



HAL
open science

Characterization of two candidate genes, NCoA3 and IRF8, potentially involved in the control of HIV-1 latency.

Sandie Munier, Delphine Delcroix-Genête, Laëtitia Carthagéna, Audrey Gumez, Uriel Hazan

► To cite this version:

Sandie Munier, Delphine Delcroix-Genête, Laëtitia Carthagéna, Audrey Gumez, Uriel Hazan. Characterization of two candidate genes, NCoA3 and IRF8, potentially involved in the control of HIV-1 latency.. *Retrovirology*, 2005, 2, pp.73. 10.1186/1742-4690-2-73 . inserm-00089307

HAL Id: inserm-00089307

<https://inserm.hal.science/inserm-00089307>

Submitted on 17 Aug 2006

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Research

Open Access

Characterization of two candidate genes, NCoA3 and IRF8, potentially involved in the control of HIV-1 latency

Sandie Munier¹, Delphine Delcroix-Genête¹, Laëtitia Carthagéna¹, Audrey Gumez¹ and Uriel Hazan*^{1,2}

Address: ¹Département des Maladies Infectieuses, Institut Cochin, INSERM U567/CNRS UMR-S 8104/Université Paris 5-René Descartes, 22 rue Méchain, 75014 Paris, France and ²UFR de Biochimie, Université Paris 7-Denis Diderot, 2 Place Jussieu, 75251 Paris, France

Email: Sandie Munier - munier@cochin.inserm.fr; Delphine Delcroix-Genête - delcroix@cochin.inserm.fr;

Laëtitia Carthagéna - carthagena@cochin.inserm.fr; Audrey Gumez - gumez@cochin.inserm.fr; Uriel Hazan* - hazan@cochin.inserm.fr

* Corresponding author

Published: 23 November 2005

Received: 28 July 2005

Retrovirology 2005, 2:73 doi:10.1186/1742-4690-2-73

Accepted: 23 November 2005

This article is available from: <http://www.retrovirology.com/content/2/1/73>

© 2005 Munier et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: The persistence of latent HIV-1 reservoirs is the principal barrier preventing the eradication of HIV-1 infection in patients by current antiretroviral therapy. It is thus crucial to understand the molecular mechanisms involved in the establishment, maintenance and reactivation of HIV-1 latency. Since chromatin remodeling has been implicated in the transcriptional reactivation of the HIV-1 promoter, we assessed the role of the histone deacetylase inhibitor sodium butyrate (NaB) on two HIV-1 latently infected cell lines (UI and ACH-2) gene expression.

Results: Analysis of microarrays data led us to select two candidate genes: *NCoA3* (Nuclear Receptor Coactivator 3), a nuclear receptor coactivator and *IRF8* (Interferon Regulatory Factor 8), an interferon regulatory factor. *NCoA3* gene expression is upregulated following NaB treatment of latently infected cells whereas *IRF8* gene expression is strongly downregulated in the promonocytic cell line following NaB treatment. Their differential expressions were confirmed at the transcriptional and translational levels. Moreover, *NCoA3* gene expression was also upregulated after treatment of UI and ACH-2 cells with phorbol myristyl acetate (PMA) but not trichostatin A (TSA) and after treatment with NaB of two others HIV-1 latently infected cell lines (OM10.1 and J1.1). *IRF8* gene is only expressed in UI cells and was also downregulated after treatment with PMA or TSA. Functional analyses confirmed that *NCoA3* synergizes with Tat to enhance HIV-1 promoter transcription and that *IRF8* represses the IRF1-mediated activation through the HIV-1 promoter Interferon-stimulated response element (ISRE).

Conclusion: These results led us to postulate that *NCoA3* could be involved in the transcriptional reactivation of the HIV-1 promoter from latency and that *IRF8* may contribute to the maintenance of the latent state in the promonocytic cell line. Implication of these factors in the maintenance or reactivation of the viral latency may provide potential new targets to control HIV-1 replication in latent viral reservoirs.

Background

The use of highly active antiretroviral therapy (HAART) in HIV-1 infected individuals has led to a significant decrease of plasma viremia to undetectable levels and has considerably improved the survival and quality of life of infected individuals (reviewed in [1]). However, the presence of cellular reservoirs that contain latent viruses capable of producing infectious particles after cellular activation lead to a rebound of the viral load after interruption of HAART (reviewed in [2]). The persistence of these latently infected viral reservoirs, despite prolonged HAART treatments, represents a major obstacle to the eradication of HIV-1 in infected patients [3-5]. Therefore, a greater understanding of the molecular mechanisms involved in establishment, maintenance and reactivation of viral latency is essential to expect the reduction of latent HIV-1 reservoirs in infected patients.

Latent HIV-1 infection can exist in many reservoirs, such as macrophages and resting memory CD4⁺ T cells (reviewed in [6]). At the cellular level, two major forms of HIV-1 latency have been described: pre- and post-integration latency [7]. CD4⁺ T cells in the post-integration state of latency represent the most stable reservoir for HIV-1 (half-life of 43 months) [8]. Several mechanisms have been proposed to account for the low level of transcription observed during post-integration latency (reviewed in [9]): the inaccessibility of the integrated provirus to the transcriptional machinery, the absence in resting cells of transcription factors involved in HIV-1 gene expression, the presence of transcriptional repressors, and the premature termination of HIV-1 transcription elongation due to the absence of the viral protein Tat and its associated cofactors. Moreover, the chromatin structure appears to be involved in the regulation of HIV-1 gene expression (reviewed in [10]). Indeed, a repressive nucleosome (nuc-1), located immediately downstream of the HIV-1 transcription start site under latency conditions, is disrupted upon transcriptional activation of the HIV-1 promoter in response to Tat, phorbol esters and histone deacetylase (HDAC) inhibitors [11]. Transcriptional activation of the HIV-1 promoter in response to PMA involves the recruitment of SWI/SNF chromatin remodeling complex [12] and cellular proteins with histone acetyltransferase (HAT) activity [13]. Therefore, chromatin remodeling plays a significant role in the transcriptional reactivation of the HIV-1 promoter from latency. Identification of host transcription factors that may regulate chromatin structure is thus critical to understand the molecular mechanisms involved in HIV-1 reactivation.

Gene expression analysis using high-density microarrays have provided a greater understanding of host-pathogen interactions (reviewed in [14]). Previous microarray studies on HIV-1 have described changes in cellular genes tran-

scription in response to HIV-1 protein expression (Nef [15,16], Tat [17,18], gp120 [19] or Vpr [20]) or following acute infection of cell lines [21-24] or Peripheral Blood Mononuclear Cells (PBMC) [25]. DNA microarrays have also been used to characterize gene expression in latently infected resting CD4⁺ T cells in viremic versus aviremic HIV-1 infected individuals [26]. Recently, global gene expression changes in cell lines latently infected with HIV-1 and induced by PMA for completion of viral replication was described by Krishnan *et al.* [27].

To complete the results obtained by Krishnan *et al.*, we used the same strategy to assess the role of the HDAC inhibitor NaB on HIV-1 latently infected cells gene expression. We performed microarray experiments on two HIV-1 latently infected cell lines (U1 and ACH-2) treated or not with NaB to induce viral reactivation. Analysis of microarrays data led us to select two candidate genes encoding transcription factors: NCoA3 (reviewed in [28]), which expression is upregulated following treatment of latently infected cells with NaB, and IRF8 (reviewed in [29]), which expression is downregulated in treated cells. Differential expression of these genes was confirmed at the transcriptional and translational levels. Moreover, NCoA3 gene expression was also upregulated after treatment of U1 and ACH-2 cells with PMA but not TSA and after treatment with NaB of two others latently infected cell lines (OM10.1 and J1.1). IRF8 gene is only expressed in U1 cells and was also downregulated after treatment with PMA or TSA. Functional analyses confirmed that NCoA3 synergizes with Tat to enhance HIV-1 promoter transcription and that IRF8 represses the IRF1-mediated activation of the HIV-1 ISRE element. Implication of IRF8 in the maintenance and NCoA3 in the reactivation of the viral latency may thus provide new insights into the control of HIV-1 replication in latent viral reservoirs.

Results

Microarray analysis

In order to understand the molecular mechanisms regulating HIV-1 latency, we studied the modifications of cellular transcription using microarrays in the promonocytic U1 and T CD4⁺ lymphocytic ACH-2 chronically HIV-1 infected cell lines after reactivation of latency. The two cell lines were treated with 10 mM of the histone deacetylase inhibitor NaB. Viral reactivation was monitored by coculture with P4 indicating cells (Figure 1A) and measuring gag viral mRNA expression (Figure 1B). Increase in both β -galactosidase activity and gag mRNA expression showed that the viral reactivation after NaB treatment was efficient. Total RNAs were extracted after 24 h and sent to the Affymetrix Microarray Facilities for subsequent hybridization on U-133A microarrays.

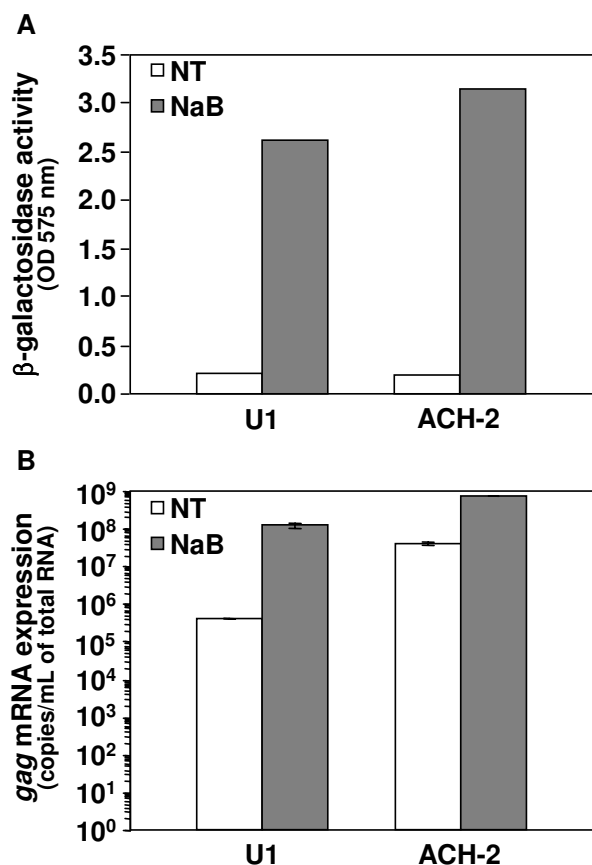


Figure 1
Analysis of viral reactivation after treatment of U1 and ACH-2 cells with NaB. U1 and ACH-2 cells were treated or not (NT) with 10 mM of NaB for 24 h and cocultured with P4 indicating cells. β -galactosidase activity was determined after 24 h coculture (A). RNA from U1 and ACH-2 cells treated or not with NaB were extracted after 24 h and *gag* viral mRNA expression was measured by real-time RT-PCR (B). Results are representative of three independent experiments.

The pattern of cellular mRNA from chronically infected cells treated with NaB was compared to that from non-treated cells. We used as specific criteria a \log_2 ratio change ≥ 1 with a change p-value ≤ 0.0001 for increased genes and a \log_2 ratio change ≤ -1 with a 1-change p-value ≥ 0.9999 for decreased genes. Hybridization experiments were performed once. We identified 740 genes that were upregulated by twofold or higher in NaB treated U1 cells and 896 genes that were downregulated, 482 genes in NaB treated ACH-2 cells that had a level increased greater than twofold and 634 genes that had a level decreased greater than twofold (data not shown). Moreover, 260 genes were commonly increased and 337 genes were decreased in

both U1 and ACH-2 NaB-treated cells (data not shown). Pathways involved in regulation of transcription, signal transduction, immune response, protein transport, metabolism, apoptosis and RNAs modifications showed altered expression following treatment with NaB. Some of the genes involved in these pathways are shown in Additional Files 1, 2, 3, 4, 5 and 6. Our analysis identified genes that have previously been associated with HIV-1 replication or latency, such as CDK9 [16], Jun [16,23], PSMB10 [27], MAPK1 [26] or OAS1 [30]. This supported the accuracy of our approach, even though, as the hybridization experiments had been performed once, the statistical relevance of the results could not be estimated.

Among the differentially expressed genes, we chose to focus on two candidate genes encoding transcription factors: *NCoA3* and *IRF8* (Tables 1 and 2). We selected these two genes based on their biological properties, their described effects on viral replication [31,32] and their differential expression observed by microarray experiments. Indeed, *NCoA3* and *IRF8* gene expression are respectively upregulated and downregulated following treatment with NaB of latently infected cells (Tables 1 and 2). Therefore, *NCoA3* and *IRF8* could be implicated respectively in the reactivation and maintenance of HIV-1 latency.

NCoA3 gene expression is upregulated following treatment with NaB of both U1 and ACH-2 latently infected cells (Tables 1 and 2). *NCoA3* is a nuclear receptor coactivator of the Steroid Receptor Coactivator (SRC) family that interacts with nuclear receptors in a ligand-dependent manner and enhances transcriptional activation *via* histone acetylation and recruitment of general transcription factors and additional cofactors (reviewed in [28]). *NCoA3* (Unigene Hs. 382168) gene expression in U1 cells is significantly upregulated by 4.9 to 22.6 fold (U1NaBvsU1 signal \log_2 ratio ranging from 2.3 to 4.5 with a change p-value < 0.00015) following treatment with NaB (Table 1). Similarly, *NCoA3* gene expression is upregulated in NaB-treated compared to non-treated ACH-2 cells by 2 to 13.9 fold but with a lower significance (ACH2NaBvsACH2 signal \log_2 ratio ranging from 1 to 3.8 with a change p-value < 0.0055) (Table 2).

IRF8 gene expression is downregulated following treatment of U1 cells with NaB (Table 1). *IRF8* is a transcription factor of the Interferon (IFN) Regulatory Factor (IRF) family that binds to IFN-stimulated response element and regulates expression of genes stimulated by IFNs (reviewed in [29]). *IRF8* (Unigene Hs. 137427) is expressed in the promonocytic cell line U1 (detection signal of 707.9 with a p-value of 0.000244) (Table 1) but is not expressed in the T CD4⁺ lymphocytic cell line ACH-2 (data not shown). Following NaB treatment, *IRF8* gene expression in U1 cells is downregulated by 16 fold

Table 1: Differential gene expression obtained for NCoA3 and IRF8 mRNAs in U1 cells treated or not with NaB.

Gene	Probe set Name ^a	U1 Signal ^b	U1 Detection p-value ^c	U1NaB Signal	U1NaB Detection p-value	U1NaBvsU1 Signal log ₂ ratio ^d	U1NaBvsU1 Change p-value ^e
<i>NCoA3</i>	207700_s_at	17.7	0.01416	98.9	0.000244	2.5	0.000035
	209060_x_at	16.9	0.171387	77.2	0.000244	2.3	0.000023
	209061_at	48.4	0.037598	166.4	0.000732	2.3	0.00002
	209062_x_at	6.3	0.72583	91.8	0.010742	4.5	0.000147
	211352_s_at	7.2	0.303711	68.6	0.00293	3.2	0.000101
<i>IRF8</i>	204057_at	707.9	0.000244	47	0.010742	-4	0.99998

^a Affymetrix U133-A reference probe set.

^b Signal intensity of hybridization.

^c Signal detection p-value < 0.048 for specific hybridization.

^d Signal log₂ ratio > 1 for increased genes and < -1 for decreased genes.

^e Change p-value < 0.0001 for significant increased genes and 1-change p-value > 0.9999 for significant decreased genes.

(U1NaBvsU1 signal log₂ ratio of -4 with a 1-change p-value of 0.99998) (Table 1).

Validation of NCoA3 and IRF8 differential transcriptional expression

Real-time RT-PCR quantifications were performed to confirm that NCoA3 and IRF8 genes were differentially expressed in the NaB-treated chronically infected cells compared to the non-treated cells. We performed quantification on RNA samples obtained from five independent NaB treatments of U1 and ACH-2 cells and real-time RT-PCR experiments were run in duplicate. NCoA3 and IRF8 expressions were normalized to the expression of Cyclophilin A. The results show in Figure 2 represent the NCoA3 expression increase fold (Figure 2A) obtained from U1 and ACH-2 cells and the IRF8 expression decrease fold (Figure 2B) obtained from U1 cells treated with NaB for 24 h and 48 h compared to non-treated cells. Concerning NCoA3, real-time RT-PCR showed an upregulation consistent with microarray data in 24 h NaB-treated U1 cells of 8.34 ± 2.42 fold compared to non-treated cells (Figure 2A). NCoA3 gene expression is also increased with a 48 h NaB treatment (upregulation of 8.40 ± 2.33 fold) (Figure 2A). Similarly, an increase of NCoA3 gene expression can be observed on ACH-2 cells following treatment with NaB (upregulation of 4.56 ± 1.28 fold in 24 h and 6.80 ± 2.34 fold in 48 h NaB-treated ACH-2 cells) (Figure 2A). Concerning IRF8, real-time RT-PCR showed a 14.96 ± 4.85 fold decrease in 24 h NaB-treated U1 cells (Figure 2B) in correlation with the microarray ratio previously obtained. Downregulation of IRF8 gene expression is also observed following 48 h NaB-treatment of U1 cells (22.06 ± 11.29 fold decrease) (Figure 2B). Taken together, results from real-time RT-PCR performed on NCoA3 and IRF8 genes corroborate with those obtained using microarray hybridizations.

We next determined whether NCoA3 and IRF8 gene expression were similarly modified in the uninfected parental cell lines. U937 and CEM cells were subjected to identical treatment and RT-PCR quantifications were performed (Figure 3). NCoA3 is upregulated both in U937 and CEM cells following treatment with NaB (upregulation of 7.32 ± 1.74 fold in 24 h and 11.45 ± 2.95 fold in 48 h NaB-treated U937 cells, upregulation of 1.93 ± 1.04 fold in 24 h and 5.59 ± 0.06 fold in 48 h NaB-treated CEM cells) (Figure 3A). IRF8 is only expressed in the promonocytic cell line U937 and, as in U1 cells, its expression was downregulated after NaB treatment (downregulation of 17.95 ± 4.15 fold in 24 h and 22.32 ± 10.82 fold in 48 h NaB-treated U937 cells) (Figure 3B). Thus, NaB treatment modify NCoA3 and IRF8 gene expression in uninfected parental cell lines U937 and CEM at a similar level than in chronically infected cells.

We then performed additional experiments to determine whether the gene expression variations observed could also be mediated by treatments with the phorbol ester PMA and another HDAC inhibitor, TSA. We thus assessed the differential regulation of NCoA3 and IRF8 gene expression in U1 and ACH-2 cells treated with PMA or TSA (Figure 4). Results indicated that NCoA3 expression is upregulated by 24 h and 48 h PMA treatment of U1 and ACH-2 cells (upregulation of 5.70 ± 1.37 fold in 24 h and 9.85 ± 0.90 fold in 48 h PMA-treated U1 cells, upregulation of 3.12 ± 1.05 fold in 24 h and 7.12 ± 1.20 fold in 48 h PMA-treated ACH-2 cells) (Figure 4A). However, TSA treatment had no significant effect on NCoA3 expression in U1 and ACH-2 cells, although TSA increased viral expression (data not shown). Concerning IRF8 expression in U1 cells, PMA and TSA treatments for 24 h induced a decrease of 3.22 ± 0.45 fold and 5.32 ± 1.09 fold, respectively (Figure 4B). These results show that NCoA3 expres-

Table 2: Differential gene expression obtained for NCoA3 mRNA in ACH-2 cells treated or not with NaB.

Gene	Probe set Name ^a	ACH-2 Signal ^b	ACH-2 Detection p-value ^c	ACH2NaB Signal	ACH2NaB Detection p-value	ACH2NaBvsACH 2 Signal log ₂ ratio ^d	ACH2NaBvsACH2 Change p-value ^e
NCoA3	207700_s_at	43.3	0.001953	99.6	0.001221	1.2	0.000241
	209060_x_at	34.5	0.01416	72.9	0.001953	1	0.000273
	209061_at	65.8	0.000732	82.6	0.000732	1.6	0.005409
	209062_x_at	20	0.466064	76.7	0.095215	2	0.000114
	211352_s_at	2.7	0.5	37	0.030273	3.8	0.004481

^a Affymetrix U133-A reference probe set.

^b Signal intensity of hybridization.

^c Signal detection p-value < 0.048 for specific hybridization.

^d Signal log₂ ratio > 1 for increased genes and < -1 for decreased genes.

^e Change p-value < 0.0001 for significant increased genes and 1-change p-value > 0.9999 for significant decreased genes.

sion is upregulated following phorbol ester but not with other HDAC inhibitor treatments in U1 and ACH-2 cells. Moreover, *IRF8* gene expression in U1 cells is downregulated with PMA or TSA treatments but at a lower extent than with NaB.

We also assessed the differential regulation of *NCoA3* and *IRF8* gene expression in others chronically HIV-1 infected cell lines. The chronically infected promonocytic OM10.1 and T CD4⁺ lymphocytic J1.1 cell lines were treated with NaB for 24 h and 48 h and real-time RT-PCR were performed to measure *NCoA3* and *IRF8* gene expression. As shown in Figure 5, *NCoA3* expression is upregulated by 4.94 ± 0.78 fold in OM10.1 and by 2.56 ± 0.64 fold in J1.1 after 24 h NaB treatment. *NCoA3* expression increased with time of NaB treatment in both cell lines (upregulation of 12.89 ± 3.10 fold in OM10.1 and 3.51 ± 0.69 fold in J1.1 cells) (Figure 5). Like ACH-2 and unlike U1 cells, the T CD4⁺ lymphocytic J1.1 and the promonocytic OM10.1 cell lines did not express *IRF8* (data not shown). Thus, the differential regulation of *NCoA3* but not *IRF8* gene expression is similar in two other related latently HIV-1 infected cell line models.

***gag* mRNA activation is correlated with NCoA3 mRNA increase and IRF8 mRNA decrease**

We performed reactivation experiments at different times, sooner than 24 h and until 48 h. Quantitative RT-PCR experiments were carried out on total RNAs. This was done using U1 cells to analyze both *NCoA3* mRNA increase (Figure 6A) and *IRF8* mRNA decrease (Figure 6B) relative to HIV *gag* mRNA along with ACH-2 cells (Figure 6C) to analyze *NCoA3* mRNA increase relative to HIV *gag* mRNA.

As observed on Figure 6C, the obtained results, both on ACH-2 and U1 cells, clearly show that *gag* mRNA activation occurs after *NCoA3* mRNA increase and accumulation. Moreover, in U1 cells, *gag* mRNA activation occurs

after *IRF8* mRNA decrease. Shorter kinetics (0 to 8 h) correlated with these results (data not shown).

Validation of NCoA3 and IRF8 differential translational expression

To confirm that the changes seen at the RNA level correlated with protein levels, we performed Western blot experiments on nuclear extract of U1, ACH-2, OM10.1 and J1.1 cells treated or not with NaB for 24 h (Figure 7). Results indicated that NaB increased the expression level of *NCoA3* protein in U1, ACH-2, OM10.1 and not in J1.1 cells (Figure 7). Moreover, *IRF8* protein expression is strongly downregulated in U1 cells following NaB treatment (Figure 7). These results correlate with the differential expression of *NCoA3* and *IRF8* genes observed with both microarray and real-time RT-PCR experiments.

Transcriptional activation of the HIV-1 promoter by NCoA3

We analyzed the functional role of *NCoA3* on viral transcription by transfection assays. HEK293 cells were cotransfected with pLTRX-luc reporter plasmid containing the luciferase gene under the control of the HIV-1 U3-R promoter region (nt -640 to +78) with or without Tat and/or *NCoA3* expression vectors. As shown in Figure 8, *NCoA3* increased Tat-stimulated HIV-1 LTR activity by 2.8 ± 1.4 fold. The presence of *NCoA3* had synergistic effect on the HIV-1 LTR activity induced by suboptimal expression of Tat. When HEK293 cells were transfected with pLTRΔTAR-luc reporter plasmid lacking the Tat-transactivation response element TAR, Tat was not able to activate the LTR transcription, as expected, and *NCoA3* had no effect on the LTR activity (Figure 8). Thus, functional analyses confirm that *NCoA3* synergizes with Tat to enhance HIV-1 promoter transcription, as expected [31], and that this effect is dependent on the presence of the TAR region.

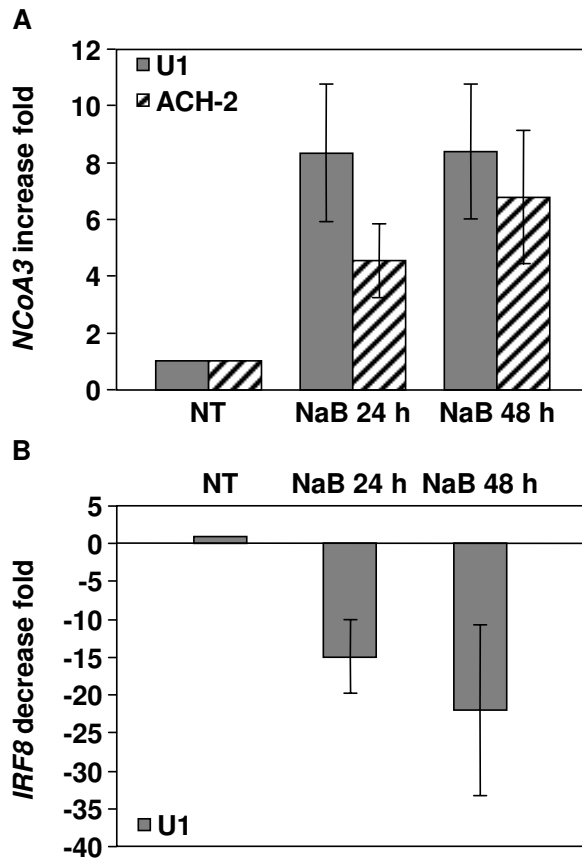


Figure 2
Real-time RT-PCR analysis of NCoA3 and IRF8 mRNAs expression in NaB-treated U1 and ACH-2 cells. Total RNAs were isolated from U1 or ACH-2 cells treated or not with NaB for 24 h and 48 h and real-time PCR were performed on cDNAs using gene specific primers for NCoA3, IRF8 or Cyclophilin A. NCoA3 and IRF8 expressions were normalized to the expression of Cyclophilin A. The NCoA3 increase fold (A) in U1 (solid bars) or ACH-2 (white bars) cells and the IRF8 decrease fold (B) in U1 cells treated with NaB for 24 h and 48 h compared to non-treated (NT) cells were determined. Results represent the means of five independent experiments performed in duplicate.

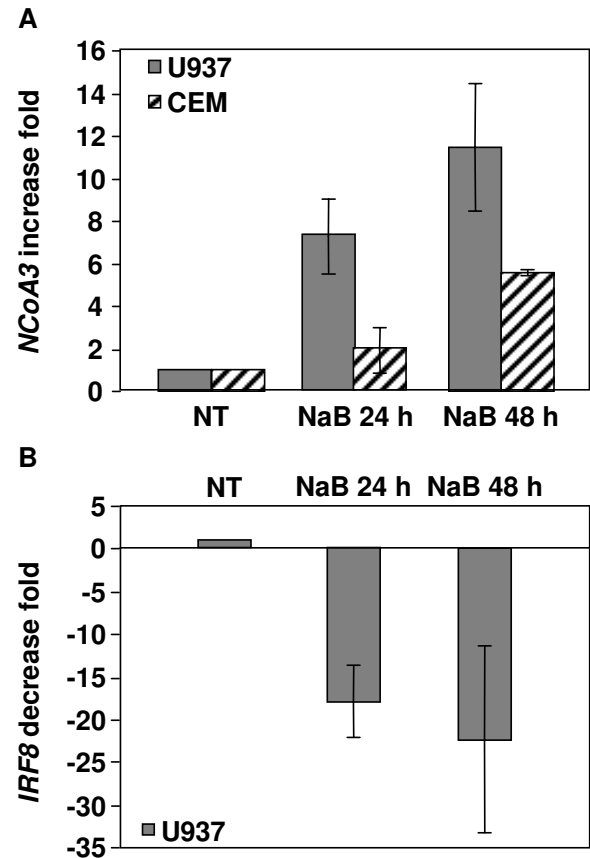


Figure 3
Real-time RT-PCR analysis of NCoA3 and IRF8 mRNAs expression in NaB-treated U937 and CEM cells. Total RNAs were isolated from U937 or CEM cells treated or not with NaB for 24 h and 48 h and real-time PCR were performed on cDNAs using gene specific primers for NCoA3, IRF8 or Cyclophilin A. NCoA3 and IRF8 expressions were normalized to the expression of Cyclophilin A. The NCoA3 increase fold (A) in U937 (solid bars) or CEM (white bars) cells and the IRF8 decrease fold (B) in U937 cells treated with NaB for 24 h and 48 h compared to non-treated (NT) cells were determined. Results represent the means of five independent experiments performed in duplicate.

Transcriptional repression of the HIV-1 ISRE element by IRF8

We analyzed the functional role of IRF8 on viral transcription by transfection assays. HEK293 cells were cotransfected with pISRE-TK-luc reporter plasmid corresponding to the HIV-1 IFN-stimulated response element, located downstream transcription start site (nt +194 to +223) [33], with or without IRF1 and/or IRF8 expression vectors. As shown in Figure 9, the basal activity of the ISRE-TK was increased by 7.4 ± 1.0 fold in the presence of IRF1 as expected [32], whereas a decrease was detected in the pres-

ence of IRF8 (21.9 ± 10.6 to 41.4 ± 9.5 %). The expression of IRF8 inhibited by 43.5 ± 10.6 to 74.7 ± 2.5 % the IRF1-mediated activation of the ISRE-TK in a dose dependent fashion (Figure 9). The expression of the dominant negative IRF8 DNA-binding domain (IRF8-DBD) inhibited by 76.4 ± 6.5 % the IRF1-mediated activation of the ISRE-TK, as expected [34] (Figure 9). The inhibitory effects of IRF8 and IRF8-DBD expression and activation effect of IRF1 expression was abolished when the ISRE sequence was mutated (pISREmut-TK-luc, Figure 9). These results show that IRF8 represses the ISRE-TK promoter transcription

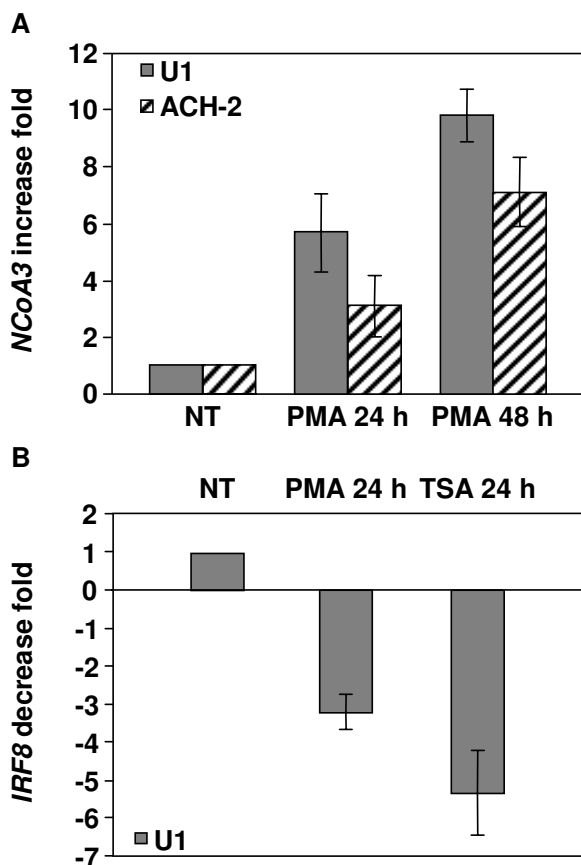


Figure 4
Real-time RT-PCR analysis of NCoA3 and IRF8 mRNAs expression in PMA- or TSA-treated U1 and ACH-2 cells. Total RNAs were isolated from U1 or ACH-2 cells treated or not with PMA for 24 h and 48 h or TSA for 24 h and real-time PCR were performed on cDNAs using gene specific primers for NCoA3, IRF8 or Cyclophilin A. NCoA3 and IRF8 expressions were normalized to the expression of Cyclophilin A. The NCoA3 increase fold (A) in U1 (solid bars) or ACH-2 (white bars) cells treated with PMA for 24 h and 48 h and the IRF8 decrease fold (B) in U1 cells treated with PMA or TSA for 24 h compared to non-treated (NT) cells were determined. Results represent the means of three independent experiments performed in duplicate.

through the ISRE element from the HIV-1 promoter, as expected [32].

Discussion

The existence of long-lasting HIV-1 reservoirs is the principal barrier preventing the eradication of HIV-1 infection in patients by current antiretroviral therapy. It is thus crucial to understand the molecular mechanisms involved in establishment, maintenance and reactivation of HIV-1 latency. In this study, the role of the HDAC inhibitor NaB

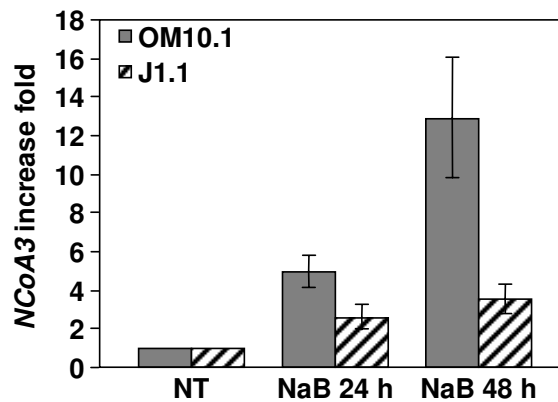


Figure 5
Real-time RT-PCR analysis of NCoA3 mRNAs expression in OM10.1 and J1.1 cells. Total RNAs were isolated from OM10.1 or J1.1 cells treated or not with NaB for 24 h and 48 h and real-time PCR were performed on cDNAs using gene specific primers for NCoA3 or Cyclophilin A. NCoA3 expression was normalized to the expression of Cyclophilin A. The NCoA3 increase fold in OM10.1 (solid bars) or J1.1 cells (white bars) treated with NaB for 24 h and 48 h compared to non-treated (NT) cells were determined. Results represent the means of two independent experiments performed in duplicate.

on HIV-1 latently infected cells gene expression was explored using microarrays. Since chromatin remodeling is involved in the regulation of HIV-1 gene expression (reviewed in [10]), differential expression of cellular genes in latently infected cells following treatment with NaB might be related to the maintenance and reactivation of latency.

Recently, Krishnan *et al.* [27] described the global gene expression changes in HIV-1 latently infected cell lines treated or not with PMA to induce viral reactivation compared to the uninfected parental cell lines treated under the same conditions. Here, we compared gene expression profiles of two HIV-1 latently infected cell lines (U1 and ACH-2) treated with NaB to that of non-treated corresponding cell lines. We thus avoided identification of genes which differential expression could result from the establishment and cloning of the chronically infected cell lines. Based on our specific criteria, we identified few hundreds of genes affected by NaB treatment implicated in biological pathways previously shown to be modulated by HIV-1 replication. For example, reactivation of latency induced an upregulation of CDK9, the catalytic component of transcription elongation factor b (P-TEFb), which acts in concert with Tat to direct the processivity of HIV-1 transcription. It was shown that CDK9 mRNA and protein levels are induced following T cell activation and Nef

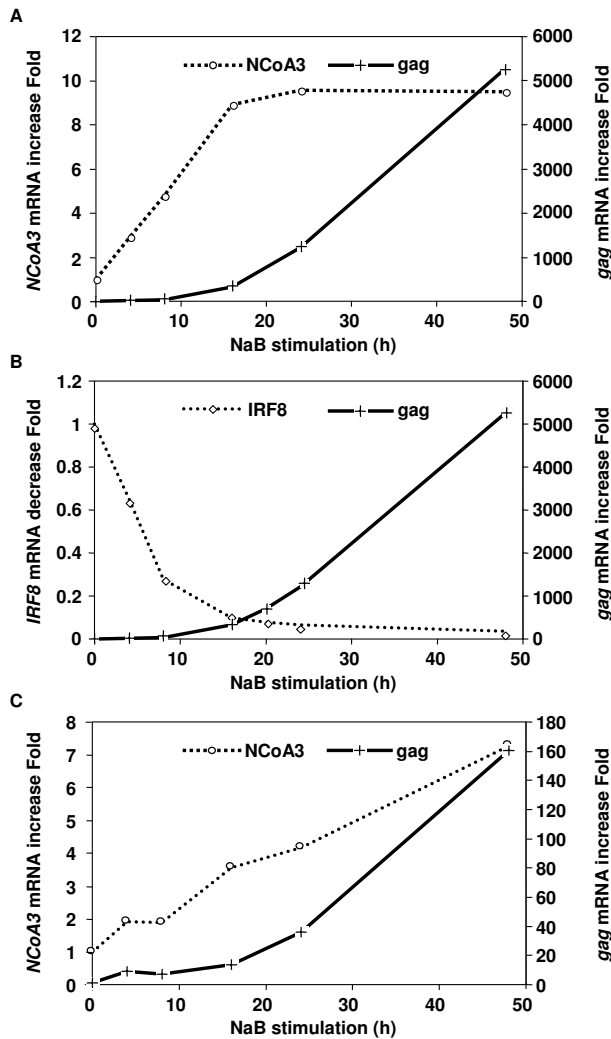


Figure 6
Analysis of HIV *gag*, *NCoA3*, and *IRF8* mRNA expression after NaB stimulation on U1 and ACH-2 cells. U1 (A and B) and ACH-2 (C) cells were stimulated with 10 mM NaB and $5 \cdot 10^6$ cells were taken at $t = 0, 4, 8, 16, 24, 48$ h for RNA extraction to perform qRT-PCR. *NCoA3* (A and C), *IRF8* (B) and *gag* (A, B and C) mRNA contents were measured. *Cylophilin A* was used as internal standard. Results represent a representative experiment performed in duplicate.

expression, and that this correlates with kinase activity, thus enhancing HIV-1 transcription [16,35].

After NaB treatment of latently infected cell lines, we observed an upregulation of genes involved in vesicular transport of protein like syntaxin and nexin. It was found by Chun *et al.* that numerous genes involved in protein/vesicle transport are upregulated in resting T CD4⁺ cells of viremic patients, strongly suggesting that enhanced activities in secretory pathways may help in the assembly and

release of viral particles [26]. Recently, it was shown that multiple genes involved in cholesterol synthesis are induced by Nef [36]. NaB treatment also induced some of these genes (*INSIG1*, *HMGCS1*, *IDI1*, *LSS* or *SREBF1*) and could thus enhanced virion infectivity and viral replication.

Krishnan *et al.* have described an increase in expression of several proteasome subunits in ACH-2 cells prior induction of lytic replication by PMA and proposed that the higher expression of proteasomes may lead to increased degradation of HIV-1 mRNA [27]. After induction of lytic replication by NaB, proteasome subunits PSMB10 and PSMB8 were downregulated in ACH-2 and U1 cells, suggesting a role in the maintenance of the latent state. Indeed, reactivation of latency was achieved with proteasome inhibitors [27]. Among the downregulated genes after NaB treatment, we identified genes involved in RNA modifications. Krishnan *et al.* have shown alterations in the expression of DEAD-box and other RNA binding proteins during HIV-1 replication [37]. Especially, *DDX18* and *DDX39* are upregulated in latently infected cells [37]. After NaB treatment of latently infected cells, we observed a decrease in the expression of these two proteins, thus providing more support for their role in maintaining HIV-1 latency.

The only purpose of our microarray analysis was to identify candidate genes potentially involved in the control of the HIV latency. For this reason, we decided to focus on two candidate genes previously described to influence viral expression and that may be involved in reactivation and maintenance of latency: *NCoA3* and *IRF8*, respectively. Hybridization experiments were performed once. Consequently, we did not further analyze the statistical relevance of the results and performed complementary approaches to confirm the mRNA variations of the selected candidate genes.

NCoA3 is a nuclear receptor coactivator that enhances ligand-induced transcriptional activation of nuclear receptors (reviewed in [28]). We show that *NCoA3* (Unigene Hs. 382168) gene expression is upregulated following treatment with NaB of U1 and ACH-2 latently infected cells. This differential transcriptional expression was confirmed by real-time RT-PCR and is also mediated by PMA but not TSA. Upregulation of *NCoA3* is thus achieved following phorbol ester but not other HDAC inhibitor treatment. However, NaB and TSA act on different pathways and at different concentrations and target different genes [38]. Transcriptional increase of *NCoA3* was observed in parental uninfected corresponding cell lines U937 and CEM and in two others latently HIV-1 infected cell lines, OM10.1 and J1.1. *NCoA3* protein level is also upregulated following treatment with NaB in the U1, ACH-2 and

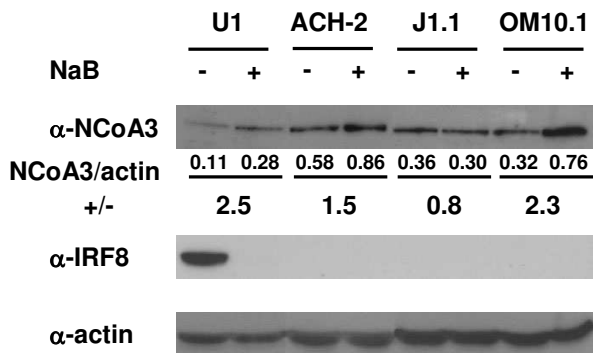


Figure 7
Western blot analysis of NCoA3 and IRF8 proteins expression. Nuclear extract (100 μ g) from U1, ACH-2, J1.1 and OM10.1 treated (+) or not (-) with NaB for 24 h were resolved by SDS-PAGE and immunoblotted with anti-NCoA3 or anti-IRF8 antibody, as indicated. The amount of protein was normalized using anti-actin antibody. Figures below NCoA3 immunoblot indicated the results of the quantification using Image Tool (Syngene) software of the ratio NCoA3/actin upon NaB-treatment (+) versus NCoA3/actin non-treated (-). Results are representative of three independent experiments.

OM10.1 cell lines. Moreover, NCoA3 increases the Tat-induced HIV-1 LTR promoter transcriptional activity through the TAR region, in accordance with other data [31]. The differential expression of NCoA3 observed led us to postulate that NCoA3 could be involved in the transcriptional reactivation of the HIV-1 promoter from latency, at low concentrations of Tat.

This hypothesis is supported by several findings. Previous microarray studies on latently infected resting CD4⁺T cells in infected individuals have shown an upregulation of NCoA3 gene expression in viremic versus aviremic patients [26]. Moreover, Kino *et al.* showed that NCoA factors improve Tat transactivation of HIV-1 LTR promoter activity and interact with Tat [31]. Tat transactivation activity is mediated by its interaction with components of the basal transcription machinery (including TBP, TAFII250, RNA polymerase II), with kinase complexes able to phosphorylate the C-terminal domain of RNA polymerase II (in particular with the P-TEFb complex composed of cyclin T1/CDK9) and with cellular proteins possessing HAT activity (p300/CBP, P/CAF and GCN5) (reviewed in [39]). Kino *et al.* showed that one member of the family, NCoA2, functions as a Tat coactivator on the HIV-1 LTR by bridging promoter-bound proteins with the Tat-P-TEFb complex through its interaction with Tat and Cyclin T1 [31]. Stimulation of Tat transactivation activity by NCoA3 could involve similar mechanisms.

Furthermore, it has been recently demonstrated that recruitment of HATs to the LTR is an early event in HIV-1 transcriptional activation [13] and that a consequence of histone acetylation is the recruitment of the ATP-dependent chromatin remodeling complex hSWI/SNF to the LTR [12]. NCoA3 could mediate chromatin remodeling by recruitment of additional cofactors with HAT activity (such as p300/CBP and P/CAF) and by an intrinsic HAT activity [40] and may thus contribute to the transcriptional reactivation of the HIV-1 promoter from latency.

IRF8 is a transcription factor that binds to ISRE and regulates expression of genes stimulated by IFNs (reviewed in [29]). IRF8 is able to both activate and repress gene transcription depending on the target gene. We show that *IRF8* (Unigene Hs. 137427) gene is only expressed in the promonocytic cell line U1 and its expression is strongly downregulated following NaB treatment of these cells. This differential transcriptional expression was confirmed by real-time RT-PCR and is also observed, albeit at lower extent, after PMA and TSA treatments of U1 cells. IRF8 protein level is similarly downregulated following treatment with NaB. Moreover, IRF8 represses the IRF1-mediated activation of the HIV-1 ISRE element of the LTR, in accordance with other data [32]. The decreased expression of IRF8 following reactivation of latency using different molecules suggest that IRF8 may contribute in the maintenance of the latent state in the promonocytic cell line.

It has been shown that binding of specific transcription factors downstream of the HIV-1 transcription start site is crucial to control HIV-1 transcription [33,41]. Among these sites is an ISRE element that recruits IRF1 and IRF2 *in vivo* [33]. Previous studies have investigated the role of IRFs on the modulation of HIV-1 replication (reviewed in [42,43]) and showed that IRF1 activates HIV-1 LTR transcription, interacts with Tat [32] and increases HIV-1 replication [44]. However, IRF8 represses IRF1-Tat-mediated transactivation of the LTR by interfering with IRF1-Tat association [32]. Moreover, it has been shown that IRF8 inhibits HIV-1 replication in T CD4⁺ lymphocytic and promonocytic cell lines [32,34]. These data support the hypothesis that repression of HIV-1 transcription by IRF8 could be implicated in the maintenance of proviral quiescence in latently infected cells.

Moreover, the result obtained after measurement of *gag*, *NCoA3* and *IRF8* mRNA after different times of NaB stimulation clearly showed a correlation between *gag* mRNA increase and *NCoA3* mRNA increase or *IRF8* mRNA decrease, respectively. These correlations support the hypothesis that IRF8 and NCoA3 factors may be involved in the control of the HIV latency.

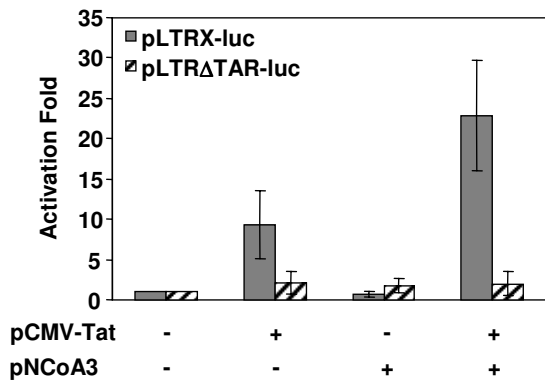


Figure 8
NCoA3 increases the Tat-stimulated HIV-1 LTR activity. HEK293 cells were cotransfected with pLTRX-luc (10 ng, grey bars) or pLTRΔTAR-luc (10 ng, white bars) with (+) or without (-) suboptimal amounts of pCMV-Tat (5 ng) and/or pNCoA3 (1 μg) expression vectors. NLI (normalized luciferase index) were measured after 24 h and the activation folds compared to the basal activity of the corresponding pLTR-luc were determined. Results represent the means of five independent experiments.

Chronically HIV-1 infected cell lines used in this study provide useful models for studying HIV-1 latency but are not in a quiescent state as cellular reservoirs *in vivo*. Moreover, it has been shown that mutations in the *tat* gene and in the TAR sequence are responsible for the latency observed in U1 and ACH-2 cells, respectively [45,46]. We thus confirmed the differential expression of *NCoA3* but not *IRF8* genes in two others chronically HIV-1 infected cell lines, OM10.1 and J1.1. We will now investigate the involvement of *NCoA3* and *IRF8* to regulate viral expression in primary cells such as resting T CD4⁺ lymphocytes or macrophages.

Conclusion

Additional experiments are currently underway to validate the biological relevance of the differential expression of *IRF8* and *NCoA3* genes in latency maintenance and reactivation. Since the persistence of integrated HIV-1 genomes despite potent suppression of viral replication is a major obstacle for current antiretroviral therapy, selective disruption of the HIV-1 proviral latency may provide good strategies to decrease latent HIV-1 reservoirs. Thus, identification of cellular genes that are differentially expressed during HIV-1 reactivation of latency is crucial to understand the molecular mechanisms involved in the control of HIV-1 latency.

Methods

Cell cultures and treatments

The chronically HIV-1 infected T CD4⁺ lymphocytic cell lines ACH-2 [47] and J1.1 [48] derived from CEM and Jurkat cells respectively, and the chronically HIV-1 infected promonocytic cell lines U1 [49] and OM10.1 [50] derived from U937 and HL-60 cells respectively, were obtained through the National Institutes of Health (NIH) AIDS Research and Reference Reagent Program. Suspension cell lines were grown in RPMI 1640 (Invitrogen) with 10% fetal bovine serum (Invitrogen), 50 U/mL penicillin, 50 μg/mL streptomycin (Invitrogen) and 2 mM glutamine (Invitrogen). Cells were treated with 10 mM of sodium butyrate (NaB; Sigma), or with 10 ng/mL of PMA (Sigma), or with 300 nM of TSA (Sigma). Cells were harvested generally 24 h and 48 h after treatment and cell viability was estimated before subsequent RNA extraction or nuclear extract preparation. P4 indicator cells are HeLa CD4⁺ cells carrying the lacZ gene under the control of the HIV-1 LTR. P4 and HEK293 cells were grown in DMEM (Invitrogen) containing 5% fetal bovine serum (Invitrogen), 50 U/mL penicillin, 50 μg/mL streptomycin (Invitrogen) and 2 mM glutamine (Invitrogen).

Plasmids

The pLTRX-luc construct contains the luciferase (*luc*) gene downstream of the HIV-1 BRU U3-R promoter region (nt -640 to +78) [51]. The pLTRΔTAR-luc construct corresponds to the pLTRX-luc plasmid in which the TAR region (nt +38 to +78) was deleted [51]. The pCMV-Tat expression vector was kindly provided by S. Emiliani (Institut Cochin, Paris, France). The pIRF8 expression vector (pcDNAmycHis-ICSBP) and dominant negative construct pIRF8-DBD, which contains the DNA binding domain of IRF8, were a kind gift of B.Z. Levi (Technion-Israel Institute of Technology, Haifa, Israel). The pNCoA3 expression vector (pcDNA3.1-AIB1) was a kind gift of P.S. Meltzer (NIH, Bethesda, USA) [52]. The pIRF1 construct was generated by cloning the fragment excised from pHuIRF-3-1 (a kind gift of T. Taniguchi, University of Tokyo, Tokyo, Japan) by HindIII/NotI digestion in the pcDNA3.1 plasmid (Invitrogen). The pISRE-TK-luc and pISREmut-TK-luc constructs were generated by cloning a wild-type (AGGGACTTGAAAAGCGAAAAGGGAAACCAGAG) or mutated (AGGGACTTGCCCCGCCCCGGAAACCA-GAG) synthetic oligonucleotide corresponding to the HIV-1 BRU ISRE sequence (nt +194 to +223) [33,53] in the pTK-luc plasmid in which the luciferase gene is under the control of the truncated HSV-1 thymidine kinase promoter minimum region [51]. The pCMV-LacZ was kindly provided by M. Alizon (Institut Cochin, Paris, France).

Total RNA extraction

Total RNAs were extracted using the RNeasy Mini Kit (Qiagen). The procedure included an "on-column"

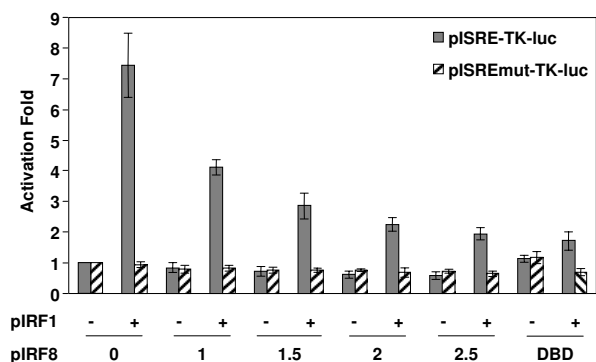


Figure 9
IRF8 represses the IRF1-mediated activation of the HIV-1 ISRE element. HEK293 cells were cotransfected with pISRE-TK-luc (250 ng, solid bars) or pISREmut-TK-luc (250 ng, white bars) with (+) or without (-) pIRF1 (250 ng), pIRF8 (1–2.5 μ g), or pIRF8-DBD (1 μ g) expression vectors. NLI (normalized luciferase index) were measured after 24 h and the activation folds compared to the basal activity of the pISRE-TK-luc or pISREmut-TK-luc were determined. Results represent the means of five independent experiments.

DNase I digestion step according to the manufacturer's instructions. RNA quality was assessed using the Agilent Bioanalyzer 2100 and spectrophotometric analysis prior to cDNA synthesis.

Microarray experiments

Microarray experiments were performed using the U133-A microarrays (Affymetrix) containing 22283 oligonucleotides spots. Total RNAs obtained from chronically infected U1 and ACH-2 stimulated or not with NaB for 24 h were sent to Dr. C. Thibault (Affymetrix Microarray Facilities, IGBMC, Strasbourg, France) for amplification, labeling and hybridization. Hybridization experiments were performed once. Results were then analyzed with the Mas5.0 Software (Affymetrix) and interpreted using the Data Mining Tool (Affymetrix) and Microsoft Excel softwares. For individual analyses, the p-value cut off was 0.048 as suggested by Affymetrix. For comparative analyses, a \log_2 ratio change ≥ 1 for increased genes and ≤ -1 for decreased genes were defined. Gene expression changes were considered to be significant when the change p-value was ≤ 0.0001 for increased genes and 1-change p-value ≥ 0.9999 for decreased genes.

Real-time RT-PCR

Quantifications of cellular RNAs were performed using a Light Cycler instrument (Roche Diagnostics). Briefly, cDNAs were synthesized from 1 μ g of total RNA with MoMLV reverse transcriptase (Superscript II, Invitrogen) and 1/10th aliquots of the corresponding samples were

used for real-time PCR in a 20 μ L reaction mixture containing 1X LightCycler FastStart DNA Master SYBR Green I (Roche Diagnostics), 4 mM MgCl₂, and 500 nM of each primer. The reactions were carried out in duplicate and the results were normalized to the expression of Cyclophilin A. Primers for quantitative PCR were designed using Oligo 6 software. All primer pairs produced single amplification product as determined by melting curve analyses. The sequences of the primers used were (5' to 3'): NCoA3 forward CTTTGGGCATTCTGAACCTGTC, NCoA3 reverse GCCTCATCACCGCAGCAC, IRF8 forward GGAGTGCG-GTCGCTCTGAAA, IRF8 reverse GTCGTAGGTGGTGTAC-CCCGTCA, Cyclophilin A forward AGTGGTTGGATGGCAAGC, Cyclophilin A reverse GAT-TCTAGGATACTGCGAGCAAA. PCR reactions were carried out with a denaturation step of 10 min at 95°C followed by forty-five cycles of 10 s at 95°C, 5 s at annealing temperature (55°C for NCoA3 and Cyclophilin A, 59°C for IRF8) and 20 s amplification at 72°C. Quantifications of cDNAs were determined in reference to a standard curve prepared by amplification of serial dilutions of PCR product containing matching sequences. Analyses were performed using the second-derivative-maximum method provided by the Light Cycler quantification software, version 3.5 (Roche Diagnostics).

Quantification of *gag* viral mRNA was performed by real-time RT-PCR as described in [54].

Nuclear extracts preparation

For nuclear extract preparation, 10.10⁶ cells were harvested, washed and nuclei were isolated by addition of 150 μ L of buffer I (50 mM Tris pH 7.9, 10 mM KCl, 10% glycerol, 1 mM EDTA, 0.2% NP40) followed by a centrifugation at 3000 g for 3 min. Nuclear extracts were prepared by addition of 15 μ L of buffer II (20 mM Hepes pH 7.9, 400 mM NaCl, 10 mM KCl, 20% glycerol, 1 mM EDTA) for 20 min at 4°C followed by a centrifugation at 15000 g for 10 min. Protein concentrations were determined by the Bio-Rad protein assay.

Western blot analysis

Nuclear extracts (100 μ g) were loaded on 8% SDS-polyacrylamide gel and the proteins were transferred to nitrocellulose membrane (Hybond-C, Amersham) that was subsequently blocked for 1 h with 5% non-fat dry milk in PBS-T (PBS, 0.05% Tween20) and incubated with antibodies directed against NCoA3 (goat polyclonal anti-ACR C-20, Santa Cruz Biotechnology, Inc.), IRF8 (goat polyclonal anti-ICSBP C-19, Santa Cruz Biotechnology, Inc.) or actin (mouse monoclonal anti-actin, Calbiochem) for 2 h. The membranes were then washed and incubated with secondary antibodies conjugated to horseradish peroxidase (HRP conjugated rabbit anti-goat (DakoCytomation) or goat anti-mouse (Calbiochem)

immunoglobulins). Hybridizations were revealed using an ECL enhanced chemiluminescence kit (ECL, Amersham). The quantification was done using the Image Tools (Syngene) software.

Transient transfection and enzymatic assays

HEK293 cells were transfected using calcium phosphate co-precipitation method. Cells were lysed 24 h after transfection with a buffer containing 60 mM Na₂HPO₄, 40 mM NaH₂PO₄, 10 mM KCl, 10 mM MgSO₄, 2.5 mM EDTA, 50 mM β-mercaptoethanol and 0.125% Nonidet P-40. Luciferase activities were measured as previously described [55]. Cotransfection with pCMV-LacZ plasmid was performed to normalize transfection efficiency and β-galactosidase activities were determined using Chlorophenol red β-D-galactopyranoside (CPRG, Roche Diagnostics) assay as previously described [55]. The normalized luciferase index (NLI) was defined as the ratio of luciferase to β-galactosidase activities.

Competing interests

The author(s) declare that they have no competing interests.

Authors' contributions

SM performed the microarray analyses, real-time RT-PCR and drafted the manuscript. DD and LC carried out real-time RT-PCR, Western blot and transfection experiments. AG participated in transfection experiments. UH conceived the study, participated in its design and coordination and helped to draft and finalize the manuscript. All authors read and approved the final manuscript.

Additional material

Additional File 1

Genes upregulated in U1 and ACH-2 cells.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1742-4690-2-73-S1.doc>]

Additional File 2

Genes specifically upregulated in U1 cells.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1742-4690-2-73-S2.doc>]

Additional File 3

Genes specifically upregulated in ACH-2 cells.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1742-4690-2-73-S3.doc>]

Additional File 4

Genes downregulated in U1 and ACH-2 cells.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1742-4690-2-73-S4.doc>]

Additional File 5

Genes specifically downregulated in U1 cells.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1742-4690-2-73-S5.doc>]

Additional File 6

Genes specifically downregulated in ACH-2 cells.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1742-4690-2-73-S6.doc>]

Acknowledgements

We thank Drs. S. Emiliani, B.Z. Levi, P.S. Meltzer, T. Taniguchi, M. Alizon for providing plasmids, and the National Institutes of Health AIDS Research and Reference Reagent Program for the kind gift of reagents. We are grateful to Dr. C. Thibault for her precious help in microarray hybridization data mining. L.C. holds a fellowship from the Ministère de l'Éducation Nationale, de l'Enseignement Supérieur et de la Recherche. S.M. is supported by a grant from Sidaction. Sidaction (AO15) supported this work. We thank S. Nisole for helpful discussion.

References

1. Pomerantz RJ, Horn DL: Twenty years of therapy for HIV-1 infection. *Nat Med* 2003, **9**:867-873.
2. Pierson T, McArthur J, Siliciano RF: **Reservoirs for HIV-1: mechanisms for viral persistence in the presence of antiviral immune responses and antiretroviral therapy.** *Annu Rev Immunol* 2000, **18**:665-708.
3. Wong JK, Hezareh M, Gunthard HF, Havlir DV, Ignacio CC, Spina CA, Richman DD: **Recovery of replication-competent HIV despite prolonged suppression of plasma viremia.** *Science* 1997, **278**:1291-1295.
4. Finzi D, Hermankova M, Pierson T, Carruth LM, Buck C, Chaisson RE, Quinn TC, Chadwick K, Margolick J, Brookmeyer R, Gallant J, Markowitz M, Ho DD, Richman DD, Siliciano RF: **Identification of a reservoir for HIV-1 in patients on highly active antiretroviral therapy.** *Science* 1997, **278**:1295-1300.
5. Chun TW, Stuyver L, Mizell SB, Ehler LA, Mican JA, Baseler M, Lloyd AL, Nowak MA, Fauci AS: **Presence of an inducible HIV-1 latent reservoir during highly active antiretroviral therapy.** *Proc Natl Acad Sci U S A* 1997, **94**:13193-13197.
6. Blankson JN, Persaud D, Siliciano RF: **The challenge of viral reservoirs in HIV-1 infection.** *Annu Rev Med* 2002, **53**:557-593.
7. Lassen K, Han Y, Zhou Y, Siliciano J, Siliciano RF: **The multifactorial nature of HIV-1 latency.** *Trends Mol Med* 2004, **10**:525-531.
8. Finzi D, Blankson J, Siliciano JD, Margolick JB, Chadwick K, Pierson T, Smith K, Lisziewicz J, Lori F, Flexner C, Quinn TC, Chaisson RE, Rosenberg E, Walker B, Gange S, Gallant J, Siliciano RF: **Latent infection of CD4(+) T cells provides a mechanism for lifelong persistence of HIV-1, even in patients on effective combination therapy.** *Nat Med* 1999, **5**:512-517.
9. Williams SA, Greene WC: **Host factors regulating post-integration latency of HIV.** *Trends Microbiol* 2005, **13**:137-139.
10. He G, Ylisastigui L, Margolis DM: **The regulation of HIV-1 gene expression: the emerging role of chromatin.** *DNA Cell Biol* 2002, **21**:697-705.

11. Verdin E, Paras PJ, Van Lint C: **Chromatin disruption in the promoter of human immunodeficiency virus type I during transcriptional activation.** *Embo J* 1993, **12**:3249-3259.
12. Henderson A, Holloway A, Reeves R, Tremethick DJ: **Recruitment of SWI/SNF to the human immunodeficiency virus type I promoter.** *Mol Cell Biol* 2004, **24**:389-397.
13. Lusic M, Marcello A, Cereseto A, Giacca M: **Regulation of HIV-1 gene expression by histone acetylation and factor recruitment at the LTR promoter.** *Embo J* 2003, **22**:6550-6561.
14. Kato-Maeda M, Gao Q, Small PM: **Microarray analysis of pathogens and their interaction with hosts.** *Cell Microbiol* 2001, **3**:713-719.
15. Shaheduzzaman S, Krishnan V, Petrovic A, Bittner M, Meltzer P, Trent J, Venkatesan S, Zeichner S: **Effects of HIV-1 Nef on cellular gene expression profiles.** *J Biomed Sci* 2002, **9**:82-96.
16. Simmons A, Aluvihare V, McMichael A: **Nef triggers a transcriptional program in T cells imitating single-signal T cell activation and inducing HIV virulence mediators.** *Immunity* 2001, **14**:763-777.
17. De la Fuente C, Santiago F, Deng L, Eadie C, Zilberman I, Kehn K, Maddukuri A, Baylor S, Wu K, Lee CG, Pumfery A, Kashanchi F: **Gene expression profile of HIV-1 Tat expressing cells: a close interplay between proliferative and differentiation signals.** *BMC Biochem* 2002, **3**:14.
18. Izmailova E, Bertley FM, Huang Q, Makori N, Miller CJ, Young RA, Aldovini A: **HIV-1 Tat reprograms immature dendritic cells to express chemoattractants for activated T cells and macrophages.** *Nat Med* 2003, **9**:191-197.
19. Cicala C, Arthos J, Selig SM, Dennis GJ, Hosack DA, Van Ryk D, Spangler ML, Steenbeke TD, Khazanie P, Gupta N, Yang J, Daucher M, Lempicki RA, Fauci AS: **HIV envelope induces a cascade of cell signals in non-proliferating target cells that favor virus replication.** *Proc Natl Acad Sci U S A* 2002, **99**:9380-9385.
20. Janket ML, Manickam P, Majumder B, Thotala D, Wagner M, Schaffer EA, Collman RG, Srinivasan A, Ayyavoo V: **Differential regulation of host cellular genes by HIV-1 viral protein R (Vpr): cDNA microarray analysis using isogenic virus.** *Biochem Biophys Res Commun* 2004, **314**:1126-1132.
21. Corbeil J, Sheeter D, Genini D, Rought S, Leoni L, Du P, Ferguson M, Masys DR, Welsh JB, Fink JL, Sasik R, Huang D, Drenkow J, Richman DD, Gingeras T: **Temporal gene regulation during HIV-1 infection of human CD4+ T cells.** *Genome Res* 2001, **11**:1198-1204.
22. Geiss GK, Bumgarner RE, An MC, Agy MB, van 't Wout AB, Hammersmark E, Carter VS, Upchurch D, Mullins JI, Katze MG: **Large-scale monitoring of host cell gene expression during HIV-1 infection using cDNA microarrays.** *Virology* 2000, **266**:8-16.
23. Van 't Wout AB, Lehrman GK, Mikheeva SA, O'Keeffe GC, Katze MG, Bumgarner RE, Geiss GK, Mullins JI: **Cellular gene expression upon human immunodeficiency virus type I infection of CD4(+)-T-cell lines.** *J Virol* 2003, **77**:1392-1402.
24. Wen W, Chen S, Cao Y, Zhu Y, Yamamoto Y: **HIV-1 infection initiates changes in the expression of a wide array of genes in U937 promonocytes and HUT78 T cells.** *Virus Res* 2005.
25. Vahey MT, Nau ME, Jagodzinski LL, Yalley-Ogunro J, Taubman M, Michael NL, Lewis MG: **Impact of viral infection on the gene expression profiles of proliferating normal human peripheral blood mononuclear cells infected with HIV type I RF.** *AIDS Res Hum Retroviruses* 2002, **18**:179-192.
26. Chun TW, Justement JS, Lempicki RA, Yang J, Dennis GJ, Hallahan CW, Sanford C, Pandya P, Liu S, McLaughlin M, Ehler LA, Moir S, Fauci AS: **Gene expression and viral production in latently infected, resting CD4+ T cells in viremic versus aviremic HIV-infected individuals.** *Proc Natl Acad Sci U S A* 2003, **100**:1908-1913.
27. Krishnan V, Zeichner SL: **Host cell gene expression during human immunodeficiency virus type I latency and reactivation and effects of targeting genes that are differentially expressed in viral latency.** *J Virol* 2004, **78**:9458-9473.
28. Xu J, Li Q: **Review of the in vivo Functions of the p160 Steroid Receptor Coactivator Family.** *Mol Endocrinol* 2003, **12**:12.
29. Levi BZ, Hashmueli S, Gleit-Kielmanowicz M, Azriel A, Meraro D: **ICSBP/IRF-8 transactivation: a tale of protein-protein interaction.** *J Interferon Cytokine Res* 2002, **22**:153-160.
30. Bosinger SE, Hosiawa KA, Cameron MJ, Persad D, Ran L, Xu L, Boulassel MR, Parenteau M, Fournier J, Rud EV, Kelvin DJ: **Gene expression profiling of host response in models of acute HIV infection.** *J Immunol* 2004, **173**:6858-6863.
31. Kino T, Slobodskaya O, Pavlakis GN, Chrousos GP: **Nuclear receptor coactivator p160 proteins enhance the HIV-1 long terminal repeat promoter by bridging promoter-bound factors and the Tat-P-TEFb complex.** *J Biol Chem* 2002, **277**:2396-2405.
32. Sgarbanti M, Borsetti A, Moscufo N, Bellocchi MC, Ridolfi B, Nappi F, Marsili G, Marziali G, Coccia EM, Ensoli B, Battistini A: **Modulation of human immunodeficiency virus I replication by interferon regulatory factors.** *J Exp Med* 2002, **195**:1359-1370.
33. Van Lint C, Amella CA, Emiliani S, John M, Jie T, Verdin E: **Transcription factor binding sites downstream of the human immunodeficiency virus type I transcription start site are important for virus infectivity.** *J Virol* 1997, **71**:6113-6127.
34. Thornton AM, Ogryzko VV, Dent A, Sharf R, Levi BZ, Kanno Y, Staudt LM, Howard BH, Ozato K: **A dominant negative mutant of an IFN regulatory factor family protein inhibits both type I and type II IFN-stimulated gene expression and antiproliferative activity of IFNs.** *J Immunol* 1996, **157**:5145-5154.
35. Herrmann CH, Carroll RG, Wei P, Jones KA, Rice AP: **Tat-associated kinase, TAK, activity is regulated by distinct mechanisms in peripheral blood lymphocytes and promonocytic cell lines.** *J Virol* 1998, **72**:9881-9888.
36. Van 't Wout AB, Swain JV, Schindler M, Rao U, Pathmajayan MS, Mullins JI, Kirchhoff F: **Nef induces multiple genes involved in cholesterol synthesis and uptake in human immunodeficiency virus type I-infected T cells.** *J Virol* 2005, **79**:10053-10058.
37. Krishnan V, Zeichner SL: **Alterations in the expression of DEAD-box and other RNA binding proteins during HIV-1 replication.** *Retrovirology* 2004, **1**:42.
38. Marks P, Rifkind RA, Richon VM, Breslow R, Miller T, Kelly WK: **Histone deacetylases and cancer: causes and therapies.** *Nat Rev Cancer* 2001, **1**:194-202.
39. Brigati C, Giacca M, Noonan DM, Albini A: **HIV Tat, its TARgets and the control of viral gene expression.** *FEMS Microbiol Lett* 2003, **220**:57-65.
40. Chen H, Lin RJ, Schiltz RL, Chakravarti D, Nash A, Nagy L, Privalsky ML, Nakatani Y, Evans RM: **Nuclear receptor coactivator ACTR is a novel histone acetyltransferase and forms a multimeric activation complex with P/CAF and CBP/p300.** *Cell* 1997, **90**:569-580.
41. El Kharroubi A, Martin MA: **cis-acting sequences located downstream of the human immunodeficiency virus type I promoter affect its chromatin structure and transcriptional activity.** *Mol Cell Biol* 1996, **16**:2958-2966.
42. Battistini A, Marsili G, Sgarbanti M, Ensoli B, Hiscott J: **IRF regulation of HIV-1 long terminal repeat activity.** *J Interferon Cytokine Res* 2002, **22**:27-37.
43. Marsili G, Borsetti A, Sgarbanti M, Remoli AL, Ridolfi B, Stellacci E, Ensoli B, Battistini A: **On the role of interferon regulatory factors in HIV-1 replication.** *Ann N Y Acad Sci* 2003, **1010**:29-42.
44. Sgarbanti M, Marsili G, Remoli AL, Ridolfi B, Stellacci E, Borsetti A, Ensoli B, Battistini A: **Analysis of the signal transduction pathway leading to human immunodeficiency virus-I-induced interferon regulatory factor-I upregulation.** *Ann N Y Acad Sci* 2004, **1030**:187-195.
45. Emiliani S, Van Lint C, Fischle W, Paras PJ, Ott M, Brady J, Verdin E: **A point mutation in the HIV-1 Tat responsive element is associated with postintegration latency.** *Proc Natl Acad Sci U S A* 1996, **93**:6377-6381.
46. Emiliani S, Fischle W, Ott M, Van Lint C, Amella CA, Verdin E: **Mutations in the tat gene are responsible for human immunodeficiency virus type I postintegration latency in the UI cell line.** *J Virol* 1998, **72**:1666-1670.
47. Clouse KA, Powell D, Washington I, Poli G, Strebel K, Farrar W, Barstad P, Kovacs J, Fauci AS, Folks TM: **Monokine regulation of human immunodeficiency virus-I expression in a chronically infected human T cell clone.** *J Immunol* 1989, **142**:431-438.
48. Perez VL, Rowe T, Justement JS, Butera ST, June CH, Folks TM: **An HIV-1-infected T cell clone defective in IL-2 production and Ca2+ mobilization after CD3 stimulation.** *J Immunol* 1991, **147**:3145-3148.
49. Folks TM, Justement J, Kinter A, Schnittman S, Orenstein J, Poli G, Fauci AS: **Characterization of a promonocyte clone chronically infected with HIV and inducible by 13-phorbol-12-myristate acetate.** *J Immunol* 1988, **140**:1117-1122.
50. Butera ST, Perez VL, Wu BY, Nabel GJ, Folks TM: **Oscillation of the human immunodeficiency virus surface receptor is regu-**

lated by the state of viral activation in a CD4+ cell model of chronic infection. *J Virol* 1991, **65**:4645-4653.

51. Hazan U, Thomas D, Alcami J, Bachelier F, Israël N, Yssel H, Virelizier JL, Arenzana-Seisdedos F: **Stimulation of human T-cell clone with anti-CD3 or tumor necrosis factor induces NF-kappa B translocation but not human immunodeficiency virus I enhancer-dependent transcription.** *Proc Natl Acad Sci USA* 1990, **87**:7861-7865.
52. Anzick SL, Kononen J, Walker RL, Azorsa DO, Tanner MM, Guan XY, Sauter G, Kallioniemi OP, Trent JM, Meltzer PS: **AIB1, a steroid receptor coactivator amplified in breast and ovarian cancer.** *Science* 1997, **277**:965-968.
53. El Kharroubi A, Verdin E: **Protein-DNA interactions within DNase I-hypersensitive sites located downstream of the HIV-I promoter.** *J Biol Chem* 1994, **269**:19916-19924.
54. Brussel A, Sonigo P: **Evidence for gene expression by unintegrated human immunodeficiency virus type I DNA species.** *J Virol* 2004, **78**:11263-11271.
55. Dumonceaux J, Nisole S, Chanel C, Quivet L, Amara A, Baleux F, Briand P, Hazan U: **Spontaneous mutations in the env gene of the human immunodeficiency virus type I NDK isolate are associated with a CD4-independent entry phenotype.** *J Virol* 1998, **72**:512-519.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

