Mutations in the gene encoding the synaptic scaffolding protein SHANK3 are associated with autism spectrum disorders

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SHANK3 (also known as ProSAP2) regulates the structural organization of dendritic spines and is a binding partner of neuroligins; genes encoding neuroligins are mutated in autism and Asperger syndrome. Here, we report that a mutation of a single copy of *SHANK3* on chromosome 22q13 can result in language and/or social communication disorders. These mutations concern only a small number of individuals, but they shed light on one gene dosage-sensitive synaptic pathway that is involved in autism spectrum disorders.

Autism spectrum disorders (ASD) affect about 6 of every 1000 children and are characterized by impairments in reciprocal social interaction and communication as well as restricted and stereotyped patterns of interests and activities¹. ASD ranges from severe (in the case of autistic disorder with moderate or severe cognitive impairment) to a milder variant (Asperger syndrome with higher cognitive ability). Although the causative genes remain largely unknown², familial and twin studies indicate that ASD is one of the most genetic neuropsychiatric disorders. Standard karyotype analyses show chromosomal rearrangements in 3%-6% of cases, the most common being deletions and duplications on chromosomes 15q, 22q and 7q³. One of the most frequent rearrangements associated with cognitive deficits, the 22q13.3 microdeletion syndrome is characterized by neonatal hypotonia, global developmental delay, normal to accelerated growth, absent to severely delayed speech, autistic behavior, and minor dysmorphic features⁴. The loss of terminal 22q13.3 can be subtle and can go undetected by routine chromosome analysis; FISH is often required to confirm the presence of this deletion.

Among the three genes (*ACR*, *RABL2B*, *SHANK3*) located in the minimal telomeric region⁵, *SHANK3* (also known as ProSAP2) is the strongest candidate for the neurobehavioral symptoms observed in patients with 22q13 deletions. SHANK3 is a scaffolding protein found in excitatory synapses directly opposite to the presynaptic active zone. Shank proteins are believed to function as master organizers of the postsynaptic density (PSD), owing to their ability to form multimeric complexes with postsynaptic receptors, signaling molecules and cytoskeletal proteins present in dendritic spines and PSDs^{6,7}. SHANK3 can bind to the cell adhesion proteins neuroligins⁸; we have previously found genes encoding neuroligins (*NLGN3* and *NLGN4*) to be mutated in individuals with autism and Asperger syndrome⁹. *SHANK3* was disrupted by a *de novo* balanced translocation in a child with all the features of the 22q13.3 deletion syndrome¹⁰. In this paper, we report evidence showing that abnormal gene dosage of *SHANK3* is associated with severe cognitive deficits, including language and speech disorder and ASD.

We used FISH analysis (n=97) and/or direct sequencing (n=227) to investigate chromosome 22q13 and *SHANK3* in patients with ASD (**Supplementary Methods**). We also sequenced all *SHANK3* exons in a minimum of 190 controls to ascertain the diversity of *SHANK3* nonsynonymous variations in the general population. *SHANK3* spans 57 kb and contains 24 exons. Seven exons are alternatively spliced, including exon 18, which is detected mostly in the brain (**Supplementary Fig.** 1). During our screening, three families with ASD showed unambiguous alteration of 22q13 or

SHANK3. In family ASD 1, the proband with autism, absent language and moderate mental retardation carried a *de novo* deletion of 22q13 (the clinical description of all patients is provided in the **Supplementary Note**). The deletion breakpoint was located in intron 8 of *SHANK3* and removed 142 kb of the terminal 22q13 (**Fig. 1a**). This deletion had been "repaired" by addition of telomeric repeats and was similar to the minimum deleted region described previously⁵. The recurrent deletions in this region may be due to the quadruplex-forming G-rich sequence (QGRS) surrounding the breakpoint (**Supplementary Fig. 2**), which provides a structural substrate for inappropriate telomere formation.

In family ASD 2, two brothers with autism were heterozygous for an insertion of a guanine nucleotide in exon 21 (**Fig. 1b**). Both brothers had severely impaired speech and severe mental retardation. The mutation was absent in an unaffected brother and the unaffected parents. Using 14 informative SNPs, we found that the mutation was located on the same maternal haplotype in the two affected brothers and that the unaffected brother did not have this haplotype (**Supplementary Fig. 3**). The mutation was absent in the DNA isolated from blood leukocytes and mouth cells of the mother. These results strongly suggest a germinal mosaicism in the mother. The guanine insertion creates a frameshift at nucleotide 3680, modifying the C-terminal sequence of the protein (**Fig. 1b**). This putative truncated protein lacks several crucial domains involved in mGluR and actin binding (Homer, AbP1, cortactin) and in the synaptic targeting and postsynaptic assembly of SHANK3 multimers^{11,12}. Consistent with the loss of these domains, when we over-expressed the truncated protein in rat hippocampal neuronal cells, we did not observe any synaptic localization compared with the wild-type sequence (**Supplementary Fig. 4**).

In family ASD 3, we identified a terminal 22q deletion in a girl with autism and severe language delay and a 22qter partial trisomy in her brother with Asperger syndrome, who demonstrated precocious language development and fluent speech (**Fig. 1c**). We found that these unbalanced cytogenetic abnormalities were inherited from a paternal translocation, t(14;22)(p11.2;q13.33). The chromosome 14p11.2 breakpoint fell within the heterochromatic DNA sequence characteristic of acrocentric chromosomes and did not contain any putative transcripts or genes. On chromosome 22q13.33, using informative SNPs and quantitative PCR, we mapped the breakpoint between *ALG12* and *MLC1* (**Fig. 1d**). The deletion and duplication rearrangement observed in both siblings involved 25 genes, including *SHANK3*, located in the 800-kb terminal sequence of 22q13. No other *SHANK3* deletions or duplications were observed after screening 155 individuals by quantitative PCR (58 with autism, 38 with Asperger syndrome and 59 controls).

In the remaining individuals with ASD, we identified seven who had rare nonsynonymous variations, which were not observed in controls (n=270-333; Fig. 2 and Supplementary Table 1). However, all these variations were inherited from healthy parents, ruling out their direct involvement as dominant mutations in the disorder. Notably, for two substitutions modifying highly conserved amino acids (R12C and R300C; Supplementary Fig. 3), we observed that the overexpressed mutated GFP Shank3 fusion proteins clustered but showed significantly less colocalization with the presynaptic marker protein Bassoon, suggesting nonsynaptic clustering (Supplementary Fig. 4). These

observations might reflect posttranslational modifications or abnormal folding of the protein. Thus, although these genetic variations cannot be considered as causal mutations, they might nevertheless modify the synaptic scaffolding and represent risk factors for ASD in interaction with other susceptibility genes.

In this study, we show that a *SHANK3* heterozygous mutation can cause ASD. Notably, in the boy with Asperger syndrome in family ASD 3, the presence of an additional copy of 22q13/*SHANK3* did not impair his language ability but seems to have led to a severe impairment in social communication. These results, together with previous reports^{13,14}, highlight the importance of a fine gene dosage for the development of speech/language and/or social communication in humans.

The mutations identified in these patients are thought to affect the function and localization of SHANK3 at PSD and dendritic spines. These results are consistent with the alterations of dendritic spines in individuals with learning disabilities¹⁵. In mice, Shank-3 promotes the maturation and the enlargement of dendritic spine heads and is even able to induce spine formation in aspiny neurons¹¹. In ASD, an abnormality of synapse formation and maintenance was first suggested by the identification of mutations in X-linked *NLGN3* and *NLGN4*⁹, and next confirmed by functional studies of the causative mutations. Therefore, we hypothesize that the protein complex including neuroligins and SHANK participates in the assembly of specialized postsynaptic structures required for the development of language and social communication.

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Figure 1. Genetic analyses of three families with ASD and *SHANK3* mutations. (a) In family ASD 1, the proband carries a *de novo* terminal deletion of the paternal chromosome 22q13. The deletion breakpoint is located in intron 8 of *SHANK3*. The breakpoint was sequenced after amplification of the proband DNA using primer 1 in *SHANK3* and primer 2 in the telomeric repeats. The heterogenous smear in the proband is likely due to the difference in telomere length from chromosome to chromosome and/or priming at different locations by the telomeric primer. (b) In family ASD 2, the two probands carry the same *de novo SHANK3* frame-shift mutation on the maternal chromosome 22q13. The mutation is absent from the mother blood and buccal cells, suggesting a germinal mosaicism. The guanine insertion is located in exon 21 of *SHANK3*, leading to a premature truncated protein. (c) In family ASD 3, the father carries a balanced translocation t(14,22)(p11.2;q13.33), proband A (Asperger syndrome) presents a partial 22qter trisomy and proband B (autism) has a 22qter deletion. (d) Using quantitative fluorescent PCR, we mapped the breakpoint between the genes *ALG12* and *MLC1*. The dosage quotient has a theoretical value of 0.5 for a deletion and 1.5 for a duplication.



Figure 2. Localization of rare nonsynonymous variations or truncating *SHANK3* mutations identified in families with ASD. ANK: ankyrin repeats; SH3: Src homology 3 domain; PDZ: postsynaptic density 95/Discs large/zona occludens-1 homology domain; SAM: sterile alpha motif domain.