lines, except for Mcl-1, and did not statistically correlate with each other. Bag-1 was also detected in all cell lines. Bfl-1/A1 expression in only one out of 11 cell lines is noteworthy because earlier studies have shown the association of Bfl-1/A1

expression with the survival of murine plasma cells³¹. We also looked for the expression of anti-apoptotic proteins. Bak and Bax were variably expressed in all cell lines, the latter weakly expressed in 5 cell lines. We detected Bid in 10 out of 11 cell lines. Bad was not detected, which may be attributed to its usual very low expression in myeloma cells²⁷.

As the susceptibility to apoptosis is determined in part by the relative concentrations of pro- and anti-apoptotic Bcl-2-family members, we studied their fluctuation in four myeloma cell lines induced into apoptosis after withdrawal of survival factors. We also studied their fluctuation when cells were rescued from apoptosis by adding three different survival factors: IL-6, IFN- α or IGF-1. In the four cell lines, Mcl-1 was the only anti-apoptotic protein whose expression dramatically decreased when cells were IL-6 starved. Removal of cytokines did not reproducibly affect the expression of other anti-apoptotic proteins (Bcl-xL, Bcl-2, Bag-1); nor did this affect the expression of the pro-apoptotic proteins Bax, Bak or Bid. Bad remained undetectable when cells underwent apoptosis. Adding IL-6 or IFN- α up-regulated Mcl-1 expression without affecting the expression of other anti-apoptotic or pro-apoptotic members. Interestingly, adding IGF-1, as potent a survival factor as IL-6 or IFN- α , did not induce Mcl-1 expression nor affect the other Bcl-2 members. Using 4 cell lines whose survival strictly depends on adding exogenous cytokines, we failed to confirm previous results obtained with the MDN IL-6-dependent myeloma cell line, showing that Bcl-xL was up-regulated by adding IL-6²⁴. In the U266 cell line, a blockage of Stat3 by a dominant-negative Stat3 induces apoptosis in association with a decrease in Bcl-xL expression²⁵. To rule out a technical artefact, this lack of Bcl-xL regulation was confirmed with a second anti-Bcl-xL antibody. We checked the specificity of these antibodies by immunoblot using the recombinant Bcl-xL protein. Another possibility to explain the lack of Bcl-xL regulation is that, contrary to Puthier et al.²⁴, we used serum-free culture conditions to comparatively study the effects of IL-6, IFN- α and IGF-1. Indeed Bcl-xL has a longer half-life compared to Mcl-1 and the duration of IL-6 withdrawal can be too short to get a Bcl-xL down-

regulation. However, we also found no Bcl-xL down-regulation when myeloma cells were starved of IL-6 and cultured with FCS for 96 hours whereas a great fraction of myeloma cells apoptosed. Addition of IL-6 did not result in Bcl-xL up-regulation. In agreement with our findings in serum-free conditions, we found that Mcl-1 was decreased after IL-6 withdrawal and increased after IL-6 addition in these long-term culture experiments. Thus, a change in the Bcl-xL level when myeloma cells are induced to survive with IL-6 is not a common characteristic and may be restricted to particular cell lines. On the other hand, the Mcl-1 level appears to be strongly regulated by IL-6 but also by IFN- α in all the cell lines we have studied so far. Previous studies have shown that Mcl-1 expression was correlated with the survival of normal B lymphocytes³² or neutrophils³³. Transfection of Mcl-1 cDNA in hematopoietic cells makes it possible to promote their survival in culture conditions favoring apoptosis³⁴. Increased survival of hematopoietic and lymphoid cells was found in Mcl-1 transgenic mice. In addition, myeloid cell lines were obtained from these Mcl-1 transgenic mice³⁵.

A common transduction pathway induced in myeloma cell lines by IL-6 and IFN- α is the activation of JAK kinases and tyrosine phosphorylation of STAT factors, mainly Stat3^{36,37}. We have extended our previous observations⁹ and show that IL-6 and IFN- α induced Stat3 phosphorylation in the 4 cytokine-dependent cell lines studied here. Stat1 was inconsistently found phosphorylated in myeloma cell lines after IL-6 or IFN- α activation, in agreement with previous observations^{9,38}. It was recently shown that blocking JAK activity and/or Stat3 activation by tyrphostin AG490 or a Stat3 mutant induced U266 cells to apoptosis in association with a down-regulation of Bcl-xL²⁵. In the MDN myeloma cell line, blocking JAK activation by tyrphostin AG490 also blocked Mcl-1 induction by IL-6³⁹. We have recently shown that inhibiting JAK2 and JAK1 activation by tyrphostin AG490 induced a strong apoptosis in the IL-6-dependent myeloma cell lines used in the current study, in association with a blockage of Stat3 activation and ERK2 activation⁴⁰. These studies argue that the myeloma cell survival activity of IL-6 or IFN- α

is linked to JAK/Stat pathway activation, yielding an up-regulation of Mcl-1 or Bcl-xL expression depending on the myeloma cell line.

The myeloma cell survival activity of IGF-1 is not associated with an up-regulation of Mcl-1 or of other anti-apoptotic proteins, a finding consistent with its lack of JAK/Stat pathway activation. These results are in agreement with our previous findings, showing that the myeloma cell survival factor activity of IGF-1 is not linked to gp130 IL-6 transducer activation through stimulation of an autocrine gp130 cytokine¹³. IGF-1 is known to activate the Ras/MAP kinase and PI-3 kinase pathway in several cell lines; however, in preliminary experiments we failed to inhibit its survival activity by adding wortmannin, a PI-3 kinase inhibitor or PD098059, a MAPK kinase inhibitor (results not shown). Apoptosis is not regulated only by the relative levels of the anti-apoptotic and pro-apoptotic proteins but also by their phosphorylation or activation of other apoptosis effectors. In addition, all the members of the Bcl-2 family have not been studied here due to the lack of commercially available reagents. As IGF-1 is present in high amounts in the bone marrow, the tissue in which myeloma cells develop, the understanding of the mechanisms involved in the survival activity of IGF-1 requires further study.

Material and methods

Cytokines and antibodies.

Purified human recombinant IL-6 was kindly provided by Sandoz Forschungsinstitut (Vienna, Austria) and human recombinant IFN- α -2b by Schering Plough (Dardilly, France). Recombinant purified IGF-1 was purchased from R&D (Minneapolis, MN, U.S.A.), monoclonal anti-Bcl-2 antibody from Dako (Copenhagen, Denmark), anti- β actin from Sigma (St Louis, MO, U.S.A.) and anti-Stat3 from Transduction Laboratories (Lexington, KY, USA). Rabbit or goat polyclonal antibodies specific for Bcl-x, Mcl-1, Bag-1, Bak, Bfl-1/A1, Bid and ERK2 were obtained from Santa Cruz Biotechnology (Santa Cruz, CA, USA), Bad, phospho-Stat3 and phospho-ERK1 and ERK2 from New England Biolabs (Beverly, MA, U.S.A.), Bax from Upstate Biotechnology (Lake Placid, NY, U.S.A.) and a second anti-Bcl-x from Transduction Laboratories. Recombinant human Bcl-xL was obtained from Santa Cruz Biotechnology.

Human myeloma cell lines (HMCL).

XG-1, XG-2, XG-3, XG-4, XG-5, XG-6, XG-7, XG-10, XG-12, XG-13 and XG-14 myeloma cell lines were obtained from 11 patients with terminal disease. The HMCL were established as described previously²⁸. These HMCLs are true myeloma cell lines: CD19⁻, CD20⁻, CD138⁺, CD38⁺⁺, cytoplasmic immunoglobulin⁺⁺. They are not infected with EBV²⁸. Their growth is dependent upon addition of exogenous IL-6, as does the growth of primary cells. Cells were free of mycoplasma contamination as assayed using the Boehringer Mannheim kit of detection (Mannheim, Germany).

Culture of myeloma cells

The HMCLs were routinely cultured with 2 ng/mL of IL-6 in RPMI 1640 supplemented with 10% fetal calf serum (FCS) and 5 x 10⁻⁵ M 2-mercaptoethanol. In order to comparatively investigate the mechanisms involved in the myeloma cell survival activity of IL-6, IFN- α or IGF-1, we used serum-free culture conditions to avoid the

HAL author manuscript inserm-00130981, version 1

confounding effects of IGF-1 and IGF binding proteins present in serum. Therefore, the HMCLs were washed twice, cultured for one hour in RPMI1640 supplemented with 1% bovine serum albumin (BSA, Sigma) and washed once again. Cells were then cultured at a concentration of 2×10^5 cells/mL in RPMI1640, 1% BSA with either no cytokine, IL-6 (2 ng/mL), IFN- α (200 U/mL) or IGF-1 (100 ng/mL) for 2, 6 and 24 hours. In other experiments, myeloma cells were starved of IL-6 for 24 hours in presence of FCS (RPMI1640 + 10 % FCS), washed twice and cells were cultured with FCS with or without IL-6 (2 ng/ml) for 24, 48, 72 or 96 hours at a concentration of 2×10^5 cells/ml.

Assays for detection of apoptotic cells.

Apoptotic cells were detected by using fluorescein isothiocyanate-labeled annexin V (FITC-annexin-V, Boehringer Mannheim). Annexin V has a high affinity for phosphatidylserine present on the outer cytoplasmic membrane of apoptotic cells²⁹. Cells were washed, labeled with FITC-annexin-V according to the manufacturer's recommendations and analyzed with a FACScan flow cytometer using Cell Quest software (Becton Dickinson, Mountain View, CA, USA). In one experiment, annexin V⁺ and annexin V⁻ cells were sorted using a Vantage cell sorter (Becton Dickinson) and the cell cycle was analyzed by propidium iodide-DNA staining as previously described⁴¹.

Western Blot Analysis

Cells were lysated in 10 mM Tris-HCl (pH 7.05), 50 mM NaCl, 50mM NaF, 30 mM sodium pyrophosphate (NaPPi), 1% Triton X-100, 5 μ M ZnCl₂, 100 μ M Na₃VO₄, 1mM DTT, 20 mM b-glycerophosphate, 20 mM p-nitrophenolphosphate (PNPP), 2.5 μ g/mL aprotinin, 2.5 μ g/mL leupeptin, 0.5 mM PMSF, 0.5 mM benzamidine, 5 μ g/mL pepstatin and 50 nM okadaik acid. The protein content of each lysate was quantified using the bicinchoninic acid kit (Sigma). Aliquots containing 50 μ g of total protein were resolved in 12% sodium dodecyl sulfate-polyacrylamide by gel electrophoresis (SDS-PAGE) and transferred to a nitrocellulose membrane (Schleicher and Schuell, Dassel, Germany). Membranes were blocked for 2h at room temperature in 140 mM NaCl, 3 mM

KCl, 25 mM Tris-HCl (pH 7.4), 0.1% tween 20 (TBS-T), 5% non-fat milk), then incubated for 1h at room temperature with primary antibodies. The primary antibodies were visualized with goat anti-rabbit (Sigma) or goat anti-mouse (Biorad SA, Yvry Sur Seine, France) peroxydase-conjugated antibodies (at 1:10 000 dilution in TBS-T) using an enhanced chemiluminescence detection system. The membranes were stripped twice with 100 mM Glycin, pH 2.2 , 0.1% NP 40 and 1% SDS for 30 minutes at room temperature. Blots were quantified by densitometry using acquisition into Adobe Photo Shop (Apple, Cupertino, CA, U.S.A) and analysing by the NIH Image softwares (National Institutes of Health, Bethesda, MD, U.S.A.). In the case of the experiments performed with exponentially growing cell lines, the level of protein expression was normalized according to β actin expression.

Statistical analysis

The mean percentages of apoptotic cells in the culture groups with cytokines were compared to corresponding control groups and the statistical significance was evaluated by using the Student *t*-test for pairs. To look for a correlation of Bcl-2-family protein expression among the various myeloma cell lines, the non-parametric Spearman correlation test was used.

Acknowledgments

This work was supported by a grant from the Ligue Contre le Cancer (Paris, France).

References

1. Greipp PR, Katzmann JA, O'Fallon WM and Kyle RA (1988) Value of beta 2-microglobulin level and plasma cell labeling indices as prognostic factors in patients with newly diagnosed myeloma. *Blood*. 72: 219-223.
2. Drewinko B and Alexanian R (1977) Growth kinetics of plasma cell myeloma. *J Nat Cancer Inst*. 58: 1247.
3. Slifka MK, Antia R, Whitmire JK and Ahmed R (1998) Humoral immunity due to long-lived plasma cells. *Immunity*. 8: 363-372.
4. Merville P, Dechanet J, Desmouliere A, Durand I, Debouteiller O, Garrone P, Banchereau J and Liu YJ (1996) Bcl-2(+) tonsillar plasma cells are rescued from apoptosis by bone marrow fibroblasts. *J Exp Med*. 183: 227-236.
5. Gu ZJ, Vos JD, Rebouissou C, Jourdan M, Zhang XG, Rossi JF, Wijdenes J and Klein B (2000) Agonist anti-gp130 transducer monoclonal antibodies are human myeloma cell survival and growth factors. *Leukemia*. 14: 188-197.
6. Zhang XG, Gu ZJ, Lu ZY, Yasukawa K, Yancopoulos GD, Turner K, Shoyab M, Taga T, Kishimoto T, Bataille R and Klein B (1994) Ciliary neurotropic factor, interleukin 11, leukemia inhibitory factor, and oncostatin M are growth factors for human myeloma cell lines using the interleukin 6 signal transducer gp130. *J Exp Med*. 179: 1337-1342.
7. Nishimoto N, Ogata A, Shima Y, Tani Y, Ogawa H, Nakagawa M, Sugiyama H, Yoshizaki K and Kishimoto T (1994) Oncostatin M, leukemia inhibitory factor, and interleukin-6 induce the proliferation of human plasmacytoma cells via the common signal transducer, gp130. *J Exp Med*. 179: 1343-1347.
8. Westendorf JJ and Jelinek DF (1996) Growth regulatory pathways in myeloma. Evidence of autocrine oncostatin M expression. *J Immunol*. 157: 3081-3088.

9. Ferlin-Bezombes M, Jourdan M, Liautard J, Brochier J, Rossi JF and Klein B (1998) IFN-alpha is a survival factor for human myeloma cells and reduces dexamethasone-induced apoptosis. *J Immunol.* 161: 2692-2699.
10. Liu P, Oken M and Van Ness B (1999) Interferon-alpha protects myeloma cell lines from dexamethasone-induced apoptosis. *Leukemia.* 13: 473-480.
11. Xu F, Gardner A, Tu Y, Michl P, Prager D and Lichtenstein A (1997) Multiple myeloma cells are protected against dexamethasone-induced apoptosis by insulin-like growth factors. *Br J Haematol.* 97: 429-440.
12. Georgii-Hemming P, Wiklund HJ, Ljunggren O and Nilsson K (1996) Insulin-like growth factor I is a growth and survival factor in human multiple myeloma cell lines. *Blood.* 88: 2250-2258.
13. Ferlin M, Noraz N, Hertogh C, Brochier J, Taylor N and Klein B (2000) Insulin-like growth factor induces the survival and proliferation of myeloma cells through an IL-6-independent transduction pathway. *Br J Haematol.* In press.
14. Gross A, McDonnell JM and Korsmeyer SJ (1999) BCL-2 family members and the mitochondria in apoptosis. *Genes Dev.* 13: 1899-1911.
15. Adams JM and Cory S (1998) The Bcl-2 protein family: arbiters of cell survival. *Science.* 281: 1322-1326.
16. Takayama S, Sato T, Krajewski S, Kochel K, Irie S, Millan JA and Reed JC (1995) Cloning and functional analysis of BAG-1: a novel Bcl-2-binding protein with anti-cell death activity. *Cell.* 80: 279-284.
17. Hamilton MS, Barker HF, Ball J, Drew M, Abbot SD and Franklin IM (1991) Normal and neoplastic human plasma cells express bcl-2 antigen. *Leukemia.* 5: 768-771.
18. Pettersson M, Jernberg-Wiklund H, Larsson LG, Sundstrom C, Givol I, Tsujimoto Y and Nilsson K (1992) Expression of the bcl-2 gene in human multiple myeloma cell lines and normal plasma cells. *Blood.* 79: 495-502.

19. Ong F, Nieuwkoop JAV, Degrootswings GMJS, Hermans J, Harvey MS, Kluin PM and Kluinnelemans JC (1995) Bcl-2 protein expression is not related to short survival in multiple myeloma. *Leukemia*. 9: 1282-1284.
20. Harada N, Hata H, Yoshida M, Soniki T, Nagasaki A, Kuribayashi N, Kimura T, Matsuzaki H and Mitsuya H (1998) Expression of Bcl-2 family of proteins in fresh myeloma cells. *Leukemia*. 12: 1817-1820.
21. Puthier D, Pellat-Deceunynck C, Barille S, Robillard N, Rapp MJ, Juge-Morineau N, Harousseau JL, Bataille R and Amiot M (1999) Differential expression of Bcl-2 in human plasma cell disorders according to proliferation status and malignancy. *Leukemia*. 13: 289-294.
22. Lichtenstein A, Tu YP, Fady C, Vescio R and Berenson J (1995) Interleukin-6 inhibits apoptosis of malignant plasma cells. *Cell Immunol*. 162: 248-255.
23. Schwarze MMK and Hawley RG (1995) Prevention of myeloma cell apoptosis by ectopic bcl-2 expression or interleukin 6-mediated up-regulation of bcl- x(L). *Cancer Res*. 55: 2262-2265.
24. Puthier D, Derenne S, Barille S, Moreau P, Harousseau JL, Bataille R and Amiot M (1999) Mcl-1 and Bcl-xL are co-regulated by IL-6 in human myeloma cells. *Br J Haematol*. 107: 392-395.
25. Catlett-Falcone R, Landowski TH, Oshiro MM, Turkson J, Levitzki A, Savino R, Ciliberto G, Moscinski L, Fernandez-Luna JL, Nunez G, Dalton WS and Jove R (1999) Constitutive activation of Stat3 signaling confers resistance to apoptosis in human U266 myeloma cells. *Immunity*. 10: 105-115.
26. Tu Y, Renner S, Xu F, Fleishman A, Taylor J, Weisz J, Vescio R, Rettig M, Berenson J, Krajewski S, Reed JC and Lichtenstein A (1998) BCL-X expression in multiple myeloma: possible indicator of chemoresistance. *Cancer Res*. 58: 256-262.
27. Xu FH, Sharma S, Gardner A, Tu Y, Raitano A, Sawyers C and Lichtenstein A (1998) Interleukin-6-induced inhibition of multiple myeloma cell apoptosis: support

for the hypothesis that protection is mediated via inhibition of the JNK/SAPK pathway. *Blood*. 92: 241-251.

28. Zhang XG, Gaillard JP, Robillard N, Lu ZY, Gu ZJ, Jourdan M, Boiron JM, Bataille R and Klein B (1994) Reproducible obtaining of human myeloma cell lines as a model for tumor stem cell study in human multiple myeloma. *Blood*. 83: 3654-3663.
29. Vermes I, Haanen C, Steffens-Nakken H and Reutelingsperger C (1995) A novel assay for apoptosis. Flow cytometric detection of phosphatidylserine expression on early apoptotic cells using fluorescein labelled Annexin V. *J Immunol.Methods*. 184: 39-51.
30. Kayalar C, Ord T, Testa MP, Zhong LT and Bredesen DE (1996) Cleavage of actin by interleukin 1 beta-converting enzyme to reverse DNase I inhibition. *Proc Natl Acad Sci U S A*. 93: 2234-2238.
31. Knodel M, Kuss AW, Lindemann D, Berberich I and Schimpl A (1999) Reversal of Blimp-1-mediated apoptosis by A1, a member of the Bcl-2 family. *Eur J Immunol*. 29: 2988-2998.
32. Lomo J, Smeland EB, Krajewski S, Reed JC and Blomhoff HK (1996) Expression of the bcl-2 homologue mcl-1 correlates with survival of peripheral blood b lymphocytes. *Cancer Res*. 56: 40-43.
33. Moulding DA, Quayle JA, Hart CA and Edwards SW (1998) Mcl-1 expression in human neutrophils: regulation by cytokines and correlation with cell survival. *Blood*. 92: 2495-2502.
34. Zhou P, Qian L, Kozopas KM and Craig RW (1997) Mcl-1, a Bcl-2 family member, delays the death of hematopoietic cells under a variety of apoptosis-inducing conditions. *Blood*. 89: 630-643.
35. Zhou P, Qian L, Bieszczad CK, Noelle R, Binder M, Levy NB and Craig RW (1998) Mcl-1 in transgenic mice promotes survival in a spectrum of hematopoietic cell types and immortalization in the myeloid lineage. *Blood*. 92: 3226-3239.

36. Ogata A, Chauhan D, Teoh G, Treon SP, Urashima M, Schlossman RL and Anderson KC (1997) IL-6 triggers cell growth via the Ras-dependent mitogen-activated protein kinase cascade. *J Immunol.* 159: 2212-2221.
37. Jelinek DF, Aagaard-Tillery KM, Arendt BK, Arora T, Tschumper RC and Westendorf JJ (1997) Differential human multiple myeloma cell line responsiveness to interferon-alpha. Analysis of transcription factor activation and interleukin 6 receptor expression. *J Clin Invest.* 99: 447-456.
38. Berger LC, Hawley TS, Lust JA, Goldman SJ and Hawley RG (1994) Tyrosine phosphorylation of JAK-TYK kinases in malignant plasma cell lines growth-stimulated by interleukins 6 and 11. *Biochem Biophys Res Commun.* 202: 596-605.
39. Puthier D, Bataille R and Amiot M (1999) IL-6 up-regulates mcl-1 in human myeloma cells through JAK / STAT rather than ras / MAP kinase pathway. *Eur J Immunol.* 29: 3945-3950.
40. De Vos J, Jourdan M, Tarte K, Jasmin C and Klein B (2000) JAK2 tyrosine kinase inhibitor tyrphostin AG490 downregulates the mitogen-activated protein kinase (MAPK) and signal transducer and activator of transcription (STAT) pathways and induces apoptosis in myeloma cells. *Br J Haematol.* In press.
41. Jourdan M, Ferlin M, Legouffe E, Horvathova M, Liautard J, Rossi JF, Wijdenes J, Brochier J and Klein B (1998) The myeloma cell antigen syndecan-1 is lost by apoptotic myeloma cells. *Br J Haematol.* 100: 637-646.

ANNEXIN V ⁺ (%)

		Hours of culture			
		0	2	6	24
XG-1	Co	34 ±10	69 ±11	74 ±8	81 ±10
	IL-6		59 ±14*	61 ±10	61 ±14
	IFN- α		59 ±14	59 ±11	60 ±13
	IGF-1		58 ±12	60 ±11	64 ±13
		0	2	6	24
XG-2	Co	25 ±6	49 ±12	67 ±17	82 ±15
	IL-6		43 ±15*	55 ±18	58 ±18
	IFN- α		42 ±14*	49 ±18	48 ±14
	IGF-1		44 ±17*	59 ±16	68 ±19
		0	6	24	48
XG-6	Co	14 ±1	28 ±6	52 ±27	58 ±24
	IL-6		21 ±7	36 ±25	36 ±23
	IFN- α		19 ±7	36 ±25	41 ±24
	IGF-1		24 ±6	46 ±28	48 ±24
		0	2	6	24
XG-13	Co	28 ±5	39 ±7	51 ±3	62 ±14
	IL-6		36 ±5*	39 ±7	45 ±17
	IFN- α		34 ±5*	35 ±6	44 ±14
	IGF-1		32 ±5	35 ±6	42 ±14

Table 1: IL-6, IFN-α and IGF-1 are myeloma cell survival factors

Myeloma cells were harvested during the exponential growth phase with IL-6 and FCS, IL-6 and FCS starved for 1 hour and then cultured for 2, 6, 24 or 48 hours with no cytokine, 2 ng/mL of IL-6, 200 U/mL of IFN-α or 100 ng/mL of IGF-1 in RPMI1640 culture medium and 1% BSA. At the end of the culture, cells were stained with FITC-annexin V to determine the percentage of apoptotic cells and analyzed with a FACScan. Results are mean values ± SD of 3–6 different experiments. In all these experiments, the percentage of viable cells assayed by trypan blue exclusion was not significantly affected during the first 24 hours of culture.

Except when indicated (*), the mean percentage of apoptotic cells in the culture group with cytokines is significantly different ($P < .05$) from corresponding control groups (Control at 2 h, 6 h, 24 h or 48 h), as determined by a Student *t*-test for pairs.

Legends of Figures

Figure 1: IL-6, IFN- α and IGF-1 are myeloma cell survival factors

XG-13 cells were harvested during the exponential growth phase with IL-6 and FCS, IL-6 and FCS starved for 1 hour and then cultured for 2, 6 or 24 hours with no cytokine, 2 ng/mL of IL-6, 200 U/mL of IFN- α or 100 ng/mL of IGF-1 in RPMI1640 culture medium and 1% BSA. At the end of the culture, cells were stained with FITC-annexin V to determine the percentage of apoptotic cells with a FACScan.

Figure 2: Sorting and propidium iodide staining of annexin V- and annexin V+ myeloma cell lines

XG-6 cells were IL-6 starved for 24 hours and stained with FITC-annexin V. In the experiment outlined in the Figure, 40% of XG-6 cells were annexin V⁺. Annexin V⁺ and annexin V⁻ cells were sorted using a FACS-Vantage cell sorter and DNA was labeled with PI. Results are the FACS profiles of annexin V staining of the unsorted and sorted populations as well as the profiles of DNA staining by PI. The percentages of annexin V⁺ cells or of cells with DNA content less than G1 are indicated in the panels.

Figure 3: Immunoblot analysis of Bcl-2-family protein expression on myeloma cells

Cell lysates were obtained from myeloma cells harvested during the exponential growth phase with IL-6 and FCS. An equal amount of protein (25 μ g per lane) was loaded for each HMCL and β actin expression is shown as a control. Expression of anti-apoptotic proteins (panel A) and of pro-apoptotic proteins (panel B).

Figure 4: Regulation of Bcl-2-family protein expression by IL-6, IFN- α or IGF-1 on XG-6 myeloma cells

XG-6 myeloma cells were IL-6 and FCS starved for 1 hour and cultured for 2, 6 and 24 hours with no cytokine (Co), 2 ng/mL IL-6 (IL-6), 200 U/mL IFN- α (IFN- α) or 100 ng/mL IGF-1 (IGF) in RPMI1640 and 1% BSA. At the end of the culture, the cell count and the percentage of viable cells were assayed by trypan blue exclusion. The percentages of apoptotic cells were determined by staining with FITC-annexin V. Cells were immediately lysated and assayed for Bcl-2-family protein expression using Western blot analysis. The amount of proteins was carefully determined and 25 μ g of total proteins from each culture group were loaded on the gel. In the experiments, MAPK expression was used as a loading protein control because it was not affected by the level of apoptosis. Western blots are for one representative experiment out of 3.

Figure 5: Regulation of Bcl-2-family protein expression by cytokines or growth factors in myeloma cell lines

XG-1, XG-2 and XG-13 myeloma cells were processed as indicated in the legend of Figure 4. Results are expression of Bcl-2-family proteins using Western blot analysis after 6 hours of culture with no cytokine (Co), 2 ng/mL IL-6 (IL-6), 200 U/mL IFN- α (IFN- α) or 100 ng/mL IGF-1 (IGF) in RPMI1640 and 1% BSA. Western blots are for one representative experiment out of 2 to 4, depending on the cell line.

Figure 6: Mcl-1 and not Bcl-xL regulation by IL-6 in FCS culture conditions

XG-1 myeloma cells were processed as indicated in the legend of Figure 4. Results are Mcl-1 and Bcl-xL protein expression using Western blot analysis after 2 or 6 hours of culture with no cytokine (Co), 2 ng/mL IL-6 (IL-6), 10% FCS or 2 ng/mL IL-6 and 10% FCS (IL-6 + FCS). Western blots are for one representative experiment out of 3.

Figure 7: Lack of Bcl-xL regulation by IL-6 in long-term culture conditions

XG-6 cells were IL-6 starved for 24 hours in RPMI1640 with 10 % FCS and cultured with or without IL-6 in presence of FCS for 24, 48, 72 or 96 hours. At the end of the culture the cells were processed as indicated in the legend of Figure 4.

Figure 8: Stat3, ERK1 and ERK2 phosphorylation in myeloma cells stimulated with IL-6, IFN- α or IGF-1

Myeloma cells were IL-6 and FCS starved for one hour and cultured with no cytokine (Co), 2 ng/mL IL-6 (IL-6), 200 U/mL IFN- α (IFN- α) or 100 ng/mL IGF-1 (IGF) in RPMI1640 and 1% BSA. Stat3 phosphorylation was analyzed by Western blot after 6 hours of culture using phospho-specific Stat3 antibody (P-Stat3), ERK1 and ERK2 phosphorylation after 15 minutes of culture using phospho-specific MAPK (P-MAPK). Blots were then stripped and reprobed for Stat3 or total MAPK. Western blots are for one representative experiment out of 3.

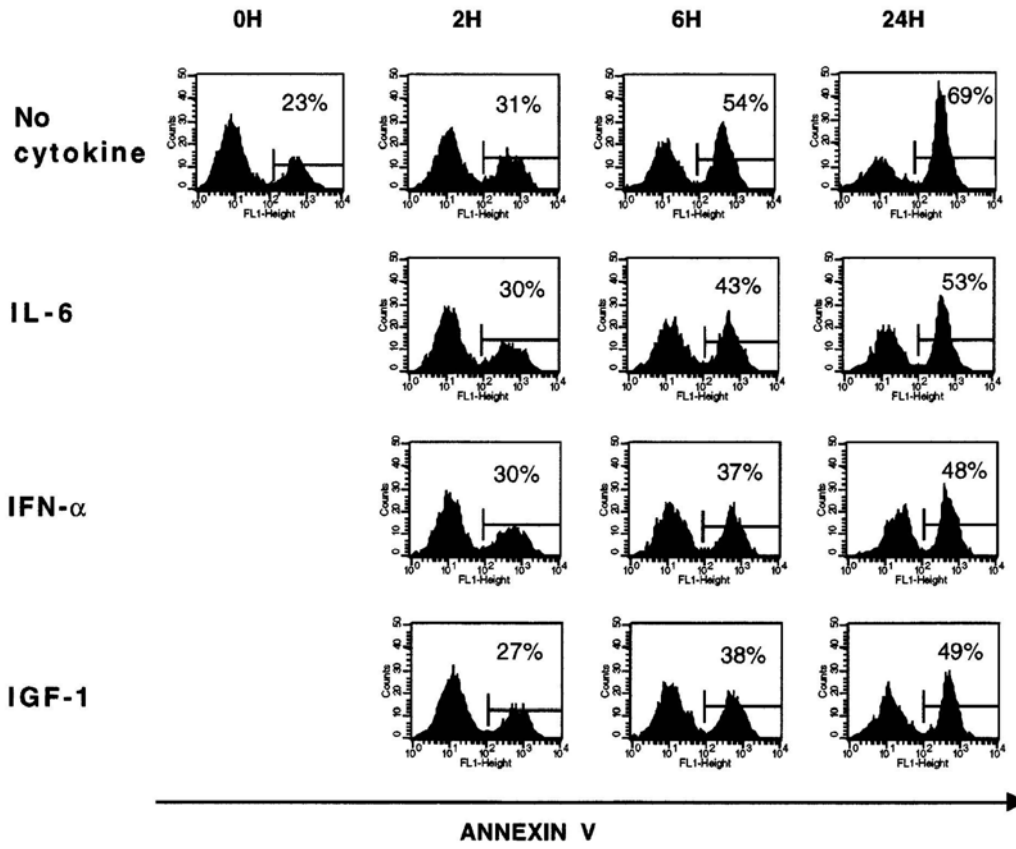


Figure 1

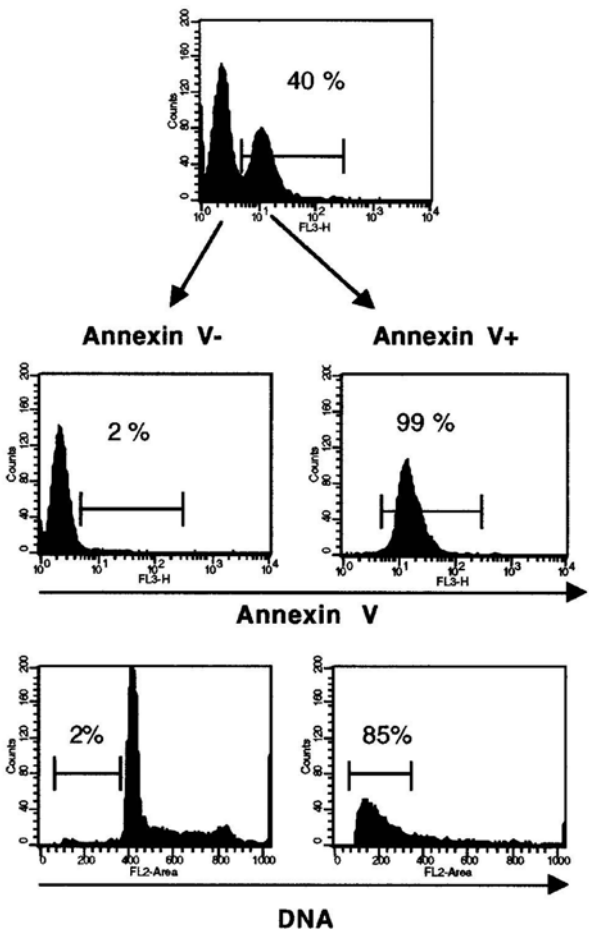


Figure 2

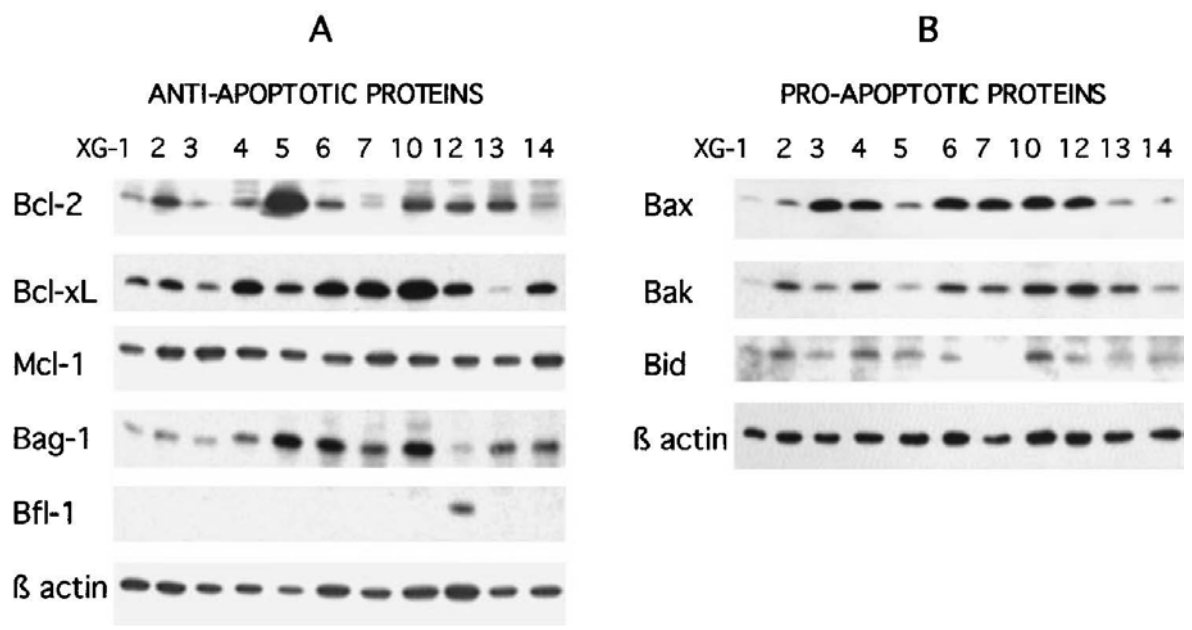


Figure 3

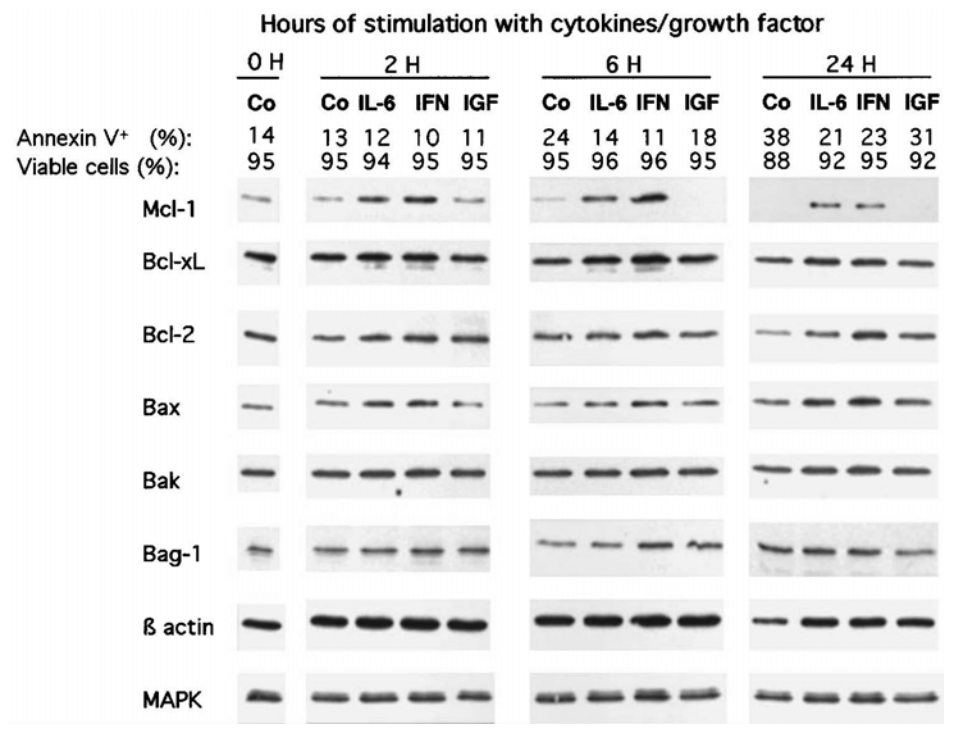


Figure 4

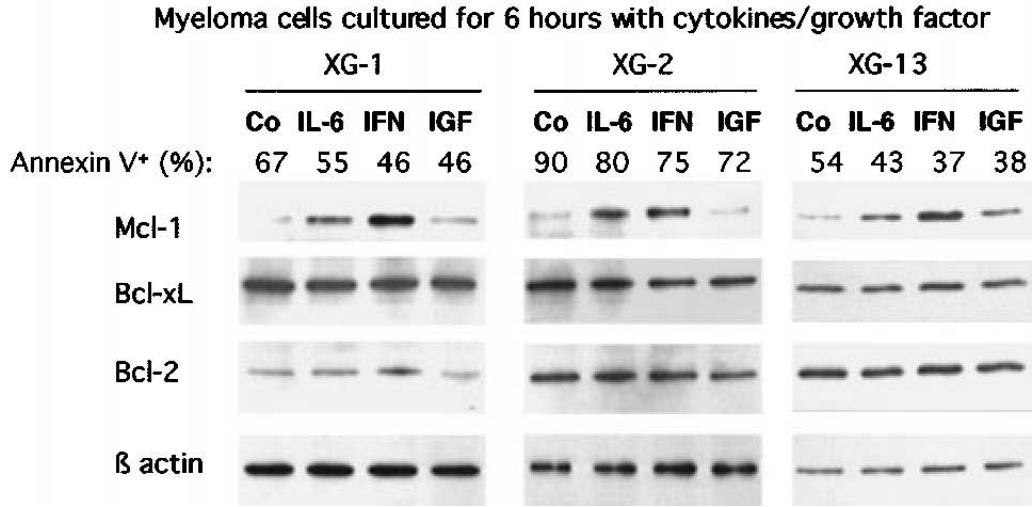


Figure 5

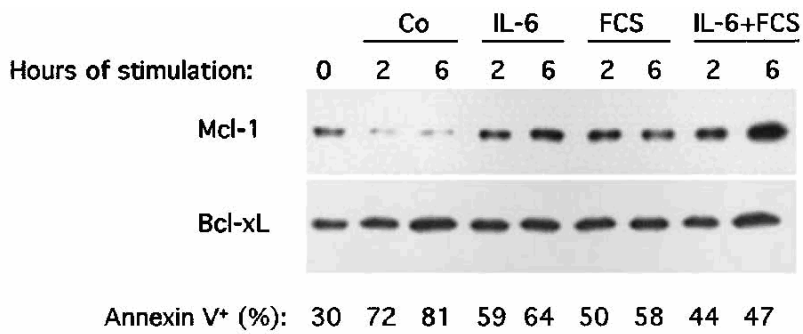


Figure 6

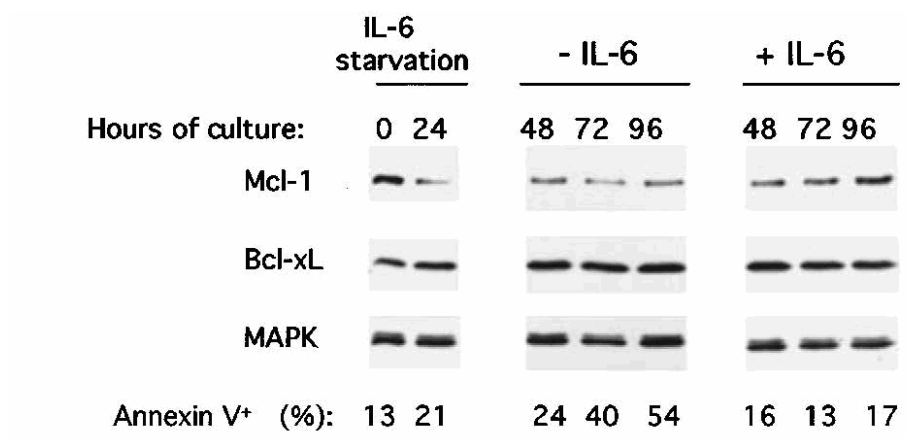


Figure 7

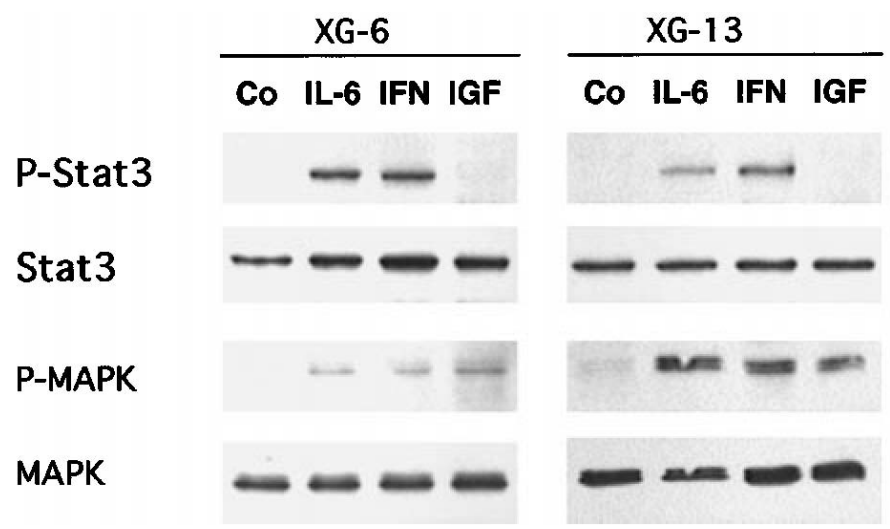


Figure 8